Re-examining the Hemiptera phylogeny using supermatrices and phylogenomic data sets

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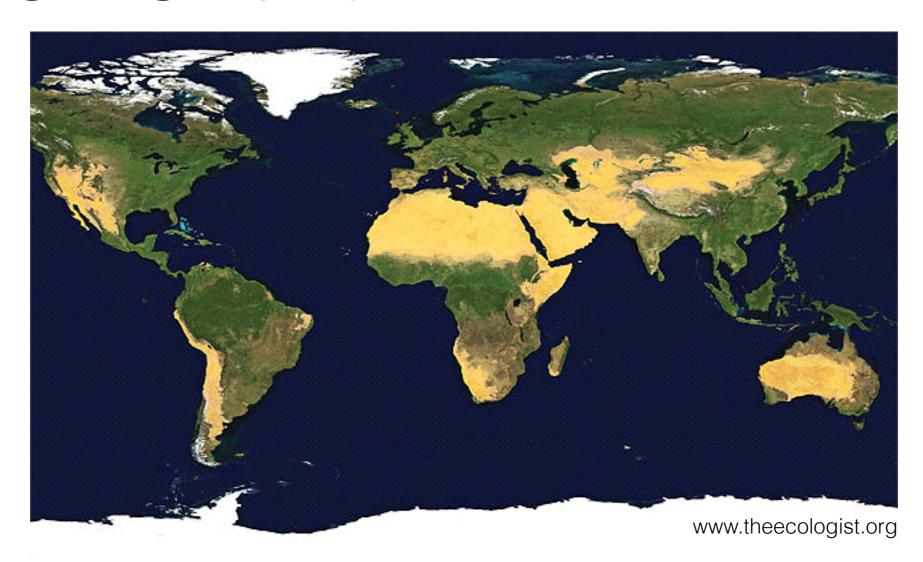




OUTLINE

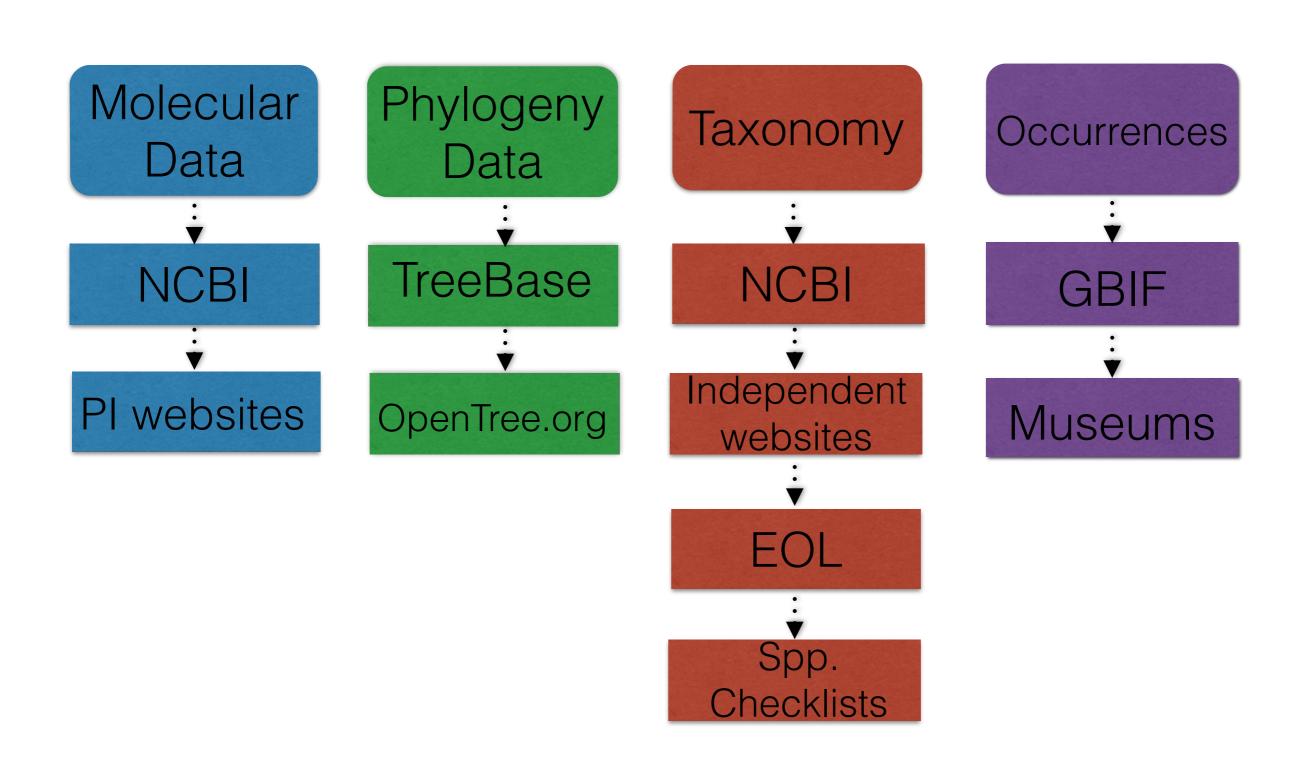
- 1. Introduction
- 2. Methods
- 3. Preliminary Results
- 4. Future directions

Introduction: Southern Hemisphere biogeography and diversification



- Three major episodes of cooling since the Eocene
- Floral turnover
- Signatures of both changes in diversification and floral turnover in Australian cicadas

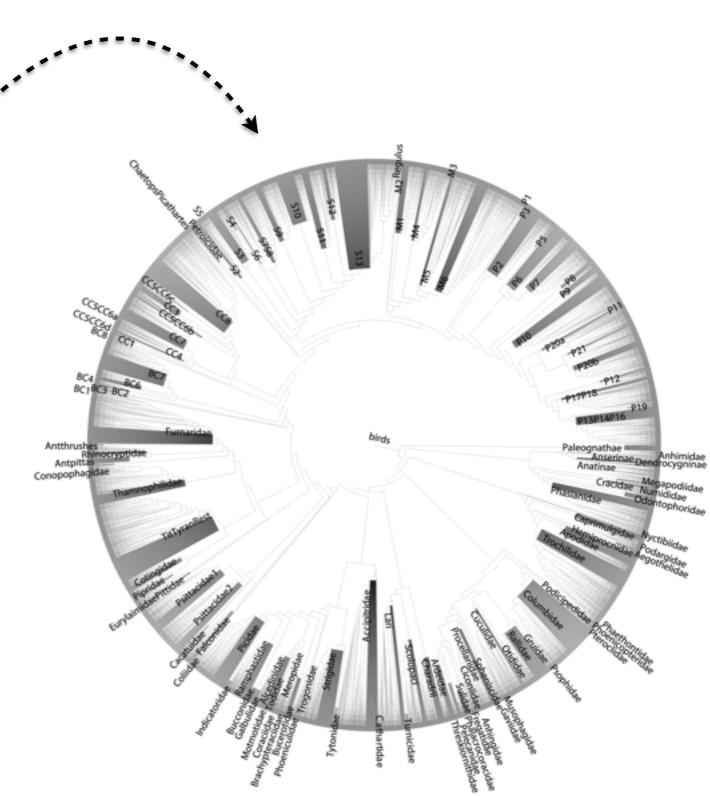
Goal: synthesize all available Hemiptera data to test hypotheses



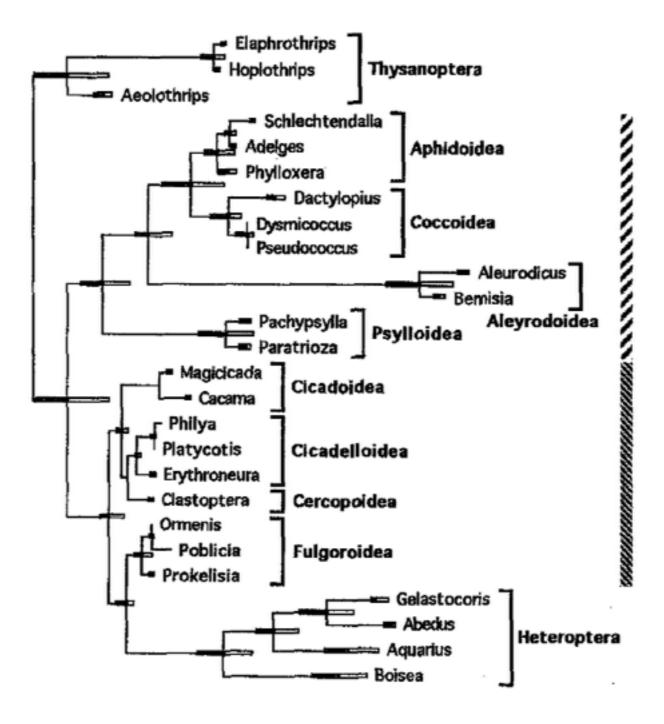
AVAToL: Open Tree of Life

Published Taxonomies Published Phylogenies

- Phylogeny of all life
- Accessible to the public
- Public curation
- Add/Extract phylogenies
- First release: early 2014
- Two additional AVAToL groups

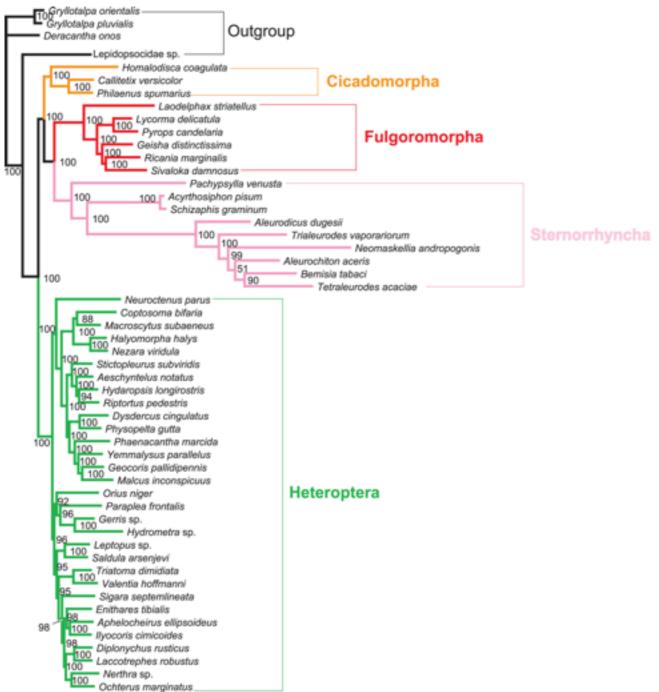


Hemiptera Phylogenies: von Dohlen & Moran 1995 J. Mol. Evol.

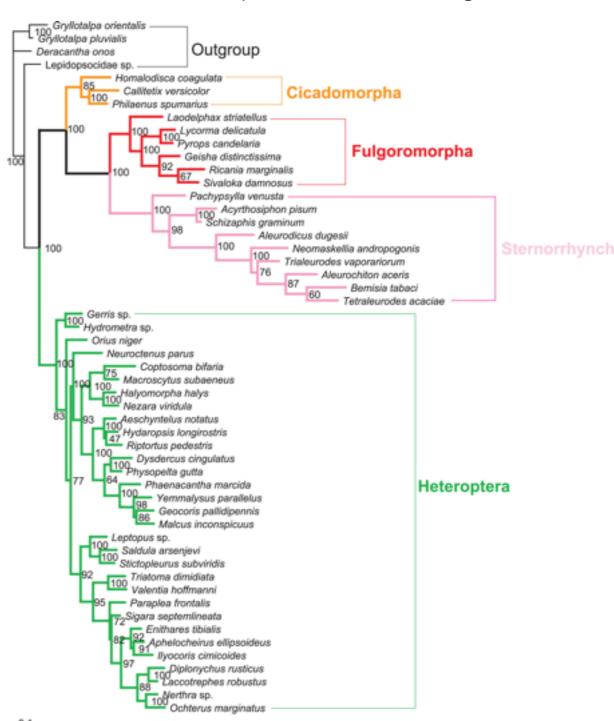


Recent Hemiptera Phylogenies: Song et al. 2012 PLOS One

BI Tree:1st & 2nd postions, 2 rRNA genes

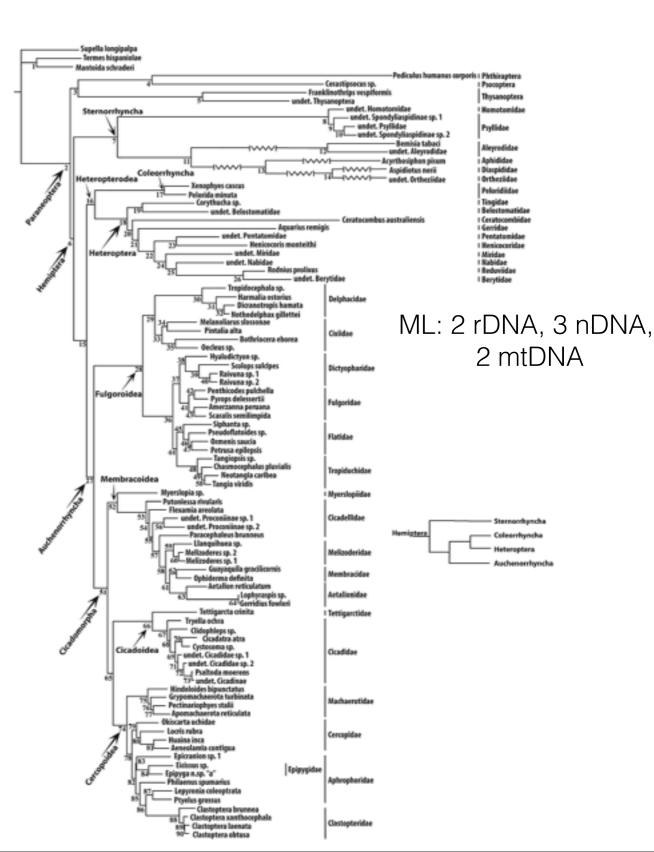


ML Tree:1st & 2nd postions, 2 rRNA genes



Recent Hemiptera Phylogenies: Cryan and Urban 2012 J. Syst. Ent.

- Well-supported backbone for most higher taxa
- Lacking support for Hemiptera
- Additional molecular evidence for long branches in Sternorrhyncha



Methods: assemble transcriptome data gene alignments

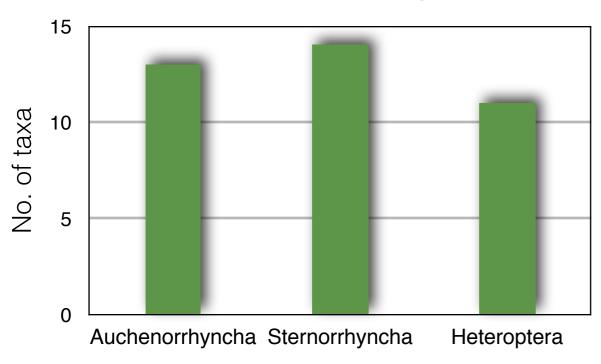
- Developed a linux and python pipeline to automate the retrieval and assembly of SRA data (gsAssembler and Trinity)
- Individual sequencing runs inspected individually using FastQC and PRINSEQ for QC (adaptors, low complexity, poly-A tails, etc.)
- Removal of bacterial, viral, and Ribosomal RNA sequences using DeconSeq and Pathoscope
- Assemblies were quickly check by mapping reads to back to assemblies using BWA
- HaMStR pipeline used to assign orthologs using 4103
 Paraneoptera single-copy orthologs from OrthoDB (*Pediculus humanus, Rhodnius prolixus**, and *Acyrthosiphon pisum*)

Methods: assemble NCBI dbEST data gene alignments

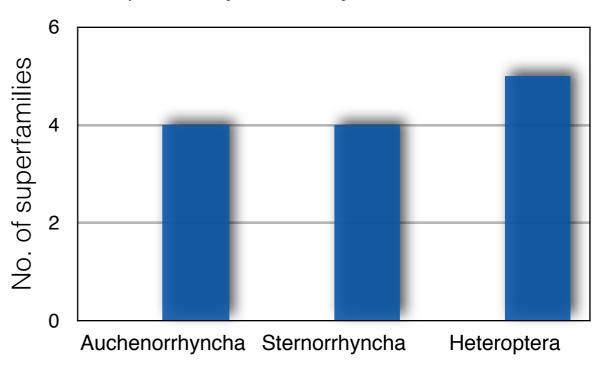
- Taxa with > 6,000 reads
- removed vector, low-complexity reads, and Ribosomal RNA
- assembled and identified orthologs the same as NGS transcriptomes

Results: taxonomic diversity of dbEST and transcriptome data

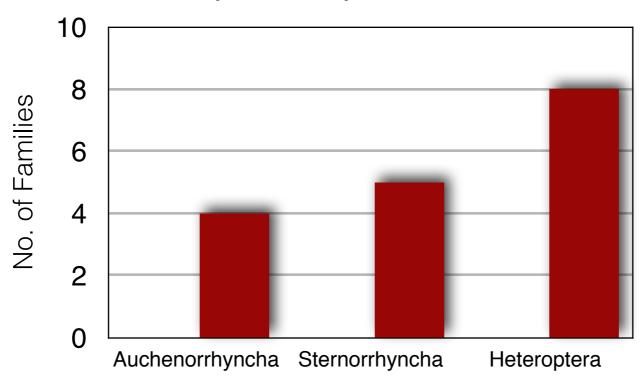




Superfamly diversity within suborders



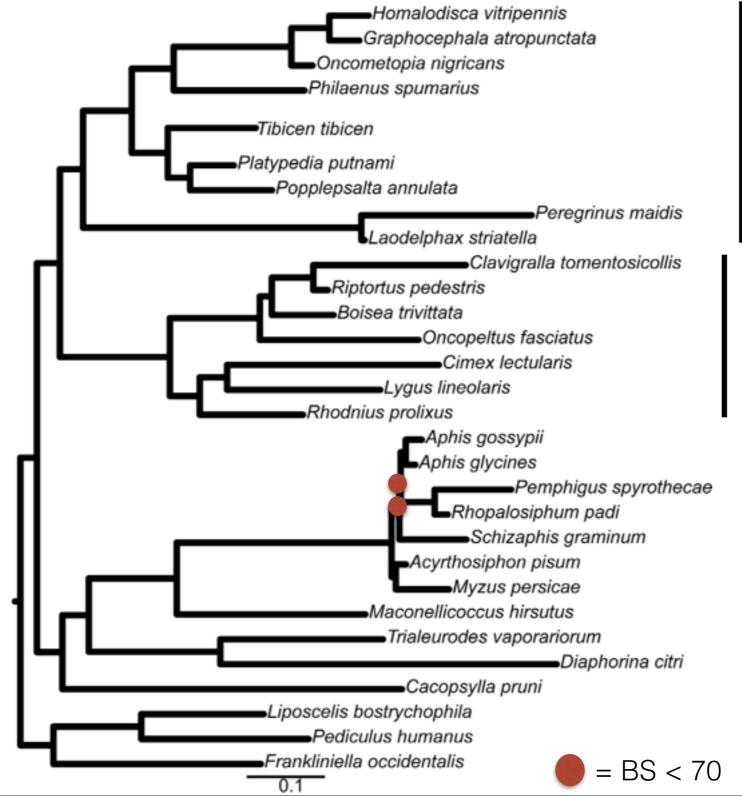
Family diversity within suborders



Hemiptera phylogeny: transcriptome & dbEST data

ML: 35 loci; 15,508AA;

partitioned; 100BS reps



Auchenorrhyncha

Graminella nigrifrons, Nilaparvata lugens, Magicicada septendecim, Sogatella furcifera

Heteroptera

Arma chinensis, Lygaeus kalmii, Lygus hesperus, Triatoma rubida

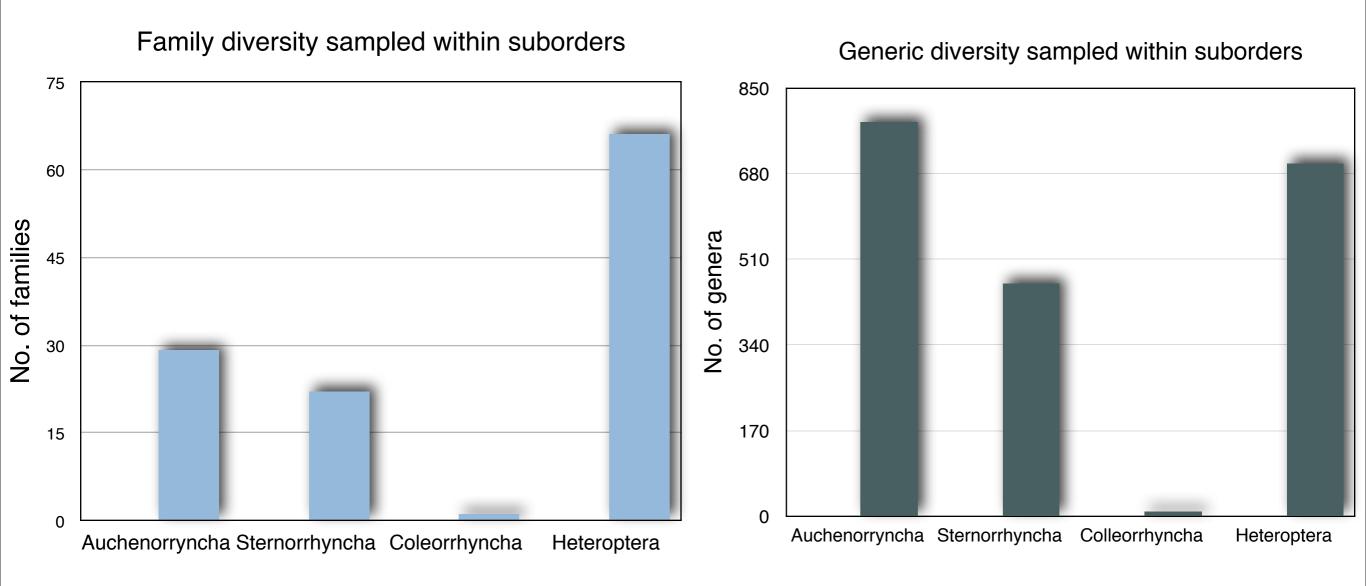
Sternorrhyncha

Aphis nerii, Pachypsylla venusta, Sitobion avenae

Methods: assemble Sanger data gene alignments

- Mega-phylogeny method using Phlawd
- GenBank release 194 (February 2013)
- 8 Loci: COI, 12S, 16S, elongation factor 1 alpha (EF1a), wingless (Wg), Histone 3 (H3), 18S, 28S
- BLAST all sequences against a non-redundant database (Genbank Release 198)
- Align protein-coding nucleotides with AA alignment and ribosomal loci by secondary structure in Mafft
- Partitions and evolutionary models estimated in PartitionFinder
- ML nucleotide and AA phylogenies estimated in RAxML and FastTree
- Outgroups: Psocoptera, Thysanoptera, and Phthiraptera

Results: taxonomic diversity of Sanger data

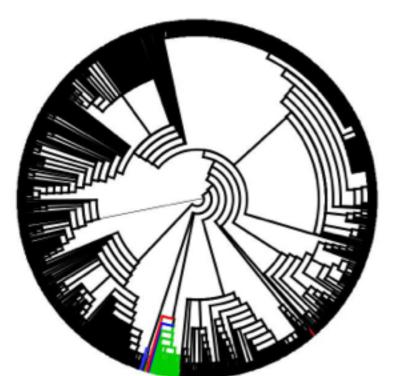


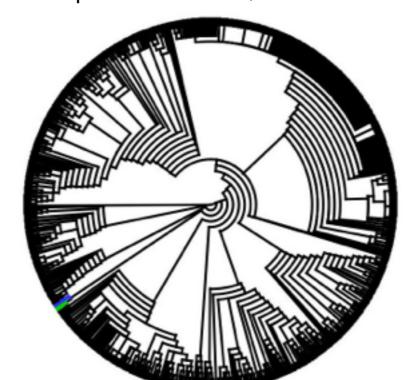
Results: Sanger data quality

- Removed ~ 5% of sequences
 - stop codons in the middle of coding regions
 - best BLAST hit to bacteria
 - incorrect locus labels

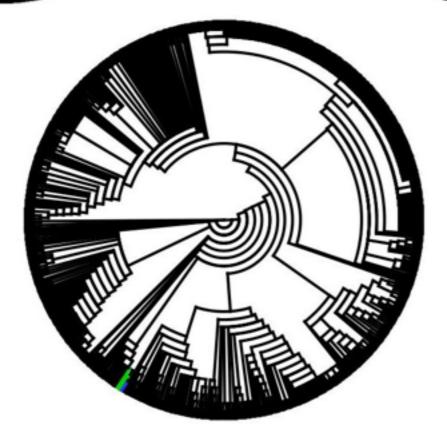
Results: gene trees and non-monophyletic suborders

Wg 1st & 2nd pos; ML partitioned; 747 taxa Wg RY-code 3rd pos; ML partitioned; 747 taxa





- Psylids
- Outgroup
- Heteroptera



Wg 1st & 2nd pos; rogue taxa removed; ML partitioned; 747 taxa

Results: concatenated phylogeny

Nucleotides: 5248 taxa, 5956 bp; 75% missing data

- Outgroup non-monphyletic
- Suborders non-monophyletic

Amino acids: 3759 taxa, 1083 AA; 70% missing data

- Outgroup non-monphyletic
- Suborders non-monophyletic

Future directions

- Continue working at the gene tree level, including the transcriptome and EST data
- Try mixture models to accommodate rate heterogeneity.
- Estimate phylogenetic decisiveness and prune taxa accordingly
- Compare nucleotide bias among loci and taxa
- Estimate a synthetic tree using Bayesian phylogeny estimates for genera, families, and tribes to use with backbone of NGS/est data

Questions?