

MYELOID NEOPLASIA

Mutational profile and benefit of gemtuzumab ozogamicin in acute myeloid leukemia

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KEY POINTS

- The benefit of GO in non-CBF AML patients with favorable or intermediate ELN 2017 risk is restricted to those with signaling mutations.
- Higher CD33 expression levels on non-CBF AML blasts correlate with the presence of activating signaling mutations.

Acute myeloid leukemia (AML) is a highly heterogeneous disease both in terms of genetic background and response to chemotherapy. Although molecular aberrations are routinely used to stratify AML patients into prognostic subgroups when receiving standard chemotherapy, the predictive value of the genetic background and co-occurring mutations remains to be assessed when using newly approved antileukemic drugs. In the present study, we retrospectively addressed the question of the predictive value of molecular events on the benefit of the addition of gemtuzumab ozogamicin (GO) to standard front-line chemotherapy. Using the more recent European LeukemiaNet (ELN) 2017 risk classification, we confirmed that the benefit of GO was restricted to the favorable (hazard ratio [HR], 0.54, 95% confidence interval [CI], 0.30-0.98) and intermediate (HR, 0.57; 95% CI, 0.33-1.00) risk categories, whereas it did not influence the outcome of patients within the adverse risk subgroup (HR, 0.93; 95% CI, 0.61-1.43). Interestingly, the benefit of GO was significant for patients with activating signaling mutations (HR, 0.43; 95% CI, 0.28-0.65), which correlated with higher CD33 expression levels. These results suggest that molecular

aberrations could be critical for future differentially tailored treatments based on integrated genetic profiles that are able to predict the benefit of GO on outcome. (Blood. 2020;135(8):542-546)

Introduction

Major advances were made over the past decades in defining underlying cytogenetic and molecular aberrations routinely used to inform acute myeloid leukemia (AML) classifications and stratify patients into prognostic subgroups.¹ Although the prognostic relevance of molecular alterations has been well investigated, translation of this information into clinical care is ongoing.² Until recently, standard induction chemotherapy in AML patients relied mostly on the "3+7" regimen combining an anthracycline plus cytarabine. However, recent studies have highlighted newly approved medications directed against AML-specific antigens or pathways.³ Given this game-changing context, there is a need to analyze the impact of these new drugs in light of the complex AML molecular landscape.

Here, we report the extensive mutational analysis of a well-annotated cohort of AML patients aged 50 to 70 years old. Patients were randomized into the Acute Leukemia French Association (ALFA)-0701 trial (ClinicalTrials.gov Identifier: NCT00927498),⁴ which evaluated the added value of fractionated doses of gemtuzumab ozogamicin (GO) to standard front-line chemotherapy.³ Results from our group⁴ and other investigators⁵⁻⁷ suggested that the addition of GO to chemotherapy might result in a substantial improvement in patient outcome, especially in AML patients with nonadverse cytogenetics⁸ and/or high CD33 expression.^{9,10} Considering these findings and the widespread use of high-throughput sequencing technologies in current clinical care, we retrospectively addressed the question of the predictive value of molecular events on the benefit of GO in the ALFA-0701 study.

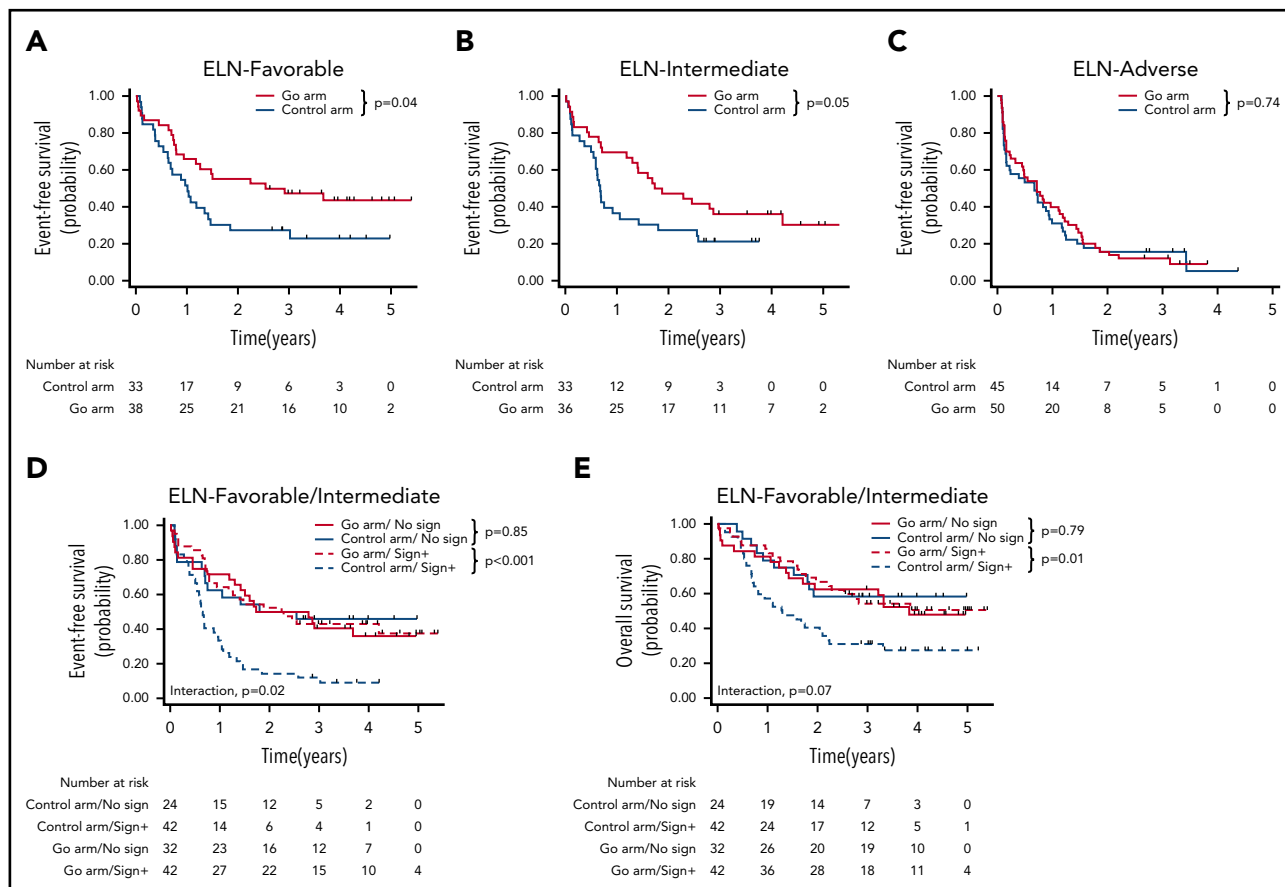


Figure 1. Benefit of GO according to ELN classification and the existence of signaling mutation. (A-C) EFS according to ELN 2017 subgroups. EFS (D) and OS (E) according to protocol arm and the existence of signaling mutations.

Study design

A total of 235 patients with non-core binding factor (CBF) AML was studied for molecular aberrations using a 43-gene targeted panel (supplemental Methods, available on the *BloodWeb* site). *FLT3*-internal tandem duplication (ITD) screening was performed by fragment analysis, as previously described,⁸ and the allelic ratio (AR) was determined as the ratio of the area under the curve *FLT3*-ITD/*FLT3* wild type. All samples were screened for recurrent gene rearrangements and *KMT2A*-partial tandem duplication using reverse transcriptase multiplex ligation-dependent probe amplification assay.¹¹ *CD33* expression was evaluated as previously described.⁹ Single-gene analyses were restricted to subgroups with >15 patients. The “signaling mutations” group was defined by the presence of ≥ 1 mutation in *FLT3*, *NRAS*, *KRAS*, *PTPN11*, *JAK2*, *RIT1*, or *CBL*, regardless of the variant allelic frequency (VAF).

Results and discussion

Mutational profiling

A total of 221 patients (94%) had ≥ 1 mutation. Eleven genes were recurrently mutated in $\geq 10\%$ of patients: *NPM1* (34%), *DNMT3A* (29%), *FLT3* (28% [*FLT3*-ITD 21%; *FLT3*-TKD 8%]), *TET2* (20%), *RUNX1* (16%), *NRAS* (14%), *IDH2* (13%), *SRSF2* (13%), *ASXL1* (13%), *TP53* (11%), and *IDH1* (11%) (supplemental Figures 1 and 2). According to the European LeukemiaNet (ELN)

2017 risk classification, 71 (30%), 69 (29%), and 95 (40%) patients had favorable, intermediate, and adverse risk, respectively. The distributions of gene mutations, ELN risk categories,¹ and *CD33* expression⁹ were well balanced between the control group and the GO group (supplemental Table 1).

Benefit of GO in molecularly defined subgroups

The global complete remission (CR) rate with or without platelet recovery was 71% (166/235), with no difference between the GO and control arms. The CR rate was 86%, 83%, and 64% in the ELN favorable-risk, intermediate-risk, and adverse-risk categories, respectively. None of the ELN risk subgroups or gene mutations benefited from GO in terms of CR rate (data not shown).

For the 235 patients, the 3-year event-free survival (EFS) was 25.7% (95% confidence interval [CI], 2.6-30.0). The benefit of GO was restricted to the ELN favorable-risk (hazard ratio [HR], 0.54; 95% CI, 0.30-0.98) and intermediate-risk (HR, 0.57; 95% CI, 0.33-1.00) categories, whereas it did not influence the outcome of patients with adverse risk (HR, 0.93; 95% CI, 0.61-1.43) (Figure 1A-C). At the molecular level, a benefit of GO was observed in patients with *NPM1* (HR, 0.48; 95% CI, 0.28-0.83), *FLT3*-ITD (HR, 0.36; 95% CI, 0.18-0.72), *FLT3*-TKD (HR, 0.20; 95% CI, 0.05-0.78), *NRAS* (HR, 0.43; 95% CI, 0.19-0.95), *KRAS* (HR, 0.27; 95% CI, 0.09-0.84), and *SRSF2* mutations (HR, 0.28; 95% CI, 0.12-0.65) (Figure 2A; supplemental Table 2; supplemental Figure 3). In contrast, no benefit was observed in patients with adverse-risk

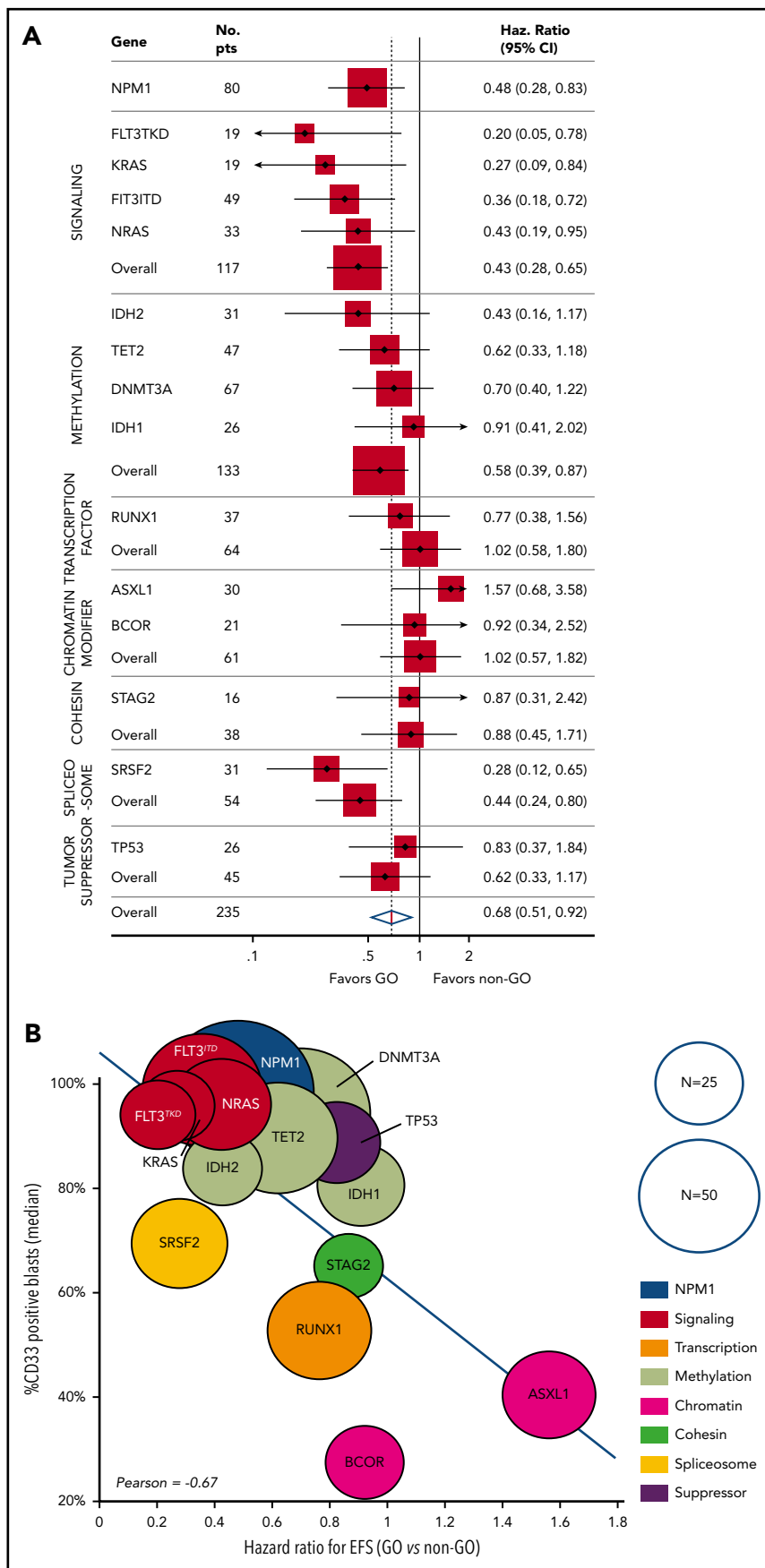


Figure 2. Benefit of GO according to mutation profile and correlation with CD33 expression. (A) The forest plot shows the HR for EFS in the GO arm vs the control arm for each mutation (with $N \geq 15$) and mutation subgroup. (B) The correlation between the HRs and CD33 expression on AML blasts. Each circle represents a subgroup of patients with a mutation. The size of each circle is proportional to the number of patients.

molecular factors¹ *RUNX1*, *ASXL1*, and *TP53*. Considering mutational classes¹² (supplemental Methods), a greater benefit of GO was observed in patients with activating signaling mutations (HR, 0.43; 95% CI, 0.28-0.65) or epigenetic (HR, 0.68; 95% CI, 0.48-0.97) or spliceosome deregulation (HR, 0.44; 95% CI, 0.24-0.80). The predominant benefit of signaling mutations led us to assess the interaction between these mutations and GO in ELN 2017 favorable/intermediate-risk patients (Figure 1D-E). With regard to EFS and overall survival (OS), patients with signaling mutations benefited from GO (EFS: HR, 0.36; 95% CI, 0.21-0.61 and OS: HR, 0.47; 95% CI, 0.26-0.83), whereas patients without signaling mutations did not (EFS: HR, 1.07; 95% CI, 0.53-2.16 and OS: HR, 1.11; 95% CI, 0.50-2.47). The interaction between a benefit of GO and the presence of a signaling mutation was significant for EFS ($P = .02$), with only a trend for OS ($P = .07$). Because many cooperating mutations may be associated in the same patients, we next assessed whether the benefit of GO observed in patients with *NPM1*, epigenetic, or spliceosome mutations was restricted to those patients with signaling mutations. In these 3 subgroups, a significant benefit in terms of EFS was only observed in patients with co-occurring signaling mutations (supplemental Figure 4). Interaction was significant for patients with *NPM1* or epigenetic mutations. Altogether, these observations suggested that a benefit of GO was restricted to patients with activating signaling mutations.

Finally, we investigated the impact of *FLT3*-ITD AR on GO benefit. An AR cutoff was defined as the median value of 0.4 (supplemental Figure 5). In patients with *FLT3*-ITD, there was no difference in HR (GO vs control) for EFS between the high-AR group (HR, 0.42; 95% CI, 0.16-1.08) and the low-AR group (HR, 0.35; 95% CI, 0.12-0.99). We then considered a composite VAF for signaling mutations as the sum of individual gene VAFs. A VAF cutoff was defined as the median value of 25%. A slightly lower HR for EFS was observed in patients with high VAF (HR, 0.36; 95% CI, 0.19-0.66) compared with patients with low VAF (HR, 0.50; 95% CI, 0.28-0.92). However, both profiles led to a benefit of GO in terms of EFS.

Correlation between benefit of GO and CD33 expression

A correlation between GO activity and CD33 target expression was suggested by our group⁹ and other investigators.¹⁰ To further explore the association between signaling mutations and benefit of GO, we compared the presence of mutations with CD33 expression levels on AML blasts. Interestingly, the benefit of GO in terms of EFS was correlated with CD33 expression levels among the different gene mutations, with particularly high levels observed on signaling mutation-positive AML blasts (Pearson correlation coefficient, -0.67 ; Figure 2B). Among patients with epigenetic mutations, the level of CD33 expression was significantly higher in patients with signaling mutations (98% vs 60%; $P < .001$). This difference was not observed in patients with *NPM1* or spliceosome mutations.

A biological mechanism linking CD33 expression and signaling mutations is still missing. Such molecular events co-occur in a wide range of hematological malignancies and are generally considered secondary or late clonal events.¹³ Signaling mutations are found in up to 75% of patients with CBF AML,¹³ a group for which a clear benefit of GO was demonstrated,⁵ despite a characteristically low expression of CD33.¹⁰ With no evidence of

direct control of activating signaling on CD33 expression, one could hypothesize that the interaction observed could reflect leukemic cell adaptation to its microenvironment. CD33 belongs to the Siglec family and is a transmembrane glycoprotein receptor committed to myeloid lineage. CD33 harbors 2 tyrosine residues on its cytoplasmic domain that function as docking sites for the tyrosine phosphatases SHP-1 and SHP-2.¹⁴ Thus, CD33 is considered an inhibitory receptor that negatively regulates cell proliferation and activation.¹⁵⁻¹⁷ In this context, activated kinase signaling could be a coevolutionary pattern that overcomes CD33 downstream effects on leukemic cell development.

In conclusion, in the context of the ALFA-0701 trial, we showed that the benefit of GO in patients is determined by underlying molecular aberrations and is primarily restricted to patients bearing signaling mutations associated with high CD33 expression. This observation could be critical for future differentially tailored treatments based on integrated genetic profiles. Also, future investigations should evaluate the relevance of the use of GO considering other targeted therapy, including FLT3 inhibitors.

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Authorship

Contribution: E.F., N.D., B.D., A.M.-R., M.C., and C. Preudhomme performed molecular analyses; E.G. reviewed flow cytometry data; C.T. reviewed cytogenetics; E.R., P.T., D.C., X.T., S. Chantepie, J.-V.M., E.L., K.C.-L., J.L., C. Pautas, H.D., S. Castaigne, C. Preudhomme, and N.B. contributed to the execution of the study and collected and assembled the data; and E.F., N.D., and N.B. wrote the manuscript, which was approved by all of the authors.

Conflict-of interest disclosure: S. Castaigne has received consultancy fees from Pfizer, and H.D. has received honoraria from Pfizer for serving on advisory boards. The remaining authors declare no competing financial interests.

A complete list of the members of the Acute Leukemia French Association appears in "Appendix."

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Footnotes

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All requests for sequence data, BAM files, and VCF files should be sent to Nicolas Boissel (nicolas.boissel@aphp.fr).

The online version of this article contains a data supplement.

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REFERENCES

- Döhner H, Estey E, Grimwade D, et al. Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. *Blood*. 2017;129(4):424-447.
- Meyer SC, Levine RL. Translational implications of somatic genomics in acute myeloid leukaemia. *Lancet Oncol*. 2014;15(9):e382-e394.
- Walasek A. The new perspectives of targeted therapy in acute myeloid leukemia. *Adv Clin Exp Med*. 2019;28(2):271-276.
- Castaigne S, Pautas C, Terré C, et al; Acute Leukemia French Association. Effect of gemtuzumab ozogamicin on survival of adult patients with de-novo acute myeloid leukaemia (ALFA-0701): a randomised, open-label, phase 3 study. *Lancet*. 2012; 379(9825):1508-1516.
- Burnett AK, Hills RK, Milligan D, et al. Identification of patients with acute myeloblastic leukemia who benefit from the addition of gemtuzumab ozogamicin: results of the MRC AML15 trial. *J Clin Oncol*. 2011;29(4):369-377.
- Burnett AK, Russell NH, Hills RK, et al. Addition of gemtuzumab ozogamicin to induction chemotherapy improves survival in older patients with acute myeloid leukemia. *J Clin Oncol*. 2012;30(32):3924-3931.
- Hills RK, Castaigne S, Appelbaum FR, et al. Addition of gemtuzumab ozogamicin to induction chemotherapy in adult patients with acute myeloid leukaemia: a meta-analysis of individual patient data from randomised controlled trials. *Lancet Oncol*. 2014;15(9):986-996.
- Renneville A, Abdelali RB, Chevret S, et al. Clinical impact of gene mutations and lesions detected by SNP-array karyotyping in acute myeloid leukemia patients in the context of gemtuzumab ozogamicin treatment: results of the ALFA-0701 trial. *Oncotarget*. 2014;5(4):916-932.
- Olombel G, Guerin E, Guy J, et al. The level of blast CD33 expression positively impacts the effect of gemtuzumab ozogamicin in patients with acute myeloid leukemia. *Blood*. 2016; 127(17):2157-2160.
- Pollard JA, Alonzo TA, Loken M, et al. Correlation of CD33 expression level with disease characteristics and response to gemtuzumab ozogamicin containing chemotherapy in childhood AML. *Blood*. 2012; 119(16):3705-3711.
- Ruminy P, Marchand V, Buchbinder N, et al. Multiplexed targeted sequencing of recurrent fusion genes in acute leukaemia. *Leukemia*. 2016;30(3):757-760.
- Döhner H, Weisdorf DJ, Bloomfield CD. Acute myeloid leukemia. *N Engl J Med*. 2015; 373(12):1136-1152.
- Itzykson R, Duployez N, Fasan A, et al. Clonal interference of signaling mutations worsens prognosis in core-binding factor acute myeloid leukemia. *Blood*. 2018;132(2):187-196.
- Paul SP, Taylor LS, Stansbury EK, McVicar DW. Myeloid specific human CD33 is an inhibitory receptor with differential ITIM function in recruiting the phosphatases SHP-1 and SHP-2. *Blood*. 2000;96(2):483-490.
- Vitale C, Romagnani C, Puccetti A, et al. Surface expression and function of p75/AIRM-1 or CD33 in acute myeloid leukemias: engagement of CD33 induces apoptosis of leukemic cells. *Proc Natl Acad Sci USA*. 2001; 98(10):5764-5769.
- Lajaunias F, Dayer J-M, Chizzolini C. Constitutive repressor activity of CD33 on human monocytes requires sialic acid recognition and phosphoinositide 3-kinase-mediated intracellular signaling. *Eur J Immunol*. 2005;35(1):243-251.
- Balaian L, Zhong RK, Ball ED. The inhibitory effect of anti-CD33 monoclonal antibodies on AML cell growth correlates with Syk and/or ZAP-70 expression. *Exp Hematol*. 2003;31(5):363-371.