BC205: Algorithms for Bioinformatics.

II. Sequence Analysis

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Sequence Analysis

- In this class we will start looking into real biological problems, focusing on sequence analysis
- · We will discuss some very basic concepts of computation such as hashing
- We will then turn to the implementation of the things we learnt last time using Brute
 Force and Divide and Conquer Approaches
- We will discuss a new approach (Binary Search) and implement it the context of a Genome Analysis

The biological problems:

- · Compare different species on the basis of DNA composition
- · Find evidence of horizontal gene transfer in a bacterial genome
- · Locate the Origin of Repication of a Bacterial Genome

Bioinformatics Warm-Up

- 1. You are given a DNA sequence
 - Can you count the number of nucleotides of each of the four bases (A, G, C, T)?
 - · How many calculations will you need?
 - · How will you implement it?
- 2. Now consider the same problem only instead of nucleotides we need to count the number of all 8-nucleotides. What do you need to consider to attack the problem?

Aspects of DNA Composition

- GC content
- genomic signatures
- · parity distributions
- k-mer frequencies

GC content

We call GC content (or GC%) the ratio of (G+C) nucleotides of a given DNA sequence

• Why is it important? G-C pairs are linked with 3 hydrogen bonds, while A-T ones with 2. High GC genomes are more stable in terms of physical chemistry.

GC is related to:

- · Biochemical level: Thermal stability
- Evolutionary level: Organism Phylogeny, Mutational pressures
- Genomic level: Genome size
- Functional level: Functional role of underlying sequences
- · and many more

GC content in Genomic Sequences

- Bacteria: GC% is highly variable between species
- Bacteria: GC% is rather homogeneous within each genome
- Bacteria: GC% can be used in their classification

GC content in Genomic Sequences

- Eukaryotes: Very homogeneous overall GC% (~40-45% in all animals)
- Eukaryotes: Fluctuation of GC contentalong the chromosomes and organization in areas of (rather) stable GC%
- Eukaryotes: Regions of stable high/low GC content that segregate mammalian genomes in isochores

Problem 1: GC content in Bacterial Genomes

- Given the DNA sequence of a Bacterial Genome, calculate its GC content:
 - Read the Sequence
 - Enumerate G
 - Enumerate C
 - Divide (G+C) over length of the sequence

GCContent. Pseudocode

- The idea is to **exhaustively** enumerate all mononucleotides, therefore our approach is a very basic Brute Force approach.
- Given that the content of the sequence is unknown we have no other choice.
- We will proceed by reading each nucleotide in the sequence and check its value. Then increment a variable each time we find a G or a C.

GCContent: Implementation (naive)

```
In [4]: # Naive GC content
        import regex as re
        f=open('files/ecoli.fa', 'r')
        seq = ""
        window=1000
        total = 0
        nG=nC=0
        GCCont=0
        times=0;
        for line in f:
            x=re.match(">", line)
            if x == None:
                length=len(line)
                total=total+length
                 seq=seq+line[0:length-1]
        for k in range(len(seq)):
            if(seq[k]=="G"):
                nG+=1
            elif(seq[k]=="C"):
                nC+=1
        GCContent=(nG+nC)/len(seq)
        print(GCContent)
```

0.5074167653333127

GCContent: Implementation (using Python's count function)

```
In [5]: import regex as re
        f=open('files/staaur.fa', 'r')
        seq = ""
        window=1000
        total = 0
        nG=nC=0
        GCCont=0
        times=0;
        for line in f:
            x=re.match(">", line)
            if x == None:
                length=len(line)
                total=total+length
                seq=seq+line[0:length-1]
        f.close()
        nC=seq.count("C")
        nG=seq.count("G")
        GCCont=(nG+nC)/len(seq);
        print(GCCont)
```

Hands on #1:

- Download a couple of bacterial genome sequences from ENSEMBL Bacteria (http://bacteria.ensembl.org/index.html)
- · Implement GC content
- · Report the results
- An example would be

Genome	GC
a-Bac1	0.334
e-Bac2	0.595
e-Bac3	0.668
g-Bac4	0.409
e-Bac5	0.551
a-Bac6	0.352
a-Bac7	0.354
g-Bac8	0.418
g-Bac9	0.434
e-Bac8	0.627

Problem 2: Variability of GC content *between* Bacterial Genomes

- Given a number of bacterial genomes:
 - Get their genome sequences
 - Calculate the GC contents
 - Calculate differences between the GC contents
 - Rank genomes based on their differences
- · Pseudocode:
 - Perform GC_content on each of the genomes you downloaded
 - Calculate D_(i,j)=|GC_i-GC_j| over all i,j
 - Sort D (i,j)

Problem 2: Approach

We could do this very easily with R but it can also be done otherwise

For example

```
In [1]: f=open('files/GCContent.tsv', 'r')
        i=0
        GCC={}
        for line in f:
            i=i+1
            if(i>1):
                species=line.split()[0]
                GC=line.split()[1]
                GCC[species]=float(GC)
        gcdistances={}
        for genome1 in GCC.keys():
            for genome2 in GCC.keys():
                pair=genome1+":"+genome2
                gcdistances[pair]=abs(float(GCC[genome1])-float(GCC[genome2]))
                gcdistances[pair]=round(gcdistances[pair],2)
                #print(pair, round(gcdistances[pair],3))
        sorted(gcdistances.items(), key=lambda x: x[1])
```

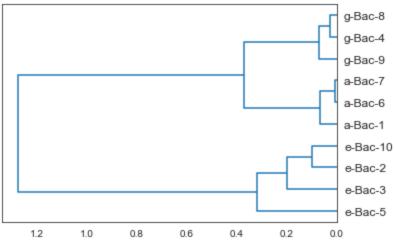
```
Out[1]: [('a-Bac1:a-Bac1', 0.0),
         ('e-Bac2:e-Bac2', 0.0),
         ('e-Bac3:e-Bac3', 0.0),
         ('g-Bac4:g-Bac4', 0.0),
         ('e-Bac5:e-Bac5', 0.0),
         ('a-Bac6:a-Bac6', 0.0),
         ('a-Bac6:a-Bac7', 0.0),
         ('a-Bac7:a-Bac6', 0.0),
         ('a-Bac7:a-Bac7', 0.0),
         ('g-Bac8:g-Bac8', 0.0),
         ('g-Bac9:g-Bac9', 0.0),
         ('e-Bac10:e-Bac10', 0.0),
         ('g-Bac4:g-Bac8', 0.01),
         ('g-Bac8:g-Bac4', 0.01),
         ('a-Bac1:a-Bac6', 0.02),
         ('a-Bac1:a-Bac7', 0.02),
         ('a-Bac6:a-Bac1', 0.02),
         ('a-Bac7:a-Bac1', 0.02),
         ('q-Bac8:q-Bac9', 0.02),
         ('g-Bac9:g-Bac8', 0.02),
         ('e-Bac2:e-Bac10', 0.03),
         ('g-Bac4:g-Bac9', 0.03),
         ('g-Bac9:g-Bac4', 0.03),
         ('e-Bac10:e-Bac2', 0.03),
         ('e-Bac3:e-Bac10', 0.04),
         ('e-Bac10:e-Bac3', 0.04),
         ('e-Bac2:e-Bac5', 0.05),
         ('g-Bac4:a-Bac7', 0.05),
         ('e-Bac5:e-Bac2', 0.05),
         ('a-Bac7:g-Bac4', 0.05),
         ('g-Bac4:a-Bac6', 0.06),
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         ('a-Bac7:g-Bac8', 0.06),
         ('g-Bac8:a-Bac7', 0.06),
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         ('e-Bac3:e-Bac2', 0.07),
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         ('a-Bac6:g-Bac9', 0.08),
         ('a-Bac7:g-Bac9', 0.08),
         ('q-Bac8:a-Bac1', 0.08),
         ('g-Bac9:a-Bac6', 0.08),
         ('g-Bac9:a-Bac7', 0.08),
         ('e-Bac5:e-Bac10', 0.09),
         ('e-Bac10:e-Bac5', 0.09),
         ('a-Bac1:g-Bac9', 0.1),
         ('g-Bac9:a-Bac1', 0.1),
         ('e-Bac5:g-Bac9', 0.11),
         ('g-Bac9:e-Bac5', 0.11),
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         ('g-Bac4:e-Bac5', 0.13),
```

```
('e-Bac5:e-Bac3', 0.13),
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('g-Bac9:e-Bac2', 0.16),
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('a-Bac7:e-Bac5', 0.19),
('g-Bac9:e-Bac10', 0.19),
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('e-Bac10:g-Bac4', 0.22),
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('g-Bac4:e-Bac3', 0.26),
('a-Bac7:e-Bac10', 0.27),
('e-Bac10:a-Bac7', 0.27),
('a-Bac6:e-Bac10', 0.28),
('e-Bac10:a-Bac6', 0.28),
('a-Bac1:e-Bac10', 0.29),
('e-Bac10:a-Bac1', 0.29),
('e-Bac3:a-Bac7', 0.31),
('a-Bac7:e-Bac3', 0.31),
('e-Bac3:a-Bac6', 0.32),
('a-Bac6:e-Bac3', 0.32),
('a-Bac1:e-Bac3', 0.33),
('e-Bac3:a-Bac1', 0.33)]
```

Notice how smaller values are obtained for same bacterial family (a-, g- and e-proteobacteria)

Let's now look how we can use this simple quantity to infer relationships between different genomes. In the following we make use of some python functions to organize the genomes in a tree structure that resembles the way evolutionary biologists try to infer phylogenetic relationships.

```
import pandas as pd
import seaborn as sns
import matplotlib.pyplot as plt
from scipy.spatial.distance import pdist, squareform
from scipy.cluster.hierarchy import linkage, dendrogram
# Load the dataframe and assign values/labels
df = pd.read csv('files/GCContent simple.csv')
dvalues = df['GCContent'].values.reshape(-1,1)
dlabels = list(df['Genome'])
# Calculate the distances
distances = pdist(dvalues)
# Convert the pairwise distances into a square distance matrix
distance matrix = squareform(distances)
# Calculate the linkage matrix using Ward's method
linkage matrix = linkage(distance matrix, method='ward')
# Plot the dendrogram
sns.set style('white')
dendrogram(linkage matrix, labels=dlabels, color threshold=0, orientation='l
# Show the plot
plt.show()
<ipython-input-4-c652b4de09d1>:21: ClusterWarning: scipy.cluster: The symme
tric non-negative hollow observation matrix looks suspiciously like an unco
ndensed distance matrix
  linkage matrix = linkage(distance matrix, method='ward')
                                             g-Bac-8
                                             g-Bac-4
                                             g-Bac-9
                                             a-Bac-7
                                             a-Bac-6
```



Problem 3: What about different regions of the genome?

- We just saw how genomic GC% values may be used to draw conclusions for bacterial phylogeny
- But: How representative is the GC% value you calculated above?

And: How efficiently can it be used to describe a genome?

Problem 3: Why should we care?

- We mentioned that GC% is stable within bacterial genomes
- But Some areas of bacterial genomes are special
- Parts of the bacterial genome have been "horizontally" (as opposed to vertically, i.e. from their "mom") transferred from other species.

Problem 3: Stability of GC content *along* Bacterial Genomes

- Regions of "strange", or "deviating" GC% values in a given genome are red flags of HGT. The problem now is:
 - Given a bacterial genome sequence:
 - Locate regions of the genome where horizontal gene transfer may have occurred.

Problem 3: Approach

- Choose a window to scan your sequence. This will be your resolution
- · Calculate GC per window
- Try to locate GC values that deviate from the genome average

Problem 3: The core

 We basically repeat the approach for GC content but now we calculate one value for each window

```
In [9]: import regex as re
    f=open('files/Staaur.fa', 'r')

seq = ""
    window=1000
    nG=nC=0
    GCCont=[]
    total=0

for line in f:
        x=re.match(">", line)
        if x == None:
            length=len(line)
            total=total+length
            seq=seq+line[0:length-1]
    f.close()

step=100
```

```
times=int(len(seq)/step);

for i in range(times):
    DNA=seq[i*step:i*step+window]
    nC=DNA.count("C")
    nG=DNA.count("G")
    GCCont.append((nG+nC)/window)

for k, value in enumerate(GCCont[1:10]):
    print(k*step, "\t", value)

0    0.321
100    0.322
```

```
100
         0.322
200
         0.322
        0.311
300
400
        0.315
        0.317
500
600
        0.312
        0.313
700
800
        0.313
```

Hands-on #2:

- Get the genome sequence of St. aureus
- · Implement Sliding GC
- · Locate positions in the genome with extreme values of GC
- The problem is: What do we mean by "extreme values"?* How do we define "extreme"?

Problem 3: Statistics Interlude

- Given a set/sample of values, how can we decide on whether a value could be part of that sample or not?
- In our problem: We know that the GC% of bacteria tends to be characteristic of the genome. Can we "spot" regions of the genome that bear GC% values that are *different* from that characteristic value?
- Q1: How will we define that characteristic value?
- Q2: How will we quantify the difference as big enough or not?

Problem 3: Theoretical basis (simplified)

- Central Limit Theorem (simplified):
 - Regardless of the underlying distribution, the mean of a large number of samples follow the normal distribution.
 - We can thus model GC values per window based on the normal distribution

Problem 3: The statistics

- We will model the "characteristic value" as the mean of GC values for all windows
- · We will also calculate the standard deviation of these values to assess the variance
- · We will then apply...

Z-transformation

- Given a value x, we can compare x to a normal distribution with mean=m and standard deviation=std with the z-score: Z(x)= (x - m)/std
 - Z(x) is thus the difference of x from m in units of standard deviation.

Knowing that in a normal distribution ~99.5% of the values fall within +/-3*std a value of Z(x)>3 or Z(x)<-3 makes it highly unlikely that x is part of our distribution.

Problem 3. Predicting HGT locations

 We can now combine sliding GC content calculations with a Z-score transformation and a filtering for |Z|>=3

```
In [16]: import regex as re
         import numpy as np
         f=open('files/Staaur.fa', 'r')
         seq = ""
         window=1000
         nG=nC=0
         GCCont=[]
         for line in f:
             x=re.match(">", line)
             if x == None:
                 length=len(line)
                  seq=seq+line[0:length-1]
         f.close()
         step=100
         times=int(len(seq)/step);
         for i in range(times):
             DNA=seq[i*step:i*step+window]
             nC=DNA.count("C")
             nG=DNA.count("G")
             GCCont.append((nG+nC)/window)
         # Calculate Z-scores
         mGC=np.mean(GCCont)
         sdGC=np.std(GCCont)
         zGC=(GCCont-mGC)/sdGC
         for i in range(len(zGC)):
             if abs(zGC[i])>=3:
                  print(i*step, zGC[i])
```

```
2764699
```

- 311700 3.3056871660938962
- 311800 3.510729542254593
- 311900 3.7450636864382467
- 312000 3.80364722248416
- 312100 3.6571883823693767
- 312200 3.422854238185723
- 446200 -3.138501798956574
- 446300 -3.109210030933617
- 610000 3.0713530219102427
- 610100 4.125856670736683
- 610200 4.272315510851466
- 610300 4.77027556724173
- 610400 5.121776783517211
- 610500 5.180360319563124
- 610600 6.000529824205912
- 610700 6.088405128274782
- 610800 6.146988664320696
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- 611000 5.561153303861561
- 611100 5.443986231769735
- 611200 5.736903911999302
- 611300 5.414694463746778
- 611400 5.268235623631994
- 011100 5.20025502505155.
- 611500 5.180360319563124
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- 613300 3.129936557956156
- 613400 3.2763953980709397
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- 613600 4.096564902713726
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- 612000 5 205402605722021
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- 615400 3.6571883823693767

```
615500 4.623816727126947
```

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- 615900 6.146988664320696
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- 616100 5.795487448045215
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- 616300 5.414694463746778
- 616400 5.590445071884518
- 616500 5.268235623631994
- 616600 5.180360319563124
- 616700 4.975317943402427
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- 616000 4 00744363033555
- 616900 4.887442639333557
- 617000 4.77027556724173
- 617100 4.77027556724173
- 617200 4.711692031195817
- 617300 4.59452495910399
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- 618100 3.0713530219102427
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- 618300 3.2471036300479827
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- 618500 3.510729542254593
- 618600 3.6864801503923337
- 618700 4.008689598644858
- 618800 4.68240026317286
- 618900 4.828859103287644
- 619000 5.356110927700864
- 619100 5.268235623631994
- 619200 5.180360319563124
- 619300 5.502569767815648
- 619400 5.356110927700864
- 619500 5.356110927700864
- 619600 5.092485015494254
- 619700 4.33089904689738
- 019/00 4.33009904009/30
- 619800 4.272315510851466
- 619900 4.301607278874423
- 620000 4.360190814920336
- 620100 4.59452495910399
- 620200 4.067273134690771
- 620300 3.832938990507117 620400 4.067273134690771
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- 620600 3.832938990507117
- 020000 3.03293699030/11/
- 620700 3.7743554544612037
- 620800 3.1006447899331993
- 659300 3.129936557956156 659400 3.422854238185723
- 659500 3.9793978306219007

```
659600 4.360190814920336
```

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- 659800 5.092485015494254
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1688900 3.2471036300479827
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- 1689700 4.68240026317286
- 1689800 4.77027556724173
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- 1802000 4.360190814920336
- 1802100 5.092485015494254
- 1802200 5.297527391654951
- 1802300 5.209652087586081
- 1802400 5.531861535838605
- 1802500 5.121776783517211
- 1802600 5.297527391654951
- 1802700 5.297527391654951
- 1802800 4.59452495910399
- 1802900 4.096564902713726
- 1803000 3.3349789341168528
- 1804000 3.2763953980709397
- 1804100 3.9501060625989437
- 1804200 4.184440206782596
- 1804300 4.096564902713726
- 1804400 4.360190814920336
- 1804500 4.389482582943293
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- 1804800 4.740983799218774
- 1804900 4.740983799218774

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1805000 4.799567335264687
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- 1805100 4.916734407356514
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- 1805400 5.473277999792691
- 1805500 5.590445071884518
- 1805600 5.502569767815648
- 1805700 5.736903911999302
- 1805800 6.088405128274782
- 1805900 6.264155736412522
- 1806000 6.146988664320696
- 1806100 5.649028607930432
- 1806200 5.209652087586081
- 1806300 5.092485015494254
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Problem 2: Revisited

- Background DNA composition has some functional role besides simply reflecting mutational pressures
- This means that in some cases we need to know why the local composition is guided by other aspects of molecular evolutio. E.g. why would rRNA genes be G+C-rich even in AT-rich genomes?
- We need to find a way to control for background nucleotide composition

Problem 2 Revisited: Distinguishing between genomes through their sequence composition

- 1. Going beyond the GC content
- 2. Going beyond simple bases (mononucleotides, k=1)
- 3. Analyzing all dinucleotide frequencies of k=2
- Pseudocode:
 - For each kmer in 4^k k-mers
 - Calculate N(kmer)
 - Create a table

Problem: How to count k-mer frequencies

- For mononucleotides we did it with a Brute Force approach. However the mononucleotides are 4. The k-mers are 4^k.
- How can we count the frequencies of k-mers?
 - 1. Do we need **all** k-mers?
 - 2. Do we need to check each k-mer at every step?
- How many calculations do we need if we answer "yes" to 1,2 above.

Solution: Hashing Strategy instead of Brute Force

- Read the sequence in chunks of *k* nucleotides
- For each subsequence increment a dictionary value with the subsequence as key

Problem 2 Revisited: K-mer frequencies

```
In [7]: import regex as re
        f=open('files/Staaur.fa', 'r')
        seq = ""
        k=2
        kmers={}
        for line in f:
           x=re.match(">", line)
           if x == None:
                length=len(line)
                seq=seq+line[0:length-1]
        f.close()
        for i in range(len(seq)-k):
            DNA=seq[i:i+k]
            if DNA not in kmers.keys():
                kmers[DNA]=1
            else:
                kmers[DNA]+=1
        {k: float(v) / len(seq) for k, v in kmers.items()}
```

```
Out[7]: {'GT': 0.052417641124766205,
         'TA': 0.09542485456825499,
         'AT': 0.11214421533772755,
         'TT': 0.12211817633673684,
         'AC': 0.0527258121046812,
         'CT': 0.04827324782914885,
         'TC': 0.05299347234545244,
         'CA': 0.06572144020018092,
         'AA': 0.12267628410904767.
         'CG': 0.02563642552046353,
         'TG': 0.06441677737793518,
         'GG': 0.02547908470325341,
         'CC': 0.025711298047273862,
         'AG': 0.04831231175618033,
         'GA': 0.05203640613318122,
         'GC': 0.033911829099659674}
```

Problem 2 Revisited: A table of 4^k frequencies of occurrence

Base	Α	Т	G	С
Α	0.090	0.112	0.048	0.053
Т	0.095	0.090	0.064	0.053
G	0.052	0.052	0.023	0.034
С	0.066	0.048	0.026	0.023

- Values may be seen as "probabilities" of finding each k-mer in the sequence
- Can we use the notion of the probability to modify the table so that we get rid of the background nucleotide composition?

Problem 2 Revisited: Removing Background Composition

- The problem stated above persists at the level of k-mers: The background DNA composition may affect our results
- At the k-mer level we can remove the background using ratios of observed/expected frequencies
- Which is the expected frequency of a given k-mer?

Problem 2 Revisited: Observed/Expected(o/e) k-mer frequencies

- · Mathematics Interlude:
 - Assume two events A, B that are linked with each other

- We then say that A and B are dependent (or conditioned) and we have a "conditional probability" of A happening given B is also happening
- We can think of k-mers the same way: a k-mer is more probable to occur if its constituent mono-mers are occurring
- Bottomline: Any given k-mer's frequency of occurrence is dependent on the frequencies of occurrence of its mononucleotides. Thus:

Given a k-mer of length k the o/e-ratio frequency is defined as:

$$R[N_1N_2..N_k] = F[N_1N_2..N_k]/(F[N_1]F[N_2]..F[N_k])$$

In this way we can define a new table of modified frequencies that is independent of mononucleotide composition

Problem 2 Revisited: Observed/Expected K-mer frequencies

```
In [8]: import regex as re
        f=open('files/Staaur.fa', 'r')
        seq = ""
        k=2
        kmers={}
        for line in f:
            x=re.match(">", line)
            if x == None:
                length=len(line)
                seq=seq+line[0:length-1]
        f.close()
        pnuc={}
        for i in range(len(seq)):
            nuc=seq[i]
            if nuc not in pnuc.keys():
                pnuc[nuc]=1
            else:
                pnuc[nuc]+=1
        pnuc={k: float(v) / len(seq) for k, v in pnuc.items()}
        for i in range(len(seg)-k):
            DNA=seq[i:i+k]
            if DNA not in kmers.keys():
                kmers[DNA]=1
            else:
                kmers[DNA]+=1
        kmers={k: float(v) / len(seq) for k, v in kmers.items()}
        rkmers=kmers
```

```
for kmer in kmers.keys():
    nuc1=list(kmer)[0]
    nuc2=list(kmer)[1]
    rkmers[kmer]=round(kmers[kmer]/(pnuc[nuc1]*pnuc[nuc2]),3)
print(rkmers)
```

```
{'GT': 0.955, 'TA': 0.848, 'AT': 0.997, 'TT': 1.088, 'AC': 0.949, 'CT': 0.872, 'TC': 0.957, 'CA': 1.183, 'AA': 1.088, 'CG': 0.946, 'TG': 1.174, 'GG': 0.949, 'CC': 0.94, 'AG': 0.878, 'GA': 0.946, 'GC': 1.252}
```

Problem 2 Revisited: A table of o/e 4^k frequencies of occurrence

Base	Α	G	С	Т
Α	0.800	0.997	0.878	0.949
G	0.848	0.799	1.174	0.957
С	0.946	0.955	0.848	1.252
Т	1.183	0.872	0.946	0.841

- Notice how values now go >1. What does this mean?
- How is this table better (or not) than the previous one?

Genomic Signatures: Comparing o/e k-mer composition

- Genomic Signatures are defined as the table of o/e k-mers for a given genome
- We can use these tables to analyze distances between genomes. (Hint: even eukaryote genomes!)

Hands-on #3:

- Get chromosome 1 from (human, mouse, fly, worm, yeast)
- Use a genomic signature approach to cluster genomic signatures from different genomes
- Calculate the distance between rho xy(p) and rho xy(s) to create a table of distances.

Problem 4: Finding the DNA Replication in a bacterial genome

- Bacterial Genomes replicated their genome starting at one point and proceeding towards the opposite point in the circular genome from both directions.
- Bacterial genomes also have a particular distribution of nucleotides along their genome

- The difference of A-T (and G-C) complementary nucleotides goes through a sort of "phase transition" that splits the genome approximately in half.
- Do you know what this split is?
- · Do you know why it is so?

How is this related to Sequence Analysis?

- Due to the pioneering work of E. Chargaff we know that A~T and G~C in singlestranded DNA
- We know that this holds for all complete genomes except very few exceptions
- The exceptions are the few genomes that **do not** replicate symmetrically
- DNA-strand parity:
 - Strand X is replicated in-continuously
 - Accumulates more substitutions
 - If substitutions are biased the strand will guide the change in both strands through base-pairing

Approaching the problem

- We thus expect (and observe) the parity to be violated and that this violation occurs symmetrically on either side of the OriC
- We are looking for a way to locate this *phase transition* in the parity violation
- We thus need:
 - A measure of the parity
 - A way to monitor this measure along the genome
 - A way to locate abrupt changes in its values

Breaking the problem into pieces

- 1. Analyze the DNA composition along the genome
- 2. Calculate a quantity that will be informative
- 3. Create a condition that will test the location of the Ori
- · Pseudocode: Given a bacterial genome:
 - Count nucleotides in windows of N base pairs
 - Calculate the scaled AT-skew as (A-T)/(A+T)
 - Create an array of the skew values along the genome
 - Locate the transition point

Problem 4: Parity Measure Implementation

```
seq = ""
for line in f:
   x=re.match(">", line)
    if x == None:
        length=len(line)
        seq=seq+line[0:length-1]
f.close()
window=1000
step=100
times=int(len(seq)/step)
for i in range(times):
        DNA=seq[i*step:i*step+window]
        A=DNA.count("A")
        T=DNA.count("T")
        C=DNA.count("C")
        G=DNA.count("G")
        print(i*step,"\t",i*step+window,"\t",float(A-T)/(A+T))
```

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0	2716200		-0.03074670571010249
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2715400		5400	-0.03468208092485549
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	2723400	-0.09798270893371758
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2723000	2724000	-0.0967741935483871
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2723500	2724500	-0.0670926517571885
2723600	2724600	-0.08469055374592833
2723700	2724700	-0.05140961857379768
	2724800	-0.03908794788273615
2723800		
2723900	2724900	-0.10289389067524116
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2729500	2730500	-0.017241379310344827
2729600	2730600	-0.0666666666666667
2729700	2730700	-0.10395314787701318
2729800	2730800	-0.14782608695652175
2729900	2730900	-0.12790697674418605
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		-0.15550225540544050
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2730300	2731300	-0.1926040061633282
2730400	2731400	-0.15789473684210525
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2730800	2731800	-0.11428571428571428
2730900	2731900	-0.10204081632653061
2731000	2732000	-0.08978328173374613
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2731300	2732300	-0.13713405238828968
2731400	2732400	-0.1354642313546423
2731500	2732500	-0.14674735249621784
2731600	2732600	-0.16314199395770393
2731700	2732700	-0.155555555555556
2731800		
~ / J ± O O O	7737800	- 0 10395804 10055 155 1
2731900	2732800 2732900	-0.16395864106351551 -0.19174041297935104

2732000	2733000	-0.2266666666666666
2732100	2733100	-0.19880418535127056
2732200	2733200	-0.1874062968515742
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2732400	2733400	-0.21036585365853658
2732500	2733500	-0.2260061919504644
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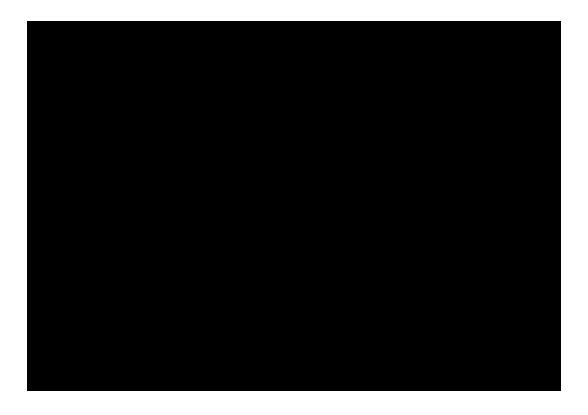
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```

Problem 4: Plotting the Values

```
In [10]: import matplotlib.pyplot as plt
import matplotlib.pyplot as plt
import regex as re

f = open('files/Staaur.fa', 'r')
```

```
seq = ""
total = 0
A=T=G=C=[]
times=0;
for line in f:
       x=re.match(">", line)
        if x == None:
                length=len(line)
                total=total+length
                seq=seq+line[0:length-1]
f.close()
x=[]
ATparity=[]
window=100000
step=10000
times=int(len(seq)/step);
for i in range(times):
   x.append(i)
    DNA=seq[i*step:i*step+window]
   A=DNA.count("A")
   T=DNA.count("T")
   C=DNA.count("C")
   G=DNA.count("G")
   ATparity.append(float(A-T)/float(A+T))
# plotting points as a scatter plot
plt.plot(x, ATparity, color= "green", linewidth=5.0)
#plt.scatter(x, ATparity, color= "green")
# x-axis label
plt.xlabel('Genome Coordinates')
# frequency label
plt.ylabel('(A-T)/(A+T)')
# plot title
plt.title('S. aureus AT parity')
# showing legend
#plt.legend()
# function to show the plot
plt.show()
```



Problem 4: Locating the breakpoint(s)

- Not a simple problem. In fact one (breakpoint detection) for which research is ongoing in many fields
- Things you could try:
 - Using derivation (checking the difference between each value and the previous one)
 - Density-based approaches: Trying to locate the region around which changes in the sign occur more robustly (i.e. given many different points around it)

Concept. Binary Searches

• Let's think of a simpler problem first:

Suppose you are given a quadratic equation: $f(x)=ax^{**}2+bx+c$ and you are asked to locate a root of the equation in an interval [k,m].

- How would you proceed?
- A fast and efficient way is to start by checking the values f(k) and f(m). If their product in f(k)f(m)<0 this means that the function "crosses" the x-axis at some point between k and m. How then can we locate that point?
- The answer is given by iterative splits of [k,m] in intervals that are always have the size of m-k. (That is [i=k, j=(k+m)/2] or [i=(k+m)/2, j=m]) and checking if the condition of f(i)f(j)<0 holds. If it does, we choose that interval and repeat.
- Question 1: What do we need to consider before starting?

• Question 2: How do we stop?

Exercises: To think about

- Use a genomic signature approach to locate possible HGT genes in the genome of *St. aureus*. Do your results of "outliers" differ from those obtained with the GC content approach?
- Write a program to locate the origin of replication for a given bacterial genome using the parity rules described in the lecture.
- The approach of the Genomic Signatures for k=2 works rather well because the k-mers are 16 but what about larger numbers of k (k=7 or more)? Would you use the same approach?

In []: