A Bayesian zero-inflated beta-binomial model for longitudinal data with group-specific changepoints

2024-08-13

Package Installed

```
library(VGAM)
                    # for rbetabinom
library(mvtnorm)
                    # for rmut proposal
library(truncnorm) # for rtruncnorm()
library(aod)
library(msm)
library(tmvtnorm)
                   # for trmut()
library(ggplot2)
library(tidyr)
library(dplyr)
library(ggplot2)
library(BayesLogit) # for rpq function
library(MCMCpack) # for Iwish update of Sigmab
library(gamlss)
                    # for ZIBB functions
```

Data Generating

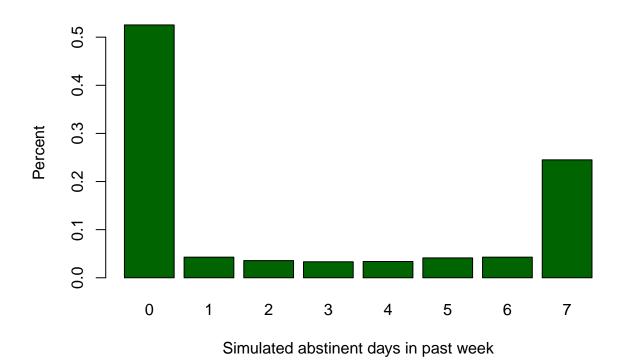
```
set.seed(2202023)
ntrial<-7
                                    # number of quest. in one ACEs score (fixed)
n<-500
                                    # total subjects
                                   # repeated measure per subject (unbalanced)
nis < -sample(1:12,n,T)
id<-rep(1:n,nis)</pre>
N<-length(id)
                                    # total observations
t < -rep(0, N)
for (i in 1:n) t[id==i] <-sort(sample(1:12,nis[i])) # time variable
                                                        # centered time
trt < -rbinom(n, 1, ...5)
                                                        # trt indicaotr
tx<-rep(trt,nis)</pre>
truekappa1<-a
truekappa1<-3</pre>
                                         # true changepoint (control)
xg1<-(t-kappa1)*(t>kappa1)*(tx==0)
                                        # 1(t>cp1)*1(trt=control)
truekappa2<-kappa2<-2</pre>
                                         # true changepoint (tx)
xg2<-(t-kappa2)*(t>kappa2)*(tx==1) # 1(t>cp2)*1(trt=treatment)
xg < -xg1 + xg2
                                         # random slope after corr. cps
Xs<-cbind(1,t,tx,t*tx)</pre>
                                         # Sub design matrix for "Binary"/"BB component"
X1<-cbind(Xs,xg1,xg2)</pre>
                                         # design matrix for *binary component*
truealpha <- alpha <- c(0.59, 0.86, 0.45, -0.35, -1.15, -0.54) # true alpha (binary)
```

```
X2<-cbind(Xs,xg1,xg2)</pre>
                                                   # design matrix for *BB component*
truebeta <- beta <- c(-0.94, 0.59, 0.89, 0.43, .68, -.66) # true beta (count)
p1 < -ncol(X1)
                                       # number of parameters (binary)
p2<-ncol(X2)
                                       # number of parameters (count)
truesigmab<-sigmab<-matrix(c(0.5,0.05,0.05,0.05,0.05,0.05,
                              0.05,0.35,0.05,0.05,0.05,0.05,
                              0.05,0.05,0.35,0.05,0.05,0.05,
                              0.05,0.05,0.05,0.5,0.15,0.05,
                              0.05,0.05,0.05,0.15,0.5,0.05,
                              0.05,0.05,0.05,0.05,0.05,0.35),6,6,byrow = T) # 6 by 6 random effect varia
trueB<-B<-rmvnorm(n,rep(0,6),sigmab)
                                                # true random effects
trueb11<-b11<-B[,1]
trueb12<-b12<-B[,2]
trueb13<-b13<-B[,3]
trueb21<-b21<-B[,4]
trueb22<-b22<-B[,5]
trueb23<-b23<-B[,6]
B11<-rep(b11,nis)
B12<-rep(b12,nis)
B13<-rep(b13,nis)
B21<-rep(b21,nis)
B22<-rep(b22,nis)
B23<-rep(b23,nis)
### Binary component ###
eta1<-X1%*%alpha+B11+B12*t+B13*xg
mu1 \leftarrow exp(eta1)/(1 + exp(eta1))
u<-rbinom(N,1,mu1)
                                    # Abs. latent variable
N1 < -sum(u)
                                    # true num of ppl at risk
### Count component ###
eta2<-X2%*%beta+B21+B22*t+B23*xg
mu2 < -exp(eta2)/(1+exp(eta2))
truerho<-rho<-0.25
                                   # Correlation parameter
y < -rep(0, N)
y[u==1]<-rbetabinom(N1,size=ntrial,prob=mu2[u==1],rho=rho) # outcome
cat("Proportion of obs at risk=",N1/N)
## Proportion of obs at risk= 0.6153846
cat("Proportion of structural zeros=",1-mean(u))
## Proportion of structural zeros= 0.3846154
cat("Proportion of zeros=",length(y[y==0])/N)
```

Proportion of zeros= 0.5254342

Histogram of the simulated data

```
tmp<-table(y)
barplot(tmp/N,col="darkgreen",xlab="Simulated abstinent days in past week",ylab="Percent")</pre>
```



MCMC Set-Up

```
#-----
# Bayesian ZIBB model #
# MCMC-MH Setting #
#----#
# Priors
alpha0<-rep(0,p1)
                        # prior mean for alpha
beta0<-rep(0,p2)
                        # prior mena for beta
T0a<-diag(0.01,p1)
                       # prior precision for alpha
T0b<-diag(0.01,p2)
                       # prior precision for beta
d0<-7
                         # prior inverse Wishart: IW(7,I_6)
CO<-diag(6)
                         # prior mean for kappa
kappa0<-0
sigmak0<-10^4
                        # prior var for kaapa
nk<-1
L0<--3
                        # CP from week -3 (2) to 6 (11)
U0<-6
# proposal
sigmar0<-0.01
                        # proposal var for rho
                       # proposal var for kappa1
sigmakap10<-0.1
sigmakap20<-0.1
                      # proposal var for kappa2
sigmab210<-0.1
                      # proposal var for b21
                       # proposal var for b22
sigmab220<-0.1
sigmab230<-0.1
                       # proposal var for b23
# Inists
alpha<-rep(0,p1)
beta < -rep(0,p2)
covb < -diag(0.01, p2)
                        # Proposal covariance
rho<-0.5
                        # over-dispersed
A1<-0
                        # Acceptance counter (beta)
A2<-0
                         # Acceptance counter (rho)
                        # Acceptance counter (kappa) -> control
Akap1<-0
                        # Acceptance counter (kappa) -> tx
Akap2<-0
                      # At risk indicator
y1<-rbinom(N,1,.5)
y1[y>0]<-1
                          # If y>0, then patient is at risk w.p. 1
n0 < -length(y[y==0])
                         # Number of observed 0's
bmat<-rmvnorm(n,rep(0,6),diag(6))</pre>
b11<-bmat[,1]
b12<-bmat[,2]
b13<-bmat[,3]
b21<-bmat[,4]
b22<-bmat[,5]
b23<-bmat[,6]
B11<-rep(b11,nis)
B12<-rep(b12,nis)
B13<-rep(b13,nis)
```

```
B21<-rep(b21,nis)
B22<-rep(b22,nis)
B23<-rep(b23,nis)
Bmat<-cbind(b11,b12,b13,b21,b22,b23)</pre>
sigmab<-cov(Bmat)</pre>
#----#
# Initial matrix #
#----#
kappa1<-0
kappa2<-0
                                      # Init changepoints
xg1<-(t-kappa1)*(t>kappa1)*(tx==0) # Initialize design matrix
xg2<-(t-kappa2)*(t>kappa2)*(tx==1) # Initialize design matrix
xg < -xg1 + xg2
X1 < -X2 < -cbind(Xs, xg1, xg2)
################
# Store Samples #
################
nsim<-30000
                                # Number of MCMC Iterations
thin < -5
                                       # Thinnisng interval
burn<-10000
                                # Burnisn
lastit<-(nsim-burn)/thin</pre>
                                  # Last stored value
Betatmp<-matrix(0,nsim,p2)</pre>
Alpha<-matrix(0,lastit,p1)</pre>
Beta<-matrix(0,lastit,p2)</pre>
Rho<-rep(0,lastit)</pre>
Kappa1<-rep(0,lastit)</pre>
Kappa2<-rep(0,lastit)</pre>
Sigmab<-matrix(0,lastit,36)</pre>
B11s<-matrix(0,lastit,n)</pre>
B12s<-matrix(0,lastit,n)
B13s<-matrix(0,lastit,n)
B21s<-matrix(0,lastit,n)
B22s<-matrix(0,lastit,n)
B23s<-matrix(0,lastit,n)
L<-matrix(0,lastit,N)</pre>
                               # Likelihood function
```

MCMC Algorithm (not run)

```
#-----#
# MCMC + MH #
#-----#
set.seed(1234)
time.start<-proc.time()
for (i in 1:nsim) {
    ### BINARY: update alpha ####
    mu<-X1%*%alpha+B11+B12*t+B13*xg
    omega<-rpg(N,1,mu)
    z<-(y1-1/2)/omega
    v<-solve(TOa+crossprod(X1*sqrt(omega)))</pre>
```

```
m<-v%*%(T0a%*%alpha0+t(sqrt(omega)*X1)%*%c(sqrt(omega)*(z-B11-B12*t-B13*xg)))
alpha<-c(rmvnorm(1,m,v))
### RANDOM: binary (b1i1) ###
priorprec < -c(1/(sigmab[1,1]-sigmab[1,-1]%*%solve(sigmab[-1,-1])%*%sigmab[-1,1]))
priormean<-Bmat[,-1]%*%t(sigmab[1,-1]%*%solve(sigmab[-1,-1]))
v<-1/(priorprec+c(tapply(omega,id,sum)))</pre>
m<-v*(priorprec*priormean+c(tapply(omega*(z-X1\%*\%alpha-B12*t-B13*xg),id,sum)))
b11<-rnorm(n,m,sqrt(v))
B11<-rep(b11,nis)
### RANDOM: binary (b1i2) ###
priorprec < -c(1/(sigmab[2,2]-sigmab[2,-2]%*%solve(sigmab[-2,-2])%*%sigmab[-2,2]))
 priormean < -Bmat[,-2] %*\%t(sigmab[2,-2]%*\%solve(sigmab[-2,-2])) 
v<-1/(priorprec+c(tapply(omega*t^2,id,sum)))</pre>
m<-v*(priorprec*priormean+c(tapply(t*omega*(z-X1%*%alpha-B11-B13*xg),id,sum)))</pre>
b12<-rnorm(n,m,sqrt(v))
B12<-rep(b12,nis)
### RANDOM: binary (b1i3) ###
priorprec <-c(1/(sigmab[3,3]-sigmab[3,-3]%*%solve(sigmab[-3,-3])%*%sigmab[-3,3]))
priormean<-Bmat[,-3]%*%t(sigmab[3,-3]%*%solve(sigmab[-3,-3]))
v<-1/(priorprec+c(tapply(omega*xg^2,id,sum)))</pre>
m<-v*(priorprec*priormean+c(tapply(xg*omega*(z-X1%*%alpha-B11-B12*t),id,sum)))</pre>
b13<-rnorm(n,m,sqrt(v))
B13<-rep(b13,nis)
### update at-risk ind ###
eta1<-X1%*%alpha+B11+B12*t+B13*xg
eta2<-X2\%*\%beta+B21+B22*t+B23*xg
pi<-pmax(0.001,pmin(0.999,1/(1+exp(-eta1)))) # at-risk probability
pr<-1/(1+exp(-eta2))
q<-dbetabinom(0,size=ntrial,prob=pr,rho=rho)</pre>
                          # Conditional prob that y1=1 given y=0 -- i.e. Pr(chance zero/observed zero
theta<-pi*q/(pi*q+1-pi)
y1[y=0] < -rbinom(n0,1,theta[y=0]) # If y=0, then draw a "chance zero" w.p. theta, if y=1, then y1=1
nis1<-tapply(y1,id,sum)
### COUNT: update beta ###
# Current likelihood
eta<-X2%*%beta+B21+B22*t+B23*xg
mu < -1/(1+exp(-eta))
lold<-sum(dbetabinom(y[y1==1],size=ntrial,prob=mu[y1==1],rho=rho,log=T))</pre>
# Draw candidate beta and compute likelihood
                                           # Draw from "symmetric" MV t_3 dist
betanew <- beta +rmvnorm(1, sigma=.15*covb)
eta<-X2\%*\%c(betanew)+B21+B22*t+B23*xg
mu < -1/(1 + exp(-eta))
lnew<-sum(dbetabinom(y[y1==1],size=ntrial,prob=mu[y1==1],rho=rho,log=T))</pre>
# Acceptance prob on log scale =log(lnew x prior) - log (lold x prior)
r1<-lnew+dmvnorm(betanew,beta0,solve(T0b),log=T)-(lold+dmvnorm(beta,beta0,solve(T0b),log=T))
if(log(runif(1))<r1) {</pre>
```

```
beta<-c(betanew)
 }
Betatmp[i,]<-beta</pre>
if (i==nsim/2) covb<-cov(Betatmp[(nsim/4+1):nsim/2,]) # Update proposal cov
### RANDOM: binary (b2i1) ###
priorprec < -c(1/(sigmab[4,4]-sigmab[4,-4]%*%solve(sigmab[-4,-4])%*%sigmab[-4,4]))
priormean<-Bmat[,-4]%*%t(sigmab[4,-4]%*%solve(sigmab[-4,-4]))
b21new<-rnorm(n,b21,sqrt(sigmab210))
eta<-X2%*%beta+rep(b21new,nis)+B22*t+B23*xg
mu < -1/(1 + exp(-eta))
                   # log-likelihood if empty block
lnew < -rep(1,n)
lnew[nis1>0] < -tapply(dbetabinom(y[y1==1],size=ntrial,prob=mu[y1==1],rho=rho,log=T),id[y1==1],sum)
eta<-X2%*%beta+B21+B22*t+B23*xg
mu < -1/(1+exp(-eta))
lold<-rep(1,n)</pre>
                # log-likelihood if empty block
lold[nis1>0] <-tapply(dbetabinom(y[y1==1],size=ntrial,prob=mu[y1==1],rho=rho,log=T),id[y1==1],sum)
ratio <- lnew + dnorm (b21 new, priormean, sqrt (1/priorprec), log=T) - (lold + dnorm (b21, priormean, sqrt (1/priorpre
utmp<-1*(log(runif(n))<ratio)
b21[utmp==1] <-b21new[utmp==1]
B21<-rep(b21,nis)
### RANDOM: binary (b2i2) ###
priorprec < -c(1/(sigmab[5,5]-sigmab[5,-5]%*%solve(sigmab[-5,-5])%*%sigmab[-5,5]))
priormean<-Bmat[,-5]%*%t(sigmab[5,-5]%*%solve(sigmab[-5,-5]))
b22new<-rnorm(n,b22,sqrt(sigmab220))
eta<-X2\%*\%beta+B21+rep(b22new,nis)*t+B23*xg
mu < -1/(1 + exp(-eta))
                   # log-likelihood if empty block
lnew<-rep(1,n)</pre>
lnew[nis1>0] < -tapply(dbetabinom(y[y1==1], size=ntrial, prob=mu[y1==1], rho=rho, log=T), id[y1==1], sum)
eta<-X2%*%beta+B21+B22*t+B23*xg
mu < -1/(1+exp(-eta))
lold<-rep(1,n)</pre>
                  # log-likelihood if empty block
lold[nis1>0] <-tapply(dbetabinom(y[y1==1],size=ntrial,prob=mu[y1==1],rho=rho,log=T),id[y1==1],sum)
ratio <- lnew + dnorm (b22new, priormean, sqrt (1/priorprec), log=T) - (lold + dnorm (b22, priormean, sqrt (1/priorpre
utmp<-1*(log(runif(n))<ratio)
b22[utmp==1] <-b22new[utmp==1]
```

```
B22<-rep(b22,nis)
### RANDOM: binary (b2i3) ###
priorprec < -c(1/(sigmab[6,6]-sigmab[6,-6]%*%solve(sigmab[-6,-6])%*%sigmab[-6,6]))
priormean<-Bmat[,-6]%*%t(sigmab[6,-6]%*%solve(sigmab[-6,-6]))
b23new<-rnorm(n,b23,sqrt(sigmab230))
eta<-X2%*%beta+B21+B22*t+rep(b23new,nis)*xg
mu < -1/(1+exp(-eta))
lnew < -rep(1,n)
                    # log-likelihood if empty block
lnew[nis1>0] < -tapply(dbetabinom(y[y1==1], size=ntrial, prob=mu[y1==1], rho=rho, log=T), id[y1==1], sum)
eta<-X2%*%beta+B21+B22*t+B23*xg
mu < -1/(1+exp(-eta))
lold<-rep(1,n)</pre>
                # log-likelihood if empty block
lold[nis1>0] <-tapply(dbetabinom(y[y1==1],size=ntrial,prob=mu[y1==1],rho=rho,log=T),id[y1==1],sum)
ratio <- lnew + dnorm (b23 new, priormean, sqrt (1/priorprec), log=T) - (lold + dnorm (b23, priormean, sqrt (1/priorpre
utmp<-1*(log(runif(n))<ratio)
b23[utmp==1] <-b23new[utmp==1]
B23<-rep(b23,nis)
### COUNT: update rho ###
# Current likelihood
eta<-X2%*%beta+B21+B22*t+B23*xg
mu < -1/(1+exp(-eta))
lold<-sum(dbetabinom(y[y1==1],size=ntrial,prob=mu[y1==1],rho=rho,log=T))</pre>
# Draw candidate rho and compute likelihood from truncated noraml
rhonew<-rtnorm(1,rho,sqrt(sigmar0),0,1)</pre>
                                                    # Draw from truncated normal
lnew<-sum(dbetabinom(y[y1==1],size=ntrial,prob=mu[y1==1],rho=rhonew,log=T))</pre>
# Acceptance prob on log scale =log(lnew x prior) - log (lold x prior)
rrho<-lnew-lold+dtnorm(rho,rhonew,sqrt(sigmar0),0,1,log=T)-dtnorm(rhonew,rho,sqrt(sigmar0),0,1,log=T)
if(log(runif(1))<rrho) {</pre>
 rho<-rhonew
 if (i> burn & i\hat{1}thin==0) A2<-A2+1</pre>
}
# Update sigmab
Bmat<-cbind(b11,b12,b13,b21,b22,b23)
sigmab<-riwish(d0+n,C0+crossprod(Bmat))</pre>
# update changepoint (kaapa) --> tx
kappa2new<-rtnorm(1,kappa2,sqrt(sigmakap20),L0,U0) # propose new cp
xg2new<-(t-kappa2new)*(t>kappa2new)*(tx==1)
xgnew<-xg1+xg2new
```

```
X1new<-cbind(Xs,xg1,xg2new)</pre>
                                                  # new design matrix (binary)
eta1<-X1new%*%alpha+B11+B12*t+B13*xgnew
mu1 < -pmax(0.001, pmin(0.999, 1/(1+exp(-eta1))))
X2new<-cbind(Xs,xg1,xg2new)</pre>
                                                  # new design matrix (BB)
eta2<-X2new%*%beta+B21+B22*t+B23*xgnew
mu2 < -pmax(0.001, pmin(0.999, 1/(1+exp(-eta2))))
lnew < -sum(dZIBB(y[tx==1], mu=mu2[tx==1], sigma=rho/(1-rho), nu=1-mu1[tx==1], bd=ntrial, log=T))
eta1<-X1%*%alpha+B11+B12*t+B13*xg
mu1 < -pmax(0.001, pmin(0.999, 1/(1+exp(-eta1))))
eta2<-X2%*%beta+B21+B22*t+B23*xg
                                                              # old design matrix (BB)
mu2 < -pmax(0.001, pmin(0.999, 1/(1+exp(-eta2))))
lold<-sum(dZIBB(y[tx==1],mu=mu2[tx==1],sigma=rho/(1-rho),nu=1-mu1[tx==1],bd=ntrial,log=T))
# (lnew-lold)+(priornew-priorold)+(proposalnew-proposalold)
rkap2<-(lnew-lold)+
  (dtnorm(kappa2,kappa2new,sqrt(sigmakap20),L0,U0,log=T)-dtnorm(kappa2new,kappa2,sqrt(sigmakap20),L0,
if(log(runif(1))<rkap2) {</pre>
 kappa2<-kappa2new
 xg2<-xg2new
                 # update slope after cp for tx
 xg<-xgnew # update slope after cp
               # update design matrix (binary)
 X1<-X1new
 X2<-X2new # update design matrix (bb)
 if (i> burn & i\hstyle=0) Akap2<-Akap2+1</pre>
}
# update changepoint (kaapa) --> control
kappa1new<-rtnorm(1,kappa1,sqrt(sigmakap10),L0,U0) # propose new cp</pre>
xg1new<-(t-kappa1new)*(t>kappa1new)*(tx==0)
xgnew<-xg1new+xg2
X1new<-cbind(Xs,xg1new,xg2)</pre>
                                                  # new design matrix (binary)
eta1<-X1new%*%alpha+B11+B12*t+B13*xgnew
mu1 < -pmax(0.001, pmin(0.999, 1/(1+exp(-eta1))))
X2new<-cbind(Xs,xg1new,xg2)</pre>
                                                  # new design matrix (BB)
eta2<-X2new%*%beta+B21+B22*t+B23*xgnew
mu2 < -pmax(0.001, pmin(0.999, 1/(1+exp(-eta2))))
lnew < -sum(dZIBB(y[tx==0], mu=mu2[tx==0], sigma=rho/(1-rho), nu=1-mu1[tx==0], bd=ntrial, log=T))
eta1<-X1%*%alpha+B11+B12*t+B13*xg
mu1 < -pmax(0.001, pmin(0.999, 1/(1+exp(-eta1))))
eta2<-X2%*%beta+B21+B22*t+B23*xg
                                                              # old design matrix (BB)
```

```
mu2 < -pmax(0.001, pmin(0.999, 1/(1+exp(-eta2))))
lold < sum(dZIBB(y[tx==0], mu=mu2[tx==0], sigma=rho/(1-rho), nu=1-mu1[tx==0], bd=ntrial, log=T))
\# (lnew-lold)+(priornew-priorold)+(proposalnew-proposalold)
rkap1<-(lnew-lold)+
  (dtnorm(kappa1, kappa1new, sqrt(sigmakap10), L0, U0, log=T)-dtnorm(kappa1new, kappa1, sqrt(sigmakap10), L0,
if(log(runif(1))<rkap1) {</pre>
 kappa1<-kappa1new
 xg1<-xg1new
                # update slope after cp (control)
 xg<-xgnew
                # update slope after cp
                  # update design matrix (binary)
 X1<-X1new
 X2<-X2new
                  # update design matrix (bb)
 ##################
# Store Results #
################
if (i> burn & i\n\tag{thin==0} {
 j<-(i-burn)/thin</pre>
 Alpha[j,]<-alpha
 Beta[j,]<-beta</pre>
 Rho[j]<-rho
 Kappa1[j]<-kappa1</pre>
 Kappa2[j]<-kappa2</pre>
 Sigmab[j,]<-c(sigmab)</pre>
 B11s[j,]<-b11
 B12s[j,]<-b12
 B13s[j,]<-b13
 B21s[j,]<-b21
 B22s[j,] \leftarrow b22
 B23s[j,]<-b23
  # likelihood function for ZIBB
 eta1<-X1%*%alpha+B11+B12*t+B13*xg
 mu1 \leftarrow pmax(0.001, pmin(0.999, 1/(1+exp(-eta1))))
 eta2<-X2<mark>%*%</mark>beta+B21+B22*t+B23*xg
 mu2 \leftarrow pmax(0.001, pmin(0.999, 1/(1+exp(-eta2))))
 L[j,]<-dZIBB(y,mu=mu2,sigma=rho/(1-rho),nu=1-mu1,bd=ntrial)
}
if (i\%5==0) {
 print(i)
 print(truealpha)
 print(alpha)
 print(truebeta)
 print(beta)
 print(truerho)
 print(rho)
```

```
print(truekappa1)
  print(kappa1)
  print(truekappa2)
  print(kappa2)
  print(truesigmab)
  print(sigmab)
}

{
time.tol<-proc.time()-time.start)</pre>
```

Save/Load MCMC Samples

```
# samples<-list(Alpha=Alpha,</pre>
#
                 Beta=Beta,
#
                 Rho=Rho,
#
                 Sigmab=Sigmab,
#
                 Kappa1=Kappa1,
#
                 Kappa2=Kappa2,
#
                 B11s=B11s,
#
                 B12s=B12s,
#
                 B13s=B13s.
#
                 B21s=B21s,
#
                 B22s=B22s,
#
                 B23s=B23s)
dir.sav<-"C:\\Users\\chech\\OneDrive - Medical University of South Carolina\\Research\\Bayes ZIBB\\Gith
\# save(samples, file=paste(dir.sav, "ZIBBCP-Simulation-N500.Rda", sep=""))
load(file=paste(dir.sav,"ZIBBCP-Simulation-N500.Rda",sep=""))
Alpha <- samples $ Alpha
Beta <- samples $Beta
Rho<-samples$Rho
Sigmab <- samples $Sigmab
Kappa1<-samples$Kappa1</pre>
Kappa2<-samples$Kappa2</pre>
B11s<-samples$B11s
B12s<-samples$B12s
B13s<-samples$B13s
B21s<-samples$B21s
B22s<-samples$B22s
B23s<-samples$B23s
```

Results

```
# Results
malpha<-colMeans(Alpha)
qalpha<-apply(Alpha,2,quantile,c(0.025,0.975))
mbeta<-colMeans(Beta)
qbeta<-apply(Beta,2,quantile,c(0.025,0.975))</pre>
```

```
mrho<-mean(Rho)
qrho \leftarrow quantile(Rho, c(0.025, 0.975))
msigmab<-colMeans(Sigmab)</pre>
qsigmab <- apply (Sigmab, 2, quantile, c(0.025, 0.975))
mkap1<-mean(Kappa1)</pre>
qkap1 \leftarrow quantile(Kappa1, c(0.025, 0.975))
mkap2<-mean(Kappa2)
qkap2<-quantile(Kappa2,c(0.025,0.975))</pre>
truealpha
## [1] 0.59 0.86 0.45 -0.35 -1.15 -0.54
print(malpha)
## [1] 0.3632604 0.9253200 0.6731569 -0.3319699 -1.1373650 -0.6780192
print(qalpha)
                       [,2]
                                                                [,6]
              [,1]
                                 [,3]
                                            [,4]
                                                      [,5]
## 2.5% -0.1221006 0.6650762 0.03597402 -0.69174539 -1.6069616 -1.0670925
## 97.5% 0.9754308 1.2056293 1.25938617 0.02731948 -0.6503024 -0.2719076
truebeta
## [1] -0.94 0.59 0.89 0.43 0.68 -0.66
print(mbeta)
print(qbeta)
##
              [,1]
                       [,2]
                               [,3]
                                        [,4]
                                                 [,5]
                                                             [,6]
## 2.5% -1.2448380 0.3807418 0.419972 0.2481761 0.1825027 -0.67161412
## 97.5% -0.4772675 0.7256284 1.195628 0.6838851 0.9877138 -0.05173769
truerho
## [1] 0.25
print(mrho)
## [1] 0.278412
print(qrho)
##
       2.5%
               97.5%
## 0.2312829 0.3223268
ind=upper.tri(truesigmab,diag = T)
print(truesigmab[ind])
## [16] 0.05 0.05 0.05 0.05 0.05 0.35
print(msigmab[c(ind)])
  [1] 1.191406237 0.015455297 0.573733561 0.013769515 -0.011645816
```

```
## [6] 0.451519022 -0.008580402 0.028673486 0.010977867 0.257743738
## [16] 0.052737247 0.029913022 0.009933817 0.032549082 -0.020688467
## [21] 0.307494560
print(qsigmab[,c(ind)])
##
            [,1]
                      [,2]
                                [,3]
                                          [,4]
                                                      [,5]
                                                               [,6]
                                                                          [,7]
## 2.5% 0.455959 -0.1667685 0.3373984 -0.1156159 -0.14550638 0.1249357 -0.2053484
## 97.5% 2.279762 0.1834361 0.8940911 0.1526152 0.09558679 1.2336610 0.1321427
                          [,9]
                                   [,10]
                                              [,11]
               [,8]
                                                         [,12]
## 2.5% -0.07734927 -0.09202636 0.08246784 -0.2076451 -0.05270105 -0.1785281
## 97.5% 0.16164710 0.12368251 0.54660632 0.2026450 0.16117973 0.1156774
              [,14]
                       [,15]
                                  [,16]
                                            [,17]
                                                      [,18]
## 2.5% -0.09147108 0.3055626 -0.1245842 -0.1215617 -0.1298095 -0.05404482
## 97.5% 0.11663806 0.6139320 0.2636779 0.2478683 0.1387012 0.20576788
##
             [,20]
## 2.5% -0.1871606 0.1020287
## 97.5% 0.1398025 0.8820946
truekappa1
## [1] 3
print(mkap1)
## [1] 3.075909
print(qkap1)
      2.5%
              97.5%
## 2.459150 3.715393
truekappa2
## [1] 2
print(mkap2)
## [1] 1.83943
print(qkap2)
              97.5%
##
      2.5%
## 1.123590 2.604459
print(truekappa1-truekappa2)
## [1] 1
mean (Kappa1-Kappa2)
## [1] 1.236479
quantile(Kappa1-Kappa2,c(.025,.975))
##
       2.5%
                97.5%
## 0.2470876 2.2039351
```

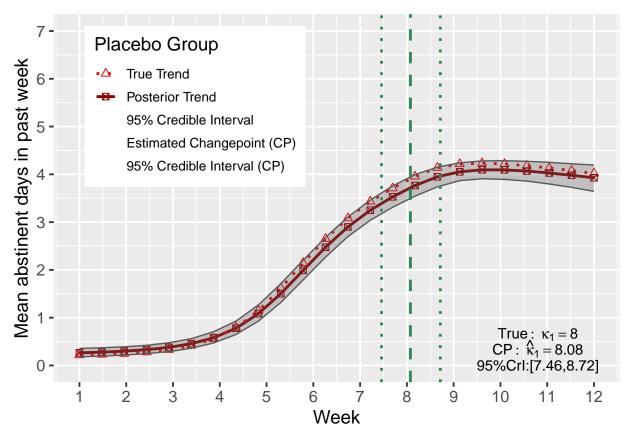
Figures

Anbsinence trends (placebo vs treatment)

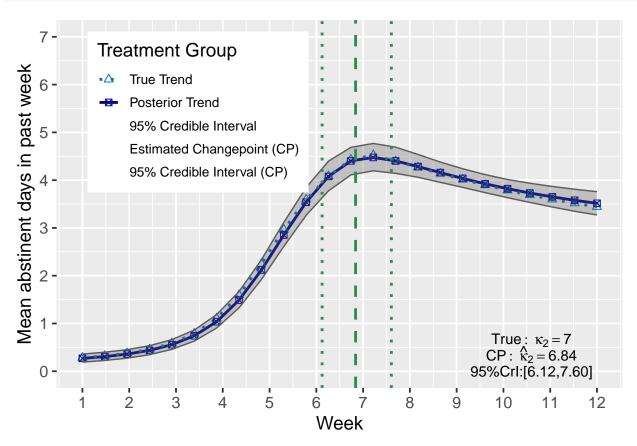
```
# Figures #
#----#
          # 24 time-point grids
num=24
grid<-seq(-4,7,length.out=num)</pre>
n1 < -sum(trt == 0)
                     # number of subjects (Placebo)
n2 < -sum(trt == 1)
                     # number of subjects (Treatment)
tp<-rep(grid,n)</pre>
nisp<-rep(num,n)</pre>
txp<-rep(trt,nisp)</pre>
# True trajectory (placebo and tx groups)
xgp1<-(tp-truekappa1)*(tp>truekappa1)*(txp==0)
xgp2<-(tp-truekappa2)*(tp>truekappa2)*(txp==1)
xgp<-xgp1+xgp2
Xp<-cbind(1,tp,txp,tp*txp,xgp1,xgp2)</pre>
eta1obs<-Xp%*%truealpha+rep(trueb11,nisp)+rep(trueb12,nisp)*tp+rep(trueb13,nisp)*xgp
eta2obs<-Xp%*%truebeta+rep(trueb21,nisp)+rep(trueb22,nisp)*tp+rep(trueb23,nisp)*xgp
piobs<-1/(1+exp(-eta1obs))</pre>
muobs < -1/(1 + exp(-eta2obs))
yobs<-piobs*muobs*ntrial</pre>
dat<-data.frame(yobs=yobs,tp=tp,txp=txp)</pre>
tmp<-dat %>%
  group_by(txp,tp)%>%
  summarise(yobs=mean(yobs))
YPOS1<-array(0,dim=c(n1,num,lastit))</pre>
YPOS2<-array(0,dim=c(n2,num,lastit))</pre>
YDIFF<-matrix(0,lastit,num)
for (j in 1:lastit){
  b11<-B11s[j,];B11<-rep(b11,nisp)
  b12<-B12s[j,];B12<-rep(b12,nisp)
  b13<-B13s[j,];B13<-rep(b13,nisp)
  b21<-B21s[j,];B21<-rep(b21,nisp)
  b22<-B22s[j,];B22<-rep(b22,nisp)
  b23<-B23s[j,];B23<-rep(b23,nisp)
  alpha<-Alpha[j,]</pre>
  beta<-Beta[j,]</pre>
  kappa1<-Kappa1[j]</pre>
  kappa2<-Kappa2[j]</pre>
  spgp1<-(tp-kappa1)*(tp>kappa1)*(txp==0)
  spgp2 < -(tp-kappa2)*(tp>kappa2)*(txp==1)
  spgp<-spgp1+spgp2
```

```
X<-cbind(1,tp,txp,tp*txp,spgp1,spgp2)</pre>
  eta1<-X%*%alpha+B11+B12*tp+B13*spgp
  pi < -1/(1+exp(-eta1))
  eta2<-X%*%beta+B21+B22*tp+B23*spgp
  mu < -1/(1 + exp(-eta2))
  mmu<-matrix(pi*mu*ntrial,n,num,byrow=TRUE)</pre>
  YPOS1[,,j]<-mmu[trt==0,]
  YPOS2[,,j]<-mmu[trt==1,]</pre>
  YDIFF[j,]<-colMeans(YPOS2[,,j])-colMeans(YPOS1[,,j])</pre>
  #print(j)
}
ypos1<-colMeans(t(colMeans(YPOS1[,,1:lastit])))</pre>
ypos2<-colMeans(t(colMeans(YPOS2[,,1:lastit])))</pre>
yposlower1<-apply(t(colMeans(YPOS1[,,1:lastit])),2,quantile,prob=0.025)</pre>
yposupper1<-apply(t(colMeans(YPOS1[,,1:lastit])),2,quantile,prob=0.975)</pre>
yposlower2<-apply(t(colMeans(YPOS2[,,1:lastit])),2,quantile,prob=0.025)</pre>
yposupper2<-apply(t(colMeans(YPOS2[,,1:lastit])),2,quantile,prob=0.975)</pre>
dplot<-data.frame(grid=rep(grid+5,2),</pre>
                   mmu1=c(tmp$yobs[1:num],ypos1),
                   lb1=c(rep(NA, num), yposlower1),
                   ub1=c(rep(NA, num), yposupper1),
                   gp=c(rep("True Trend", num), rep("Posterior Trend", num)),
                   mmu2=c(tmp$yobs[((num+1):(2*num))],ypos2),
                   1b2=c(rep(NA,num),yposlower2),
                   ub2=c(rep(NA,num),yposupper2))
ggplot(dplot,aes(x=grid,y=mmu1,col=gp,shape=gp))+
  geom_line(linetype=c(rep("solid",num),rep("dotted",num)),size=1)+
  geom point(size=2)+
  geom_ribbon(aes(ymin = lb1, ymax = ub1,col="95% Credible Interval",
                  fill="95% Credible Interval"),
              linetype=1,alpha=0.3,show.legend = F)+
  geom_vline(aes(xintercept=mkap1+5,col="Estimated Changepoint (CP)"),
             linetype="dashed", size=1, show.legend = F)+
  geom_vline(aes(xintercept=qkap1[1]+5,col="95% Credible Interval (CP)"),
             linetype="dotted",size=1,show.legend = F)+
  geom_vline(aes(xintercept=qkap1[2]+5,col="95% Credible Interval (CP)"),
             linetype="dotted",size=1,show.legend = F)+
```

```
scale_x_continuous(breaks = 1:12,limits=c(1,12))+
scale_y_continuous(breaks = 0:7,limits=c(0,7))+
scale_color_manual(breaks = c("True Trend", "Posterior Trend",
                              "95% Credible Interval",
                              "Estimated Changepoint (CP)",
                              "95% Credible Interval (CP)"),
                   values = c("red2","red4","grey36","seagreen","seagreen"))+
scale shape manual(breaks = c("True Trend", "Posterior Trend"),
                   values = c(2,7)+
scale fill manual(breaks = c("95% Credible Interval"),
                  values = c("grey36"))+
xlab("Week")+ylab("Mean abstinent days in past week")+
guides(color = guide_legend(title="Placebo Group",
                            override.aes = list(
                              linetype = c(3,1,1,5,3),
                              shape=c(2,7,NA,NA,NA)),
                            reverse = F), fill="none",
       shape="none",linetype="none")+
theme_gray(base_size = 14)+
theme(legend.key = element_rect(fill = "white"),
      legend.position = c(0.25,0.75),legend.text=element_text(size=10),
      axis.text=element_text(size=12),
     axis.title=element_text(size=14))+
annotate('text', x = 10.8, y = .65,
         label = "True:~kappa[1]==8.0",parse = TRUE,size=4)+
annotate('text', x = 10.8, y = 0.35,
         label = "CP:~hat(kappa)[1] == 8.08", parse = TRUE, size=4)+
annotate(geom="text", x=10.8, y=0, label="95%CrI:[7.46,8.72]",
         color="black",size=4)
```



```
ggplot(dplot,aes(x=grid,y=mmu2,col=gp,shape=gp))+
  geom_line(linetype=c(rep("solid",num),rep("dotted",num)),size=1)+
  geom point(size=2)+
  geom_ribbon(aes(ymin = 1b2, ymax = ub2,col="95% Credible Interval",
                  fill="95% Credible Interval"),
              linetype=1,alpha=0.3,show.legend = F)+
  geom vline(aes(xintercept=mkap2+5,col="Estimated Changepoint (CP)"),
             linetype="dashed",size=1,show.legend = F)+
  geom_vline(aes(xintercept=qkap2[1]+5,col="95% Credible Interval (CP)"),
             linetype="dotted",size=1,show.legend = F)+
  geom_vline(aes(xintercept=qkap2[2]+5,col="95% Credible Interval (CP)"),
             linetype="dotted",size=1,show.legend = F)+
  scale_x_continuous(breaks = 1:12,limits=c(1,12))+
  scale_y_continuous(breaks = 0:7,limits=c(0,7))+
  scale_color_manual(breaks = c("True Trend", "Posterior Trend",
                                "95% Credible Interval",
                                "Estimated Changepoint (CP)",
                                "95% Credible Interval (CP)"),
                     values = c("#0072B2","darkblue","grey36",
                                "seagreen", "seagreen"))+
  scale_shape_manual(breaks = c("True Trend", "Posterior Trend"),
                     values = c(2,7)+
  scale_fill_manual(breaks = c("95% Credible Interval"),
                    values = c("grey36"))+
  xlab("Week")+ylab("Mean abstinent days in past week")+
  guides(color = guide_legend(title="Treatment Group",
```



Traceplots

```
par(mfrow=c(3,2))
# binary part
plot(1:lastit,Alpha[,1],type="l",col="darkgreen",
     xlab="Iteration",ylab=expression(alpha[0]))
abline(h=malpha[1],col="blue4")
legend(cex=1.2, "topleft", legend=c("p = 0.10", "ESS = 119"))
plot(1:lastit,Alpha[,2],type="1",col="darkgreen",
     xlab="Iteration",ylab=expression(alpha[1]))
abline(h=malpha[2],col="blue4")
legend(cex=1.2, "topleft", legend=c("p = 0.67", "ESS = 143"))
plot(1:lastit,Alpha[,3],type="l",col="darkgreen",
     xlab="Iteration",ylab=expression(alpha[2]))
abline(h=malpha[3],col="blue4")
legend(cex=1.2,"topleft",legend=c("p = 0.72", "ESS = 163"))
plot(1:lastit,Alpha[,4],type="l",col="darkgreen",
     xlab="Iteration",ylab=expression(alpha[3]))
abline(h=malpha[4],col="blue4")
legend(cex=1.2, "topleft", legend=c("p = 0.08", "ESS = 276"))
plot(1:lastit,Alpha[,5],type="l",col="darkgreen",
     xlab="Iteration",ylab=expression(alpha[41]))
abline(h=malpha[5],col="blue4")
legend(cex=1.2,"topleft",legend=c("p = 0.16","ESS = 269"))
plot(1:lastit,Alpha[,6],type="1",col="darkgreen",
     xlab="Iteration",ylab=expression(alpha[42]))
abline(h=malpha[6],col="blue4")
legend(cex=1.2,"topleft",legend=c("p = 0.07","ESS = 268"))
```

