Package 'xMWAS'

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Type Package

Title xMWAS: R package for data-integration and differential network analysis
Version 0.54
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Description xMWAS allows integration of omics data using multivariate methods such as PLS. The package performs community detection and network analysis to allow visualization of positive or negative associations between different datasets generated using samples from the same individuals.
License GPL2.0
LazyLoad no
Depends mixOmics, WGCNA, snow, igraph, graph, RBGL, plyr, plsgenomics
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xMWAS_0.54-package xMWAS
Description
xMWAS allows integration of omics data using multivariate methods such as PLS. The package performs community detection and network analysis to allow visualization of positive or negative associations between different datasets generated using samples from same individuals. The run_xmwas() function performs the integrative, community detection, and network analysis. The

output files include .gml files that can be used with software like Cytoscape.

Details

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License: gpl2.0
LazyLoad: yes

Author(s)

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run_xmwas run_xmwas

Description

The function uses sPLS or PLS and network functions in mixOmics package to perform pairwise integrative and correlation analysis. The pairwise graphs are merged using igraph and community detection is performed using the Multilevel clustering algorithm. Association networks can be visualized in R or using Cytoscape.

Usage

```
run_xmwas(xome_fname = NA, yome_fname = NA, zome_fname = NA, wome_fname = NA,
  outloc = NA,
  class_fname = NA, Xome_data = NA, Yome_data = NA, Zome_data = NA, Wome_data = NA,
  classlabels = NA, xmwasmethod = "spls", plsmode = "canonical", max_xvar = 5000,
  max_yvar = 5000, max_zvar = 5000, max_wvar = 5000, rsd.filt.thresh = 1,
  all.missing.thresh = 0.1, missing.val = 0, corthresh = 0.4, keepX = 1000,
  keepY = 1000, keepZ = 1000, keepW = 1000, pairedanalysis = FALSE,
  optselect = TRUE, rawPthresh = 0.05, numcomps = 10,
  net_edge_colors = c("blue", "red"), net_node_colors = c("orange", "green", "blue", "gold"),
        Xname = "X", Yname = "Y", Zname = "Z", Wname = "W",
  net_node_shape = c("circle", "rectangle", "triangle", "star"), seednum = 100,
        label.cex = 0.3, vertex.size = 6, max_connections = NA,
        centrality_method = "eigenvector", use.X.reference = FALSE,
        removeRda = TRUE, compare.classes = TRUE, class.comparison.allvar = TRUE, ...)
```

Arguments

xome_fname	Full path with filename for dataset A. Default: NA; The software uses the value provided for Xome_data when this is set to NA.
yome_fname	Full path with filename for dataset B Default: NA; The software uses the value provided for Yome_data when this is set to NA.
zome_fname	Full path with filename for dataset C Default: NA; The software uses the value provided for Zome_data when this is set to NA.

wome_fname	Full path with filename for dataset D Default: NA; The software uses the value provided for Wome_data when this is set to NA.	
Xome_data	Data matrix for dataset A. Run: data(exnci60); head(exnci60\$mrna) to see how to format data matrices.	
Yome_data	Data matrix for dataset B	
Zome_data	Data matrix for dataset C	
Wome_data	Data matrix for dataset D	
outloc	Output directory	
classlabels	Data matrix with phenotype information. Set to NA if this information is not available. see: data(classlabels_casecontrol) for case vs control design or data(classlabels_repeatmeas for repeat measures	
class_fname	File with phenotype information. Set to NA if this information is not available. see: data(classlabels_casecontrol) for case vs control design or data(classlabels_repeatmeasures) for repeat measures	
xmwasmethod	Method for data integration. eg: "pls": partial least squares regression "spls": sparse partial least squares regression "o1pls": orthogonal partial least squares regression	
plsmode	"canonical" for bi-directional relationships; "regression" for regression/predictive relationships	
max_xvar	Maximum number of X variables to select based on relative standard deviation (RSD). e.g. 10000	
max_yvar	Maximum number of Y variables to select based on relative standard deviation (RSD). e.g. 10000	
max_zvar	Maximum number of Z variables to select based on relative standard deviation (RSD). e.g. 10000	
max_wvar	Maximum number of W variables to select based on relative standard deviation (RSD). e.g. 10000	
rsd.filt.thresh		
Relative standard deviation (sd/mean) threshold		
all.missing.thr	Remove variables (rows) that do not meet the minimum threshold for presence of non-missing values. e.g. 0.8	
missing.val	How are the missing values represented in the input data files? Default: 0	
corthresh	Correlation threshold. eg: 0.7	
keepX	Maximum number of X variables to select in sPLS. Note: keepX, keepY, keepZ, and keepW are only used when xmwasmethod is set to "spls"	
keepY	Maximum number of Y variables to select in sPLS. Note: keepX, keepY, keepZ, and keepW are only used when xmwasmethod is set to "spls"	
keepZ	Maximum number of Z variables to select in sPLS. Note: keepX, keepY, keepZ, and keepW are only used when xmwasmethod is set to "spls"	
keepW	Maximum number of W variables to select in sPLS. Note: keepX, keepY, keepZ, and keepW are only used when xmwasmethod is set to "spls"	
pairedanalysis	Are their repeated measurements? TRUE or FALSE	
optselect	Find optimal number of PLS components. TRUE or FALSE	
rawPthresh	p-value threshold calculated using Student's t-test. eg: 0.05	

Number of components to use in PLS model. eg: 3 numcomps

net_edge_colors

Colors for edges.

net_node_colors

Colors for nodes.

Name for X dataset. eg: "Genes" Xname Name for Y dataset. eg: "Proteins" Yname Name for Z dataset. eg: "Metabolites" Zname

Wname Name for W dataset. eg: "EnvironmentalExposures"

net_node_shape Shapes for nodes.

seednum Seed for random number generator used for plotting the network.

Size of the labels. eg: 0.8 label.cex

Size of the nodes. vertex.size

max_connections

Maximum number of associations to include in the network. The connections between nodes are ranked based on the strength of association (+ve and -ve). Only the top "max_connections" connections are shown and used for centrality and community detection analyses. Set max_connections=NA if you want to use all connections. e.g. 1e5, 1e6, or NA

centrality_method

Method for centrality analysis. Options: 1) "eigenvector" for eigenvector centrality, which is based on the number and quality of connections - centrality scores range from 0 to 1, where 1 means high centrality and 0 means low or no centrality. Nodes/vertices with high centrality scores are connected to many other nodes, which are in turn connected to many other nodes. Please see igraph::eigen_centrality function for more details.

- 2) "betweenness" for betweenness centrality, which is based on the number of shortest paths going through a node/vertex - centrality scores are normalized and scaled to 0 to 1 range, where 1 means high betweenness centrality and 0 means low or no betweenness centrality. Please see igraph::betweenness function for more details.
- 3) "degree.count" based on the number of connections of a node. -centrality scores are normalized and scaled to 0 to 1 range. High centrality means more connections. Please see igraph::degree function for more details.
- 4) "degree.weight" is based on the sum of absolute weights of connections of a node. -centrality scores are normalized and scaled to 0 to 1 range. High centrality means stronger connections.
- 5) "closeness" based on the reciprocal of the sum of the distances of a node/vertex to all other nodes. -centrality scores are normalized. High centrality means the node is closer to all other nodes.

use.X.reference

TRUE or FALSE if you want to use Xome_data as reference. If TRUE, only X<->Y, X<->Z, and X<->W pairwise analysis will be performed.

removeRda compare.classes

TRUE or FALSE; set to TRUE if you want to remove the intermediate files.

TRUE or FALSE; set to TRUE if you want to compare individual classes as provided in class labels file.

class.comparison.allvar

TRUE or FALSE; set to TRUE if all nodes shown

Author(s)

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