CEE6580

Spring 2018

HW #2

Due 3/6/2018

Can be done in groups up to 3 people or individually.

**Problem 1. (20 pts)**

One of the promising applications of Microbial Fuel Cells (MFCs) is in the treatment of Wastewater.

a) Calculate how much electricity (kW) could theoretically be generated by the Ithaca Area WWTP if they were to install MFCs for electricity generation to destroy influent BOD. Assume all the electrons in the WW organic matter could be harnessed (transferal of the eeqs from the WW organics through an MFC circuit to O2 to form H2O) and that a 1 Volt voltage drop per electron could be attained. SHOW all Work.

NOTE:The Ithaca WWTP treats an average of 6.5 million gallons per day of wastewater. This WW corresponds to 7200 pounds (3273 kg) per day of BOD (biochemical oxygen demand).

b) Why would this theoretical level never be reached? What are the potential drawbacks of applying MFCs to wastewater treatment?

**Problem 2 (20 points).**

For the salicylate monooxygenase (EC:1.14.13.1) of P. putida find its entry on the BRENDA enzyme database.

(a) write out the full stoichiometry of the reaction it catalyzes.

(b) using parameters on the EC entry page write out a generalized kinetic rate expression for the disappearance of salicylic acid over time in a batch reactor in the presence of benzoate.

(c) when benzoate and salicylate are both present at 0.1μM, and NADH and O2 are both much greater than their respective Ks values, what is the specific salicylate utilization rate (mass/(L\*time)? Assume a level of 50 mg (0.05 g) of enzyme per L.

**Problem 3 (20 pts)**

Bioenergetics of growth on naphthalene.

Naphthalene is the lightest of the polynuclear aromatic hydrocarbons (PAHs).

Using web and print resources,

a) Calculate how much free energy is theoretically available from the complete aerobic “mineralization” (i.e. oxidation completely to CO2) of one eeq of naphthalene with oxygen as the EA. (Hint: b/c naphthalene doesn’t appear in the Bioenergetics half rxn table, youll need to write out the balance redox reaction with O2 as the EA, then use free energies of formation of all products & reactants (available through the Knovel link at the website) to calculate ΔG0’r )

b) Determine the fraction of that energy that actually gets captured in the form of ATP based upon known pathways for naphthalene catabolism (assume the pathway through catechol is used). Consider ATP produced by both substrate level phosphorylation and by oxidative phosphorylation (OP) via the electron transport chain. For each NADH, 3 ATP are produced via OP and for each FADH, 2 ATP are produced. (At biologically relevant concentrations of ATP, ADP and Phosphate, ΔG for the formation of ATP from ADP & Pi is +12.5 kcal/mole ATP – 52.5 kJ/mole ATP).

c) How does this compare to the E=0.6 value used by McCarty (in the Gossett bioenergetics packet)

**Problem 4 (20 pts)**

Exploring Reactor performance for TCE degradation in a CSTR. Let’s assume the following parameters apply to *Dehalococcoides* growing anaerobically on TCE.

K = 0.1 umole/L

qmax = 5.28x 10-9 umoles/cell/day

Y= 1.6 x 108 cells X/umole TCE

b= 0.05/d

V= 1 L

So = 200 umoles/L

Assume Monod kinetics apply, there is no other limiting substrate besides TCE, and that there is no biomass in the influent.

1. First let’s consider steady state operation predictions.
   1. What is the minimum θc (residence time) (and maximum Q in L/day) to avoid cell washout at this So?
   2. If you operate at twice this θc, what will be the steady state effluent concentrations of S and X?
   3. If you operate at 10 times this θc, what will be the effluent concentrations of S and X?
2. Using Stella or Excel to model X and S in this system during startup. Above we solved for the steady state concentrations of S and X under different θc. Now we will explore the dynamics of the effluent S & X during “startup” of a reactor.
   1. Assume an initial inoculum level in the bioreactor of X,initial = 1x106 cells /L and S,init = 200 umoles/L. Then run the model with Q =0.25 L/day with 200 umoles/L influent (as in part “a” above). Plot X and S over time. What are the steady state levels ultimately? How long did it take for the system to come to 95% of the steady state values for X & S?
   2. Chose one kinetic parameter (qmax, Y, b, K) to change the value of. Vary from the set values down to 20% (1/5th the value) and up to 500% (5x the value). What is the effect on steady state X & S? Present plots.
   3. Hand in your model electronically via Blackboard.

**Problem 5: (20 pts)**

a) Create an excel spreadsheet and plot ΔG versus electron acceptor concentration for the following respirations linked to glucose catabolism. Assume 25°C and the following concentrations of other chemical species: HCO3- = 70 mM; CO2 = 0.02 atm; glucose = 10 mM; Fe+2 = 0.1 mM; H+ = 1E-7 M (pH=7); N2 = 0.8 atm; H2S = HS- = 0.0001 mM.

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| Electron Acceptor | Range of concentration to consider |
| O2 | 0.000002 atm – 0.2 atm |
| SO4-2 | 0.000001 M – 0.1 M |
| Fe+3 | 0.000001 M – 0.5 M |
| NO3- | 0.000001 M – 0.1 M |

b) If a growth media contains 10 mM glucose as the electron donor and the following mixture of Electron Acceptors, predict which type of organism has the thermodynamic advantage – aerobes, nitrate reducers, sulfate-reducers, or iron reducers.

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| O2 = 0.00005 atm |
| SO4-2 = 100 mM |
| Fe+3 = 1 mM |
| NO3- = 1 mM |