Basic ggplots

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R tutorial 1: Using ggplot to create basic plots

Here, we will use the popular **iris** dataset, collected by Edgar Anderson in 1936, to illustrate how ggplot can be used to create beautiful graphics. It contains sepal and petal measurements for 50 individuals from three *Iris* species; namely *I. setosa*, *I. versicolor*, and *I. virginica*.

Check the R dataset package for other interesting data sets to tinker with.

Some notes on syntax:

- package_name:: specifies the package name, where the two colons access all the functions associated with that package
- Hashes (#) denote comments, that are not read by R. Use these to annotate your code
- In ggplot, the plus sign (+) means that you are adding additional "layers" to your plot
- An equal sign (=) is synonymous with the arrow (->). You can use either to assign names to objects
- Dollar signs (\$) are like keys that access features of an object, such as columns or lists
- Use a question mark (?) to access the help file for a particular package (e.g. **?ggplot**)

Let's get started!

[1] "setosa"

if (!require("pacman"))

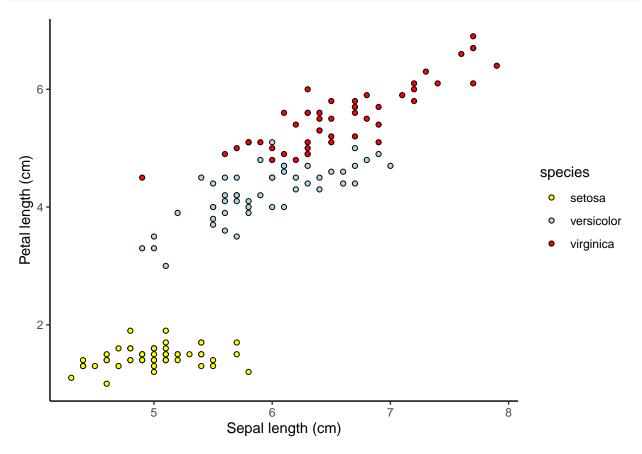
```
install.packages("pacman")
## Loading required package: pacman
pacman::p_load(datasets, xlsx, janitor, ggplot2, Rmisc, dplyr, ggpubr)
# access the embedded dataset
iris data = datasets::iris
iris_data = janitor::clean_names(iris_data)
# for practice, write this data to the project folder as a .csv file
write.csv(iris_data, file = "data/iris_data.csv", quote = FALSE, row.names = FALSE)
# now read it back in
iris_data_readin = read.csv("data/iris_data.csv")
# now as an Excel file
xlsx::write.xlsx(iris_data, file = "data/iris_data.xlsx",
                 sheetName = "iris_data", row.names = FALSE)
iris_data_readin_xlsx = xlsx::read.xlsx(file = "data/iris_data.xlsx", sheetIndex = 1)
# make iris species a factor/grouping variable
iris_data$species = as.factor(iris_data$species)
levels(iris data$species)
```

"versicolor" "virginica"

ggplot

Using geom_point()

geom_point() generates dot plots, like this:



Here, we create a subset for each *Iris* species, using the **filter()** function in the **dplyr** package:

```
setosa = dplyr::filter(iris_data, species=="setosa")
versicolor = dplyr::filter(iris_data, species=="versicolor")
virginica = dplyr::filter(iris_data, species=="virginica")
```

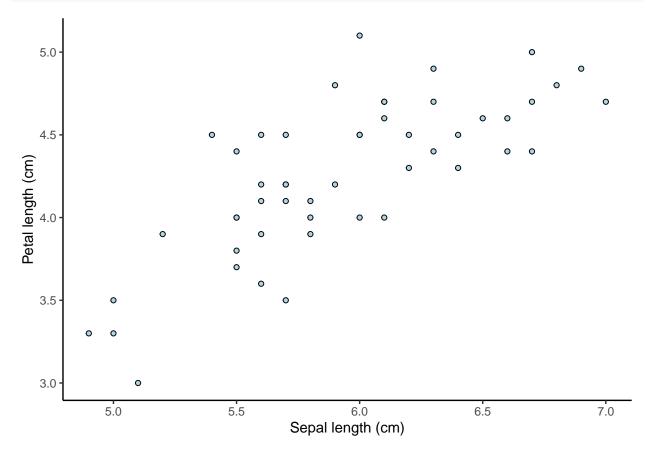
Now we can run a quick correlation test to see if there is a significant correlation between sepal and petal length in $I.\ versicolor$:

```
cor.test(versicolor$sepal_length, versicolor$petal_length, method = "pearson")
```

```
##
## Pearson's product-moment correlation
##
## data: versicolor$sepal_length and versicolor$petal_length
## t = 7.9538, df = 48, p-value = 2.586e-10
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
## 0.6020680 0.8532995
## sample estimates:
## cor
## 0.754049
```

Here we see a significant correlation (p < 0.05), with a correlation coefficient of r = 0.75. Typically, a value greater than 0.7 is considered to be strong.

Let's plot only *I. versicolor*, using **dplyr::filter()**:



Let's run a quick linear model, where we check whether there is a relationship between petal and sepal length:

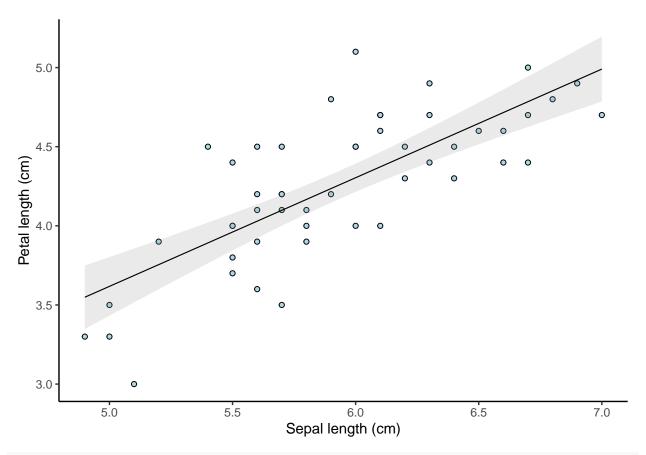
```
versicolor_lm = lm(data = versicolor, petal_length~sepal_length)
summary(versicolor_lm)
##
## Call:
## lm(formula = petal_length ~ sepal_length, data = versicolor)
##
## Residuals:
##
        Min
                      Median
                  1Q
                                     3Q
                                             Max
## -0.68611 -0.22827 -0.04123 0.19458 0.79607
##
## Coefficients:
                Estimate Std. Error t value Pr(>|t|)
##
## (Intercept)
                 0.18512
                             0.51421
                                       0.360
                                                 0.72
                                       7.954 2.59e-10 ***
## sepal_length 0.68647
                             0.08631
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## Residual standard error: 0.3118 on 48 degrees of freedom
## Multiple R-squared: 0.5686, Adjusted R-squared: 0.5596
## F-statistic: 63.26 on 1 and 48 DF, p-value: 2.586e-10
If we plot the trend line for this linear regression, the intercept = 0.19, and the gradient = 0.69. The R^2
value is 0.56 (p < 0.05). Let's add this line to our dot plot, with a confidence interval band:
ggplot2::ggplot(data = versicolor,
                aes(x=sepal_length, y=petal_length)) +
  geom_point(shape = 21, fill = "lightblue") +
```

alpha = 0.2, color = "black", linewidth = 0.4, se = TRUE)

xlab("Sepal length (cm)") +
ylab("Petal length (cm)") +

stat_smooth(method = "lm", formula = y~x, geom = "smooth",

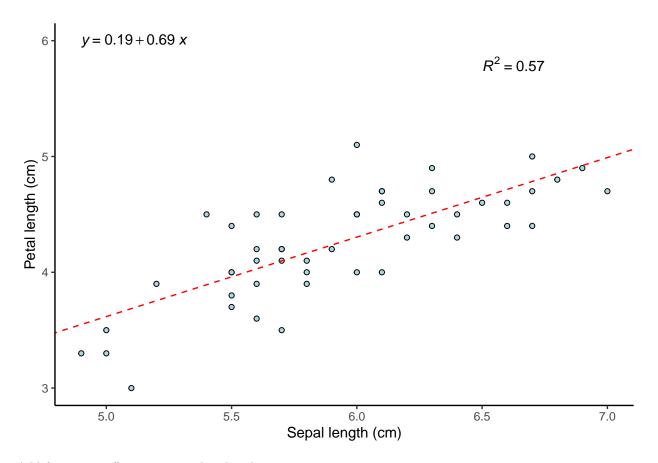
theme_classic() +



set se = FALSE to remove the confidence interval band

We can also add this line using the **geom_abline()** function:

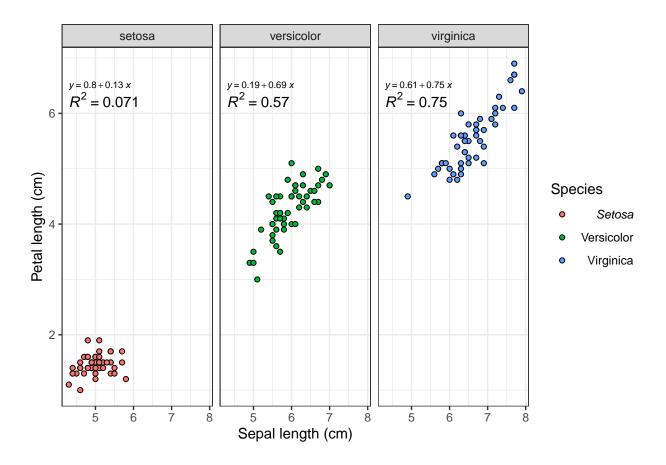
```
coeff = coefficients(versicolor_lm)
intercept = coeff[1]
gradient = coeff[2]
ggplot2::ggplot(data = versicolor,
                aes(x=sepal_length, y=petal_length)) +
  geom_point(shape = 21, fill = "lightblue") +
  # add the abline
  geom_abline(intercept = intercept, slope = gradient, linewidth = 0.5,
              color = "red", linetype = "dashed") +
  xlab("Sepal length (cm)") +
  ylab("Petal length (cm)") +
  theme_classic() +
  # the stat_regline_equation() function in the ggpubr package adds the equation as text to the plot
  ggpubr::stat_regline_equation(label.y = 6, aes(label = after_stat(c(eq.label)))) +
  # the stat_regline_equation() function in the ggpubr package adds the R-squared value as text to the
  ggpubr::stat_regline_equation(label.y = 5.8, label.x = 6.5,
                                aes(label = after_stat(c(rr.label))) )
```



Add facet_wrap() to separate the plots by species:

Scale for fill is already present.

Adding another scale for fill, which will replace the existing scale.

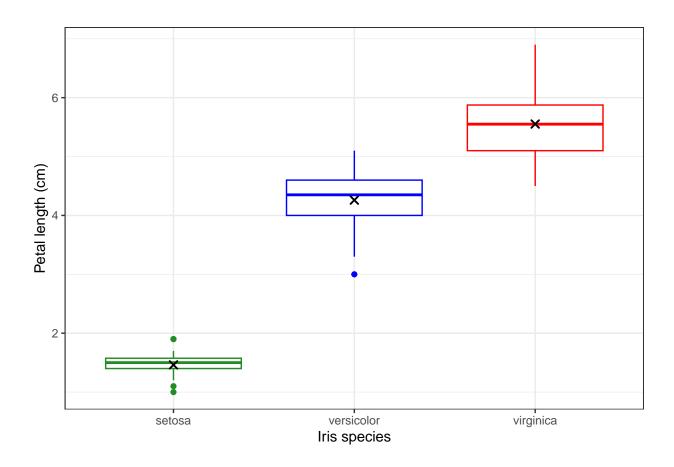


Using geom_boxplot()

The **geom_boxplot()** function creates box-and-whisker summary plots for your data:

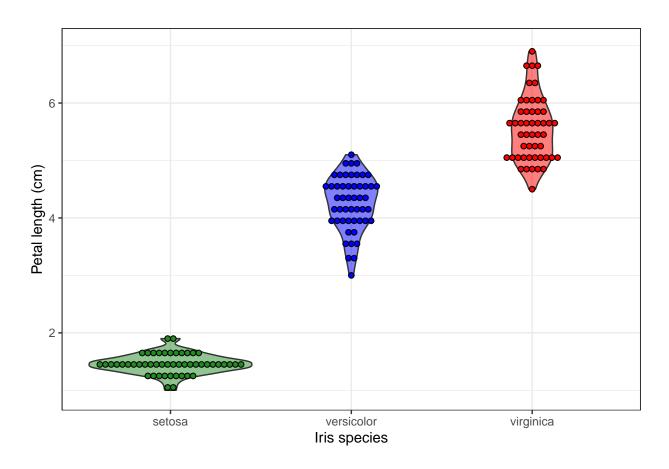
```
ggplot2::ggplot(data = iris_data, aes(x=species, y=petal_length, color=species)) +
  geom_boxplot() +
  scale_color_manual(values = c("forestgreen", "blue", "red")) +
  xlab("Iris species") +
  ylab("Petal length (cm)") +
  theme_bw() +
  theme(legend.position="none") +
  # add points to denote means
  stat_summary(fun = "mean", colour = "black", shape = 4)
```

Warning: Removed 3 rows containing missing values (`geom_segment()`).



Using geom_violin()

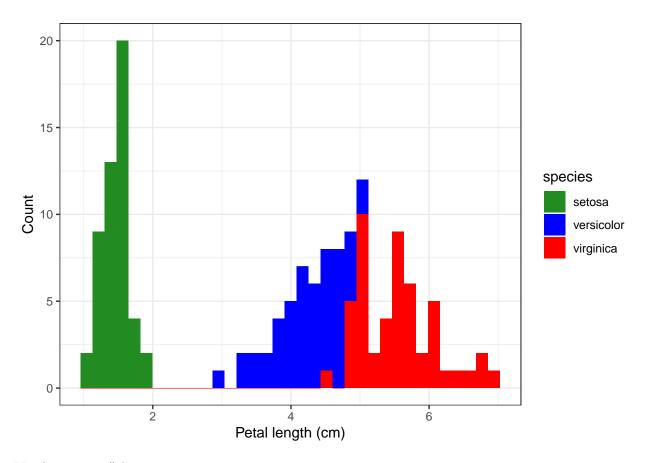
Bin width defaults to 1/30 of the range of the data. Pick better value with ## `binwidth`.



Using geom_histogram()

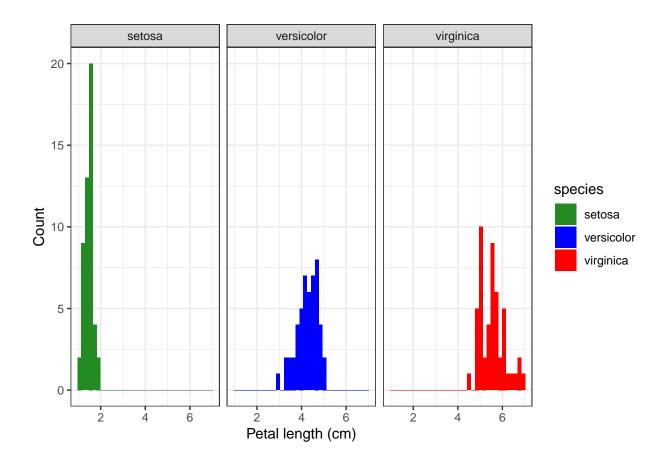
Use $\mathbf{geom_histogram}$ to generate histogram plots. Set an appropriate bin value:

```
ggplot2::ggplot(data = iris_data, aes(x=petal_length, fill=species)) +
  geom_histogram(bins = 35) +
  scale_fill_manual(values = c("forestgreen", "blue", "red")) +
  xlab("Petal_length (cm)") +
  ylab("Count") +
  theme_bw()
```



Use facet_wrap() here:

```
ggplot2::ggplot(data = iris_data, aes(x=petal_length, fill=species)) +
  geom_histogram(bins = 35) +
  scale_fill_manual(values = c("forestgreen", "blue", "red")) +
  xlab("Petal length (cm)") +
  ylab("Count") +
  theme_bw() +
  facet_wrap(~species)
```



Saving your plots as .PNG, .JPG, .PDF, or .SVG $\,$

To save a plot, first assign it a name. We will call the last plot we made **petal_length_hist**:

Now we can use the ggsave() function to save it in any format we like. Play around with the dimensions to get an ideal size: