### SpidermiR:Application Examples

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2016-10-21

#### Introduction

n this vignette, we demonstrate some applications of SpidermiR as tool for the study of miRNA network. For basic use of the SpidermiR package, please refer to the vignette Working with SpidermiR package.

### SpidermiR Downstream Analysis: Case Studies

## Case Study n.1: Role of miRNAs in shared protein domains network in Prostate Cancer

In this case study, we downloaded shared protein domains network in Homo Sapiens, using SpidermiRquery, SpidermiRprepare, and SpidermiRdownload with the function Case\_Study1\_loading\_1\_network. This function downloads the shared proteind network in HomoSapiens as provided by GeneMania. Then preprocessing the network give us the measures of the network.

```
Case_Study1_loading_1_network<-function(species){</pre>
org<-SpidermiRquery species(species)</pre>
net_shar_prot<-SpidermiRquery_spec_networks(organismID = org[6,],</pre>
                                                 network = "SHpd")
out_net<-SpidermiRdownload_net(net_shar_prot)</pre>
geneSymb_net<-SpidermiRprepare_NET(organismID = org[6,],data = out_net)</pre>
ds<-do.call("rbind", geneSymb net)</pre>
data2<-as.data.frame(ds[!duplicated(ds), ])</pre>
m<-c(data2$gene symbolA)</pre>
m2<-c(data2$gene_symbolB)
s < -c(m, m2)
fr<- unique(s)</pre>
network = "SHpd"
print(paste("Downloading of 1 ",network, " network ",
             "in ",org[6,]," with number of nodes: ",
             length(fr)," and number of edges: ",nrow(data2),
             sep = ""))
return(geneSymb net)
```

Then, we focused on role of miRNAs in this network. We integrated miRNA information using SpidermiRanalyze in the fucntion Case\_Study1\_loading\_2\_network.

In order to understand the underlying biological process of a set of biomarkers of interest (e.g. from differentially expressed genes, DEGs) we performed an analysis to identify the DEGs connected between them in the shared protein domains network.

```
Case Study1 loading 3 network<-function(data,dataFilt,dataClin){</pre>
highstage <- dataClin[grep("7|8|9|10", dataClin$gleason_score), ]</pre>
highstage <- highstage [,c("bcr patient barcode", "gleason score")]
highstage<-t(highstage)
samples_hight<-highstage[1,2:ncol(highstage)]</pre>
dataSmTP <- TCGAquery_SampleTypes(barcode = colnames(dataFilt),</pre>
                                    typesample = "TP")
dataSmNT <- TCGAquery_SampleTypes(barcode = colnames(dataFilt),</pre>
                                    typesample ="NT")
colnames(dataFilt) <-substr(colnames(dataFilt),1,12)</pre>
se<-substr(dataSmTP, 1, 12)</pre>
common<-intersect(colnames(dataFilt),samples_hight)</pre>
dataSmNT<-substr(dataSmNT, 1, 12)</pre>
sub_net2<-SpidermiRanalyze_DEnetworkTCGA(data,
                                            TCGAmatrix=dataFilt,
                                            tumour=common,normal=dataSmNT)
ft<-sub_net2$V1
ft1<-sub net2$V2
fgt<-c(ft,ft1)
miRNA NET <- SpidermiRanalyze mirna network (sub net2,
                                             disease="prostate cancer",miR trg="val")
TERZA NET<-rbind(miRNA NET, sub net2)</pre>
print(paste("In the 3 network we found",length(unique(miRNA_NET$V1)),
             " miRNAs and ".
            length(unique(fgt)), " genes with ", nrow(TERZA_NET),
             " edges " ))
return(TERZA_NET)
```

The function Case\_Study1\_loading\_4\_network is able to reveal the communities based on density metrics. We focused on the community with the higher number of elements.

}

# Case Study n.2: miRNAs regulating degree centrality genes in physical interactions network in breast cancer

In this case study, we downloaded physical interactions network in Homo Sapiens, using SpidermiRquery, SpidermiRprepare, and SpidermiRdownload with the function Case\_Study2\_loading\_1\_network. This function downloads the physical interactions network in HomoSapiens as provided by GeneMania. Then preprocessing the network give us the measures of the network.

```
Case Study2 loading 1 network<-function(species){</pre>
org<-SpidermiRquery_species(species)</pre>
net_PHint<-SpidermiRquery_spec_networks(organismID = org[6,],</pre>
                                            network = "PHint")
out_net<-SpidermiRdownload_net(net_PHint)</pre>
geneSymb_net<-SpidermiRprepare_NET(organismID = org[6,],data = out_net)</pre>
ds<-do.call("rbind", geneSymb_net)</pre>
data1<-as.data.frame(ds[!duplicated(ds), ])</pre>
sdas<-cbind(data1$gene_symbolA,data1$gene_symbolB)</pre>
sdas<-as.data.frame(sdas[!duplicated(sdas), ])</pre>
m<-c(data1$gene_symbolA)</pre>
m2<-c(data1$gene_symbolB)
s < -c(m, m2)
fr<- unique(s)</pre>
network="PHint"
print(paste("Downloading of 1 ",network,
             " network ","in ",org[6,],
             " with number of nodes: ",length(fr),
             " and number of edges: ",nrow(sdas), sep = ""))
return(geneSymb_net)
}
```

A network of miRNA-protein PI was found using Case\_Study2\_loading\_2\_network.

Statistical results showed that proteins with higher centrality are effectively targets of miRNAs with higher centrality.

```
Case_Study2_loading_3_network<-function(sdas,miRNA_NET){
ds<-do.call("rbind", sdas)
  data1<-as.data.frame(ds[!duplicated(ds), ])
  sdas<-cbind(data1$gene_symbolA,data1$gene_symbolB)</pre>
```

```
sdas<-as.data.frame(sdas[!duplicated(sdas), ])</pre>
topwhol<-SpidermiRanalyze degree centrality(sdas)
topwhol_mirna<-SpidermiRanalyze_degree_centrality(miRNA_NET)</pre>
miRNA_degree<-topwhol_mirna[grep("hsa",topwhol_mirna$dfer),]
seq_gd<-as.data.frame(seq(1, 15400, by = 50))
even<-seq_gd[c(F,T),]</pre>
even2<-even
odd<-seq_gd[c(T,F),]
odd2 < -odd[-1]
odd2[154]<-15400
f<-cbind(even2,odd2-1)
SQ<-cbind(odd,even-1)
h<-as.data.frame(rbind(f,SQ))
SQ <- as.data.frame(h[order(h$even2,decreasing=FALSE),])
table_pathway_enriched <- matrix(1, nrow(SQ),4)
colnames(table_pathway_enriched) <- c("interval min",</pre>
                                         "interval max", "gene", "miRNA")
table_pathway_enriched <- as.data.frame(table_pathway_enriched)</pre>
j=1
for (j in 1:nrow(SQ)){
  a<-SQ$even2[j]
  b<-SQ$V2[i]
  d < -c(a,b)
gene_degree10<-topwhol[a:b,]</pre>
vfg<-rbind(miRNA_degree[1:10,],gene_degree10)</pre>
subnet<-SpidermiRanalyze_direct_subnetwork(data=miRNA_NET,BI=vfg$dfer)</pre>
table_pathway_enriched[j,"interval min"] <- d[1]</pre>
table_pathway_enriched[j,"interval max"] <- d[2]</pre>
s<-unique(subnet$V1)</pre>
x<-unique(subnet$V2)
table_pathway_enriched[j,"miRNA"]<-length(s)</pre>
table_pathway_enriched[j, "gene"] <-length(x)
df<-cbind(table_pathway_enriched$gene,table_pathway_enriched$miRNA)
rownames(df)<-table_pathway_enriched$`interval max`</pre>
categories <- c("protein", "miRNA")</pre>
colors <- c("green", "magenta")</pre>
op \leftarrow par(mar = c(5, 5, 4, 2) + 0.1)
matplot(df, type="l",col=colors,xlab = "N of Clusters",
        main = "",ylab = "Interactions",cex.axis=2,cex.lab=2,cex.main=2)
legend("topright", col=colors, categories, bg="white", lwd=1,cex=2)
a<-SQ$even2[j]
b<-SQ$V2[j]
d < -c(a,b)
gene_degree10<-topwhol[a:b,]</pre>
```

#### References

- 1. Csardi G, Nepusz T. The igraph software package for complex network research. InterJournal, Complex Systems. 2006;1695(5), 1-9.
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- 3. Hegyi H, Gerstein M: Annotation transfer for genomics: measuring functional divergence in multi-domain proteins. Genome Res 2001, 11:1632-1640.