Carbon limitation patterns are linked to spatio-temporal changes in dissolved organic matter quality in an urban stream

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Abstract

Introduction

As suburban sprawl converts farmland and forests to urban infrastructure, and as the global trend of urbanization continues, the biological function of urban streams and its role in water quality degradation has received increased attention. Relatively small increases in impervious surface cover through urbanization can have measurable effects on the hydrology and biology of stream ecosystems (Booth and Jackson 1997; Klein 1979), making a “flashy” hydrologic regime that reinforces entrenchment and channel incision in streams that are often already channelized to promote storm water drainage (Dunne and Leopold 1979). These channelized streams are less retentive of particulate organic carbon, and flashy hydrology also promotes organic matter export (Paul and Meyer 2001) which, in combination with greater nutrient loads (Carpenter et al. 1998) and reduced riparian canopies (Griffiths et al. 2013) common in urban streams, can alter the contribution of heterotrophic and autotrophic processes to stream metabolism. For example, canopy opening and nutrient enrichment can increase autotrophy (Bernot et al. 2010; Griffiths et al. 2013), but stream burial can increase the importance of heterotrophy relative to autotrophy (Beaulieu et al. 2014; Pennino et al. 2014). Further, open and buried stream reaches often alternate in an urban hydrological network so that stream metabolism can be vastly different in alternating stream reaches. Carbon and nutrient cycles are linked, and while recent work has advanced our knowledge of nutrient uptake and use in streams with open and buried reaches, there has been little research on dissolved organic matter (DOM) characteristics and use in open and buried streams.

The expansion of urban infrastructure to facilitate development by covering mostly low order drainages (Elmore and Kaushal 2008) fundamentally alters a stream ecosystem by shutting out light and inducing net heterotrophic conditions. Buried streams support little to no gross primary production due to the severe reduction or absence of PAR, but they also support lower ecosystem respiration (Beaulieu et al. 2014; Pennino et al. 2014). Because buried stream reaches are often optimized to convey water quickly and efficiently for drainage purposes, they have increased water flow which reduces the ability to use water column nutrients in conjunction with net reductions in overall metabolism (Beaulieu et al. 2014; Pennino et al. 2014) which together promote nutrient export to downstream reaches and ecosystems (Beaulieu et al. 2015). Burial also alters the kinds of organic matter available to stream biota. Buried reaches have lower overall coarse and fine benthic organic matter, periphyton, and chlorophyll a standing stocks compared to open reaches (Beaulieu et al. 2014). In contrast to open reaches, which had seasonality in all organic matter standing stocks, buried reaches also have little seasonality except for higher coarse benthic organic matter (CBOM) in the fall (Beaulieu et al. 2014). Given hydrologic conditions that promote particulate matter export, and stream burial that reduces biological activity and alters organic matter standing stocks, alternating buried and open reaches of an urban stream are likely to use dissolved organic matter differently.

Rivers are conduits for carbon export from the landscape to the ocean (Regnier et al. 2013), and they are important processors of carbon in the hydrosystem as ecosystem respiration transforms organic carbon to inorganic forms (Smith and Kaushal 2015). The downstream export of inorganic carbon in rivers is similar in size to the terrestrial carbon sink (Cole et al. 2007), and elevated nutrient concentrations common to urban streams stimulates decomposition, thus accelerating ecosystem respiration and possibly intensifying the demand for organic carbon (Rosemond et al. 2015). Streams depend on allochthonous organic carbon inputs from the terrestrial landscape including direct inputs from the riparian zone and DOM exported from soil by groundwater, as well as autochthonous sources from in-stream production of algae and/or macrophytes. These sources leach DOM into the carbon pool used by microbes with different sources having different quality characteristics. Allochthonous inputs are generally more recalcitrant than autochthonous sources (McKnight et al. 2001), so the DOM pool is likely to vary seasonally in conjunction with autumn leaf inputs and vernal algal blooms. Moreover, urban infrastructure likely adds variability to the DOM pool, with open reaches having more labile DOM than buried reaches due to greater light availability and associated higher levels of primary production. These seasonal and reach-scale differences in organic matter dynamics in urban streams are likely to alter microbial demand for DOM with consequences for respiration rates and inorganic carbon production.

We used a nutrient diffusing substratum (NDS) approach coupled with extracellular enzyme activity assays and DOM characterization via fluorescence techniques to understand how organic carbon demand varies seasonally in buried and open stream reaches of an urban stream. We made three hypotheses based on temporal differences in organic matter dynamics and spatial differences in light availability caused by alternating open and buried reaches in the urban stream network. We hypothesized that spring would have higher quality DOM than other seasons, and that open reaches would have higher quality DOM than buried reaches due to more algal production in open reaches and less algal production in buried reaches. Consequently, we hypothesized that spring would have fewer extracellular enzyme indicators associated with recalcitrant carbon acquisition, and that open reaches would have less effort to acquire recalcitrant carbon compared to buried reaches. Finally, we hypothesized that microbial respiration would be more carbon limited in the fall due to the pulse of terrestrial organic matter from the riparian zone compared to the spring, and that buried reaches would be more carbon limited than open reaches due to less primary production. We made a fourth hypothesis that microbial respiration would respond more strongly to the higher quality carbon compared to the lower quality carbon added to the NDS, regardless of season or reach.

Methods

Study Sites and Experimental Design

We studied three urban streams in or near Cincinnati, Ohio (USA), and each stream consisted of paired buried and open study reaches separated by a 30-100 m buffer reach. Two buried reaches flowed through corrugate pipe and one through concrete, and buried stream widths ranged from 0.5-4.5 m. Open reaches were generally incised with restricted riparian zones, contained mobile sandy sediments, and ranged in width from 2.1-3.9 m. A more detailed site description can be found in Beaulieu et al. (2014).

We collected samples to characterize dissolved organic matter quality in summer and autumn 2011 and in spring 2012. Concurrently, we deployed tiles to measure extracellular enzyme activity and nutrient diffusing substrata to measure carbon limitation patterns. This design allowed us to compare how carbon quality, microbial enzyme activity, and the biofilm response to added carbon varied in space (buried versus open stream reaches) and time (summer, autumn, and spring). We also collected a suite of other environmental data including water chemistry, hydrologic parameters, organic matter standing stocks, and whole stream metabolism and nitrate (NO3-) uptake to understand how those factors relate to the microbial response to variations in DOM quality. Nitrate uptake was measured with 15N-NO3- release in conjunction with bromide (Br-) as a conservative tracer to calculate and model hydrologic parameters. Methods that describe the processing of isotope samples, calculating NO3- uptake rate, and modeling one- and two-station whole-stream metabolism are beyond the scope of this paper, but they are detailed in Beaulieu et al. (2014).

DOM Characterization

Dissolved organic matter quality was characterized using fluorescence excitation-emission matrices (EEMs) (Coble et al. 1990, Coble 1996, Cory et al. 2010) measured on a MODEL INFORMATION. This technique allows the differentiation of humic-like, fulvic-like, and protein-like fractions of the bulk DOM pool, which in turn are generally related to the lability or recalcitrance of the DOM available to microbial consumers in the stream. To produce the indices to distinguish among these fractions of DOM, water samples were analyzed on a spectrofluorometer that measured excitation between 240-450 nm at 10 nm intervals and emission from 290-600 nm at 2 nm intervals. The three-dimensional EEMs were then instrument corrected, blank substracted, and normalized by the water Raman signal (Cory et al. 2010), but we did not measure absorbance so we did perform the standard inner-filter correction. Therefore these results will be most useful for relative differences across sites and time rather than for comparison to literature values.

The EEMs we produced allowed us to calculate a variety of indices to characterize the quality of the DOM pool, including the humification index (HIX) (Zsolnay et al. 1999; Huguet et al. 2009), the biological freshness index (BIX) (Huguet et al. 2009), the fluorescence index (FI) (McKnight et al. 2001), and the protein-to-humic ratio (P/H) (Coble 1996; Stolpe et al. 2010). HIX characterizes the humic or autochthonous fractions of DOM (Zsolnay et al. 1999; Ohno 2002), and it is calculated as the ratio of integrated fluorescence emission intensity between 300-345 nm and between 435-480 nm at 254 nm excitation. Higher HIX values indicate DOM with humic character whereas lower values indicate either less humic or more autochthonous DOM. BIX was calculated from the ratio of emission at 380 and 430 nm at excitation of 310 nm (Huguet et al. 2009). Values <0.7 are associated with allochthonous DOM, values 0.8-1.0 are associated with autochthonous DOM, and values >1.0 are associated with aquatic bacterial sources, higher values indicate greater lability than lower values. FI is calculated from the ratio of the fluorescence intensity at 450 nm and 400 nm at excitation of 370 nm. FI values of about 1.9 indicate fulvic acids from microbes and values of about 1.4 indicate terrestrial fulvic acids. Finally, P/H was calculated from the EEMs whereby excitation at 275 nm and emission at 340 nm is associated with protein-like organic matter and excitation at 350 and emission at 480 is associated with humic-like organic matter (Coble 1996; Stolpe et al. 2010). We did not perform inner filter corrections on the data so HIX cannot be compared to literature values or among seasons, but the P/H ratio will allow us to make inferences about relative abundance of labile and recalcitrant DOM among streams and seasons.

Extracellular enzyme activities (EEA)

Periphyton cultured on tiles that we deployed in the buried and open reaches was analyzed for extracellular enzyme activities (EEA). Microbial assemblages produce extracellular enzymes to degrade organic matter and to acquire nutrients from their environment, and the activity of those enzymes serves as an index of environmental resource availability (Sinsabaugh and Foreman 2001). Acquisition of labile carbon compounds was measured as -D-glucosidase activity, acquisition of recalcitrant carbon compounds was measured as polyphenol oxidase (POX) and peroxidase activity. The ratio of recalcitrant carbon acquisition total carbon acquisition (as -D-glucosidase + polyphenol oxidase) characterizes the overall quality of the DOM pool (LCI) whereby larger values represent more recalcitrant carbon (Sinsabaugh and Shah 2011). An alternate metric of recalcitrant carbon acquisition was measured as the activity of L-3,4-dihydroxyphenylalanine (DOPA) + H2O2 as a substrate. Nitrogen acquisition was measured as the activity of 3-N-acetylglucosaminidase (NACE: EC 3.2.1.50).

All EEA assays used microplate protocols developed by Sinsabaugh and colleagues (Sinsabaugh et al. 1997; Sinsabaugh and Foreman 2011) and subsequently modified by Hill et al. (2010). Microplate arrays were run with quadruplicate assays for each tested enzyme and reference standard, where were prepared in sterile deionized water. Fluorescence quenching, or the decrease of emissions caused by interaction between target enzyme substrates and non-reactant chemicals, was measured by comparing fluorescence of standard solutions mixed with sample to that of standard solution mixed with buffer. We measured fluorescence (Model FLX800T, BioTek Instruments, Winooski, VT, USA) at excitation wavelength of 350 nm and emission wavelength of 450 nm.

Nutrient diffusing substrata (NDS)

NDS arrays were deployed in the open reaches, and at the upstream and downstream end of the buried reaches. Each NDS array had one of four 0.5 M carbon amendments (glucose, arabinose, cellobiose, or a no-carbon control (n=8 each)) to represent differences in bioavailability. The NDS were supplemented with 0.5 M N as NH4Cl and 0.5 M P as KH2PO4 to alleviate any potential nutrient limitation that could confound interpretation of the heterotrophic response to added carbon, and we used porous glass disks rather than cellulose sponges to eliminate the heterotrophic response to the sponge as a particulate carbon source. NDS arrays were installed in PVC to shade them and reduce the potential for autotrophic biofilms to colonize the glass disks, and they were deployed for two weeks. Upon collection, the samples were sent overnight on ice for laboratory analysis within 24 h.

Laboratory analysis consisted of submerging the disks in site water, incubating the disks in the dark for 3.5 h, and recording net oxygen change from the start to the end of the incubation. The glass disks were saved for calculation of biomass after weighing oven-dried (60 °C) samples before and after combustion in a muffle furnace (500 °C). The respiration response was scaled by disk area (g O2 cm-2 h-1) and by biomass (mg O2 gAFDM-1 h-1), and in order to compare the respiration response among streams and seasons, we calculated the nutrient response ratio (NRR) as respiration response for an individual NDS cup divided by the mean control response for that particular deployment.

Water chemistry and hydrologic parameters

We collected filtered (0.45 m) water samples in the field and stored them ice for transport to the laboratory where they were acidified or frozen depending on the analyte. We used standard colorimetric methods to measure nitrate+nitrate (hereafter, NO3-), dissolved reactive phosphorus (DRP), ammonium (NH4+), and bromide (Br-) on a flow injection analyzer (Lachat Instruments, Loveland, CO USA). Dissolved organic carbon (DOC) concentration was measured with a total organic C analyzer with high-temperature Pt-catalyzed combustion and NDIR detection (Shimadzu TOC-VCPH, Columbia, MD, USA).

The breakthrough curve of Br- released in conjunction with the 15N-NO3- release was used in OTIS-P (Runkel, 1998), a one-dimensional advection, dispersion and transient storage model, to estimate solute hyporheic exchange parameters such as the cross-sectional area of the transient storage zone (As) and the storage zone exchange coefficient (). From these parameters, we calculated the storage zone residence time (Tsto)

Tsto = As/ \* A

where A is the cross-sectional area of the stream channel calculated from the bromide breakthrough curve and channel measurements. We calculated the storage exchange flux (qs)

qs = \*A

which represents the average water flux through the storage zone per unit length. We also calculated fraction of the median travel time due to transient storage, F200med (Runkel, 2002).

Organic matter standing stocks

We collected 10-20 samples of organic matter from different habitat units in a stratified-random sampling design. Samples for coarse (>1 mm), fine (<1 mm), and attached (i.e., periphyton) organic matter were collected from a 0.052 m2 area isolated by an open-ended plastic cylinder placed no more than 5 cm into the sediment. Coarse benthic organic matter (CBOM) was removed by hand, and the sediments were agitated before taking a fine benthic organic matter (FBOM) sample. We collected periphyton by scraping a known area (0.006-0.04 m2) of a rock with a wire brush. We calculated sample dry mass and ash-free dry mass of samples by weighing oven-dried (60 °C) samples before and after combustion in a muffle furnace (500 °C). We used a subsample of periphyton to measure chlorophyll a using the trichromatic method (APHA 2005) following hot ethanol extraction (Sartory and Grobbelaar 1984).

We deployed unglazed clay tiles for six weeks at all sites to provide a standardized surface for algae and bacteria to colonize in order to minimize any potential among site differences. Tiles were collected with the rest of the samples, and periphyton was removed with a toothbrush and razor blade, rinsed into a bottle with site water, and held on ice until arrival at the laboratory. A subset of tiles was analyzed for algal abundance using a Palmer-Maloney counting cell (Charles et al. 2002), a subset of tiles was analyzed for total bacterial counts using qPCR, and a subset of tiles was used for laboratory extracellular enzyme activity assays. Detailed methods for these analyses are described in Beaulieu et al. (2014).

Statistical Analysis

We used multivariate generalized least squares linear models (GLS) to test how DOM quality (HIX, BIX, FI, P/H) differed among seasons (summer, autumn, spring), between reaches (buried, open), and among streams. We also used GLS to test for differences in extracellular enzyme activity (POX, DOPA-H2O2, LCI, NACE) and carbon limitation patterns among seasons, between reaches, and among streams. We used linear modeling to test relationships between carbon limitation patterns and water chemistry, hydrologic parameters, organic matter standing stocks, and whole stream metabolism and nitrate (NO3-) uptake. We used permutational multivariate analysis of variance using distance matrices (adonis in the vegan package for R, Oksanen et al. 2016) to test simultaneously how CBOM and FBOM standing stocks affect the response to glucose, arabinose, and cellobiose. All statistical analyses were done using R (R Core Team 2016)

Results

Patterns in DOM Variability

We examined differences in dissolved organic matter quality among seasons (summer, autumn, spring) and between reaches (buried, open). HIX, the humification index derived from EEMs, differed by season (GLS, p=0.0005), with autumn having higher HIX than spring or summer, which were not different from each other, and also differed by reach (GLS, p=0.021) with open reaches having higher HIX than buried (Figure 1). Because we did not perform the standard inner-filter corrections on these samples, these values cannot be compared to literature values, and using these data alone, we cannot determine whether our sites or seasons have more allochthonous or autochthonous organic content. Rather they can be used to show relative difference between reach and among seasons whereas other metrics from the EEMs can clarify allochthonous/autochthonous DOM content.

For example, the biological freshness index (BIX) varied by season (GLS, p<<0.0001) but did not differ between buried and open reaches (Figure 2A). Although BIX did not differ between spring and summer, both were significantly higher compared to autumn. Typically BIX values between 0.6-0.7 are associated with DOM having low autochthonous content whereas BIX >0.9 are associated with high autochthonous content. The fluorescence index (FI) characterized whether fulvic acids in the DOM pool are microbially derived (~1.9) or terrestrially derived (~1.4), and FI was higher in summer and spring compared to autumn (GLS, p<<0.0001) but did not differ between buried and open reaches. Summer and spring did not differ from each other.

A fourth index, the protein to humic ratio (P/H), compares autochthonous and labile tryptophan-like and protein-like content (more autochthonous and labile) to more terrestrial and recalcitrant humic-like content. This ratio varied by season (GLS, p<<0.001), with spring and summer having a higher ratio (more protein) compared to fall, and also by reach (GLS, p<<0.0002) with open reaches having lower ratio (more humic-like) than buried reaches.

Patterns in extracellular enzyme activity

We deployed standard tiles for microbes to colonize for 6 weeks prior to collecting all our samples, and we measured extracellular enzyme activity to characterize microbial effort to acquire nutrients and use different carbon sources available in the environment. Extracellular enzymes that degrade L-3,4-dihydroxyphenylalanine (DOPA) + H2O2 (DOPAH2) as a substrate correlate to lignin degradation, so it is a metric of recalcitrant carbon use. While we found no significant differences in DOPAH2 among seasons, we did find that buried reaches had higher DOPAH2 than open reaches (GLS, p=0.024) when we expressed DOPAH2 per unit dry mass (Figure 4a) or per unit carbon (data not shown). We found a similar pattern in the polyphenol oxidase (POX) extracellular enzyme activity. POX is an alternate metric of recalcitrant carbon use, and we found higher POX in buried reaches compared to open reaches (GLS, p=0.0043) (Figure 4b).

We found no evidence of spatio-temporal differences in extracellular enzyme activity (EEA) associated with labile carbon use. However, when we composited metrics of carbon use together to calculate the LCI, an index of carbon lability that compares recalcitrant carbon use to total carbon use, we found that buried reaches had higher use of recalcitrant carbon (GLS, p=0.014), and we also found that summer had greater use of recalcitrant carbon than autumn (GLS, p=0.027). There were no differences between spring and autumn (Figure 5). The LCI was also correlated to the CQI, an alternate carbon quality index derived from EEA on different substrates (data not shown).

Because carbon uptake and use is often linked to the acquisition of nitrogen from the environment, we also analyzed differences in N uptake as activity of -N-acetylglucosaminidase. We measured highest values in the autumn, intermediate values in summer, and lowest values in spring with all seasons significantly different from each other (GLS, p<<0.0001), but there were no differences between reaches (Figure 6).

Carbon limitation

We deployed NDS amended with different carbon sources (glucose, arabinose, cellobiose, and a no-carbon control) to see if patterns in carbon limitation differed between buried and open stream reaches or among seasons. Unfortunately, the NDS we deployed during summer were washed away by thunderstorms that produced very flashy runoff in these urban streams dominated by impervious surface cover. Therefore, we focus our analysis on autumn and spring to contrast the carbon limitation response to a time when leaf inputs dominate compared to when vernal algae blooms dominate.

When respiration on NDS disks was scaled by biomass (mg O2 gAFDM-1 h-1), we found no differences among carbon amendments including the no carbon control. However, when the respiration response was scaled by disk area (g O2 m-2 h-1), all NDS carbon amendments were significantly different than the control in all streams, seasons, and reaches (GLS, p<<0.001), and we found no instances where the respiration response differed among the three carbon amendments during any deployment (GLS, p>0.05). Therefore, we analyzed the nutrient response ratio (NRR) of all carbon types together to detect differences between seasonal and/or reach-scale responses. Furthermore, although we deployed NDS arrays at the top and bottom of the buried reaches, there was no difference in the response, so we categorized them all as “buried.” We found a significant interaction (GLS, p=0.0009) between season (autumn versus spring) and reach (buried versus daylight) whereby the respiration response to added carbon was stronger in autumn compared to spring, but open reaches had the strongest response in the fall and buried reaches had strongest response in the spring (Figure 7).

To see what factors might predict the areal NRR response to added carbon among streams and between seasons and reaches, we analyzed a suite of reach-scale variables including standing stocks (e.g., chl a, periphyton biomass, bacterial cell counts, FBOM, CBOM etc.), water chemistry (e.g., NH4+, NO3-, SRP, DOC, etc.), hydrologic variables (e.g., Q, As/A, travel time, etc.), ecosystem-scale functional attributes (e.g., NO3- uptake, whole-system metabolism), metrics of microbial effort to acquire nutrients using EEA assays, and metrics of dissolved organic matter quality derived from excitation-emission matrices. We found no relationships between the NRR response and water chemistry, hydrology, or ecosystem-scale functional attributes. Although EEA and DOM quality metrics often differed between seasons and reaches, there was no direct relationship between NRR and those metrics. Further, most standing stock metrics were also unrelated to the NRR response, but we did find weak positive relationships between reach-scale standing stocks of CBOM (adonis, p=0.036) and FBOM (adonis, p=0.053).

Discussion

Seasonal patterns of DOM characteristics

As we hypothesized, BIX and FI, metrics of labile DOM, show a clear pattern of less labile carbon during autumn and more labile carbon during spring, possibly due to a large influx of recalcitrant terrestrial DOM in the fall, which is typically lower quality than aquatic autochthonous DOM sources (McKnight et al. 2001), and a greater contribution of autochthonous DOM in the spring. HIX, which measures the recalcitrant humic fraction of DOM, is similar to BIX and FI with autumn having higher humic character than spring or summer. This pattern is also seen in the P/H (protein/humic) ratio, which shows more humic-like components in the autumn compared to the spring whereas summer was not distinctly different. Collectively, these patterns are consistent with the reach-scale standing stock data showing higher CBOM in autumn compared to other seasons and higher chlorophyll a in spring than in other seasons (get Beaulieu et al. 2014 Fig 3). This seasonal pattern is seen in temperate forested mountain streams (Villanueva et al. 2016), ephemeral Mediterranean streams that flow during the autumn-spring wet season (Catalan et al. 2013), and in other urbanized streams (Hosen et al. 2014). Therefore, temperate zone seasonality of autumn riparian leaf inputs and spring algal blooms imparts the dominant seasonal signature to the DOM pool of these temperate urban streams even though they have limited riparian zones due to channelization.

Despite the strong and consistent seasonal differences seen across multiple DOM optical properties, the low absolute values of BIX and FI show that the DOM pool in all seasons has a weak autochthonous component and a strong signature of terrestrially-derived fulvic acids despite higher productivity and higher algal standing stocks in the spring. Urban streams could exhibit generally stronger terrestrial or humic DOM signature than streams with other land use cover. For example, microbial humic-like DOM compounds were associated with higher population density and greater proportion of developed land across nearly 200 catchments in southeast Canada (Williams et al. 2016). In contrast to the higher microbial humic signature in Williams et al. (2016), we found more terrestrial humic-like DOM compounds instead, suggesting a terrestrial DOM source. Terrestrial DOM sources include upwelling ground water, leaking stormwater infrastructure (Kaushal and Belt 2012), and runoff from impervious surfaces (Hope et al. 2014). DOM derived from these sources may overwhelm any *in situ* aquatic signature. Alternatively, the year-round stronger terrestrial/recalcitrant characteristics could indicate that heterotrophic biofilms rapidly remove high quality DOM from the water column. For example, Franke et al. (2013) found that labile carbon, such as that from autochthonous sources, stimulated water column carbon use for energy metabolism and/or assimilation, and algal biofilms enhanced the EEA of heterotrophic biofilms, suggesting the rapid use of labile DOM in the presence of autochthonous production (Rier et al. 2014). Rapid use of high quality DOM would be consistent with systemic carbon limitation, which we found in all reaches and seasons.

Spatial patterns of DOM characteristics

Our hypothesis that open reaches would have more labile carbon than buried reaches was incorrect. Although reach-scale standing stock data showed lower overall chlorophyll and CBOM in buried compared to open reaches, reach was not a significant predictor of BIX or FI, metrics that indicate labile DOM. Because buried reaches have no avenue for direct riparian inputs, chlorophyll a is nearly absent, and primary production is undetectable (Beaulieu et al. 2014), this implies that organic matter inputs to open reaches impart the BIX and FI characteristics to the DOM pool in the buried reaches. Biological activity in buried reaches, which is lower than in the open reaches (Beaulieu et al. 2014), likely does little to alter those seasonal aspects of the DOM signature due to the absence of DOM production via photosynthesis and limited DOM consumption via ecosystem respiration (Beaulieu et al. 2014). Alternatively, BIX and FI could be determined by processes at the larger stream segment or catchment scale, rather than the reach scale. For example, at the stream network scale and across a range of discharges in urbanized catchments, BIX never had a strongly autochthonous character despite many instances of net ecosystem productivity in the spring across 30 months of continuous sampling (Smith and Kaushal 2015). Further, a cross-system study that included streams found that catchment scale land use was a good predictor of DOM composition (Williams et al. 2016), which implies that catchment urbanization could have overwhelmed reach-scale differences in organic matter dynamics at our highly urbanized streams (16-34% impervious surface cover; Beaulieu et al. 2014).

In contrast to BIX and FI, which were not affected by burial, HIX was higher (more humic) in open reaches compared to buried reaches, and this also did not support our hypothesis that buried reaches would have lower quality DOM. This pattern was reflected in the P/H (protein-to-humic) ratio, which was likely driven by the relative abundance of humic-like compounds (denominator of the ratio) rather than patterns in aquatic production that affected low molecular weight protein fractions of the DOM pool (numerator of the ratio), consistent with the year-round humic nature of DOM in these urban streams. However, it is unclear if the pattern in HIX is driven by differences in the supply of organic matter to the reaches, use of DOM in the reaches, or both. More humic DOM and low P/H in the open reaches during autumn is consistent with open reaches receiving and retaining more leaf inputs than the buried reaches, making the humic character stronger in the open reaches. Assuming the HIX value at the top of the buried reach is identical to that of the upstream open reach, a lower HIX value further down the buried reach implies that the DOM character has changed as water flows through the buried reach. Several abiotic mechanisms could account for this including dilution of the DOM pool by lower HIX groundwater or sewage sources that leak into the buried reaches (Smith and Kaushal 2015), or by sorption of humic compounds during transport through the buried reaches (Ohno 2002; Zsolnay et al. 1999). Alternatively, lower HIX in the buried reach could imply consumption of high HIX compounds, and we found evidence for greater use of recalcitrant carbon in buried reaches. Higher HIX in the open reach in spring is counter-intuitive given algae blooms and high GPP, but this is likely from terrestrial organic matter leached from greenfall inputs during leaf out and/or flower or seed production, and greenfall can be an important source of carbon in forested streams (Lewis and Likens 2007). However, in the overall context of this study, the median spring HIX values are lower than autumn in open and buried reaches, which is still consistent with an overarching seasonal affect driven by terrestrial sources in the autumn and aquatic sources in the spring. (Questions for Pennino – 1. Could this also suggest that the P/H ratio is more sensitive to biological processing of the DOM pool in different reaches given that there is more effort to degrade recalcitrant carbon sources in the buried reaches, which would drive this ratio toward protein in the buried reach? 2. Can we assume that more HIX does not imply less BIX since they measure different fractions of the DOM? In other words, could we simultaneously have both higher BIX from more algal production and higher HIX from more terrestrial inputs? ).

Taken together, the DOM data show that these urban streams are dominated by terrestrial humic sources despite spatio-temporal differences in the DOM composition driven by seasonal differences in CBOM and algae. These data also show secondary control over DOM quality due to spatial differences in organic matter inputs and possibly DOM use? that alter the characteristics of the DOM pool in buried versus open reaches.

Patterns in Carbon Use – Extracellular Enzyme Activities (EEA)

Extracellular enzyme activity (EEA) reflects the composition of the DOM pool, as perceived by microbial activity. Although P/H and HIX indicated more humic recalcitrant DOM in open reaches, buried reaches had higher DOPAH2 and POX than open reaches. This supports our hypothesis that the microbial community in buried reaches would allocate more energy toward acquiring recalcitrant carbon sources than in the open reach, regardless of season. This pattern is consistent with differential biological processing of DOM in buried versus open reaches similar to what has been seen in experiments showing greater POX in low light conditions (Wagner et al. 2015). Lower values of DOPAH2 and POX in the open reach indicate less effort to acquire recalcitrant carbon in parallel with higher levels of chlorophyll a in autumn, winter, and spring that suggest more labile carbon. Conversely, the greater effort to acquire recalcitrant carbon in buried reaches is consistent with low chlorophyll, limited periphyton cover, and an extremely low reach scale GPP estimates that suggest more recalcitrant carbon. This suggests rapid use of high quality carbon produced in the open reaches and little export to downstream buried reaches, and it is consistent with generally greater EEA in the presence of algal biofilms (Rier et al. 2014).

The LCI, which aggregates several EEA measures into a composite index of carbon use, also shows greater use of recalcitrant carbon in buried reaches. However, LCI shows a seasonal effect whereby summer has greater use of recalcitrant carbon than autumn, but that autumn and spring were not different. This pattern may be driven by low CBOM, low chlorophyll a, and high FBOM in open reaches during the summer. The lack of chlorophyll and reliance on FBOM, a highly processed carbon source, may explain the high use of recalcitrant carbon sources in summer. Furthermore, the lack of difference in enzyme activity between spring and autumn despite the major differences in CBOM and chlorophyll may reflect the overall terrestrial signature of the DOM pool, which is dominated by terrestrial sources even in the spring.

While the EEA results about carbon use are entirely consistent with expectations, they do not match the optical properties we measured in the DOM pool. The water column grab sample we took for EEMs might not reflect the DOM characteristics the biofilm experiences at the boundary layer, which could be different if labile carbon produced in the benthos of the open reaches was rapidly and selectively processed by microbes. Alternatively, the long incubation times of the tiles (5-6 weeks) to culture microbes for the EEA assays might integrate microbial exposure to a different DOM pool than was reflected in the single grab sample we used for EEMs at the end of the incubation period. The mismatch between EEA and EEMs might also be related to specific substrates used in the EEA assays not corresponding to the compounds that determine the optical properties of DOM.

Although some EEA metrics did not conform to the DOM characteristics, others did. In our streams, N-acquiring enzymes had the lowest abundance in the spring, coincident with higher quality algal DOM, and highest values in summer and autumn, when overall chlorophyll is low and the system is dominated by lower quality FBOM and CBOM standing stocks respectively. For example, organic matter C:N ratio was lower during spring in forested Mediterranean streams (Villanueva et al. 2016), and higher quality spring DOM in temperate rainforest streams was likely used as a source of labile C and N (Fellman et al. 2009). We found no significant seasonal differences between NO3- or NH4+ concentrations (data not shown), suggesting that higher quality spring DOM acted as a nitrogen source as well as a carbon source. The combined approach of using EEA and EEMs provides complementary information about the characteristics of, and microbial use of, the DOM pool.

Patterns in Carbon Use – NDS

We assessed overall carbon limitation in autumn and spring, and we found an interaction between season and reach when expressed on an areal basis. Biofilms in autumn were always more limited by carbon than biofilms in spring, which supported our hypothesis, but the pattern of which reach was more limited in a given season was reversed. Open reaches were more strongly limited by carbon than buried reaches in the autumn, but they were less limited by carbon than the buried reaches in the spring. The overall seasonal effect can be explained as autumn having a pulse of recalcitrant DOM from terrestrial leaves whereas spring has a pulse of labile DOM from algal sources, and DOM optical properties confirm this. Total DOC concentration did not vary between seasons, suggesting that the pulse of autumn leaves and spring algae blooms changed DOM composition rather than quantity. The differential response might be related to secondary reach-scale factors. For example, EEA assays confirm that biofilms in buried reaches always invested more effort to acquire recalcitrant carbon, so they might have been better able to use the autumn pulse of terrestrial DOM compared to the open reaches. In contrast, biofilms in open reaches always invested less in recalcitrant carbon acquisition which, when compounded by the fact that the pulse of autumn leaves was delivered directly to the open reaches, could have led to more intensified carbon limitation. Similarly, in the spring, open reaches responded less to the simple carbon sources in the NDS because the system had higher levels of high quality algal DOM, but P/H ratio shows that buried reaches appear to get less of this higher quality DOM, so they responded more strongly to the NDS. Less high quality DOM exported to buried reaches is consistent with the potential for rapid use of algal DOM *in situ* by heterotrophic biofilms (Franke et al. 2013; Rier et al. 2014) and is reflected in the carbon acquisition effort devoted to recalcitrant carbon sources.

We found different results when we expressed carbon limitation by area or biomass (i.e., gAFDM-1). When expressed by area, results were highly significant, but when expressed by biomass, there were no differences. This implies that the biofilm response to added carbon is not to increase the per cell carbon use rate, but simply to accumulate greater biomass. Given the fact that we relieved N and P limitation to focus on the response to added carbon, these results might not apply to carbon additions in low nutrient streams, but they may be reasonably applicable to agricultural and urban streams which tend to have chronically high background nutrient concentrations. The rapid processing of added carbon could also be a function of generally high inorganic nutrient concentrations in these urban streams in combination with the nutrients added to the NDS (Rosemond et al. 2015).

Although we hypothesized different responses to added carbon in the NDS arrays, biofilms responded the same to all carbon sources (glucose, arabinose, cellobiose). Although arabinose has been used as a less labile form of carbon in some studies (e.g., Newbold et al. 2006), our results show that it is just as bioavailable as glucose in this study system. Similarly, we used cellobiose as a breakdown product of cellulose that we predicted would be less bioavailable than glucose or arabinose, yet it was equally bioavailable to those more simple carbon sources. It is unclear if arabinose and cellobiose bioavailability is equally high as glucose in most streams or if it was high in these urban streams because of the systemic dominance of recalcitrant carbon and/or the presence of N and P in the NDS agars.

Interestingly, although we measured distinct differences in the DOM pool and microbial use of different carbon and nitrogen sources, none of those metrics were directly related to the NDS response to added carbon. The only relationship we found between the carbon response and any other environmental metric was a positive response to CBOM and FBOM, and although this relationship is sensible, it is very weak and probably driven by the fall NDS response to significantly higher CBOM stocks. The lack of a relationship with nearly all of the environmental data collected, despite the NDS response being consistent with the spatio-temporal patterns of DOM quality and EEA, may reflect a significant difference in the carbon sources we used for the NDS. We used very simple carbon sources with respect to the sources in the overall bulk DOM pool, so the biofilm response to these highly labile sources could be different than the EEA response to the environmental DOM pool the biofilms were exposed to during incubation. The NDS could also have exerted high selection pressure for a particular subset of microbes that responded differently than the natural community as a whole. Alternatively, the fact that the NDS had added N and P that was unavailable to the environmental biofilms could have induced a different response to the carbon amendments. Despite not being able to link DOM characteristics or EEA metrics in a linear relationship with the NDS response, the NDS response was still consistent with those metrics suggesting this can be a good tool to explore patterns of carbon use in streams. Overall, these results suggest that spatio-temporal variation in biofilm carbon use patterns are related primarily to seasonal changes in the DOM pool and secondarily to reach scale patterns such as stream burial that can alter the microbial effort to acquire different carbon sources. Differential carbon use in an urban stream continuum can have consequences for biogeochemical cycling of other nutrients and for downstream export of DOM, nutrients, and inorganic carbon.

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Tables (part of me wants to have another table with site descriptions, but then again we can just refer to your 2014 paper)

Table 1. Coefficients from adonis, a permutational multivariate analysis of variance using distance matrices, show weak relationships between nutrient response and particulate carbon standing stocks

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | Glucose NRR | Arabinose NRR | Cellobiose NRR | P value |
| CBOM | 0.072 | 0.060 | 0.064 | 0.036 |
| FBOM | 0.014 | 0.011 | 0.01 | 0.053 |

Figure Captions

Figure 1. Spatio-temporal variation in the humification index (HIX) derived from excitation-emission matrices.

Figure 2. Seasonal variation in the (A) biological freshness index (BIX) and (B) fluorescence index (FI) derived from excitation-emission matrices.

Figure 3. Spatio-temporal variation in the protein-to-humic ratio (P/H) derived from excitation-emission matrices.

Figure 4. Reach-scale variation in ecoenzyme activity on (A) L-3,4-dihydroxyphenylalanine (DOPA) + H2O2 (DOPAH2) and (B) polyphenol oxidase (POX), both metrics of recalcitrant carbon use.

Figure 5. Spatio-temporal variation in the LCI, and index of carbon quality where larger values indicate more recalcitrant carbon in the dissolved organic matter pool.

Figure 6. Seasonal variation in -N-acetylglucosaminidase (NACE) activity of stream biofilms.

Figure 7. Spatio-temporal variation in the nutrient response ratio (NRR: respiration/mean control) to added carbon when measured on an areal basis (g O2 cm-2 h-1).

Figures

Figure 1.

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Figure 2. (Make a 2 panel graphic eventually)



Figure 3.

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Figure 4.



Figure 5.

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Figure 6.



Figure 7.

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