Host allometry influences the evolution of host range: theory and meta-analysis

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Summary

Parasites vary widely in host specificity – some parasite species infect just a single host species, while others are capable of infecting many, thereby being generalists. Understanding the factors that drive parasites' host specificity is of basic biological interest, but also directly relevant to predicting disease emergence in new host species, identifying parasites that are likely to have unidentified additional hosts, and assessing transmission risk. Here, we use mathematical models to investigate how variation in host body size, and environmental temperature affect the evolution of parasite host generalism. Parasites are more likely to evolve a generalist strategy when hosts are large-bodied, when variation in host body size is small, and in cooler environments. We then explore these predictions using a database of over 20,000 fish-macroparasite associations. Within the database we see some evidence supporting these predictions, but also highlight mismatches between theory and data. By combining these two approaches, we both establish a theoretical basis for interpreting empirical data on parasites' host specificity and identify key areas for future work.

Introduction

The range of hosts infected by a parasite species is a key factor affecting transmission. Parasites that have a diversity of host species may be able to persist in a population, where certain reservoir hosts are crucial for the maintenance of transmission [1,2]. In addition, parasites' host range is predicted to affect parasite virulence through mechanisms including relative host availability, maladaptive virulence, and fitness costs associated with infecting novel hosts [3]. The ability of a parasite to infect multiple host species, particularly across taxonomic orders, is a risk factor for emerging infectious diseases (EID) of humans and livestock [4].

Despite evidence for the importance of a parasite’s host range, it remains unclear whether most parasites are generalists or specialists, and what factors might influence the evolution of host range. EID studies, ranging from microbes to macroparasites, suggest that most parasites are generalists: 60% of human infectious diseases are zoonotic and 80% of pathogens of domestic animals infect multiple host species [4–6]. On the other hand, theory suggests that fitness trade-offs between host range and parasite performance on each host can lead to the evolution of host specialization [7], and such trade-offs are common (although not universal) for parasites [8-10]. Ecological specialization is often thought to represent an “evolutionary dead-end,” such that specialist parasites have a reduced potential to adapt to novel hosts, with correspondingly higher extinction and lower speciation rates than generalist parasites [11]. However, empirical evidence suggests that evolutionary transitions between specialism and generalism are bidirectional [12,13]. Rather few studies have examined whether specialism or generalism is the ancestral state for macroparasites of animals, but for both feather lice (arthropods) of doves and gill monogeneans (platyhelminthes) of African freshwater fish, host generalism appears to have derived from ancestral specialism [14,15].

Attempts to understand the ecological drivers of host range evolution have focused on a number of parasite or host traits and environmental factors (Table 1). For example, environmental change could lead to geographical isolation of a population, driving host/parasite co-evolution and so specialization towards a single host species, or to geographical expansion, driving opportunities for shifts to new hosts, and thus a wider host range [16] (and see Cable et al. this volume). In this study, we focus on understanding how host body size and environmental temperature affect host range.

The relationship between host size and parasitism has been explored in depth, often with reference to island biogeography theory (IBT). IBT predicts that the number of parasite species infecting a host will increase with host body size, as larger-bodied hosts represent larger habitat patches with more niches [17-19]. However, most work on this relationship has been host-centric. Few studies have considered the question from the parasite perspective, that is, do parasites that infect large-bodied hosts infect a greater diversity of hosts [20-22]? There are, however, several reasons to suspect that host body size might influence host range. Larger hosts support higher within-host parasite abundances [23], which may influence between-host transmission, for example, by positively or negatively affecting parasite shedding [24]. Host body size also affects key host characteristics, such as longevity and carrying capacity [25], that may affect host availability to parasites.

Temperature may also influence host range through a number of processes. For example, species diversity of both hosts and parasites tends to increase near the tropics [26], an increase that can be explained by increased temperature [27]. Host range may therefore increase with environmental temperature simply because there are more host species available to parasitize. Temperature can also affect parasite survival and infectivity during free-living stages, with different species and stages preferring different temperatures [28]. Given the importance of free-living stages to transmission for many parasites, temperature may then have an important affect on host range evolution. Finally, as with body size, temperature can affect important host characteristics that might affect host availability [25].

General predictions regarding correlations between host, parasite, and environmental characteristics and parasites' host specificity largely come from simple verbal models, and empirical tests of these predictions are often equivocal [10,29]. Here,,we use evolutionary invasion analysis [30] to predict when specialism/generalism is the evolutionarily stable strategy for parasites infecting a host population consisting of different host species. We use allometric scaling relationships to characterize the body size- and temperature-dependence of key host traits, and use the model to predict how host body size, temperature, and transmission mode affect the evolution of host range. We then calculate structural and phylogenetic generalism metrics [31] from an extensive data set of macroparasites of fish [32] to test these predictions. With this approach, we aim to improve our understanding of the ecological and evolutionary factors that contribute to parasite generalism.

Table 1: Examples of host and parasite traits predicted to affect the evolution of parasite generalism. Traits investigated in this paper are indicated by \*.

The one thing we don’t have in the table are phylogenetic hypotheses: parasites infecting

|  |  |  |
| --- | --- | --- |
| **Trait** | **Levels** | **Previous Hypotheses or Observations** |
| Host seeking behaviour\* | *Active*: e.g., mobile parasites that seek out hosts;  *Passive*: e.g., parasites transmitted during host-host contact or via ingestion | Parasites that actively seek out hosts should be more specific than parasites that are transmitted by direct contact between hosts. Parasites transmitted via ingestion should be less specific than parasites infecting through other routes [33]. |
| Infection site\* | *Endoparasite*:lives inside the host; *Ectoparasite*: lives on the surface of the host | Infection site will give different opportunities for transmission mode, for example the mobility of infective stages may affect the evolution of generalism [33].  Higher number of host species per parasite and network connectance observed for endoparasites compared to ectoparasites of fish [34]. |
| Life cycle | *Complex* - Transmission involves one or more intermediate hosts  *Direct* – no intermediate hosts | Parasites with complex life cycles exhibit more range in acceptable hosts and may be more likely to evolve generalism [33].  Direct life cycle parasites of primates are less host-specific than complex life cycle parasites [35]. |
| Trophic transmission\* | *Yes* - For parasites that have complex life cycles, trophic transmission occurs when the intermediate host is ingested by the terminal host  *No* - Transmission to the terminal host does not involve ingestion | Trophic transmission will restrict exposure of life stages to guilds within trophic levels, such that host-parasite associations track broadly and predictably across trophic levels because the completion of transmission in a complex system is dependent on the structure of food webs [36]. |
| Host geographic range as proxy for temperature\* | Geographic regions: Africa; Antarctica; Australia; Indopacific; Nearctic; Neotropical; Palearctic. | Allometric relationships exist between temperature and life history parameters [25]  Higher species diversity in the tropics [26,27]  Digenean parasites of marine fish in tropical seas infect fewer hosts than those that parasitize fish in colder seas [26].  No relationship is observed between latitude and generalism for Monogeneans [37]. |
| Host body size\* | Continuous (here, maximum length of fish host) | Specialist Monogenean parasites tend to be found on large-bodied fish hosts [20,21,38].  Variance in phylogenetic diversity of host species infected by fleas is negatively correlated with mean host body size [22]. |

Model Derivation and Predictions*:* **Effect of host body size and temperature on generalism**

We begin by defining a simple host-parasite system with two hosts and two environmentally transmitted parasites, since transmission is through the environment for macroparasites of fish. For ease of presentation, the model assumes a parasite with a direct life cycle; Appendix A shows a similar analysis for a trophically transmitted parasite. The first parasite is a specialist, infecting only the first (primary) host. The second parasite in a generalist, infecting both the primary host and the secondary host. In Appendix A, we also consider the case where there are two specialist parasites and a generalist parasite. We follow the dynamics of susceptible primary and secondary hosts , primary hosts infected by the specialist parasite , primary and secondary hosts infected by the generalist parasite , and specialist and generalist parasites in the environment . The dynamics of the system can be described by the following system of equations:

In the absence of any infection, the population sizes for primary and secondary hosts will be equal to the host-specific carrying capacities and . Infection occurs by parasites in the environment contacting hosts at the *per capita* rate , which is assumed to be equal across both hosts and parasites. Contact results in the removal of parasites from the environment – note that we assume that specialist parasites do not contact the secondary host, implying that contact is parasite-controlled. Infected primary and secondary hosts die at the host-specific rates and . Parasites are shed from infected hosts at the host-specific rates and . Note that the mortality and shedding rates are assumed to be independent of which parasite strain the host is infected with. However, we assume that the cost of parasite generalism is that shedding rate by generalists is a fraction *a* of the shedding rate of specialist parasites.

We use evolutionary invasion analysis [26] to determine the conditions under which a generalist parasite can invade a system where the specialist parasite is present at equilibrium. Mathematically, this corresponds to investigating the stability of the equilibrium of the full system where and where all other state variables are at the equilibrium set by the specialist parasite. The Jacobian for that equilibrium is the block-diagonal matrix,

The stability of this system is given by the eigenvalues of and . Because we have assumed that the specialist-only system reaches a stable equilibrium, all of the eigenvalues of have negative real part, so stability is determined entirely by the eigenvalues of .

Applying the next-generation matrix [27], the specialist-only system will be unstable (i.e., generalism will evolve) whenever the invasion fitness of the generalist (which we will express as ) satisfies,

where and are the equilibrium host abundances when only the resident parasite is present. These terms have intuitive biological meanings: is the probability that a parasite in the environment infects the primary host and is the expected number of new generalist parasites produced per infected primary host; the second set of terms have an analogous interpretation for the secondary host. Thus, for generalists to be able to invade, each generalist parasite in the environment must be expected to produce more than one new generalist parasite in the environment; that is a successfully invading generalist’s *R*0 will be >1.

While the expression in Eq. (1) is easy to understand, analytically it will be easier to work with a slightly different expression. Substituting in the equilibrium abundance of the primary and secondary host, the expression for simplifies to

At this point, we could specify different parameter values (corresponding to different host and parasite traits) and ask whether . For this simple model, this analysis makes very intuitive predictions (see Appendix A), as the model in its current state does not include any biologically relevant trade-offs that constrain the relationships between parameters. It is also challenging to connect the parameters of such a general model with empirical data on host-parasite associations.

To facilitate a comparison between the model and data, we take advantage of the fact that many key parameters of the model are likely to be allometric functions of host body size and temperature. In particular, host carrying capacities and mortality rates will scale with host body size [28] as

and

where is the Boltzmann factor, which describes how temperature affects reaction kinetics (e.g., metabolic rate), is the body mass of host *i,* and are proportionality constants. *E* is the average activation energy of rate-limiting biochemical metabolic reactions, *k* is Boltzmann’s constant, and *T* is temperature. Since our dataset deals with parasites of ectotherms, we assume that *T* is the same for both hosts. Increasing mass will decrease the carrying capacity and mortality rate, whereas increasing temperature decreases carrying capacity and increases mortality rate.

Host body size and temperature should also affect parasite abundance, either within-host (for endoparasites) or on host surfaces (for ectoparasites), though the scaling of abundance with body size differs in these two cases [23]. We assume that shedding rate scales linearly with parasite abundance, giving

We add these expressions into the expression above to attain host body size-, temperature-, and life cycle-dependent criteria for the evolution of generalism. In particular, it is immediately clear that, all else equal, will be larger for endoparasites than ectoparasites because shedding rate will be higher. **This makes it easier for a generalist endoparasite to invade than a generalist ectoparasite.**

The mass of the secondary host is given by , where is the mass of the primary host, and we assume that the secondary host is smaller than the primary host (). This essentially assumes that the specialist parasite has evolved to exploit the host that maximizes its fitness, since the fitness of the specialist () is an increasing function of host body size. To investigate how the evolution of generalism is affected by host body size (*W*), the difference in body size between the two hosts (*f*), and the temperature of the environment (*T*), we ask how changes with changes in these parameters (that is, we look at the derivatives of with respect to *W*, *f*, and *T*). We will consider these derivatives for both endoparasites and ectoparasites.

For endoparasites, is an increasing function of host body size:

Thus we predict that parasites infecting large-bodied hosts are more likely to be generalists than parasites infecting small-bodied hosts. That is, when looking across a large number of host-parasite associations, **there should be positive correlations between the generalism index of each parasite and both the body size of its largest host.** Moreover, **there will be a positive correlation between generalism index and mean body size** () as , and for all *W*. However, because , the correlation between generalism index and maximum body size should be larger than that between generalism and mean body size.

Similarly, is an increasing function of *f*, the relative difference in body size between hosts:

This result is intuitive: increasing *f* increases the size of the secondary host; is the sum of terms dealing with infection in the primary and secondary host, and, as we have already shown, increasing host mass increases . Note that if we allowed (secondary host larger than primary host), this would make invasion by the generalist even easier. Thus we predict that **there should be a strong positive correlation between generalism index and the coefficient of variation in host body size.** The coefficient of variation is a better metric for this prediction than the raw variance because the variance in body sizes among hosts will be positively correlated with mean body size among hosts.

Temperature likewise has a consistent effect on , but in the opposite direction: increasing temperature decreases :

Thus we predict that **generalism should be more likely in colder environments than in warmer ones**. A corollary of this (which we cannot address in our current dataset) is that generalism should be more common among parasites of ectotherms than endotherms.

We point out that the majority of endoparasites in the fish-macroparasite database are actually trophically transmitted. Thus, in Appendix B we show that these predictions hold under a model of trophic transmission as well. This trophic transmission model is more challenging to analyse, so we are forced to rely on numerical exploration to validate this prediction.

For ectoparasites (the majority of which have direct life cycles), the response of to changes in traits is more complicated. For example, the effects of increasing host mass or increasing the difference in mass between hosts are given by the derivatives

and

For both of these derivatives, the sign is determined by . Plugging in the scaling functions for and , this expression will be negative, making both derivatives positive, as they were for endoparasites, whenever

That is, it will be easier for a generalist ectoparasite to invade when host body size increases, but only up to a point. Put another way, **this predicts that there should be few generalist parasites of either very small bodied or very large bodied hosts**. If the primary host is very large, then it will be easier for a generalist to invade if the secondary host is much smaller (i.e., *f* is small). However, it is important to note that both of these predictions now depend on the values of the parameters, making these predictions somewhat more challenging to address.

The effect of temperature will be the same for both endo- and ectoparasites: **generalists will have an easier time invading when temperatures are colder.**

In Appendix A, we consider several variants of this model, to investigate the sensitivity of our predictions to the assumptions made by this model. In particular, we considered models that differed in () key ways: the number of specialist parasites; the effect of parasitism on host population growth; the control of parasite transmission; and the parasite’s life cycle. In the model presented above, we assumed that the secondary host was unexploited by any parasite, so we considered a model variant that assumed that both the primary and secondary hosts were infected by a generalist parasite and investigated when a generalist parasite could invade the system. The model above also assumes that the host population size is regulated by the parasite (for example, we showed that the size of the susceptible population of the primary host is when only the specialist parasite is present). We therefore also investigated a model where total host population size is constant. In the model above, we assumed that the parasite has complete control of the infection process. For example, the specialist parasite is removed from the environment only by susceptible primary hosts. This assumes active host seeking by the parasite and that the parasite can detect and avoid already-infected hosts. We relaxed the assumption that the parasite can detect the infection status by assuming that parasites are removed from the environment by infected hosts, but that this has no effect on the infected host (i.e., we replaced in the equation with 𝛽(𝑆1+𝐼1,𝑠)𝑃𝑔). We relaxed the assumption that an active host-seeking parasite by assuming that the parasites are removed from the environment by all hosts, regardless of whether they are susceptible that parasite or not (i.e., we replaced in the equation with 𝛽(𝑆1+𝐼1,𝑠+𝑆2+𝐼2,𝑔)𝑃𝑔). Finally, we also considered how the predictions change for a trophically transmitted parasite, when there is a single intermediate host that consumes parasites in the environment, and then transmits those parasites to either of two definitive hosts.

For all of these models, it is possible to write down the generalist’s in a form analogous to Eq. (1) above. The models differ in how that expression simplifies when the various equilibria are substituted. Table 2 shows these simplified expressions as well as our predictions for how host body size and temperature affect the likelihood that a generalist parasite can invade.

Direct life cycle

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| Number of specialist parasites | Parasite regulates population growth? | Active or passive host seeking? | Avoidance of already infected hosts? | Generalist | Effect of increased body size on | Effect of increased temperature |
| 1 | Yes | Active | Yes |  | Increase | Decrease |
| 2 | Yes | Active | Yes |  | Increase | None |
| 2 | No | Active | Yes |  | Increase | None |
| 2 | No | Active | No |  | Increase | Increase |
| 2 | No | Passive | No |  | None | None |

Trophic transmission

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| Number of specialist parasites | Parasite regulates population growth? | Active or passive host seeking? | Avoidance of already infected hosts? | Generalist | Effect of increased body size on | Effect of increased temperature |
| 1 | Yes | Active | Yes | No simple expression | Increase | Decrease |
| 2 | Yes | Active | Yes | No simple expression | Variable | Variable |
| 2 | No | Active | Yes | No simple expression | Variable | Variable |
| 2 | No | Active | No |  | None | None |
| 2 | No | Passive | No |  | None | None |

What these analyses reveal is that, for most cases, the effect of host body size remains the same - increasing host size makes it easier for generalists to invade because larger hosts support larger parasite populations, thereby increasing shedding, and larger hosts have lower mortality rates, reducing parasite virulence. The effect of temperature, however, is more complicated, and depends on the modelling assumptions.

Methods

*Data collection*

The Fish Parasite Ecology Database contains more than 38,000 records of associations between 4,650 host fish species and 11802 helminth parasites, as well as ecological, biogeographical, and phylogenetic information on the host species, including host body size and geographic region [20]. As the number of ectoparasite species was low, additional parasite-host records were included for 105 crustacean parasite species, and we included data on parasite life history traits including reproductive strategy, life cycle stages, and transmission routes from a range of primary literature sources. If there was any ambiguity regarding the taxonomic status of the parasites they were excluded from the database. To remove synonyms and other inconsistencies, host species names were quality-checked by Entrez Direct queries ([www.ncbi.nlm.nih.gov/books/NBK179288/)](http://www.ncbi.nlm.nih.gov/books/NBK179288/)) to the NCBI taxonomy database and FishBase [29]. Parasite species names were checked against NCBI taxonomy database in the same way and also checked against the NHM Host-parasite database (<http://www.nhm.ac.uk/research-curation/scientific-resources/taxonomy-systematics/host-parasites/database)> using a custom script and the World Register of Marine Species (WoRMS), Catalogue of Life (CoL), Integrated Taxonomic Information System (ITIS) and Global Names Index (GNI) databases through the Lifewatch Taxonomic Backbone (<http://www.lifewatch.be/data-services/)>. All intermediate hosts were excluded from the calculation of generalism metrics, such that generalism in parasites with complex life cycles was based on the definitive hosts only. After data cleaning, we were left with 23,331 unique host-parasite associations between 8,846 parasite species and 4,237 fish hosts.

*Host genetic distances*

Host mitochondrial DNA sequences (complete mitochondrial genomes and full or partial sequences from mitochondrial loci) were downloaded in fasta format from the NCBI nucleotide database using an Entrez Direct query. Sequences were discarded if the sequence header did not contain the species name (either full or abbreviated scientific name). They were then sorted by locus (limited to cytochrome oxidase 1, cytochrome B, 12s and 16s) based on regular expression matches to the sequence header and the assignment to loci checked by local BLASTN [30] searches (default settings, version 2.2.29) to databases of representative sequences from the relevant locus. When multiple sequences were available for a given locus and host, a consensus sequence was generated using the EMBOSS program *cons* [31]*.* The consensus sequence was used in downstream analyses except in cases when just two sequences had contributed to the consensus and the result contained more than 1% of variable positions. In such cases the first sequence was used instead of the consensus if a megaBLAST [30] search against the NCBI nucleotide database hit a member of the same genus with percentage identity greater than 90%, and the second sequence used otherwise. For hosts without sequence data for a particular locus, sequences were extracted from mitochondrial complete genomes when available using BLASTN. Consensus sequences for all available host species were combined and aligned - adjusting for direction - using MAFFT for each locus. Alignments were then trimmed with trimAl [32] to include only those columns where less than 50% of taxa had a gap and those taxa where 50% of the nucleotide positions had the same ‘element’ (e.g. a gap or a residue) as more than half of the other taxa in the alignment.

Trimmed alignments were used to compute pairwise genetic distances using the dist.dna function and the K80 model of DNA evolution [33] in the R package *ape* [34]. Since different taxa were represented among loci, the consistency of pairwise distances among loci across a range divergence times was assessed. The pairwise genetic distances between those host taxa with sequences for all loci were extracted. For each locus these pairwise genetic distances were plotted against their corresponding pairwise distance generated from cytochrome oxidase 1 sequences. The point patterns were compared to the one to one line (complete correspondence), and correspondence to this line used to select the loci to be concatenated using *ape*. Pairwise genetic distances were recalculated on this final concatenated alignment. Missing pairs were imputed using a custom R script by averaging according to the following relationships between taxa: 1) pairs from different genera – the mean of the genetic distances of one member of the pair (determined by the data available) to congeners of the remaining pair member was calculated, 2) pairs from the same genera – the mean genetic distance of all the other pairwise comparisons within that genus was calculated. Where no data were available for any member of a genus, the mean within genus average for all genera was used. If suitable data were unavailable the same principals were applied at increasing taxonomic levels (family, order, class) until values were obtained for all pairwise comparisons.

*Generalism metrics*

No information on abundance or prevalence of parasites within hosts was available within the original database (ref), so parasites’ host generalism metrics were defined according to structural and phylogenetic metrics, with phylogeny based on the pairwise genetic distances between hosts (Table 2).

Table 2: Generalism metrics calculated from host-parasite database [19]

|  |  |  |
| --- | --- | --- |
| Metric | Description | Facet |
| Degree | Number of hosts | structural |
| G | Binary measure, G=1 if degree > 1 | structural |
| SPD | Mean pairwise phylogenetic distance between all hosts [35], SPD = 0 for G=0. | phylogenetic |
| PD | Faith’s phylogenetic distance, minimum total length of all the phylogenetic branches required to span all hosts on the phylogenetic tree [36], PD = NA for G=0 | phylogenetic |
| SES-PD | Standardized Effect Size of PD based on 1000 runs, with a negative value indicating that the observed tree length is smaller (the hosts are more closely related) than what you might find by chance | phylogenetic |

A UPGMA tree was calculated from the full distance matrix using *phangorn* in R [37] and the tree reordered from root to tip so that edges from the root node were listed first (‘cladewise’ reordering). Our final list of host-parasite associations was converted to a binary ‘community data matrix’ with parasites as rows and hosts as columns (1 indicates a recorded association, 0 indicates no record).

Faith’s phylogenetic diversity index (PD; here, the length of the host tree with the root excluded) [19,36] was generated for all parasites using *picante* in R [38]. We used the ses.pd function implemented in *picante* to generate the standardized effect size of phylogenetic diversity (SES-PD) based on 1000 runs. SES-PD compares the actual Faith’s PD value for each parasite to a summary of the metric calculated after repeatedly shuffling taxa labels of all taxa in the phylogeny in order to assess if phylodiversity is high or low for a given number of hosts.

*Data analysis*

The generalism metrics for each parasite species were compared to parasite traits to test the model predictions that generalism should be positively correlated with host body size and negatively correlated with temperature.

Metrics for parasites with direct or trophic life cycles were compared to summary measures (mean, maximum, and coefficient of variation (CV)) of the maximum length reported for each of their hosts. Note that CV of the host length is only calculated for parasites with more than one host. y fparasites and ectoparasites were assessed separately, with an additional categorical length measure used for ectoparasites, whereby the length summary measures were divided into a categorical variable according to quartiles. This was done to facilitate comparison with the model prediction that generalism should be unlikely for parasites of very large, or very small, hosts.

As a rough proxy for temperature, the effect of geographic region on parasites with direct or trophic life cycles was calculated for endoparasites and ectoparasites together. Regions were assessed as defined in Table 1 and also divided into two groups, where Antarctica, Nearctic, and Palearctic were assumed to be colder than Africa, Australia, Indopacific, and Neotropical regions. Some host-parasite associations were reported in more than one region, so for the analysis based on geographic regions the generalism metrics were calculated separately for each region.

Because the generalism metrics come from very different distributions, we used GLMs with different error distributions for statistical analyses. For degree, we used negative binomial regression with a log link function (glm.nb() in R); for G, logistic regression (glm(family="binomial") in R); and for PD, SPD, and SES-PD, linear regression (lm() in R), with PD log transformed (log PD+0.01).

Results

*Pairwise genetic distances between hosts*

The total number of host species in our dataset was 4621. Pairwise genetic distances calculated using the cytochrome oxidase 1 gene corresponded to those from cytochrome B (fig. S1) and these loci were selected for concatenation and subsequent calculation of pairwise genetic distances. 3253 hosts were represented by cytochrome oxidase 1 sequences and 2193 hosts by cytochrome B. 1915 hosts had sequences from both loci whilst a total of 3531 hosts had representative sequences from one or both loci. The mean pairwise genetic distance between hosts was 0.263 (standard deviation=0.034; fig. S2).

*Infection site*

The allometric scaling model predicts that for parasites with a direct life cycle, generalism should be higher in endoparasites compared to ectoparasites. In the fish data set of macroparasites, there are 4226 parasites with a direct life cycle, of which only 10 (0.2%) are endoparasites. Due to the small sample size for endoparasites, no significant difference is found for generalism metrics by infection site (not shown).

*Host body size*

The model predicts that, for endoparasites, there should be a positive correlation among parasites’ generalism metrics and both the maximum and mean host body size, with a particularly strong positive correlation between generalism and the coefficient of variation in host body size.

For some measures we do see a positive correlation (Table 3), and in particular there is a strong positive correlation between the coefficient of variation of host length and the mean phylogenetic distance between hosts (SPD).

The model also predicts that for ectoparasites, there should be few generalist parasites of either very small bodied or very large bodied hosts. This seems to be the case for degree, PD, and PS, but not so clear for SPD (Table 3).However, because we categorized by quartiles, in some cases relatively low host body size metric values end up in the 4th quartile because the distribution of metric values is skewed with a long tail.

*Temperature/geographic range of all parasites*

The model predicts generalism is more likely in colder environments. We see higher degree, G, and SPD in cool regions.

Table 3: Model coefficients and 95% confidence intervals from generalised linear models of metrics by traits

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| **Infection site** | **Response (metric)** | **Predictor (trait)** | | **Coefficient** | **Confidence Interval** | **Unit** |
| Endoparasite | degree | Mean host body length | | -0.00152 | -0.00211, -0.000931 | unit change in log degree per cm increase in length |
| Max host body length | | 0.00545 | 0.00507, 0.00583 |
| CV host body length | | 1.43589 | 1.27125, 1.60134 |
| G | Mean host body length | | -0.00105 | -0.00194, -0.000183 | log odds ratio per cm increase in length |
| Max host body length | | 0.00652 | 0.00567, 0.00740 |
| PD | Mean host body length | | -0.00180 | -0.00264, -0.000956 | unit change in log(PD+0.01) per cm increase in length |
| Max host body length | | 0.00209 | 0.00170, 0.00249 |
| CV host body length | | 1.16029 | 1.02587, 1.29472 |
| SPD | Mean host body length | | -0.000093 | -0.000138, -0.0000477 | unit change in SPD per cm increase in length |
| Max host body length | | 0.000240 | 0.000209, 0.000271 |
| CV host body length | | 0.06276 | 0.05289, 0.07262 |
| SES-PD | Mean host body length | | -0.0108949 | -0.0150664, -0.0067235 | unit change in SES-PD per cm increase in length |
| Max host body length | | -0.0102986 | -0.0122554, -0.0083417 |
| CV host body length | | 0.7280691 | -0.0079981, 1.4641362 |
| Ectoparasite | degree | Mean host body length | Q2 | 0.2235999 | 0.1423384, 0.3049629 | unit change in log degree compared to 1st quartile category |
| Q3 | 0.406262 | 0.334761, 0.4780085 |
| Q4 | 0.0995918 | -0.0302969, 0.2285828 |
| Max host body length | Q2 | 0.2568647 | 0.1742667, 0.339628 |
| Q3 | 0.5386372 | 0.4648116, 0.6128243 |
| Q4 | 1.019527 | 0.9210028, 1.1183049 |
| CV host body length | Q2 | 0.1901761 | 0.0808269, 0.2996433 |
| Q3 | 0.674365 | 0.5707044, 0.7783339 |
| Q4 | 0.2631751 | 0.1548948, 0.3716116 |
| G | Mean host body length | Q2 | 0.3274065 | 0.1475052, 0.5078821 | log odds ratio compared to 1st quartile category |
| Q3 | 0.466491 | 0.3068503, 0.6274147 |
| Q4 | -0.2616669 | -0.5719861, 0.0389363 |
| Max host body length | Q2 | 0.7788821 | 0.590882, 0.968715 |
| Q3 | 1.0020415 | 0.8291696, 1.1774999 |
| Q4 | 1.0857277 | 0.8338062, 1.3381528 |
| PD | Mean host body length | Q2 | 0.0292508 | -0.0882624, 0.1467641 | unit change in log(PD+0.01) compared to 1st quartile category |
| Q3 | 0.0802839 | -0.0241731, 0.1847409 |
| Q4 | -0.4862968 | -0.6968441, -0.2757497 |
| Max host body length | Q2 | 0.0143388 | -0.1141987, 0.1428764 |
| Q3 | 0.110301 | -0.0082226, 0.2288246 |
| Q4 | 0.3628297 | 0.2027306, 0.5229288 |
| CV host body length | Q2 | 0.11333 | 0.0033182, 0.2233418 |
| Q3 | 0.5111788 | 0.4013089, 0.6210488 |
| Q4 | 0.5073291 | 0.3973884, 0.6172698 |
| SPD | Mean host body length | Q2 | 0.010031 | 0.0022415, 0.0178204 | unit change in SPD compared to 1st quartile category |
| Q3 | 0.012963 | 0.0060482, 0.0198778 |
| Q4 | -0.0244058 | -0.0366981, -0.0121134 |
| Max host body length | Q2 | 0.0255724 | 0.0179175, 0.0332273 |
| Q3 | 0.0291506 | 0.0221589, 0.0361422 |
| Q4 | 0.0384315 | 0.0276104, 0.0492526 |
| CV host body length | Q2 | -0.0019701 | -0.012041, 0.0081007 |
| Q3 | 0.0210478 | 0.01099, 0.0311057 |
| Q4 | 0.0550058 | 0.0449414, 0.0650701 |
| SES-PD | Mean host body length | Q2 | -0.9680191 | -1.6079538, -0.3280843 | unit change in SES-PD compared to 1st quartile category |
| Q3 | -1.4574457 | -2.0262805, -0.8886109 |
| Q4 | -3.6327163 | -4.77928, -2.4861527 |
| Max host body length | Q2 | -0.8936799 | -1.5891787, -0.1981812 |
| Q3 | -2.2616954 | -2.90301, -1.6203809 |
| Q4 | -2.3225065 | -3.1887804, -1.4562327 |
| CV host body length | Q2 | -0.7854338 | -1.394371, -0.1764966 |
| Q3 | -0.4861553 | -1.0943073, 0.1219967 |
| Q4 | 1.9586667 | 1.350123, 2.5672105 |
| Both | degree | Geographic group (ref=”warm”) | | 0.4052638 | 0.361268, 0.4492741 | unit change in log degree for cool group |
| G | 0.2775012 | 0.187575, 0.3676597 | Log odds ratio for cool group |
| PD | 0.2072072 | 0.1407433, 0.2736711 | unit change in log(PD+0.01) for cool group |
| SPD | 0.0027182 | -0.0027517, 0.008188 | unit change in SPD for cool group |
| SES-PD | -0.7006804 | -1.0391793, -0.3621815 | unit change in SES-PD for cool group |

Discussion

The number of hosts a parasite can infect has important epidemiological and evolutionary implications [1–4]. Previous authors have approached the study of host range using a comparative approach, analysing groups of closely related parasites that differ in the number of hosts infected by species within the group to attempt to identify the key host, parasite, and environmental traits that influence host range [14–18]. These studies have suggested a number of important factors thought to influence the evolution of host range (Table 1).

Here, we take a different approach, deriving simple mathematical models that incorporate host, parasite, and environmental characteristics that are likely to be generally important to the evolution of parasite generalism. In particular, we focus on variation in host body size, temperature, and parasite life cycle (endo- vs. ectoparasite).

there is another study which uses allometric scaling to think about parasites (abundance and size but not diversity, would be good to add this as contrast to our approach but supporting use of allometry) [http://dx.doi.org/10.1017/S0031182015001444](http://dx.doi.org/10.1017/S0031182015001444" \t "_blank)

The model investigates the influence of host body size, temperature, and host infection site (endo- vs. ectoparasitism, making several simple predictions. First, for parasites with a direct life cycle, endoparasites should have a wider host range than ectoparasites. Second, for endoparasites (be they directly or trophically transmitted), increasing host body size and increasing the similarity of body size between hosts both increase the likelihood of generalism evolving. Third, for endoparasites, generalism is most likely when the hosts have intermediate body size, or when the variation in body size is very large. Fourth, regardless of transmission mode or life cycle, generalism is more likely to evolve in colder environments. In general, these predictions match hypotheses proposed in previous work that did not use explicit allometric modelling (Table 1), but allow for more nuanced predictions such as the non-linear relationship between ectoparasite richness and host body size.

The model uses evolutionary invasion analysis to determine when a parasite that infects two hosts will be able to invade a system where another parasite infects only one host. This theoretical invasion possibility was assumed to translate to observed generalism in a database of host-parasite interactions. We measured structural generalism based on number of hosts (degree and G), and also looked at phylogenetic generalism in which a parasite which infects more distantly related hosts is considered more generalist than a parasite infecting closely related hosts. Therefore, we assumed that phylogenetic generalism is more likely under the same conditions as structural generalism. Phylogenetic generalism tends to correlate with number of hosts [19]. In addition, we included a measure of phylogenetic generalism that is scaled to remove the association with number of hosts (SES-PD), which tests whether phylogenetic generalism in itself matches the model predictions.

In general, the observed relationships were fairly consistent across all the generalism metrics. However, for geography/temperature, SES-PD had the opposite relationship to the other metrics, perhaps revealing a difference between phylogenetic generalism and parasite richness.

Some other limitations of the model:

* + for ectotherms doesn’t the immune response vary with temperature, such that they are immune supressed when cold, immune active when warm. If so, increased temperature might (via immunity) protect from infection and so, comparatively, reduce mortality
  + Issue with interpretation host traits model: in reality, what the model says is that, in a community with more species of similar body size, generalist parasites are likely to be able to invade. This is not the information that we can glean from the host-parasite association database, because it gives no information about the *potential* hosts within an environment.
  + Difficulty of testing predictions for ectoparasites because those predictions are more sensitive to other parameter values.
  + Limitations of endo/ecto division when GI is possibly kind of ecto
    - But Poulin also found a difference in the relationship between parasite richness and ecological traits for endos vs ectos (ectos showing no relationship with the traits examined) http://www.jstor.org/stable/2937061?seq=1#page\_scan\_tab\_contents

When model predictions were tested in a large database of host-parasite interactions, we found generally good support for the predictions (Table 3).

As expected in the model, for endoparasites, the coefficient of variation of host body size had the strongest relationship with generalism. The effect of maximum host body size was also positively correlated with generalism, while mean body size tended to have a small negative correlation with generalism. This may be due to mean body size decreasing as generalism increases in the dataset because there are more small-bodied hosts than large-bodied hosts.

There are a number of potential limitations or confounders affecting the observed relationships in the data.

* Potential limitations/confounders for data
  + Amy: Also, are there interactions going on, i.e., endoparasites (wider host range) in warmer waters (narrower host range) – does this types of thing partly explain why the results are a bit noisy?
  + (identifying what is a species of parasite is difficult to begin with which would obviously affect measures of parasite host range, and see Walker et al in this issue)

Phenotypic plasticity is important characteristic of generalist parasites

Smith et al 2008 Costa Rica 171 morphs but COI revealed another 142 spp – appreciation of species diversity switched perception on the level of host specialism within a taxa <http://www.ncbi.nlm.nih.gov/pubmed/18716001> [39]

* + Could temperature effect be confounded with available hosts and higher host diversity in warm regions? Obvious roughness of using region as a proxy for temperature.
  + We have a problem of very little data for endoparasites with direct life cycles. Correlation between traits – so generally ectos have direct and endos have complex.
  + There is also a ton of missing data on the parasite traits – many of these parasites have hardly been studied. Perhaps this introduces a bias if parasites that are better studied are more likely to be included, and better studied parasites are more likely to have been found in more hosts.
  + Generalism indices would be more informative if had abundance or prevalence data. What is the meaning of phylogenetic vs structural diversity/generalism metrics in relation to the model?
  + Only include fish hosts.
  + Categorizing ectos by quartiles potentially limiting.

The model predictions are somewhat supported by a dataset of fish parasites, but also demonstrate what additional data would be useful to collect in these types of datasets. If we had information on parasite abundance then this could be investigated alongside generalism as they are likely to be linked, and we could also then look at the relative differences in abundance as part of the measure of generalism.

Overall: using an allometric scaling model, we find support for previous hypotheses that parasite richness increases with body size, but contradict the hypothesis that parasite richness increases with temperature. The interactions between temperature and parasite infection are likely to be complex and may have different effects on different life stages, or interact with other factors like latitudinal differences in host availability (similar argument about complexity of effect of temperature is outlined in review on host-parasite cospeciation [11]). By using a model which uses allometric scaling theory, we can demonstrate a mechanistic effect of certain traits on parasite host range and strengthen or challenge existing hypotheses that are based on other areas of ecological theory.

How does this relate to transmission? The evolution of host range is closely linked to transmission, particularly in regards to reservoir hosts, spillover/emergence, and changes in virulence. As outlined in the beginning of the introduction.

CEC: DISCUSSION

At the most general level, a parasite’s host range is likely to be shaped by trade-offs between its ability to infect multiple hosts and its fitness on each host [40](Gudelj et al. 2004). Here we quantified that trade-off using the parameter *a*, which reduced the shedding rate of a generalist parasite to a fraction of that of a specialist parasite. Such a reduction in shedding might be caused by a reduction in infection intensity, as other studies have shown that generalist parasites often have lower infection intensities than specialists (Poulin 1998, Garamszegi 2006). Indeed, many experimental evolution studies have shown that, as a parasite is forced to adapt to a novel host, it gradually loses its infectiousness and/or replication ability in the original host, such that, when the parasite is able to infect both the original and novel host, its fitness is lower in each than when it is specialized (Ebert 1998). However, fitness trade-offs are notoriously challenging to measure, so assessing the importance of such trade-offs to the evolution of host range in any large host-parasite dataset is practically impossible.

As such, many other more easily quantifiable factors have been put forward as plausible determinants of host range (see reviews in Poulin 2011, Schmid-Hempel 2011, such as phylogenetic constraint, transmission mode, virulence, host availability, and host immunity. Each of these factors likely influences the trade-off between host range and fitness in each host. For example, host range might be influenced by phylogeny if there the fitness cost of infecting multiple hosts is lower when the hosts are closely related (Poulin 1992). And while each of these factors has some empirical support, it is unclear how general any one of them might be.

However, in almost all cases, the models explaining how a proposed factor is thought to affect host range evolution are verbal. While these verbal models are often intuitively appealing, it is difficult for such models to consider the dynamical feedbacks that can occur between factors, which can ultimately impact the validity and generality of the model predictions. Thus, an important benefit of studying host range evolution using mathematical models is that the analysis can help illuminate the strengths and weaknesses of such verbal theories.

For example, previous authors have suggested both that host specificity (a narrow range) is more likely to evolve when hosts are abundant because increased abundance increases the probability that a specialist will encounter its host (Combes 2001,Ward 1992). Our model analyses reveal that host abundance is unlikely to be directly relevant to the evolution of host range. This is because parasite fitness depends not on the total abundance of hosts, but on the abundance of *susceptible* hosts. The dynamical feedbacks between the host and parasite in these models cause the abundance of susceptible hosts to depend on parasite traits, rather than host traits like carrying capacity. Thus whether a generalist parasite can invade does not depend directly on host abundance, which can be seen from the fact that carrying capacity rarely appears in the generalist values presented in Table 2. The exception is when the parasite actively seeks out hosts, but cannot discriminate between infected hosts (which are a dead-end for those parasites) and susceptible hosts. In that case, generalists are more easily able to invade when there is a high probability that a parasite will be lost from the system (i.e., is large). Turning that prediction around, specialist parasites will be favoured when it is likely that they are able to come in contact with a host, as suggested by the verbal theory.

There are few host-parasite datasets with the reliable estimates of host abundance needed to directly analyse the relationship between host abundance and parasite specialization. Vzquez et al. [] analysed host-parasite interaction networks from 32 ecological communities (7 fish-metazoan parasite datasets, 25 mammal-flea datasets) to show that specialists are more likely to be infecting abundant hosts. Specifically, they showed that the empirically observed distribution of specialism across the networks could be reproduced using a null model of network assembly that assumed that the probability of a link between a randomly chosen host-parasite pair was proportional to the host abundance. In other words, Vazquez et al. showed that abundant hosts are likely to be infected by a large number of parasites, which will include many specialists, whereas less abundant hosts will only be infected by generalists.

It is important to point out that this analysis is difficult to connect with our modelling results, as Vazquez et al. is an analysis of an existing network, whereas our model is better thought of as an attempt to understand how such a network might assemble. Moreover, we did not look at the fates of the specialist parasite in our analysis. Even when a generalist parasite can invade, it does not necessarily displace the specialist parasites – it is possible that the generalist and specialist could coexist, which would then raise questions about the dynamics of coinfection. Analysis of such models would likely be a useful avenue for future research, but is beyond the scope of this work.

However, our model analyses do support the general prediction that specialists are more likely to be infecting abundant species. Almost all of the models we analysed predicted that generalists are more likely to be able to invade the system when hosts are large-bodied, and there is strong empirical evidence for a negative relationship between abundance and body size [[28]]. However, the reason for this prediction has nothing to do with the abundance of the host, but instead with the fact that shedding rates are reduced (and mortality rates elevated) in smaller hosts, making it more difficult for a generalist to succeed.

Interestingly, previous verbal models for host range evolution have suggested the correlation between host range and host body size should work in the opposite direction, with high host specificity evolving when hosts are large-bodied (Krasnov et al. 2006). The explanation given for this prediction is that large-bodied species are more predictable in their availability because they are longer-lived than smaller-bodied hosts. However, the predictability of a resource (in this case, the host) depends on the probability of encountering that resource (Ward 1992), which is determined not by resource lifespan but by abundance. Thus the observed allometric relationship between body size and abundance would seem to run counter to this verbal model. Nevertheless, not really what these show: they show that specialist parasites tend to be found infecting large-bodied hosts. One several studies have shown a negative correlation between mean or maximum host body size and generalism (Sasal et al. 1999, Desdevises et al. 2002, Krasnov et al. 2006). However, there are some challenges in comparing these data to our own study. For two of these studies, most of the parasites considered were specialists (Sasal et al.: 51/74; Desdevises et al: 12/21) and the measures of generalism were coarser than in our analysis (Sasal et al: degree only; Desdevises et al.: qualitative index of generalism). Krasnov et al. 2006 showed a negative correlation only between mean host body size and the variance in SPD, a metric we did not calculate, as it is considered less informative than SPD [35].

Virulence is another factor that is thought to affect the evolution of host range. Simple verbal models would suggest that more virulent parasites are more likely to be specialists, as the fitness trade-off for infecting multiple hosts should be steeper (Schmid-Hempel 2013). We see evidence for that prediction with some of our models as well (Table 2), as increasing the value of host mortality would often act to decrease the generalist . However, models for the evolution of virulence often illustrate how feedbacks between host specificity and parasite fitness can drive non-intuitive outcomes (Kirchner and Roy 2002). In our modelling framework, if shedding and virulence are linked (a possibility that we did not explore), then a more virulent generalist parasite might actually be more likely to invade. For example, from Table 2, the of a generalist when there are two specialist parasites is

If shedding rate is a function of virulence, then whether increased virulence increases or decreases the generalist’s depends on the signs of and . If the increase in shedding with virulence is large enough, then the virulent generalist can invade.

One thing we have ignored here is the body size dependence for the parasite. Several studies have shown that large-bodied parasites tend to be found on large-bodied hosts [20,21,38], and parasite body size may also have a large effect on the evolution of host range.

Additional Information

**Information on the following should be included whenever relevant.**

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**Ethics**

NA

**Data Accessibility**

All manuscripts which report primary data (usually research articles) should include a Data Accessibility section which states where the article's supporting data can be accessed. This section should also include details, where possible, of where to access other relevant research materials such as statistical tools, protocols, software etc. If the data has been deposited in an external repository this section should list the database, accession number and link to the DOI for all data from the article that has been made publicly available, for instance:

DNA sequences: Genbank accessions F234391-F234402 (**http://dx.doi.org/xxxxx**)  
Phylogenetic data, including alignments: TreeBASE accession number S9123 (**http://dx.doi.org/xxxxx**)  
Climate data and MaxEnt input files: Dryad doi:10.5521/dryad.12311 (**http://dx.doi.org/xxxxx**)  
  
If the data is included in the article’s Supplementary Material this should be stated here, for instance:  
The datasets supporting this article have been uploaded as part of the Supplementary Material.

**Authors' Contributions**  
All authors contributed to the conception and design of the article. CEC and AH contributed model derivation and analysis. JGW, JC, ARE acquired the data. JGW and SJP calculated the phylogenetic metrics and cleaned and analysed the data. JGW and CEC wrote the first version of the article and all authors contributed to revisions and final editing.

**Competing Interests**

We have no competing interests.

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References

1. Authors. Year. Title. *Abbreviated Journal title* **Volume**, page range. (doi)

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6. Authors. Year. Title. *Abbreviated Journal title* **Volume**, page range. (doi)

1. Streicker, D. G., Fenton, A. & Pedersen, A. B. 2013 Differential sources of host species heterogeneity influence the transmission and control of multihost parasites. *Ecol. Lett.* **16**, 975–984. (doi:10.1111/ele.12122)

2. Viana, M., Mancy, R., Biek, R., Cleaveland, S., Cross, P. C., Lloyd-Smith, J. O. & Haydon, D. T. 2014 Assembling evidence for identifying reservoirs of infection. *Trends Ecol. Evol.* **29**, 270–279. (doi:10.1016/j.tree.2014.03.002)

3. Leggett, H. C., Buckling, A., Long, G. H. & Boots, M. 2013 Generalism and the evolution of parasite virulence. *Trends Ecol. Evol.* **28**, 592–596. (doi:10.1016/j.tree.2013.07.002)

4. Cleaveland, S., Laurenson, M. K. & Taylor, L. H. 2001 Diseases of humans and their domestic mammals: pathogen characteristics, host range and the risk of emergence. *Philos. Trans. R. Soc. B Biol. Sci.* **356**, 991–999. (doi:10.1098/rstb.2001.0889)

5. Woolhouse, M. E. J., Taylor, L. H. & Haydon, D. T. 2001 Population Biology of Multihost Pathogens. *Science (80-. ).* **292**, 1109–1112. (doi:10.1126/science.1059026)

6. Taylor, L. H., Latham, S. M. & Woolhouse, M. E. J. 2001 Risk factors for human disease emergence. *Philos. Trans. R. Soc. B Biol. Sci.* **356**, 983–9. (doi:10.1098/rstb.2001.0888)

7. Futuyma, D. A. & Moreno, G. 1988. The evolution of ecological specialization. *Ann. Rev. Ecol. Syst.* **19**: 207-233.

8, Poulin, R. 1998. Large-scale patterns of host use by parasites of freshwater fishes. *Ecol. Lett.* **1**: 118-128.

9. Garamszegi, L. Z. 2006. The evolution of virulence and host specialization in malaria parasites of primates. *Ecol. Lett.* **9**: 933-940.

10. Poulin, R. 2011. *Evolutionary Ecology of Parasites*. 2nd edition. Princeton University Press.

11. Day, E. H., Hua, X., & Bromham, L. 2016. Is specialization an evolutionary dead-end? Testing for differences in speciation, extinction, and trait transition rates across diverse phylogenies of specialists and generalists. *J. Evol. Biol.* **29**: 1257-1267.

12. Agosta, S. J., Janz, N. & Brooks, D. R. 2010 How specialists can be generalists: resolving the ‘parasite paradox’ and implications for emerging infectious disease. *Zoologia* **27**, 151–162. (doi:10.1590/S1984-46702010000200001)

13. Nosil, P. & Mooers, A. Ø. 2005 Testing hypotheses about ecological specialization using phylogenetic trees. *Evolution (N. Y).* **59**, 2256. (doi:10.1554/05-169.1)

14. Johnson, K. P., Malenke, J. R. & Clayton, D. H. 2009 Competition promotes the evolution of host generalists in obligate parasites. *Proc. Biol. Sci.* **276**, 3921–6. (doi:10.1098/rspb.2009.1174)

15. Mendlová, M. & Šimková, A. 2014 Evolution of host specificity in monogeneans parasitizing African cichlid fish. *Parasit. Vectors* **7**, 69. (doi:10.1186/1756-3305-7-69)

16. Hoberg, E. P. & Brooks, D. R. 2008 A macroevolutionary mosaic: episodic host-switching, geographical colonization and diversification in complex host-parasite systems. *J. Biogeogr.* **35**, 1533–1550. (doi:10.1111/j.1365-2699.2008.01951.x)

17. Kuris, A. M., Blaustein, A. R. & Alio, J. J. 1980 Hosts as Islands. *Am. Nat.* **116**, 570–586.

18. Ezenwa, V. O., Price, S. A., Altizer, S., Vitone, N. D. & Cook, K. C. 2006 Host traits and parasite species richness in even and odd-toed hoofed mammals, Artiodactyla and Perissodactyla. *Oikos* **115**, 526–536.

19. Guégan, J.-F., Lambert, A., Lévêque, C., Combes, C. & Euzet, L. 1992 Can host body size explain the parasite species richness in tropical freshwater fishes? *Oecologia* **90**, 197–204. (doi:10.1007/BF00317176)

20. Sasal, P., Trouve, S., Muller-Graf, C. & Morand, S. 1999. Specificity and host predictability: a comparative analysis among monogenean parasites of fish. *J. Anim. Ecol.* **68**: 437-444.

21. Desdevises, Y., Morand, S, & Legendre, P. 2002. Evolution and determinants of host specificity in the genus *Lamellodiscus* (Monogenea). *Biol. J. Linn. Soc.* **77**: 432-443.

22. Krasnov, B. R., S. Morand, D. Mouillot, G. I. Shenbrot, I. S. Khokhlova, and R. Poulin. 2006. Resource predictability and host specificity in fleas: the effect of host body mass. *Parasitology* 133: 81-88.

23. Hechinger, R. 2013. A metabolic and body-size scaling framework for parasite within-host abundance, biomass, and energy flux. *Am. Nat.* 182: 234-248.

24. Cattadori, I. M., Wagner, B. R., Wodzinski, L. A., Pathak, A. K., Poole, A. and Boag, B. 2014. Infections do not predict shedding in co-infections with two helminths from a natural system. Ecology, 95: 1684–1692. doi:10.1890/13-1538.1

25.

26. Allen, A. P., Brown, J. H., & Gillooly, J. F. 2002. Global biodiversity, biochemical kinetics, and the energetic-equivalence rule. *Science* **297**: 1545-1548.

27. Pietrock, M. & Marcogliese, D. J. 2003. Free-living endohelminth stages: at the mercy of environmental conditions. *Trends Parasitol.* **19**: 293-299.

28. Schmid-Hempel, P. 2011. *Evolutionary Parasitology.* Oxford University Press.

29.

30. Poulin, R., Krasnov, B. R. & Mouillot, D. 2011 Host specificity in phylogenetic and geographic space. *Trends Parasitol.* **27**, 355–361. (doi:10.1016/j.pt.2011.05.003)

31. Strona, G., Palomares, M. L. D., Bailly, N., Galli, P. & Lafferty, K. D. 2013 Host range, host ecology, and distribution of more than 11 800 fish parasite species. *Ecology* **94**, 544. (doi:10.1890/12-1419.1)

21. Poulin, R. 1998 Host specificity. In *Evolutionary ecology of parasites: from individuals to communities*, London, UK: Chapman and Hall.

22. Bellay, S., Oliveira, E. F., Almeida-Neto, M., Mello, M. A. R., Takemoto, R. M. & Luque, J. L. 2015 Ectoparasites and endoparasites of fish form networks with different structures. *Parasitology* **142**, 901–909. (doi:10.1017/S0031182015000128)

23. Poulin, R. & Morand, S. 2004 *Parasite Biodiversity*. Washington, D.C.: Smithsonian Institution.

24. Molnár, P. K., Kutz, S. J., Hoar, B. M. & Dobson, A. P. 2013 Metabolic approaches to understanding climate change impacts on seasonal host-macroparasite dynamics. *Ecol. Lett.* **16**, 9–21. (doi:10.1111/ele.12022)

25. Morand, S., Simková, A., Matejusová, I., Plaisance, L., Verneau, O. & Desdevises, Y. 2002 Investigating patterns may reveal processes: evolutionary ecology of ectoparasitic monogeneans. *Int. J. Parasitol.* **32**, 111–119. (doi:10.1016/S0020-7519(01)00347-2)

26.

27. Hurford, A., Cownden, D. & Day, T. 2010 Next-generation tools for evolutionary invasion analyses. *J. R. Soc. Interface* **7**, 561–571. (doi:10.1098/rsif.2009.0448)

28.

29. Froese, R., Pauly, D. & Editors 2015 FishBase.

30. McGinnis, S. & Madden, T. L. 2004 BLAST: at the core of a powerful and diverse set of sequence analysis tools. *Nucleic Acids Res.* **32**, W20–W25. (doi:10.1093/nar/gkh435)

31. Rice, P., Longden, I. & Bleasby, A. 2000 EMBOSS: The European Molecular Biology Open Software Suite. *Trends Genet.* **16**, 276–277. (doi:10.1016/S0168-9525(00)02024-2)

32. Capella-Gutiérrez, S., Silla-Martínez, J. M. & Gabaldón, T. 2009 trimAl: a tool for automated alignment trimming in large-scale phylogenetic analyses. *Bioinformatics* **25**, 1972–1973. (doi:10.1093/bioinformatics/btp348)

33. Kimura, M. 1980 A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *J. Mol. Evol.* **16**, 111–120.

34. Paradis, E., Claude, J. & Strimmer, K. 2004 APE: Analyses of phylogenetics and evolution in R language. *Bioinformatics* **20**, 289–290. (doi:10.1093/bioinformatics/btg412)

35. Poulin, R. & Mouillot, D. 2003 Parasite specialization from a phylogenetic perspective: a new index of host specificity. *Parasitology* **126**, 473–480. (doi:10.1017/S0031182003002993)

36. Faith, D. P. 1992 Conservation evaluation and phylogenetic diversity. *Biol. Conserv.* **61**, 1–10. (doi:10.1016/0006-3207(92)91201-3)

37. Schliep, K. P. 2011 phangorn: phylogenetic analysis in R. *Bioinformatics* **27**, 592–593. (doi:10.1093/bioinformatics/btq706)

38. Kembel, S. W., Cowan, P. D., Helmus, M. R., Cornwell, W. K., Morlon, H., Ackerly, D. D., Blomberg, S. P. & Webb, C. O. 2010 Picante: R tools for integrating phylogenies and ecology. *Bioinformatics* **26**, 1463–1464. (doi:10.1093/bioinformatics/btq166)

39. Smith, M. A., Rodriguez, J. J., Whitfield, J. B., Deans, A. R., Janzen, D. H., Hallwachs, W. & Hebert, P. D. N. 2008 Extreme diversity of tropical parasitoid wasps exposed by iterative integration of natural history, DNA barcoding, morphology, and collections. *Proc. Natl. Acad. Sci. U. S. A.* **105**, 12359–12364. (doi:10.1073/pnas.0805319105)

40. Gudelj, I., Van den Bosch, F. & Gilligan, C. A. 2004 Transmission rates and adaptive evolution of pathogens in sympatric heterogeneous plant populations. *Proc. R. Soc. London B Biol. Sci.* **271**, 2187–2194. (doi:10.1098/rspb.2004.2837)

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Figure S1. Comparison of pairwise genetic distances between hosts generated from cytochrome oxidase 1 (CO1) sequences and cytochrome B (cytB) sequences plotted against one to one line.



Figure S2. Distribution of genetic distances from all pairwise comparisons of hosts.

* Appendix A & B: Additional model derivation detail
* Appendix C: data
* Appendix D: additional results???