Experiments to Learn Primer Sequence Bias of Reverse Transcriptase

Overview

The purpose of this set of experiments is to learn the reverse transcriptase priming efficiency of all 7mers. This will be done with primers that contain degenerate nucleotides (e.g. Random_P5 = GATCTCCGAGTTGCNNNNNNN), and the goal is to evaluate the priming efficiency of all 4⁷ combinations of NNNNNNN. Unfortunately, strong primer hairpins will form for certain NNNNNNN, and these NNNNNNN will not prime RT and will falsely indicate very poor RT priming efficiency. To overcome this problem, there are two sets of experiments ("RT Experiments 1" and "RT Experiments 2", below) distinguished by having different "random primers" that have been designed such that the hairpins that form for one random primer will not form in the other random primer. The combined results of the two experiments will be used to quantify the RT priming efficiency of all 7mers.

```
Primer Sequences
Random P5 = GATCTCCGAGTTGCNNNNNNN
Random P7 = CGATCCGCTCAACTNNNNNNN
Common P5 = CGACGCTCTTCCGATCT ACTCTGTCCGATGT CCTGCCTTGCTTTTGGTCCG
Common P7 = GTGTGCTCTTCCGATC ACTCTGTCCGATGT CCTGCCTTGCTTTTGGTCCG
Univ P5+ = CGACGCTCTTCCGATCTCCGAG
Univ P7+ = GTGTGCTCTTCCGATCCGCTC
Primer Tm's
Random P5 = 60-70C (for NNNNNNN = AAAAAAA,...,GGGGGGG)
Random P7 = 60-70C (for NNNNNNN = AAAAAAA,...,GGGGGGGG)
Common P5 = 60C
Common P7 = 60C
Univ P5+ = 30C (for partial hybridization)
            62C (for complete hybridization)
Univ P7+ = 38C (for partial hybridization)
         = 60.5C (for complete hybridization)
Universal primers (Used to construct Univ P5+ and Univ P7+; included here to confirm correct sequences used)
>UDTD5
ACACTCTTTCCCTACACGACGCTCTTCCGATCT
Universal P5
               CGACGCTCTTCCGAT
>UDTD7
GTGACTGGAGTTCAGACGTGTGCTCTTCCGATC
Universal P7 GTGTGCTCTTCCGAT
```

RT Experiments 1

Experiments to Perform

- 1.1) RT_Experiment_1.Back:
 Standard PCR using RT template 1 as input DNA and the two primers Univ P5+ and Common P7
- 1.2) RT_Experiment_1.Bias
 RT-qSeq using RT template 1 as "input RNA" and the two primers Random P5 and Common P7

Template and Primers

Note: RNA bases (ribonucleotides) are indicated in red.

POTENTIAL PROBLEM: RNAse treatment in the RT-qSeq for RT_Experiment_1.Bias would only partially degrade the "input RNA" RT_template_1. This could cause problems. Possible solutions: 1) include other RNA bases so that template is degraded to smaller pieces that would be removed in PCR cleanup; 2) no RNAse treatment.

RT Experiment 1.Back

- "Background" product after PCR with Univ_P5+ & Common_P7 primers is 112bp:

RT Experiment 1.Bias

"Bias" product after "Qiagen 1-Step" step of RT-qSeq with Random P5 & Common P7 primers is 100bp:

RT Experiments 2

Experiments to Perform

- 1) RT Experiment 2.Back:
 - Standard PCR using RT template 2 as input DNA and the two primers Univ P7+ and Common P5
- 2) RT Experiment 2.Bias

RT-qSeq using RT template 2 as "input RNA" and the two primers Random P7 and Common P5

Template and Primers

Note: RNA bases (ribonucleotides) are indicated in red

POTENTIAL PROBLEM: RNAse treatment in the RT-qSeq for RT_Experiment_1.Bias would only partially degrade the "input RNA" RT_template_1. This could cause problems. Possible solutions: 1) include other RNA bases so that template is degraded to smaller pieces that would be removed in PCR cleanup; 2) no RNAse treatment.

RT Experiment 2.Back

"Background" product after PCR with Univ P7+ & Common P5 primers is 112bp:

RT Experiment 2.Bias

- "Bias" product after "Qiagen 1-Step" step of RT-qSeq with Random P7 & Common P5 primers is 101bp: