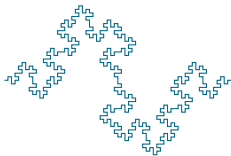


# High-Throughput Sequencing Course

## Unsupervised Learning

Biostatistics and Bioinformatics



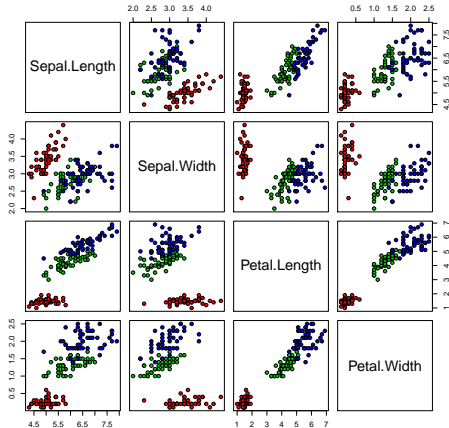
Summer 2019

## SCOPE

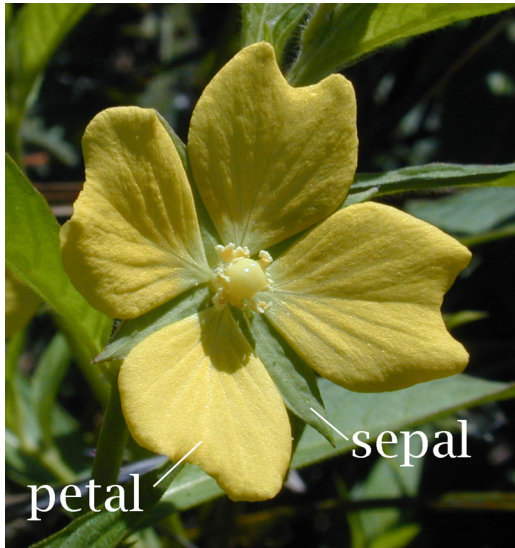
- ▶ Let  $X$  denote the genetic/genomic profile of a sample
- ▶ Often we would like to discover groups, clusters or outliers based on the genetic profiles of the samples
- ▶ These are *unsupervised* methods in the sense that the algorithm knows nothing about the grouping/clustering
- ▶ The method is only aware of the genetic profile ( $X$ ) and not the outcome  $Y$

# FISHER'S IRIS DATA

Anderson's Iris Data — 3 species



## ON PETALS AND SEPAL

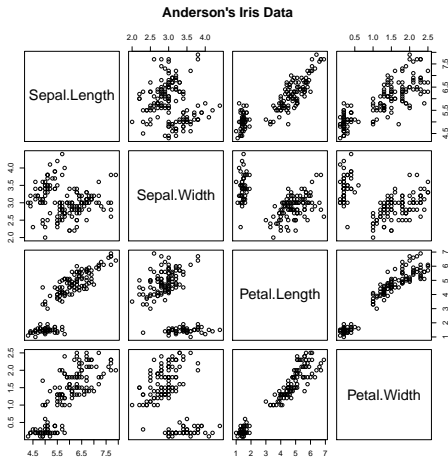


petal

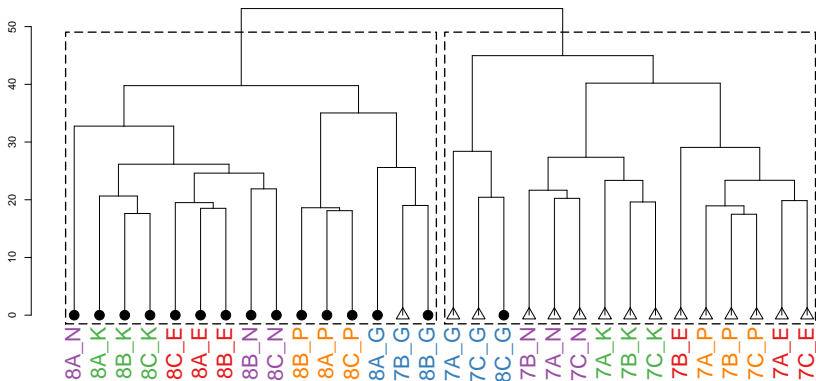
sepal

<https://en.wikipedia.org/wiki/Sepal>

# FISHER'S IRIS DATA



# 2015 DATA: AGGLOMERATIVE HIERARCHICAL CLUSTERING



## A SELF-FULFILLING PROPHECY

- ▶ Statistical methods for unsupervised learning guarantee one thing
- ▶ They will return a clustering of your data
- ▶ What they do not guarantee and are invariably unable to verify, is the biological relevance or reproducibility of the clustering
- ▶ In light of this Self-fulfilling Prophecy, these methods should be used with utmost care

## METHODS TO BE DISCUSSED

- ▶ There are many methods for unsupervised class discovery.
- ▶ We will consider three types of methods:
  - ▶ Hierarchical Clustering
  - ▶  $k$ -means Clustering
  - ▶ Ordination Methods (e.g., Multi-Dimensional Scaling (MDS) and Principal Components (PC))
- ▶ Note that there are many variations of these methods
- ▶ Most mathematical details will be left out
- ▶ We focus on discovering classes among samples (not genes)



## DISTANCE BETWEEN TWO POINTS

- ▶ Many class discover methods aim to quantify the similarity (or dissimilarity) among patients
- ▶ For each patient, the vector of gene expression can be thought of a "point" in an  $m$ -dimensional space
- ▶ For many class discovery methods, one has to be able to quantify the "distance" between two points (the expression profiles between two individuals)
- ▶ A common distance measure is the Euclidean distance

# DISTANCE (TWO POINTS ON THE PLANE)

•

•

# DISTANCE (COORDINATES)

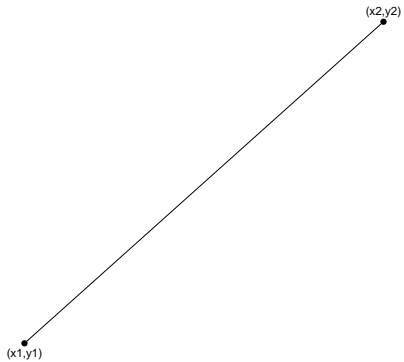
$(x_2, y_2)$



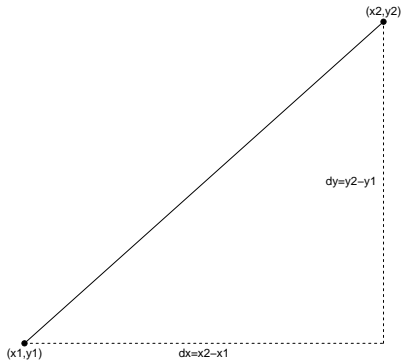
$(x_1, y_1)$



# DISTANCE



# DISTANCE (HORIZONTAL/VERTICAL SHIFTS)



# PYTHAGOREAN THEOREM (ON THE PLANE)

- ▶ According to the Pythagorean theorem

$$h^2 = dx^2 + dy^2 = (x_2 - x_1)^2 + (y_2 - y_1)^2$$

- ▶  $h$  is called the hypotenuse
- ▶ The distance between  $(x_1, y_1)$  and  $(x_2, y_2)$  is given by

$$h = \sqrt{dx^2 + dy^2} = \sqrt{(x_2 - x_1)^2 + (y_2 - y_1)^2}$$

# PYTHAGOREAN THEOREM (ON THE PLANE)

- ▶ Can be extended to higher dimensions
- ▶ In a three-dimensional space the distance between  $(x_1, y_1, z_1)$  and  $(x_2, y_2, z_2)$  is given by

$$\sqrt{(x_1 - x_2)^2 + (y_1 - y_2)^2 + (z_1 - z_2)^2}$$

- ▶ For any given dimension, the distance is obtained as the square root of the sum of the square of the coordinate-wise differences

## GOLUB *et al* LEUKEMIA DATA

- ▶ 47 patients with acute lymphoblastic leukemia (ALL)
- ▶ 25 patients with acute myeloid leukemia (AML)
- ▶ Platform: Affymetrix Hgu6800
- ▶ 7129 probe sets
- ▶ Golub *et al.* (1999). Molecular classification of cancer: class discovery and class prediction by gene expression monitoring, Science, Vol. 286:531-537.



# GOLUB *et al* LEUKEMIA DATA

Expression data from first three features and 5 patients

```
dim(exprs(Golub_Merge))
```

```
## [1] 7129 72
```

```
exprs(Golub_Merge)[1:3, 1:5]
```

```
##           39  40  42  47  48
## AFFX-BioB-5_at -342 -87  22 -243 -130
## AFFX-BioB-M_at -200 -248 -153 -218 -177
## AFFX-BioB-3_at  41  262  17 -163 -28
```

# GOLUB *et al* LEUKEMIA DATA: DISTANCE

Expression vector for patients 39 and 40

```
x <- exprs(Golub_Merge)[, "39"]  
y <- exprs(Golub_Merge)[, "40"]
```

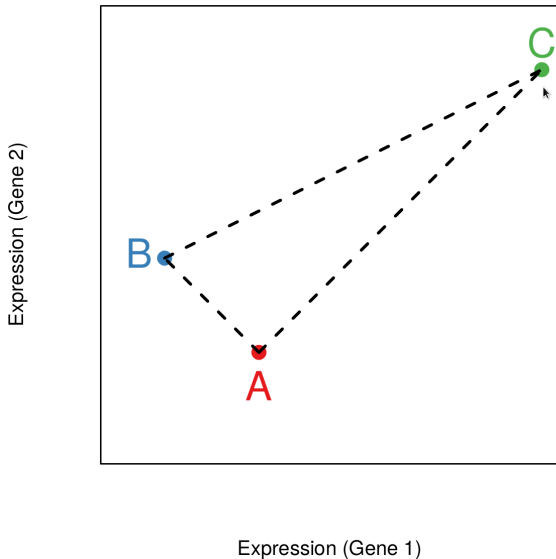
Lengths of these vectors

```
length(x)  
## [1] 7129  
  
length(y)  
## [1] 7129
```

Distance between these two vectors

```
sqrt(sum((x - y)^2))  
## [1] 101530.8
```

# RELATIVE DISTANCE (FROM CST 2011 PAPER)



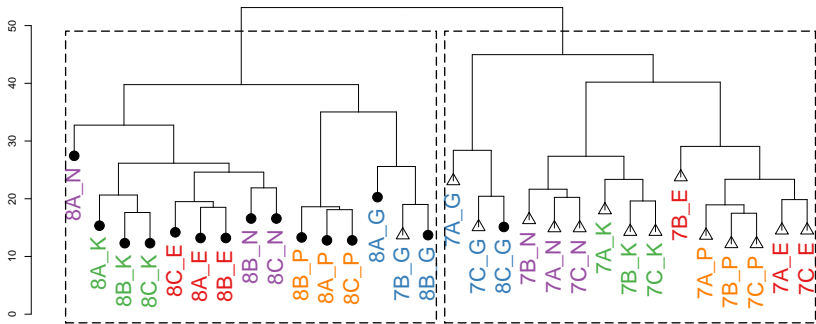
## DISSIMILARITY MATRIX

- ▶ Use pairwise distances to quantify similarity (or dissimilarity) among patients
- ▶ Construct a matrix containing all pairwise distances
- ▶ Take the first three patients in the Golub data set

```
dist(t(exprs(Golub_Merge[, 1:3])))  
##           39           40  
## 40 101530.75  
## 42 94405.04 89502.29
```

- ▶ Patient 42 is more similar (closer) to patient 39 than patient 40 (distance of 94405.04 vs 101530.75)
- ▶ Patient 39 is more similar (closer) to 42 than patient 40 (distance of 94405.04 vs 101530.75)

# 2015 DATA: AGGLOMERATIVE HIERARCHICAL CLUSTERING



# CLUSTERS

- ▶ Let  $c_1, c_2, \dots, c_n$  denote the  $n$  samples
- ▶ Define a cluster to be a set of patients
  - ▶  $(c_1)$  is a cluster with one member:  $c_1$
  - ▶  $(c_1, c_3)$  is a cluster of two members:  $c_1$  and  $c_3$
  - ▶  $(c_1, c_2, c_3)$  is a cluster of three members of  $c_1, c_2$  and  $c_3$
- ▶ Note that  $c_1$  and  $(c_1)$  are different entities

## NOTION OF A LINKAGE

- ▶ The distance measure quantified the distance between two points
- ▶ In clustering, you need to think about the criterion to link (merge) the clusters
- ▶ maximum distance (aka complete linkage)
- ▶ average distance (aka average linkage)
- ▶ minimum distance (aka single linkage)

# AGGLOMERATIVE HIERARCHICAL CLUSTERING

- ▶ Agglomerate: To form clusters
- ▶ Let each of the  $n$  points be its own cluster ( $n$  clusters each with one single member)
- ▶ Find the pair of clusters that is most similar
- ▶ Merge these two
- ▶ Now you have  $n - 1$  clusters (1 cluster with two members and  $n - 2$  clusters each with a single member)
- ▶ Compute the similarities between the  $n - 2$  "old" clusters with the new cluster
- ▶ Repeat the last two steps until all members have been merged into a single cluster.



## CLUSTERING CITIES BY DISTANCES

	ATL	BOS	ORD	DCA
ATL	0	934	585	542
BOS	934	0	853	392
ORD	585	853	0	598
DCA	542	392	598	0

## CLUSTERING CITIES BY DISTANCES (SINGLE LINKAGE)

	ATL	BOS	ORD	DCA
ATL	0	934	585	542
BOS	934	0	853	392
ORD	585	853	0	598
DCA	542	392	598	0

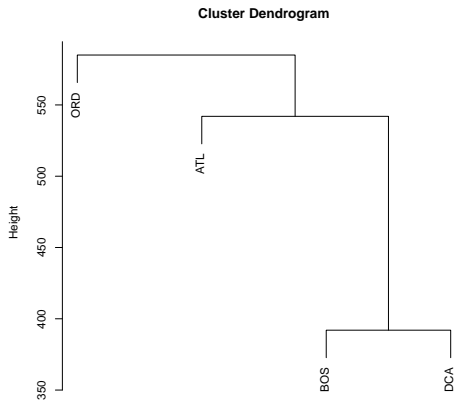
	DCA-BOS	ATL	ORD
DCA-BOS	0	542	598
ATL	542	0	585
ORD	598	585	0

## CLUSTERING CITIES BY DISTANCES (SINGLE LINKAGE)

	DCA-BOS	ATL	ORD
DCA-BOS	0	542	598
ATL	542	0	585
ORD	598	585	0

	DCA-BOS-ATL	ORD
DCA-BOS-ATL	0	585
ORD	585	0

# FOUR AIRPORTS (SINGLE LINKAGE)

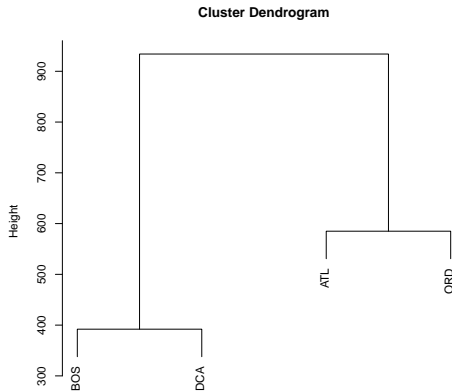


```
as.dist(cities[1:4, 1:4])  
hclust (*, "single")
```

# CLUSTERING CITIES BY DISTANCES (COMPLETE LINKAGE)

	ATL	BOS	ORD	DCA
ATL	0	934	585	542
BOS	934	0	853	392
ORD	585	853	0	598
DCA	542	392	598	0
<hr/>				
	DCA-BOS	ATL	ORD	
DCA-BOS	0	934	853	
ATL	934	0	585	
ORD	853	585	0	
<hr/>				
	DCA-BOS	ATL-ORD		
DCA-BOS	0	934		
ATL-ORD	934	0		

# FOUR AIRPORTS (COMPLETE LINKAGE)



```
as.dist(cities[1:4, 1:4])  
hclust ("", "complete")
```

## FOUR AIRPORTS (SIDE BY SIDE)

	ATL	BOS	ORD	DCA
ATL	0	934	585	542
BOS	934	0	853	392
ORD	585	853	0	598
DCA	542	392	598	0
<hr/>				
	DCA-BOS	ATL	ORD	
DCA-BOS	0	934	853	
ATL	934	0	585	
ORD	853	585	0	
<hr/>				
	DCA-BOS	ATL-ORD		
DCA-BOS	0	934		
ATL-ORD	934	0		

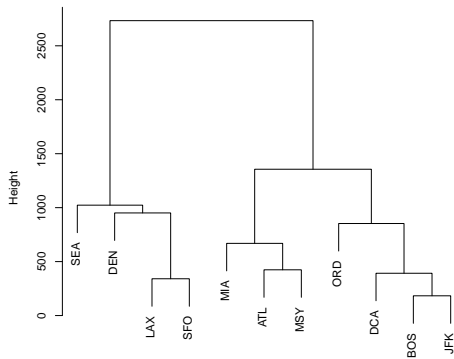
Table: Complete Linkage

	ATL	BOS	ORD	DCA
ATL	0	934	585	542
BOS	934	0	853	392
ORD	585	853	0	598
DCA	542	392	598	0
<hr/>				
	DCA-BOS	ATL	ORD	
DCA-BOS	0	542	598	
ATL	542	0	585	
ORD	598	585	0	
<hr/>				
	DCA-BOS-ATL	ORD		
DCA-BOS-ATL	0	585		
ORD	585	0		

Table: Single Linkage

# ALL AIRPORTS (COMPARISON)

Cluster Dendrogram



```
as.dist(cities)
hclust ("", "complete")
```

Cluster Dendrogram



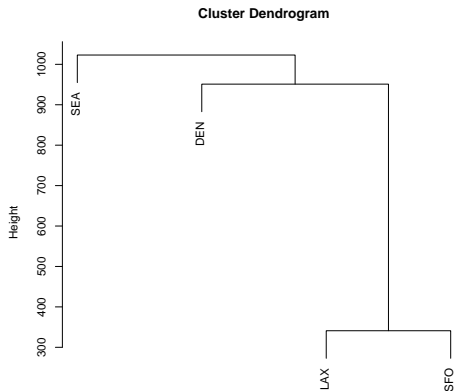


# WESTERN AIRPORTS: EXERCISE

Carry out hierarchical clustering with complete linkage

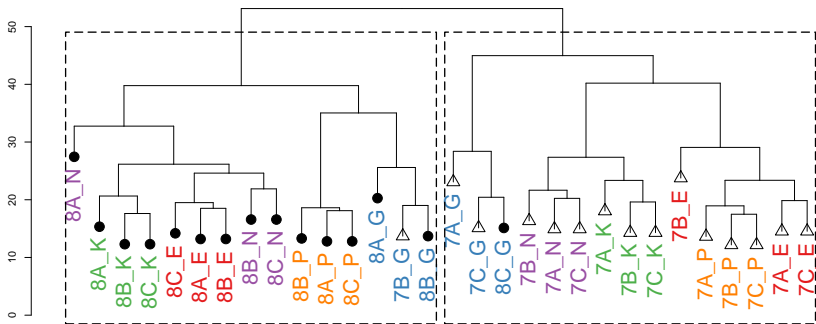
##		DEN	LAX	SEA	SFO
## DEN		0	836	1023	951
## LAX		836	0	957	341
## SEA		1023	957	0	681
## SFO		951	341	681	0

# WESTERN AIRPORTS: SOLUTION

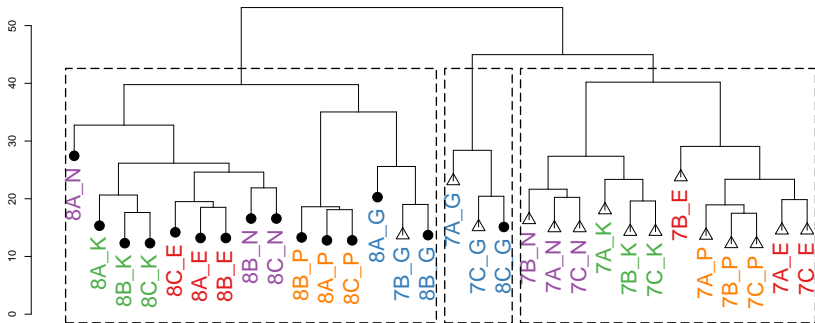


Four western airports  
hclust ("complete")

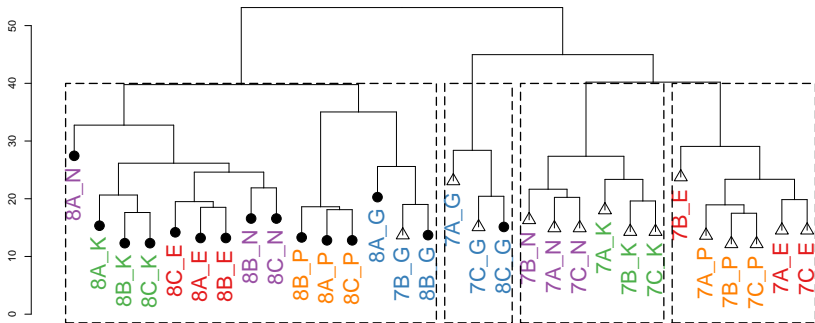
# 2015 DATA: AGGLOMERATIVE HIERARCHICAL CLUSTERING COMPLETE LINKAGE



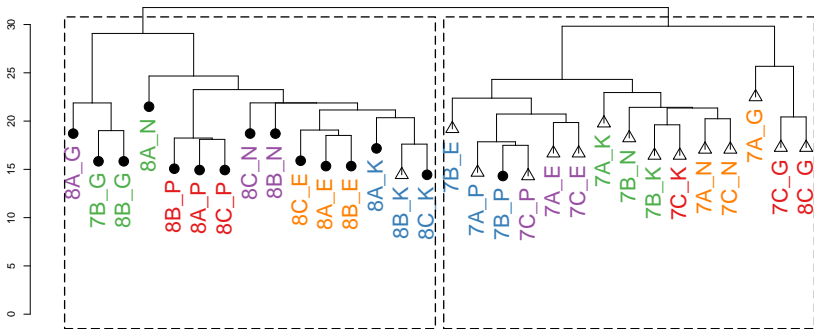
# 2015 DATA: AGGLOMERATIVE HIERARCHICAL CLUSTERING COMPLETE LINKAGE



# 2015 DATA: AGGLOMERATIVE HIERARCHICAL CLUSTERING COMPLETE LINKAGE



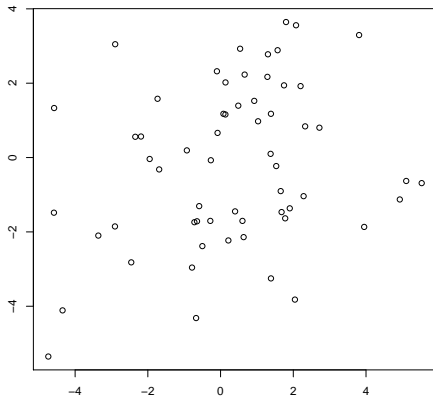
# 2015 DATA: AGGLOMERATIVE HIERARCHICAL CLUSTERING SINGLE LINKAGE



## $k$ -MEANS CLUSTERING

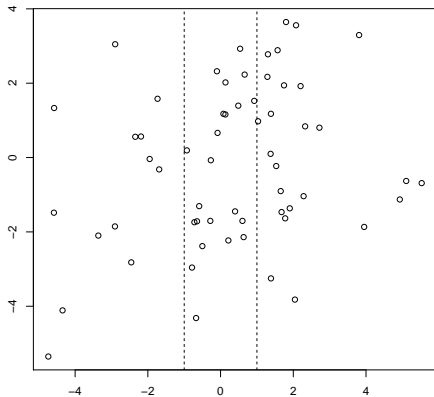
- ▶ Specify a number of potential clusters ( $k$ )
- ▶ Split of the data (either randomly or based on some previous results) into  $k$  partitions
- ▶ Compute the mean (aka centroid) for each partition
- ▶ For the first point (sample) determine the *nearest* centroid
- ▶ The closeness is typically quantified using the Euclidean distance
- ▶ Assign that point to that center
- ▶ Repeat for points 2 through  $n$
- ▶ Assess the fit using the intra-cluster variance
- ▶ Repeat as needed.

# $k$ -MEANS CLUSTERING: DATA

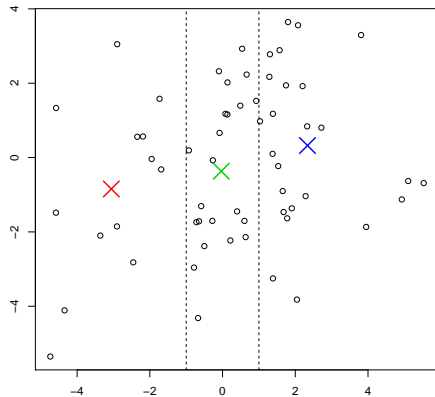




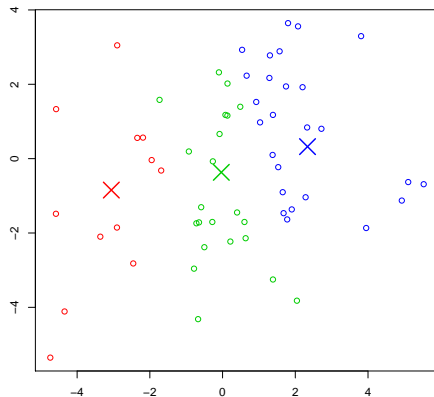
## $k$ -MEANS CLUSTERING: INITIAL CLUSTERS



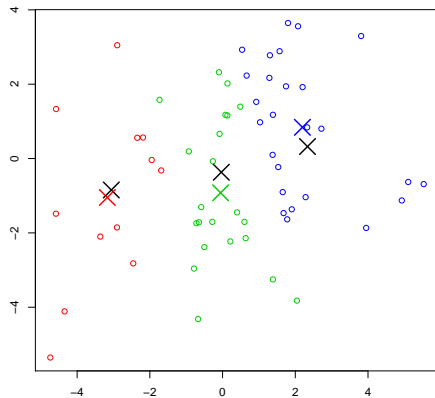
# $k$ -MEANS CLUSTERING: INITIAL CENTERS



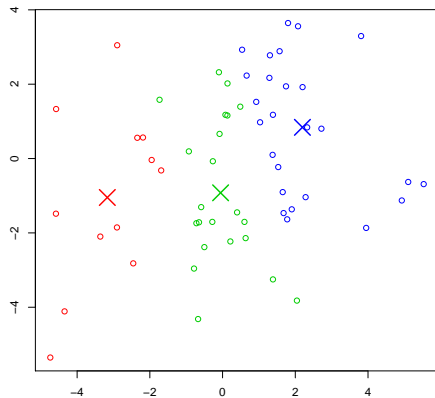
# $k$ -MEANS CLUSTERING: LABEL POINTS ACCORDING TO CENTERS



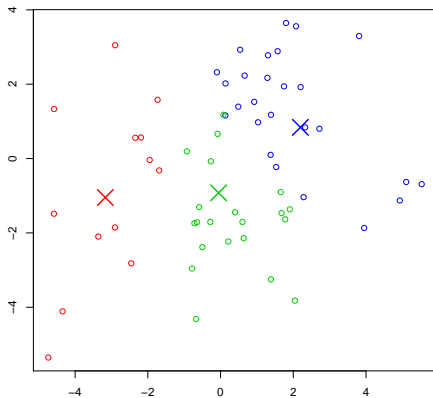
## $k$ -MEANS CLUSTERING: UPDATE CENTERS



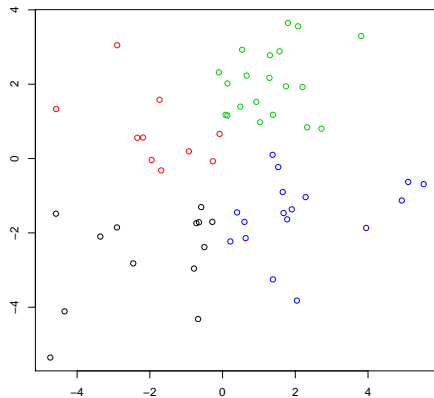
## $k$ -MEANS CLUSTERING: UPDATE CENTERS



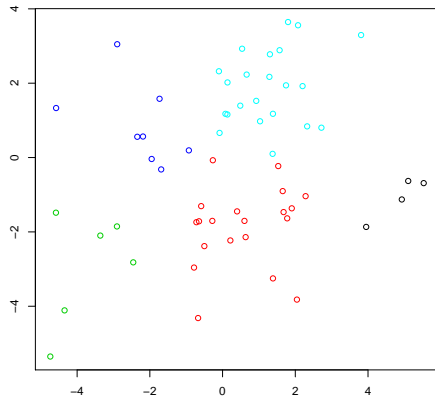
## $k$ -MEANS CLUSTERING: UPDATE POINTS



# WHY NOT 4 CLUSTERS?



# WHY NOT 5 CLUSTERS?





## $k$ -MEANS

- ▶ This is an example of *non-hierarchical* clustering
- ▶ Need to specify the number of clusters up front
- ▶ Need to specify (deterministically or randomly) the centers of the clusters up front
- ▶ Results are sensitive to the choice of  $k$  and initial partitions
- ▶ Note: All the data points were simulated from a single cluster!
- ▶ 3-means divides this single cluster into 3 subclusters
- ▶ 5-means divides this single cluster into 5 subclusters

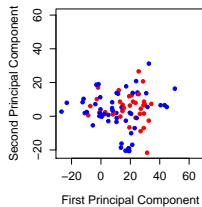
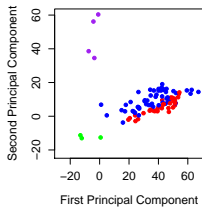
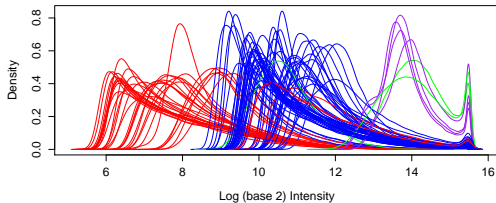
# ORDINATION ANALYSIS AND DIMENSION REDUCTION

- ▶ Genome-wide profiling platforms are high-dimensional ( $m$  is large)
- ▶ Visualization beyond  $m = 3$  not possible (for mortals)
- ▶ Representing the data by a lower dimensional format without losing too much information is desired.
- ▶ Two guiding principles:
  - ▶ Keep variables with highest variability
  - ▶ Reduce redundancy
- ▶ Two commonly used ordination analysis methods
  - ▶ Principal Components (plural) Analysis (PCA)
  - ▶ Multidimensional Scaling (MDS)

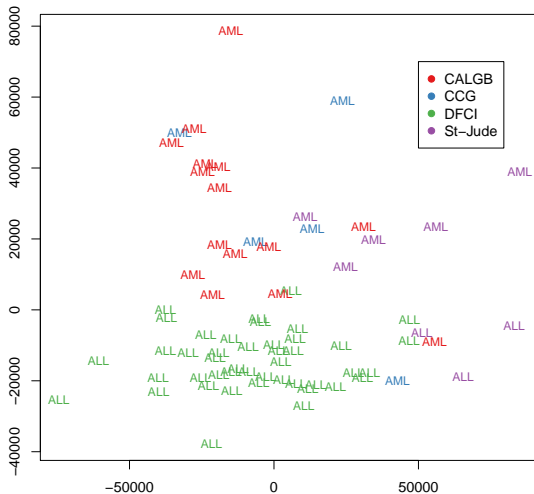
# BATCH EFFECT DISCOVERY

- ▶ PCA and MDS methods are very useful for detecting batch effects
- ▶ Batch effects tend to be stronger than biological effects
- ▶ They also affect most probe sets (the biological effect may only be captured by a few)
- ▶ This can be an effective weapon in your QC arsenal (this is how I start any new analysis)

# FROM CCR 2008 PAPER



# ALL/AML DATA (GOLUB *et. al*; SCIENCE, 1999)



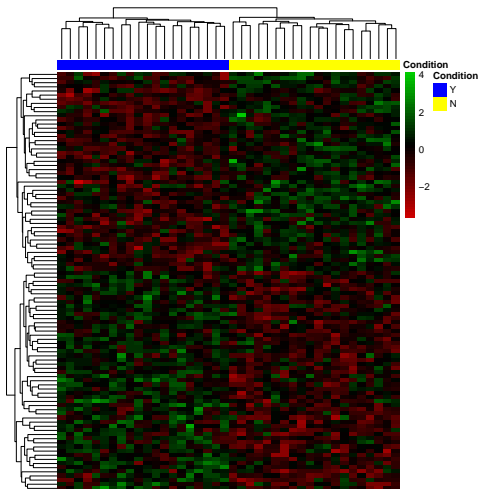
# SEMI-SUPERVISED LEARNING

- ▶ Heatmap illustration:
  - ▶ Select a panel of probe-sets based on the two-sample  $t$ -test
  - ▶ Carry out hierarchical clustering with respect to the patients (the columns)
  - ▶ Carry out hierarchical clustering with respect to the probe sets in the panel (the rows)
  - ▶ Present the results using a heatmap
- ▶ Some consider this an *unsupervised* analysis as the hierarchical clustering algorithm is unaware of the classes
- ▶ This is not an accurate assessment: It is semi-supervised in the sense that we are picking genes based on the phenotype
- ▶ A procedure is *unsupervised* if the class info is only used for annotation

# R CODE TO SIMULATE HEATMAP

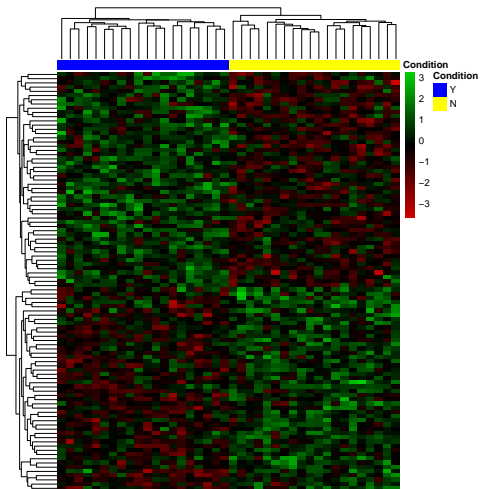
```
simulate.noise.heatmap = function(n, m, alpha) {  
  # Simulate Expression Matrix  
  EXPRS = matrix(rnorm(2 * n * m), m, 2 * n)  
  grp = factor(rep(0:1, c(n, n)))  
  rownames(EXPRS) = paste("Gene", 1:m, sep = "")  
  colnames(EXPRS) = paste("patient id", 1:(2 * n), sep = "")  
  
  # Get the two sample t-statistics  
  pvals = rowttests(EXPRS, grp)$p.value  
  topgenes = which(pvals < alpha)  
  EXPRS = EXPRS[topgenes, ]  
  annodat = data.frame(Condition = ifelse(grp == 0, "N", "Y"), row.names = colnames(EXPRS))  
  pheatmap(EXPRS, border_color = NA, show_rownames = FALSE, show_colnames = FALSE,  
    annotation_col = annodat, color = colorRampPalette(c("red3", "black",  
      "green3"))(50), annotation_colors = list(Condition = c(Y = "blue",  
        N = "yellow")))  
  return(length(topgenes))  
}
```

HEATMAP EXAMPLE:  $m = 20,000, n = 20, \alpha = 0.005$

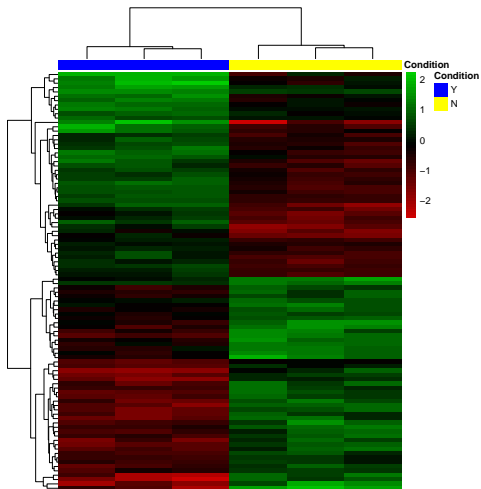




HEATMAP EXAMPLE:  $m = 40,000, n = 20, \alpha = 0.0025$



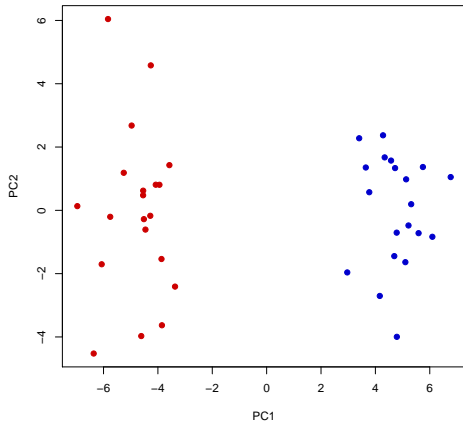
HEATMAP EXAMPLE:  $m = 20,000, n = 3, \alpha = 0.005$



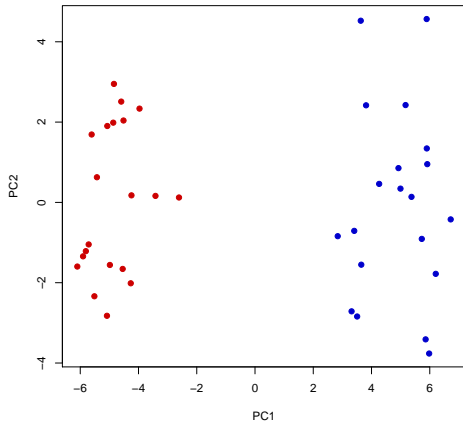
# R CODE TO SIMULATE PC

```
simulate.noise.PC = function(n, m, alpha) {  
  # Simulate Expression Matrix  
  EXPRS = matrix(rnorm(2 * n * m), m, 2 * n)  
  grp = factor(rep(0:1, c(n, n)))  
  # Get the two sample t-statistics  
  pvals = rowttests(EXPRS, grp)$p.value  
  topgenes = which(pvals < alpha)  
  EXPRS = EXPRS[topgenes, ]  
  annodat = data.frame(Condition = ifelse(grp == 0, "N", "Y"), row.names = colnames(EXPRS))  
  PC = cmdscale(dist(t(EXPRS)))  
  plot(PC, xlab = "PC1", ylab = "PC2", col = ifelse(grp == 0, "red3", "blue3"),  
        pch = 19)  
  return(length(topgenes))  
}
```

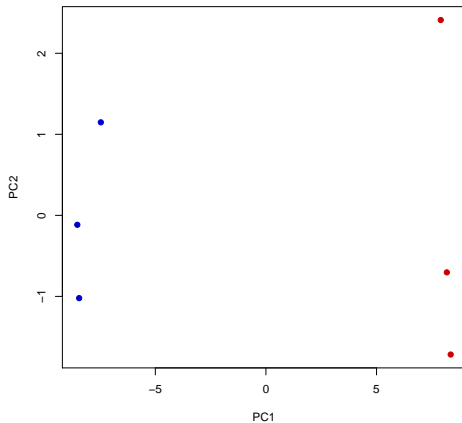
HEATMAP EXAMPLE:  $K = 20000, n = 20, \alpha = 0.005$



HEATMAP EXAMPLE:  $K = 40000, n = 20, \alpha = 0.0025$



HEATMAP EXAMPLE:  $K = 20000, n = 3, \alpha = 0.005$



## REMINDER: A SELF-FULFILLING PROPHECY

- ▶ Statistical methods for unsupervised learning guarantee one thing
- ▶ They will return a clustering of your data
- ▶ What they do not guarantee and are invariably unable to verify, is the biological relevance or reproducibility of the clustering
- ▶ In light of this Self-fulfilling Prophecy, these methods should be used with utmost care