

# Poisonous Mushroom Classification

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## Imports

*Never forget to run imports first.*

```
import requests
import zipfile
import pandas as pd
import numpy as np
import os
import altair as alt
import pandera.pandas as pa
alt.renderers.enable("png")

from sklearn.preprocessing import OneHotEncoder
from sklearn.model_selection import train_test_split
from sklearn.dummy import DummyClassifier
from sklearn.model_selection import cross_validate
from sklearn.pipeline import make_pipeline
from sklearn.svm import SVC
from sklearn.metrics import confusion_matrix, ConfusionMatrixDisplay
from scipy.stats import chi2_contingency
import itertools
from pathlib import Path
```

## Summary

Here we attempt to build a classification model using a Support Vector Machine algorithm which can use physical characteristics of mushrooms to predict whether a North American

mushroom from the *Agaricus* and *Lepiota* (Agaricaceae) family is edible or poisonous for human consumption. Our final classifier performed excellently on an unseen test data set, with precision and recall of 1.0 and an overall accuracy calculated to be 1.0. On the 2,438 test data observations, the model correctly predicted all 2,438. Although the model performs extremely well it has practical limitations, because it requires the correct identification of 20 different physical characteristics of a given mushroom in order for the corresponding prediction to be correct. We believe the model could be a powerful tool for preventing the misidentification of the *Agaricus* and *Lepiota* family of mushrooms, but further efforts to improve the practical implementation of the model for usage by the public would be very useful.

## Introduction

The *Agaricus* and *Lepiota* (Agaricaceae) family of mushrooms, includes many of the common gilled mushrooms found and cultivated in urban and suburban North America (Lincoff, 1981, 500). The family includes the common cultivated mushroom (*Agaricus bisporus*), and some of the best edible mushrooms, but it also includes poisonous mushrooms and a few that are deadly(500). These mushrooms are sometimes found in forests, but many grow on lawns, in the compost, or along roadsides and grassy areas where they are readily accessible for human consumption (500). Accurate assessment of the edibility of these mushrooms is extremely important prior to consumption, however there is no simple rule for determining whether any example is poisonous or edible (500-505).

Here we ask if we can use a supervised machine learning algorithm to predict whether a given mushroom from the *Agaricus* and *Lepiota* family of mushrooms is edible or poisonous, based upon it's physical characteristics. Answering this question is important because mushroom poisonings are common and the greatest risk factor for mushroom poisoning is the misidentification of poisonous species as edible, in particular by amateur mushroom foragers and immigrants (Diaz, 2016) If made accessible to the public in North America, accurate prediction of mushrooms in this family could prevent poisoning and support the safe consumption of common edible mushrooms.

## Methods

### Data

The data set used in this project includes hypothetical mushroom samples drawn from The Audobon Society Field Guide to North American Mushrooms by Gary Lincoff (1981, 500-525). It was sourced from the UCI Machine Learning Repository, and can be found [here](#), specifically the file *agarius-lepiota.data*. It includes 8124 descriptions of hypothetical mushroom samples that correspond to 23 species of mushrooms in the *Agaricus* and *Lepiota* Family (Mushroom,

1981). Each row includes 22 categorical variables that are physical characteristics of the mushroom sample, including the odor, gill-spacing, gill-color, gill-size, population, habitat, etc, and is labelled as edible or poisonous.

## Analysis

```
#adding the downloaded data to the data directory by code
# download data as zip and extract
url = "https://archive.ics.uci.edu/static/public/73/mushroom.zip"
response = requests.get(url)
with open("../data/raw/mushroom.zip", 'wb') as f:
    f.write(response.content)
with zipfile.ZipFile("../data/raw/mushroom.zip", 'r') as zip_ref:
    zip_ref.extractall("../data/raw")
```

## Data Validation Checks

```
'''Correct Data File Format Check'''
data_path = "../data/raw/agaricus-lepiota.data"
correct_exts = {".csv", ".data"}
ext = os.path.splitext(data_path)[1].lower()
assert ext in correct_exts, (
    f"[File format] '{data_path}' has extension '{ext}', "
    f"expected one of {sorted(correct_exts)}."
)

'''All the features in this dataset are categorical, so we can't check numeric outliers. With
```

"All the features in this dataset are categorical, so we can't check numeric outliers. With 1

## Data cleaning and wrangling

```
# Expected data column names
dataset_col_names = [
    "class", "cap_shape", "cap_surface", "cap_color", "bruises", "odor",
```

```

    "gill_attachment", "gill_spacing", "gill_size", "gill_color",
    "stalk_shape", "stalk_root",
    "stalk_surface_above_ring", "stalk_surface_below_ring",
    "stalk_color_above_ring", "stalk_color_below_ring",
    "veil_type", "veil_color",
    "ring_number", "ring_type",
    "spore_print_color", "population", "habitat"
]

```

```

df = pd.read_csv(data_path, names=dataset_col_names)
df.head()

```

	class	cap_shape	cap_surface	cap_color	bruises	odor	gill_attachment	gill_spacing	gill_size
0	p	x	s	n	t	p	f	c	n
1	e	x	s	y	t	a	f	c	b
2	e	b	s	w	t	l	f	c	b
3	p	x	y	w	t	p	f	c	n
4	e	x	s	g	f	n	f	w	b

A preview of the first five rows of the raw dataset is shown in Table Table 1, which illustrates the categorical structure of all features prior to cleaning.

## Data Validation Checks

```

'''No Duplicate Observations Check'''

```

```

duplicates = df.duplicated().sum()
assert duplicates == 0, (
    f"Found {duplicates} full-row duplicates."
)

```

```

# Data Validation Check: Target follows expected distribution
actual_distribution = (
    df["class"]
    .value_counts(normalize=True)
    .round(3)
)

```

```

        .to_dict()
    )

    # As per dataset documentation, expected proportions for each target
    expected_distribution = {'e': 0.518, 'p':0.482}

    for cls, expected_prop in expected_distribution.items():
        observed_prop = actual_distribution.get(cls, 0.0)
        assert np.isclose(observed_prop, expected_prop, atol=0.01), (
            f"Class '{cls}': observed proportion {observed_prop} "
            f"differs from expected {expected_prop}."
        )

    print(
        "[OK] Target distribution matches expected proportions "
        "(within tolerance)."
    )

```

[OK] Target distribution matches expected proportions (within tolerance).

```
'''Correct category levels check (i.e., no string mismatches or single values)'''
```

```

allowed_values = {

    "class": ["e", "p"],

    "cap_shape": ["b", "c", "x", "f", "k", "s"],

    "cap_surface": ["f", "g", "y", "s"],

    "cap_color": ["n", "b", "c", "g", "r", "p", "u", "e", "w", "y"],

    "bruises": ["t", "f"],

```

```
"odor": ["a", "l", "c", "y", "f", "m", "n", "p", "s"],

"gill_attachment": ["a", "d", "f", "n"],

"gill_spacing": ["c", "w", "d"],

"gill_size": ["b", "n"],

"gill_color": ["k", "n", "b", "h", "g", "r", "o", "p", "u", "e", "w", "y"],

"stalk_shape": ["e", "t"],

"stalk_root": ["b", "c", "u", "e", "z", "r", "?"], # ? = missing

"stalk_surface_above_ring": ["f", "y", "k", "s"],

"stalk_surface_below_ring": ["f", "y", "k", "s"],

"stalk_color_above_ring": ["n", "b", "c", "g", "o", "p", "e", "w", "y"],

"stalk_color_below_ring": ["n", "b", "c", "g", "o", "p", "e", "w", "y"],

"veil_type": ["p", "u"],

"veil_color": ["n", "o", "w", "y"],

"ring_number": ["n", "o", "t"],
```

```

    "ring_type": ["c", "e", "f", "l", "n", "p", "s", "z"],

    "spore_print_color": ["k", "n", "b", "h", "r", "o", "u", "w", "y"],

    "population": ["a", "c", "n", "s", "v", "y"],

    "habitat": ["g", "l", "m", "p", "u", "w", "d"]

}

for col, allowed in allowed_values.items():

    invalid = set(df[col].dropna().unique()) - set(allowed)

    assert not invalid, f"{col} has invalid values: {invalid}"

```

```

# Basic missingness summary (raw data)
df.isna().sum()

```

```

class                                0
cap_shape                            0
cap_surface                          0
cap_color                            0
bruises                              0
odor                                 0
gill_attachment                      0
gill_spacing                         0
gill_size                            0
gill_color                           0
stalk_shape                          0
stalk_root                           0
stalk_surface_above_ring             0
stalk_surface_below_ring             0
stalk_color_above_ring               0
stalk_color_below_ring               0

```

```

veil_type          0
veil_color         0
ring_number        0
ring_type          0
spore_print_color  0
population         0
habitat           0
dtype: int64

```

```

# Encoded missingness via '?':
(df == "?").sum()

```

```

class              0
cap_shape          0
cap_surface        0
cap_color          0
bruises           0
odor              0
gill_attachment    0
gill_spacing       0
gill_size          0
gill_color         0
stalk_shape        0
stalk_root         2480
stalk_surface_above_ring  0
stalk_surface_below_ring  0
stalk_color_above_ring  0
stalk_color_below_ring  0
veil_type          0
veil_color         0
ring_number        0
ring_type          0
spore_print_color  0
population         0
habitat           0
dtype: int64

```

```

# Convert "?" markers to proper NaN in case any other should occur apart from the known `stalk`
df = df.replace("?", np.nan)

```

```
# Changing the target to numeric values: poisonous=1, edible=0
```

```
df["is_poisonous"] = df["class"].map({"p": 1, "e": 0})
```

```
# Drop rows with NaN in the target column (none expected, but safe)
```

```
df = df.dropna(subset=["is_poisonous"])
```

```
# Drop stalk_root column due to missing values and original target column "class" because not needed  
df.drop(columns=['stalk_root', 'class'], inplace=True)
```

```
# Check remaining missing values
```

```
df.isna().sum()
```

```
cap_shape      0  
cap_surface    0  
cap_color      0  
bruises        0  
odor           0  
gill_attachment 0  
gill_spacing   0  
gill_size      0  
gill_color     0  
stalk_shape    0  
stalk_surface_above_ring 0  
stalk_surface_below_ring 0  
stalk_color_above_ring 0  
stalk_color_below_ring 0  
veil_type      0  
veil_color     0  
ring_number    0  
ring_type      0  
spore_print_color 0  
population     0  
habitat        0  
is_poisonous   0  
dtype: int64
```

```
# Validate cleaned column names
```

```
# Given that the dataset we read in for this model did not include headers, we assigned the expected column names  
expected_cols_cleaned = [
```

```

    col for col in dataset_col_names if col not in ["stalk_root", "class"]
] + ["is_poisonous"]

assert list(df.columns) == expected_cols_cleaned, \
    "The dataframe does not have the expected column names after cleaning."

print("[OK] Cleaned column names validated.")

```

[OK] Cleaned column names validated.

```

# Pandera: validate cleaned dataframe schema
# - All feature columns are strings
# - Target column is integer and only takes values {0, 1}

schema = pa.DataFrameSchema(
    {
        # All predictor/feature columns: stored as strings
        **{
            col: pa.Column(str)
            for col in expected_cols_cleaned
            if col != "is_poisonous"
        },
        # Target column: must be integer and in the set {0, 1}
        "is_poisonous": pa.Column(
            int,
            pa.Check.isin([0, 1]),
        ),
    }
)

# Validate the cleaned dataframe against the schema
schema.validate(df, lazy=True)

print(
    "[OK] Pandera schema validation passed:\n"
    "  - all feature columns are string-typed\n"
    "  - 'is_poisonous' contains only 0/1 as expected."
)

```

[OK] Pandera schema validation passed:

- all feature columns are string-typed
- 'is\_poisonous' contains only 0/1 as expected.

## Interpretation of Outlier / Anomalous Value Checks

All features in this dataset are categorical, so traditional numeric outlier checks (e.g., detecting extreme continuous values) do not apply. Instead, the relevant “anomalous values” for categorical data are unexpected or invalid category levels.

We addressed this by validating every categorical column against the official list of allowed levels from the UCI dataset documentation. Since all observed category values matched their expected sets, we conclude that no anomalous values are present in this dataset.

```
# Pandera: validate cleaned dataframe schema
# - Ensure there are no completely empty rows
# - all columns must be present
# - enforces no missing in `is_poisonous`
# - enforces <= 5% missingness in any other column

mushroom_schema = pa.DataFrameSchema(
    {
        "cap_shape": pa.Column(str),
        "cap_surface": pa.Column(str),
        "cap_color": pa.Column(str),
        "bruises": pa.Column(str),
        "odor": pa.Column(str),
        "gill_attachment": pa.Column(str),
        "gill_spacing": pa.Column(str),
        "gill_size": pa.Column(str),
        "gill_color": pa.Column(str),
        "stalk_shape": pa.Column(str),
        "stalk_surface_above_ring": pa.Column(str),
        "stalk_surface_below_ring": pa.Column(str),
        "stalk_color_above_ring": pa.Column(str),
        "stalk_color_below_ring": pa.Column(str),
        "veil_type": pa.Column(str),
        "veil_color": pa.Column(str),
        "ring_number": pa.Column(str),
        "ring_type": pa.Column(str),
        "spore_print_color": pa.Column(str),
        "population": pa.Column(str),
        "habitat": pa.Column(str),
        "is_poisonous": pa.Column(
            int,
            pa.Check(
                lambda s: s.isna().mean() == 0.0,
```

```

        element_wise=False,
        error="Target 'is_poisonous' contains missing values.",
    ),
    nullable=False,
),
},
checks=[
    # No empty observations: no row where *all* columns are missing
    pa.Check(
        lambda df: ~(df.isna().all(axis=1)).any(),
        element_wise=False,
        error="Empty rows found.",
    ),
    # Missingness not beyond expected threshold (up to max 5% per column)
    pa.Check(
        lambda df: df.isna().mean().max() <= 0.05,
        element_wise=False,
        error="One or more columns exceed 5% missingness.",
    ),
],
)

# Validate the cleaned dataframe (will raise if check fails)
df = mushroom_schema.validate(df, lazy=True)

```

## EDA Before Data Splitting

```
df.head()
```

	cap_shape	cap_surface	cap_color	bruises	odor	gill_attachment	gill_spacing	gill_size	gill_color
0	x	s	n	t	p	f	c	n	k
1	x	s	y	t	a	f	c	b	k
2	b	s	w	t	l	f	c	b	n
3	x	y	w	t	p	f	c	n	n
4	x	s	g	f	n	f	w	b	k

```
# Basic information
df.info()
```

```
<class 'pandas.core.frame.DataFrame'>
RangeIndex: 8124 entries, 0 to 8123
Data columns (total 22 columns):
#   Column                                Non-Null Count  Dtype
---  -
0   cap_shape                             8124 non-null   object
1   cap_surface                           8124 non-null   object
2   cap_color                             8124 non-null   object
3   bruises                               8124 non-null   object
4   odor                                  8124 non-null   object
5   gill_attachment                       8124 non-null   object
6   gill_spacing                          8124 non-null   object
7   gill_size                             8124 non-null   object
8   gill_color                            8124 non-null   object
9   stalk_shape                           8124 non-null   object
10  stalk_surface_above_ring              8124 non-null   object
11  stalk_surface_below_ring              8124 non-null   object
12  stalk_color_above_ring                8124 non-null   object
13  stalk_color_below_ring                8124 non-null   object
14  veil_type                             8124 non-null   object
15  veil_color                            8124 non-null   object
16  ring_number                           8124 non-null   object
17  ring_type                             8124 non-null   object
18  spore_print_color                     8124 non-null   object
19  population                            8124 non-null   object
20  habitat                               8124 non-null   object
21  is_poisonous                          8124 non-null   int64
dtypes: int64(1), object(21)
memory usage: 1.4+ MB
```

```
# Data dimensions
df.shape
```

```
(8124, 22)
```

## Data Splitting

We are trying to classify whether a mushroom is edible (0) or poisonous (1). Going to do a 70/30 train/test split due to the large and roughly equal proportions of edible and poisonous mushrooms.

A random state of 123 will ensure reproducibility when ran by subsequent users.

```
# Train-test split at the row level
#   - Keeps all columns together in each split
#   - Stratify on the target to preserve class balance

train_df, test_df = train_test_split(
    df,
    test_size=0.3,
    random_state=123,
    stratify=df["is_poisonous"],
)

print(
    "Train df shape: ", train_df.shape,
    "\nTest df shape: ", test_df.shape
)
```

```
Train df shape: (5686, 22)
Test df shape: (2438, 22)
```

```
# Build X/y for model training and evaluation from splits
X_train = train_df.drop(columns=["is_poisonous"])
y_train = train_df["is_poisonous"]

X_test = test_df.drop(columns=["is_poisonous"])
y_test = test_df["is_poisonous"]
```

## EDA After Data Splitting

```
# Sanity check: class balance in the training data

train_target_counts = train_df["is_poisonous"].value_counts()
```

```

train_target_prop = train_df["is_poisonous"].value_counts(normalize=True)

print("Training set class counts:")
print(train_target_counts.rename(index={0: "edible (0)", 1: "poisonous (1)"}))

print("\nTraining set class proportions:")
print(train_target_prop.rename(index={0: "edible (0)", 1: "poisonous (1)"}).round(3))

```

```

Training set class counts:
is_poisonous
edible (0)      2945
poisonous (1)   2741
Name: count, dtype: int64

```

```

Training set class proportions:
is_poisonous
edible (0)      0.518
poisonous (1)   0.482
Name: proportion, dtype: float64

```

```

# Data Validation: check training target distribution roughly matches UCI proportions

```

```

expected_dist = {"e": 0.518, "p": 0.482}
# but now we're working with numeric target: 0 = edible, 1 = poisonous
expected_poisonous = expected_dist["p"] # 0.482

train_poisonous = train_df["is_poisonous"].mean()
tol = 0.02

assert abs(train_poisonous - expected_poisonous) <= tol, (
    f"Training set poisonous proportion ({train_poisonous:.3f}) "
    f"differs from expected ({expected_poisonous:.3f}) by more than {tol:.2f}."
)

print(
    f"[OK] Training set poisonous proportion ({train_poisonous:.3f}) "
    f"is close to expected ({expected_poisonous:.3f})."
)

```

```

[OK] Training set poisonous proportion (0.482) is close to expected (0.482).

```

```

# Feature column names
feature_cols = X_train.columns

# Key mushroom feature summary
summary_rows = []
for col in feature_cols:
    if not col == 'is_poisonous':
        vc = X_train[col].value_counts()
        summary_rows.append({
            "feature": col,
            "n_categories": vc.shape[0],
            "most_frequent_category": vc.index[0],
            "most_frequent_count": int(vc.iloc[0])
        })

feature_summary = (
    pd.DataFrame(summary_rows)
    .sort_values("n_categories", ascending=False)
    .reset_index(drop=True)
)

feature_summary.index = feature_summary.index + 1
feature_summary

```

Table 3: Summary of mushroom feature characteristics

	feature	n_categories	most_frequent_category	most_frequent_count
1	gill_color	12	b	1202
2	cap_color	10	n	1567
3	odor	9	n	2439
4	stalk_color_above_ring	9	w	3131
5	spore_print_color	9	w	1655
6	stalk_color_below_ring	9	w	3071
7	habitat	7	d	2195
8	cap_shape	6	x	2534
9	population	6	v	2813
10	ring_type	5	p	2778
11	cap_surface	4	y	2291
12	stalk_surface_below_ring	4	s	3434
13	stalk_surface_above_ring	4	s	3630
14	veil_color	4	w	5542

Table 3: Summary of mushroom feature characteristics

	feature	n_categories	most_frequent_category	most_frequent_count
15	ring_number	3	o	5246
16	bruises	2	f	3330
17	gill_size	2	b	3939
18	stalk_shape	2	t	3211
19	gill_attachment	2	f	5534
20	gill_spacing	2	c	4766
21	veil_type	1	p	5686

We summarize all 21 categorical predictors in Table Table 3, which reports the number of levels and the dominant category for each feature. Given that `veil_type` only has one level, the values are the same for all observations so we drop that column.

```
# Drop veil_type from the feature matrices used for modeling. If check added to prevent error
if "veil_type" in X_train.columns:
    X_train = X_train.drop(columns=["veil_type"])
    X_test = X_test.drop(columns=["veil_type"])

# Redefine feature columns
feature_cols = X_train.columns
```

`veil_type` was dropped because it contains only a single category for all observations. A feature with no variation cannot help the model distinguish edible from poisonous mushrooms, provides no predictive signal, and only adds redundant columns after one-hot encoding. Therefore, it is removed to simplify the feature set and prevent unnecessary model complexity.

```
def get_poison_rate_by(feature: str) -> pd.DataFrame:
    """
    Compute the proportion of poisonous and edible mushrooms for each category
    of a given feature.

    Parameters
    -----
    feature : str
        The name of the categorical column in `df` for which the poison rates
        should be calculated.

    Returns
    -----
```

```

pandas.DataFrame
    A DataFrame indexed by the feature's categories with two columns:
    `edible_frac` and `poisonous_frac`, sorted in descending order of
    `poisonous_frac`.
    """
    category_poisonous = pd.crosstab(train_df[feature], train_df['is_poisonous'], normalize='i

# By construction, column 0 = edible, 1 = poisonous
category_poisonous.columns = ['edible_frac', 'poisonous_frac']
return category_poisonous.sort_values('poisonous_frac', ascending=False)

# Feature category maps
odor_map = {
    "a": "almond",
    "c": "creosote",
    "f": "foul",
    "l": "anise",
    "m": "musty",
    "n": "none",
    "p": "pungent",
    "s": "spicy",
    "y": "fishy",
}

habitat_map = {
    "g": "grasses",
    "l": "leaves",
    "m": "meadows",
    "p": "paths",
    "u": "urban",
    "w": "waste",
    "d": "woods",
}

gill_size_map = {
    "b": "broad",
    "n": "narrow",
}

bruises_map = {
    "t": "bruises",
    "f": "no bruises",
}

```

```

}

population_map = {
    "a": "abundant",
    "c": "clustered",
    "n": "numerous",
    "s": "scattered",
    "v": "several",
    "y": "solitary",
}

ring_type_map = {
    "c": "cobwebby",
    "e": "evanescent",
    "f": "flaring",
    "l": "large",
    "n": "none",
    "p": "pendant",
    "s": "sheathing",
    "z": "zone",
}

```

```

def stacked_poison_chart(
    feature: str,
    feature_label: str | None = None,
    category_labels: dict[str, str] | None = None,
):
    """
    Create a stacked bar chart showing edible vs poisonous fractions
    for each category of a given feature.

    Parameters
    -----
    feature : str
        Column name in df (e.g., "odor", "gill_size", "habitat").
    feature_label : str, optional
        Pretty label for axes / title. If None, derived from the column name.
    category_labels : dict[str, str], optional
        Optional mapping from raw category codes to human-readable labels.
        For example, for odor: {"a": "almond", "n": "none", ...}.
        If provided, these labels will be used on the y-axis and in tooltips.
    """

```

Returns

-----

alt.Chart

Altair chart object with stacked bars.

"""

# Use a nicer label if not provided

if feature\_label is None:

feature\_label = feature.replace("\_", " ").title()

# Table of edible / poisonous fractions by category

ct = get\_poison\_rate\_by(feature).reset\_index()

# If a category label map is provided, add a display column

if category\_labels is not None:

display\_col = f"{feature}\_label"

ct[display\_col] = ct[feature].map(category\_labels).fillna(ct[feature])

else:

display\_col = feature # use the raw feature values

# Long format for stacked bars

long\_df = ct.melt(

id\_vars=display\_col,

value\_vars=["edible\_frac", "poisonous\_frac"],

var\_name="class",

value\_name="fraction",

)

# Human-readable class labels for legend

long\_df["class\_label"] = long\_df["class"].map(

{"edible\_frac": "Edible", "poisonous\_frac": "Poisonous"}

)

# Build stacked bar chart

chart = (

alt.Chart(long\_df)

.mark\_bar()

.encode(

y=alt.Y(f"{display\_col}:N", title=feature\_label, sort="-x"),

x=alt.X("fraction:Q", title="Fraction", axis=alt.Axis(format=".2f")),

color=alt.Color(

"class\_label:N",

title="Mushroom class",

```

        scale=alt.Scale(
            domain=["Edible", "Poisonous"],
            range=["#17BECF", "#7E1E9C"],
        ),
        legend=alt.Legend(title="Mushroom class"),
    ),
    tooltip=[
        alt.Tooltip(f"{display_col}:N", title=feature_label),
        alt.Tooltip("class_label:N", title="Class"),
        alt.Tooltip("fraction:Q", title="Fraction", format=".2f"),
    ],
)
.properties(
    width=450,
)
)
return chart

```

```

# Visualize the edible vs poisonous class proportions for each odor category
stacked_poison_chart("odor", "Odor Category", odor_map)

```

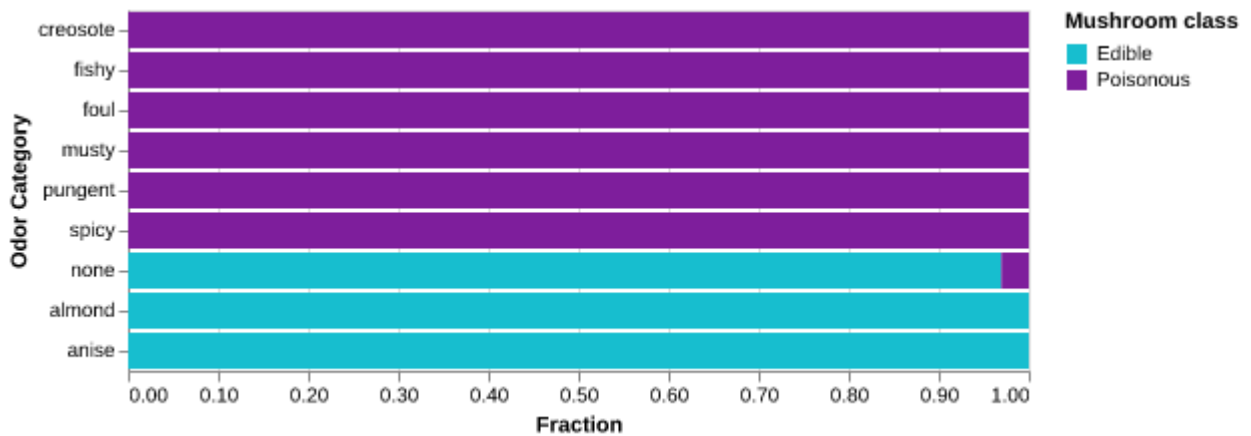


Figure 1: Class Proportions (edible vs poisonous) by Odor

```

# Visualize the edible vs poisonous class proportions for each gill size
stacked_poison_chart("gill_size", "Gill Size", gill_size_map)

```

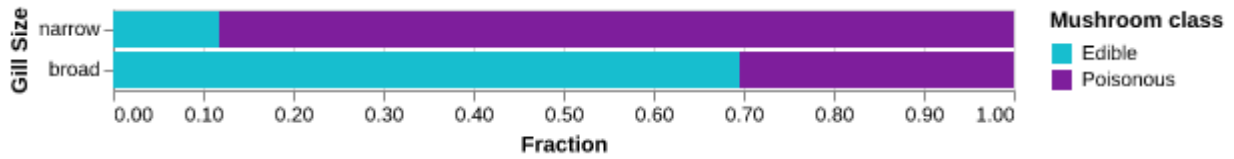


Figure 2: Class Proportions (edible vs poisonous) by Gill Size

```
# Visualize the edible vs poisonous class proportions for each habitat
stacked_poison_chart("habitat", "Habitat", habitat_map)
```

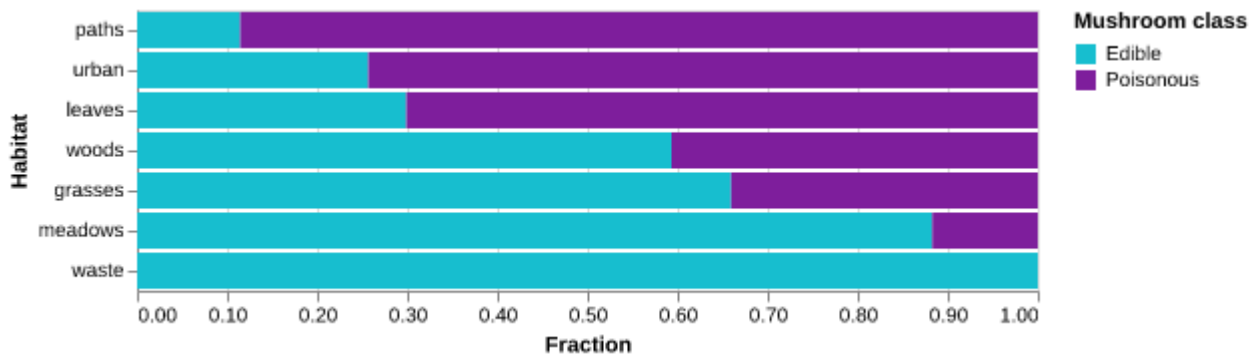


Figure 3: Class Proportions (edible vs poisonous) by Habitat

```
# Visualize the edible vs poisonous class proportions for each bruise category
stacked_poison_chart("bruises", "Bruises", bruises_map)
```

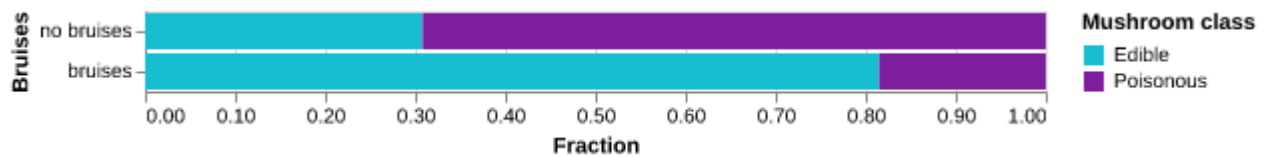


Figure 4: Class Proportions (edible vs poisonous) by Bruises

```
# Visualize the edible vs poisonous class proportions for each population type
stacked_poison_chart("population", "Population", population_map)
```

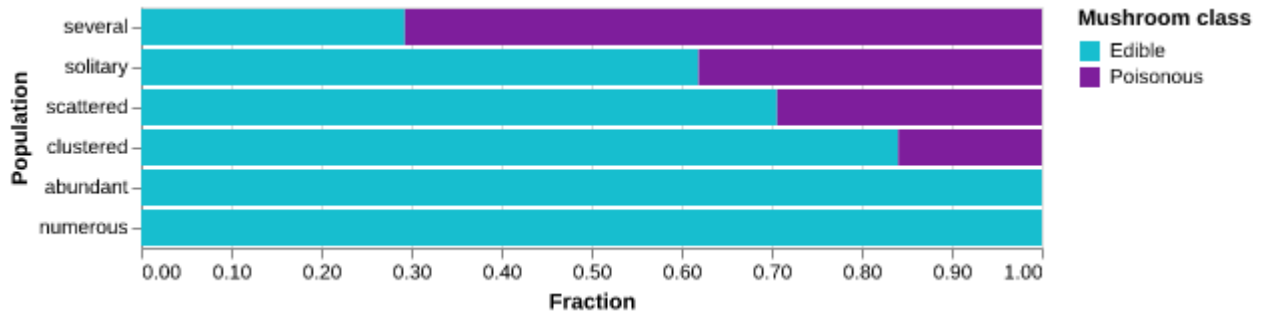


Figure 5: Class Proportions (edible vs poisonous) by Population Type

### Relationship Between Physical Features and Toxicity: An Exploratory Overview

To explore how individual mushroom characteristics relate to toxicity, we compared the proportion of poisonous and edible mushrooms across several categorical features. Odor showed the strongest separation between classes (Figure Figure 1), with foul-, creosote-, and pungent-smelling mushrooms being almost exclusively poisonous, whereas almond and anise odors were strongly associated with edibility. Gill size also demonstrated a meaningful distinction (Figure Figure 2), with broad gills far more common among edible mushrooms. Habitat exhibited notable patterns as well (Figure Figure 3): mushrooms found in wooded or grassy environments were more frequently poisonous, while those growing in urban or path environments tended to be edible. Bruising was another strong indicator (Figure Figure 4), as mushrooms that bruised were overwhelmingly edible, whereas those that did not bruise were frequently poisonous. Lastly, population type revealed clear group-level differences (Figure Figure 5), with solitary or scattered mushrooms more often poisonous, while abundant or clustered populations were more commonly edible. Together, these figures illustrate that several features, odor in particular, provide strong discriminatory signals relevant to the classification task.

```
# Compute the variance in poisonous fraction across the categories of each feature
# Show strongly the feature predicts toxicity

feature_scores = []

for col in feature_cols:
    ct = get_poison_rate_by(col)
    score = ct['poisonous_frac'].var()
    feature_scores.append({'feature': col, 'poison_variance': round(score, 2)})

feature_importance_eda = pd.DataFrame(feature_scores).sort_values(
    'poison_variance', ascending=False
).reset_index(drop=True)
```

```
feature_importance_eda.index = feature_importance_eda.index + 1
feature_importance_eda
```

Table 4: Ranking of features by how well they distinguish poisonous mushrooms

	feature	poison_variance
1	odor	0.25
2	veil_color	0.23
3	stalk_color_above_ring	0.22
4	stalk_color_below_ring	0.21
5	ring_type	0.21
6	ring_number	0.20
7	spore_print_color	0.20
8	gill_size	0.17
9	gill_color	0.14
10	cap_shape	0.14
11	bruises	0.13
12	gill_spacing	0.11
13	habitat	0.11
14	stalk_surface_below_ring	0.11
15	stalk_surface_above_ring	0.11
16	cap_surface	0.08
17	gill_attachment	0.08
18	population	0.07
19	cap_color	0.06
20	stalk_shape	0.00

A ranking of features by their discriminatory power is shown in Table Table 4, using the variance in poisonous fraction across categories as an association measure.

### Data Validation Check: Any Anomalous Correlation Between Features and/or Target?

```
# create list of all pairwise feature and feature-target combinations
train_df_cols = list(train_df.columns)
pairs = list(itertools.combinations(train_df_cols, 2))

# compute correlation for all feature pairs and feature-target pairs
```

```

def cramers_v(df, feature1, feature2):
    """ manually computes correlation for feature pairs from the pearsons chi-squared test s

    table = pd.crosstab(df[feature1], df[feature2])

    #get chi squared test statistic
    chi2, _, _, _ = chi2_contingency(table)

    #normalize chi-squared test statistic
    n = table.sum().sum()
    phi2 = chi2 / n
    r, k = table.shape

    # compute cramers v
    return np.sqrt(phi2 / min(k-1, r-1))

# create empty df with diagonal equal to 1
cramers_matrix = pd.DataFrame(np.eye(len(train_df_cols)), columns=train_df_cols, index=train_df_cols)

for feature1, feature2 in pairs:
    v = cramers_v(train_df, feature1, feature2)
    crammers_matrix.loc[feature1, feature2] = v
    crammers_matrix.loc[feature2, feature1] = v

# convert matrix to long format for Altair visualization
cramers_long = crammers_matrix.reset_index().melt(id_vars='index')
cramers_long.columns = ['feature_1', 'feature_2', 'cramers_v']

correlation_heatmap = alt.Chart(cramers_long).mark_rect().encode(
    x=alt.X('feature_1:N',
            sort=feature_cols,
            title='Feature 1'),
    y=alt.Y('feature_2:N',
            sort=feature_cols,
            title="Feature 2"),
    color=alt.Color('cramers_v:Q',
                    scale=alt.Scale(domain=[0,1], scheme='purpleorange'),
                    title="Cramer's V"
                    )
).properties(
    width=300,
    height=300
)

```

```
correlation_heatmap
```

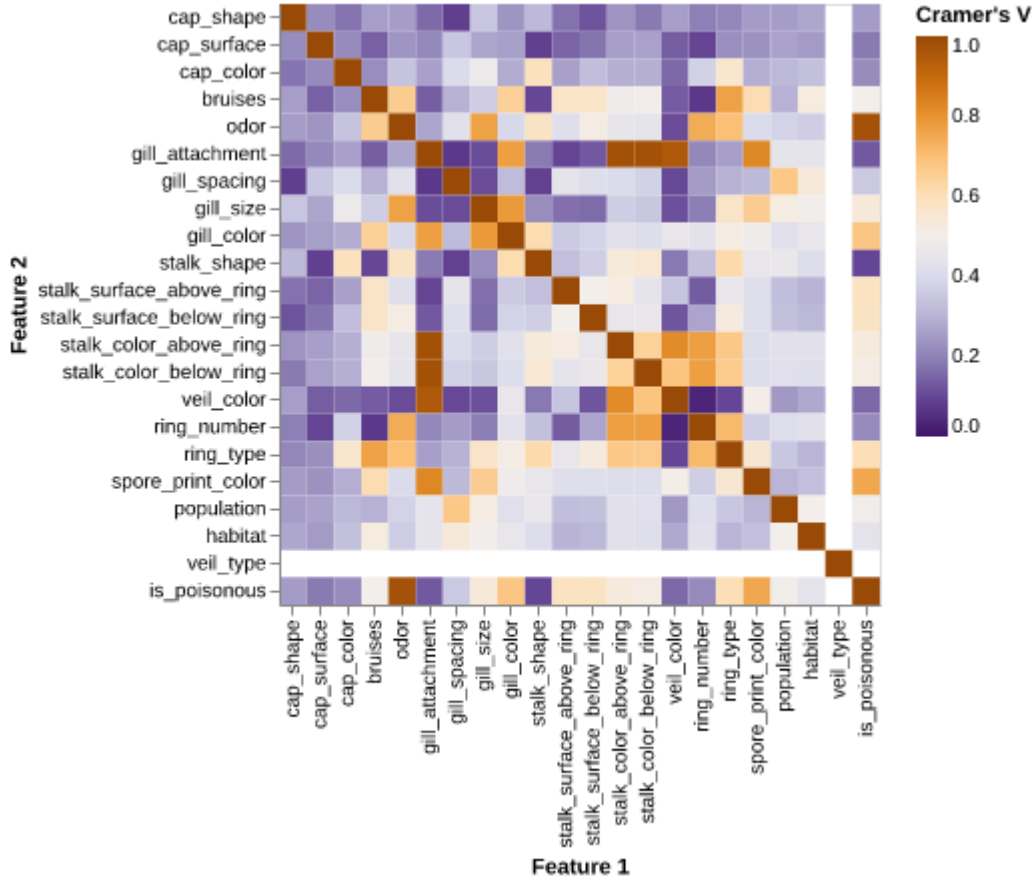


Figure 6: Correlation heatmap (Cramér's V) for all feature and feature-target pairs in the training data.

### Summary of Anomalous Correlation Between Features:

Figure 6 shows a high correlation between the feature `gill_attachment` and the three features `stalk_color_above_ring`, `stalk_color_below_ring`, and `veil_color`. I expect this is a feature of the dataset because it includes a high number of observations (5686 in our training data) and only 23 species, so there are many repeated instances of each species. If a particular species has fixed color and gill attachment characteristics than it is expected that we would find a high correlation between these features. It is not likely to be a sign of data leakage,

but does suggest that there are weaknesses in the biological representativeness of the dataset. Therefore, the dataset may not be the strongest choice for generalizing predictions about the poisonousness of mushrooms across the full Agaricus and Lepiota (Agaricaceae) family, since it may not capture the true diversity and independence of these features found in nature.

## Summary of Anomalous Correlation Between Feature “Odor” and Target:

Figure 6 shows a high correlation between the feature “odor” and the target, and a moderately high correlation between “spore\_print\_color” and the target. However, the dataset we are using includes background information on the logical rules discovered by prior researchers Duch, Adamczak, and Grabczewski (1996), and they found that “odor” is often the first split for identifying poisonous mushrooms, and “spore\_print\_color” is often the second. Hence both of these features are expected to be strong predictors of the target and the seeming anomalous correlation is simply an accurate reflection of biological reality as captured by the dataset, and not likely to be a sign of data leakage in this case.

## Model

Before we continue, we must preprocess our data such that the features work with our models. Since all of our features are strings, we can opt to use one-hot encoding for them all.

```
# Preprocessor
preprocessor = OneHotEncoder(handle_unknown="ignore")
```

We fit a `DummyClassifier` to our data as a baseline to compare our actual model with. Random state used again for reproducibility.

```
dc_pipe = make_pipeline(
    preprocessor,
    DummyClassifier(random_state=123)
)

dc_pipe.fit(X_train, y_train)

cross_val_dc = pd.DataFrame(
    cross_validate(
        dc_pipe, X_train, y_train, cv=10, n_jobs=-1, return_train_score=True
    ).mean().to_frame().rename(columns={0: "mean_value"})
)

cross_val_dc
```

Table 5: Mean 10-Fold Cross-Validation Metrics for DummyClassifier Baseline

	mean_value
fit_time	0.015795
score_time	0.004075
test_score	0.517939
train_score	0.517939

In our classification problem, we opt to use **SVC** (Buitinck, L., Louppe, G., Blondel, M., Pedregosa, Fabian, Mueller, A., Grisel, O., ... Ga"el Varoquaux. (2013)) as our model of choice for its effective usage in this scenario. We do not have an extreme amount of data (< 10000 observations) on top of only being a binary classification task.

```
svc_pipe = make_pipeline(
    preprocessor,
    SVC(random_state=123)
)

svc_pipe.fit(X_train, y_train)

cross_val_svc = pd.DataFrame(
    cross_validate(
        svc_pipe, X_train, y_train, cv=10, n_jobs=-1, return_train_score=True
    ).mean().to_frame().rename(columns={0: "mean_value"})
)

cross_val_svc
```

Table 6: Mean 10-Fold Cross-Validation Metrics for SVC Model

	mean_value
fit_time	0.545692
score_time	0.042646
test_score	1.000000
train_score	1.000000

```
y_pred = svc_pipe.predict(X_test)

cm = confusion_matrix(y_test, y_pred)
ConfusionMatrixDisplay(cm).plot()
```

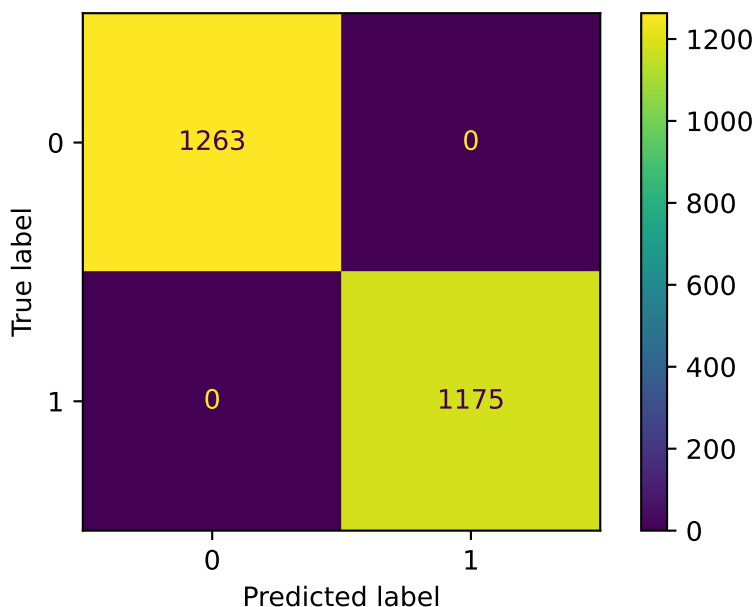


Figure 7: Confusion matrix for the SVC classifier on the test set.

## Results & Discussion

Our prediction model performed excellently on test data, classifying all of the mushroom examples correctly as poisonous or edible, with no error. Given that our model predicted perfectly, no optimization was required. As shown in Figure Figure 7, the SVC correctly classified every mushroom in the test set, yielding no false positives or false negatives. Furthermore, the SVC model shows consistent performance across folds, as summarized in Table Table 6.

This performance is identical to the results published on the [UCI website](#) from which the data was sourced (Mushroom). As shown in the confusion matrix, the model has a recall and precision of 1, due to no false positivies or negatives. The model also shows robustness, with no difference in the accuracy of training and test scores, so it is promising that this model could be deployed effectively to protect mushroom consumers from eating poisonous mushrooms. That said, there are some practical concerns for public implementation, namely the model is fit on 20 different features that correspond to specific categorical physical characteristics of a mushroom, and thus accuracy requires accurate identification of physical characteristics.

Hence, if an amateur mushroom enthusiast aimed to utilize the model to predict edibility but misidentified the physical characeristics of the mushroom they were attempting to classify, then the accuracy of the models predictions would be effectively moot as it would not be returning a prediction for the true foraged mushroom. One might consider creating a similar predictor with a smaller number of features to increase the practical efficacy of the model, though they

would need to consider balancing the models accuracy with efficacy, because the risk of falsely predicting a mushroom as edible could be fatal. It is also worth noting that the hypothetical mushroom observations in this model are based upon a book from 1981 (Lincoff), so it would be wise to consider whether the data could be updated to be sourced from a more recent source of mushroom science.

## References

- Buitinck, L., Louppe, G., Blondel, M., Pedregosa, Fabian, Mueller, A., Grisel, O., ... Ga"el Varoquaux. (2013). API design for machine learning software: experiences from the scikit-learn project. In ECML PKDD Workshop: Languages for Data Mining and Machine Learning (pp. 108–122).
- Diaz, J. H. (2016). Mistaken mushroom poisonings. *Wilderness & Environmental Medicine*, 27(2), 330-335.
- Duch, W., Adamczak, R., & Grabczewski, K. (1996). Extraction of logical rules from training data using backpropagation networks. In *Proceedings of the 1st Online Workshop on Soft Computing*, 19-30 August 1996, pp. 25-30. [Online]. Available: <http://www.bioele.nuee.nagoya-u.ac.jp/wsc1/>
- Lincoff, G. H. (1981). *The Audubon Society Field Guide to North American Mushrooms* (1981). New York: Alfred A. Knopf
- Mushroom. (1981). UCI Machine Learning Repository. <https://doi.org/10.24432/C5959T>