# easyMF User Mannual

(version 1.0)

easyMF is a user-friendly web platform that aims to facilitate biological discovery from large-scale transcriptome data through matrix factorization (MF). It offers several functional tools for gene expression matrix generation, expression matrix factorization, and metagene-based exploratory analysis including sample clustering, signature gene identification, functional gene discovery, subtype cell detection, and pathway activity inference.

- easyMF project is hosted on <a href="https://github.com/cma2015/easyMF">https://github.com/cma2015/easyMF</a>.
- easyMF docker image is available in <a href="https://hub.docker.com/r/malab/easymf">https://hub.docker.com/r/malab/easymf</a>.
- easyMF demo server can be accessed via <a href="http://easymf.omicstudio.cloud">http://easymf.omicstudio.cloud</a>.
- The following part shows installation of easyMF docker image and detailed documentation for each function in easyMF.

# 0. Metagene-based Deep Mining Using PM

Pattern matrix (AM), a matrix with samples in rows and metagenes in columns, describes sample-level relationships. In current version of easyMF, users can make use of PM for sample clustering, temporal and spatial transcriptome analysis, and subtype cell detection.

This module consists of three functions: **Sample Clustering Analysis**, **Temporal-spatial Analysis**, and **Subtype Cell Detection**.

Functions/Tools	Description	Inputs	Outputs	Time (test data)	Program	References
	Cluter samples		Cluster	~ 15s	mclust	<u>Scrucca et al.,</u> <u>2016</u>
					apcluster	Bodenhofer et al., 2011
Sample Clustering	automatically based on the	Pattern matrix	information; Cluster		SSE	This study
Analysis	pattern	matrix	visualization;		fpc	<u>Hennig, 2013</u>
	matrix				vegan	<u>Dixon, 2003</u>
					gap	<u>Maechler et</u> <u>al., 2012</u>
Temporal-spatial Analysis	' ' matriy' analysis of	expression matrix; Amplitude matrix;	genes of each module; GO enrichment analysis of	~ 5 mins	In-house scripts	This study
					cogaps	Stein-O'Brien et al., 2017
			topGO	Alexa and Rahnenführer, 2009		
	Identity a cell type of unknown single-cell	Gene expression matrix;	Identification results of cell type; Single cell type	~ 30s	prcomp	This study
type of Gene Subtype Cell unknown expression Detection single-cell matrix; RNA-Seg Spec score					ica	<u>Helwig, 2015</u>
					bignmf	<u>Pan et al.,</u> 2012
	visulization		In-house scripts	This study		

# 1. Sample Clustering Analysis

In the current version, easyMF provides six optional algorithms (<u>mclust</u>, <u>apcluster</u>, SSE, <u>fpc</u>, <u>vegan</u>, and <u>gap</u>) to cluster samples using PM coefficients. The cluster result is visualized in dot plots and tables, providing a quick overview of the relationships among samples.

## **Inputs**

• **Pattern matrix**: A pattern matrix with samples in rows and metagenes in columns. Here is an example:

	Metagene 1	Metagene 2	•••	Metagene 4
Sample 1	-2.081	0.663		-0.711
Sample 2	-2.114	0.711		-0.757
Sample 4	-2.185	0.671		-0.719

• **Cluster algorithms**: easyMF provides six cluster algorithms to be cluster samples including mclust, apcluster, SSE, fpc, vegan, and gap.

**mclust**: mclust implements model-based clustering using parameterized finite Gaussian mixture models. Models are estimated by Expectation Maximization (EM) algorithm initialized through hierarchical model-based agglomerative clustering. The optimal model is then selected according to Bayesian information criterion (BIC).

**apcluster**: affinity propagation clusters data using a set of real-valued pairwise data point similarities as input. It iterates and searches for clusters maximizing an objective net similarity function.

**SSE**: SSE calculates sum of square error based on k-means algorithm, and selects the best number of clusters based on inflection point of the residuals.

**fpc**: fpc optimums average silhouette width, partitioning around medoids with estimation of cluster number.

**vegan**: vegan is a cascade k-means partitioning using a range of *k* values. It generates the optimal cluster based on Calinski-Harabasz criterion.

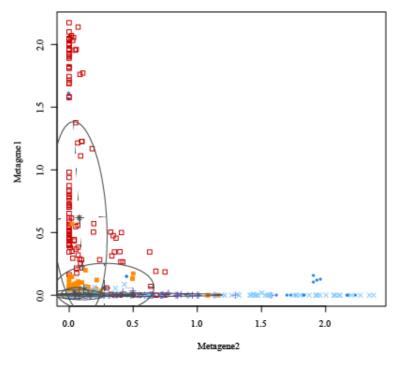
**gap**: gap calculates a goodness of clustering measure of the gap statistic for estimating the number of clusters.

NOTE: easyMF supports multi-selection for different algorithms.

### **Outputs**

• **cluster visulization**: A dot plot of the clustering results.

### mclust :6 clusters



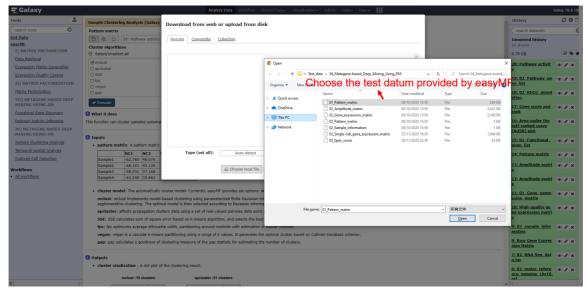
• **cluster information**: Sample cluster results for specific algorithms.

	mclust	apcluster
Sample1	1	18
Sample7	7	18
Sample11	3	21
Sample15	7	14

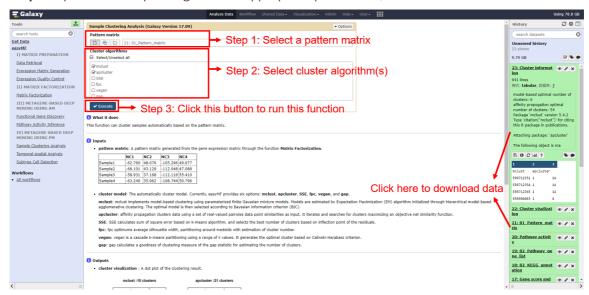
### How to use this function

- Test datum for this function is 01\_Pattern\_matrix in directory Test\_data/04\_Metagene-based\_Deep\_Mining\_Using\_PM.
- The following screenshots show us how to cluster samples using easyMF.

**Step 1**: upload test datum in directory Test\_data/01\_Matrix\_Preparation to history panel;



**Step 2**: input the corresponding files and appropriate parameters, then run the function.



# **Running time**

This step will cost ~ 15s for the test data.

# 2. Temporal-spatial Analysis

easyMF can be used to determine the extent to which genes change over time in response to perturbations (e.g., developmental time) and identify signature genes dominated at specific compartments with spatial resolution in individual tissue samples (spatial transcriptomes).

### **Inputs**

#### In **Data** section

- **Gene expression matrix**: A gene expression matrix generated by the module **Matrix Preparation**.
- **Amplitude matrix**: An amplitude matrix with genes in rows and metagenes in columns decomposed by the input gene expression matrix .
- **Pattern matrix**: A pattern matrix with samples in rows and metagenes in columns decomposed by the input gene expression matrix .
- Sample information: Sample information containing development stages or spatial compartments.

#### In Parameters section

- Threshold of Pearson Correlation Coefficient: A Pearson Correlation Coefficient value used for signature gene identification.
- **Threshold of P-value**: A *P*-value used for signature gene identification.
- **Select a species**: Species name which can be selected from drop-down menu is used to annotate gene information.

### **Outputs**

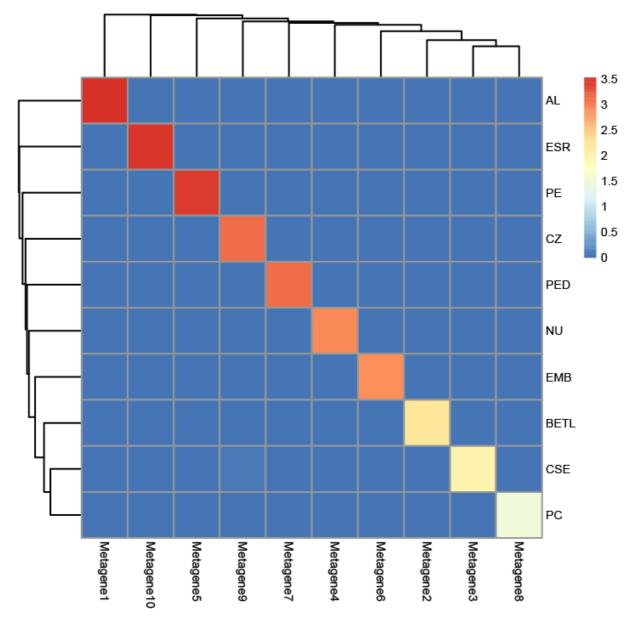
• **Signature genes of each metagene**: Summary of signature genes in each metagene. Columns represent **Metagene ID**, **Gene ID**, and **Gene description**, respectively.

Metagene ID	Gene ID	Gene description
Metagene1	Zm00001d020505	DNA glycosylase superfamily protein
Metagene2	Zm00001d012083	Thioredoxin F-type chloroplastic
Metagene3	Zm00001d033585	Leaf permease1

• **GO enrichment analysis of each metagene**: Summary of GO enrichment results of signature genes in each metagene.

Metagene ID	Туре	GO	Term	Annotated	Significant	Expected	elimFisher	Significant gene
Metagene1	BP	GO:0042445	hormone metabolic process	91	6	1.65	0.00624	Zm00001d011117;Zm00001d032223;Zm00001d039174;Zm00001d039650

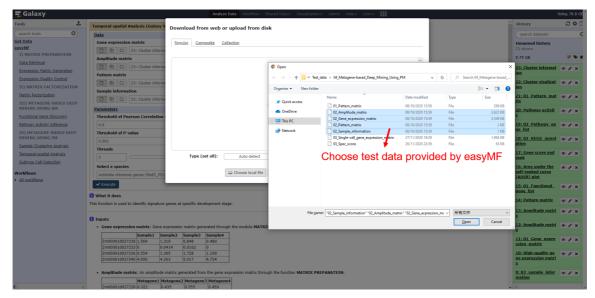
• Visualization of metagenes: Hierarchical clustering analysis of pattern matrix.



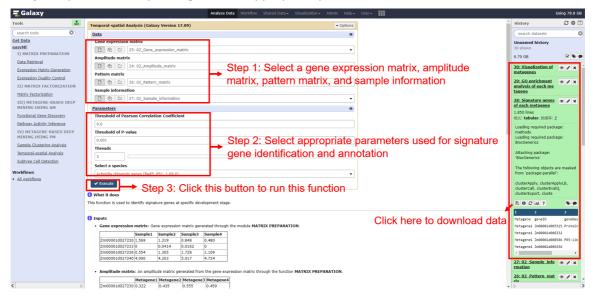
## How to use this function

- Test data for this function are in directory Test\_data/04\_Metagene-based\_Deep\_Mining\_Using\_PM including 02\_Gene\_expression\_matrix, 02\_Amplitude\_matrix, 01\_Pattern\_matrix, and 02\_Sample\_information.
- The following screenshots show us how to implement temporal-spatial transcriptome analysis using easMF.

**Step 1**: upload test data in directory <code>Test\_data/04\_Metagene-based\_Deep\_Mining\_Using\_PM</code> to history panel;



**Step 2**: input the corresponding files and appropriate parameters, then run the function.



## **Running time**

This step will cost ~ 5 mins for the test data.

# 3. Subtype Cell Detection

Single-cell RNA-Seq, which measures gene expressions at the level of a single cell, has been developed as a powerful technique to investigate the function of individual cells. easyMF can be used to identify cell types of unknown cells, which is one of key steps in the process of single-cell transcriptome analysis.

## **Inputs**

• **Gene expression matrix**: Gene count matrix of the single cell.

	AAGATGTTTTAA	CCTCACCGATAA	•••	CAATCGTTTGTG
AT1G01010	118	36		7
AT1G01020	5	2		3
AT1G01030	0	0		0
AT1G01040	1	2		1

- **Decomposition options**: Currently, easyMF provides three algorithms to decompose single-cell gene expression matrix including **PCA**, **ICA**, and **NMF**.
- **Spec score**: Spec score of the known type of tissues. Here's an example:

	Atrichoblast	Columella	Cortex	Endodermis
ORF25	0	0.127	0	0
AT2G26550	0	0	0	0
AT2G26570	0	0	0.102	0.591
PSBA	0	0	0.105	0.105

In addition, **Spec score** can also be obtained through a known tissue expression matrix.

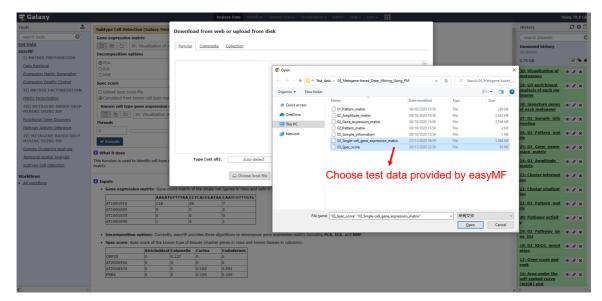
## **Outputs**

- t-SNE dimensional reduction of single cells
- Cell type detection result:

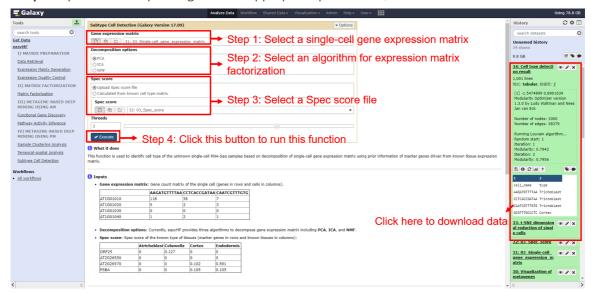
Cell_name	type
AAGATGTTTTAA	Trichoblast
CCTCACCGATAA	Trichoblast
GCGTTTGCCCTC	Cortext
AACCTGGTATTG	Atrichoblast

# How to use this function

- Test data for this function are in directory Test\_data/04\_Metagene-based\_Deep\_Mining\_Using\_PM
  including [03\_Single-cell\_gene\_expression\_matrix, and 03\_Spec\_score.
- The following screenshots show us how to detect cell type for single-cell RNA-Seq data using easMF.
   Step 1: upload test data in directory Test\_data/04\_Metagene-based\_Deep\_Mining\_Using\_PM to history panel;



**Step 2**: input the corresponding files and appropriate parameters, then run the function.



# **Running time**

This step will cost ~ 30s for the test data.