mlPEA User Manual

(version 1.0)

- mlPEA is a user-friendly, full-functionality pipeline specifically designed to enhance the processing, analysis, and interpretation of m6A-Seq in non-model plants by leveraging machine learning.
- mlPEA comprises four functional modules: **Data Preprocessing, Transcriptome Construction, m^{6}A Calling, and Functional Exploration**.
- mlPEA was powered with an advanced packaging technology, which enables compatibility and portability.
- mlPEA project is hosted on http://github.com/cma2015/mlPEA
- mlPEA docker image is available at http://hub.docker.com/r/malab/mlpea
- mlPEA server can be accessed via http://mlpea.omstudio.cloud

m^{6}A Calling Module

This module provides step-by-step functions required for epitranscriptome reads mapping and identification of RNA modifications.

Align Reads to Genome

Bowtie2 is wrapped to align epitranscriptome reads to genome.

Tools	Description	Input	Output	Time (test data)	Reference
Bowtie2	Bowtie2 is a short read aligner which achieves a combination of high speed, sensitivity and accuracy by combining the strengths of the full-text minute index with the flexibility and speed of hardware-accelerated dynamic programming algorithms, therefore bowtie2 is suitable for large genomes	Epitranscriptome sequencing reads in FASTQ format and reference genome sequences in FASTA format	Read alignments in SAM/BAM format	~5 mins	(Grabherr *et al*., 2011)

Identify RNA Modifications

Identify RNA Modifications implements three pipelines.

Tools	Description	Input	Output	Time (test data)	Reference
Peak Calling	used the SlidingWindow method with default options. To call m6A peaks, each transfrag was scanned using a 100-bp sliding window with 25-bp steps.	Input the transcripts in FASTA format;Input the ReadsCoverage file in IP sample;Input the ReadsCoverage file in input sample	The enriched peak region matrix in BED format	~5 mins	(Zhai *et al*, 2018)
Merge Biological Replicates	obtain consistent RNA modifications among three biological replicates.	Peak regions for biological Replicates	Consistent peak regions among multiple biological replicates	~10 mins	in-house scripts
High Confidence Peak	All peaks derived from the HC transcripts are designated as positive samples and utilized as input for the weakly supervised learning framework to facilitate model training.	Transcripts in FASTA format;The enriched peak region matrix in BED format	High condfidence Peak region in BED format	~20mins	(Huang *et al*, 2021; Song *et al*, 2024)

Align Reads to Genome

Input

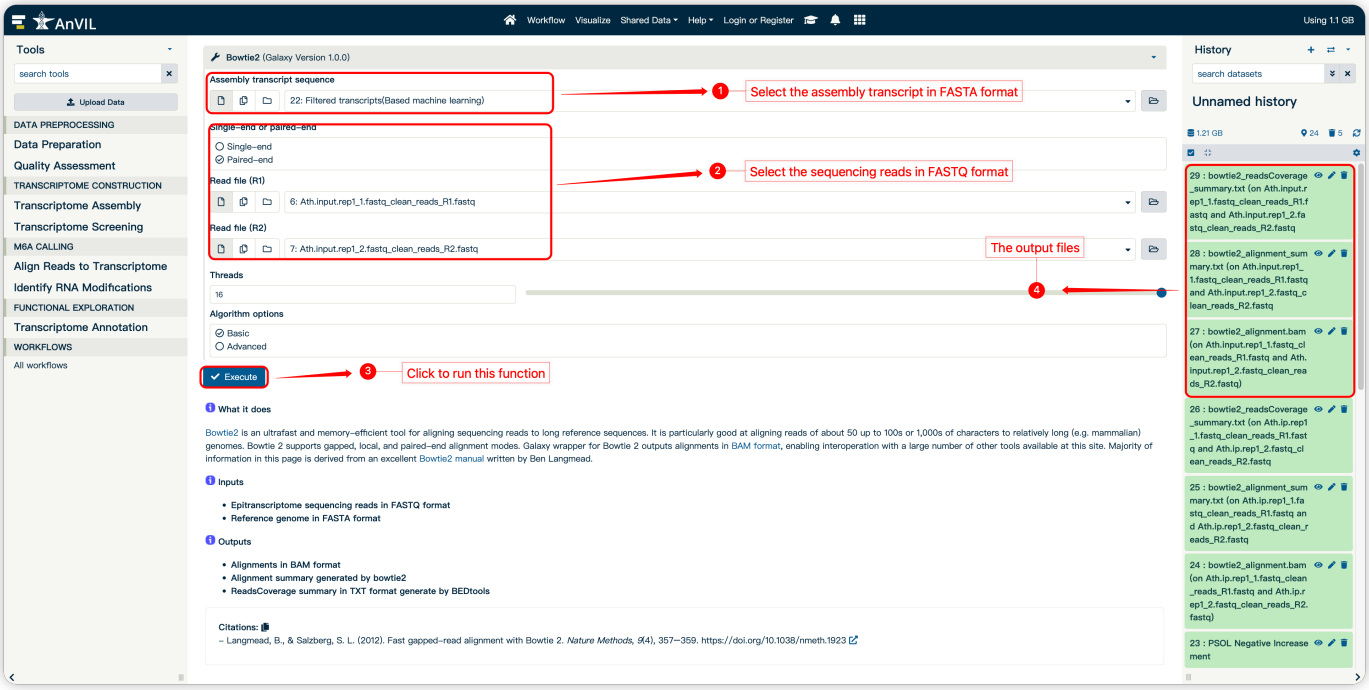
- **Epitranscriptome sequencing reads in FASTQ format**
- **Assembled transcripts in FASTA format**

Output

- **Alignments in BAM format**
- **Alignment summary generated by bowtie2**
- **ReadsCoverage summary in TXT format generate by BEDtools**

How to use this function

- The following screenshot shows us how to use this function



Peak Calling

In this function, Peak calling was performed using PEA (Zhai *et al*, 2018), which used the SlidingWindow method with default options. To call m6A peaks, each transfrag was scanned using a 100-bp sliding window with 25-bp steps.

Input

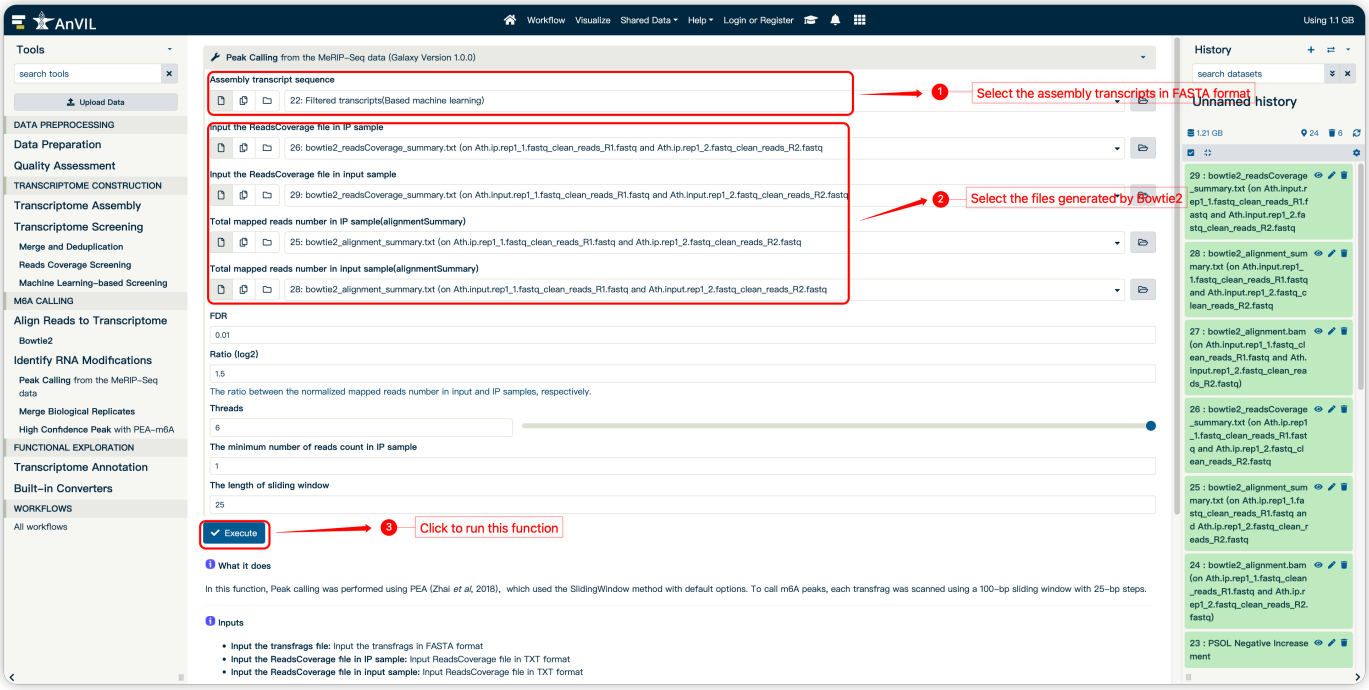
- **Input the transfrags file:** Input the transfrags in FASTA format
- **Input the ReadsCoverage file in IP sample:** Input ReadsCoverage file in TXT format
- **Input the ReadsCoverage file in input sample:** Input ReadsCoverage file in TXT format

Output

- **The enriched peak region matrix in BED format**

How to use this function

- The following screenshot shows us how to use this function



Merge Biological Replicates

In this function, mlPEA obtained consistent RNA modifications among three biological replicates.

Input

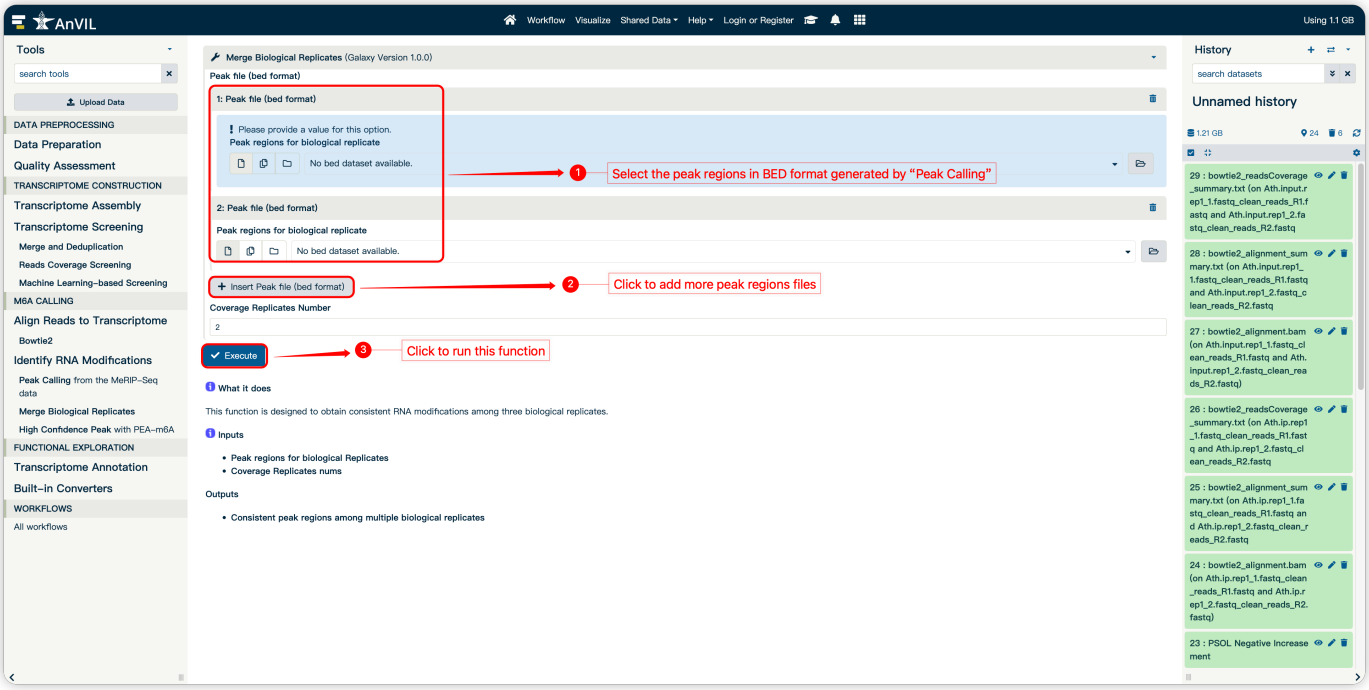
- **Peak regions for biological Replicates**

Output

- **Consistent peak regions among multiple biological replicates**

How to use this function

- The following screenshot shows us how to use this function



High Confidence Peak

All peaks derived from the HC transcripts are designated as positive samples and utilized as input for the weakly supervised learning framework to facilitate model training.

Input

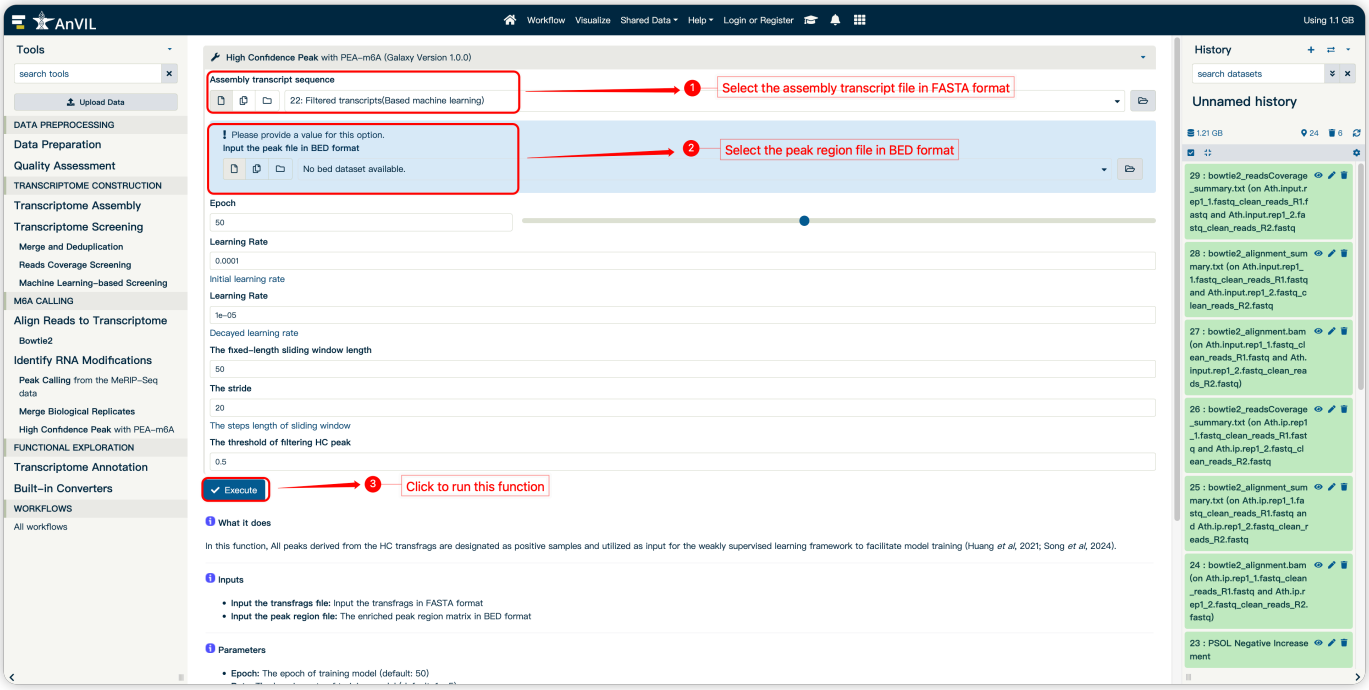
- **Input the transfrags file:** Input the transfrags in FASTA format
- **Input the peak region file:** The enriched peak region matrix in BED format

Output

- **High condfidence Peak region in BED format**

How to use this function

- The following screenshot shows us how to use this function


AnVIL
Workflow
Visualize
Shared Data
Help
Login or Register
Using 1.1 GB
Tools
search tools
Upload Data
DATA PREPROCESSING
Data Preparation
Quality Assessment
TRANSCRIPTOME CONSTRUCTION
Transcriptome Assembly
Transcriptome Screening
Merge and Deduplication
Reads Coverage Screening
Machine Learning–based Screening
M6A CALLING
Align Reads to Transcriptome
Bowtie2
Identify RNA Modifications
Peak Calling from the MeRIP–Seq data
Merge Biological Replicates
High Confidence Peak with PEA–m6A
FUNCTIONAL EXPLORATION
Transcriptome Annotation
Built–in Converters
WORKFLOWS
All workflows
High Confidence Peak with PEA–m6A (Galaxy Version 1.0.0)
Assembly transcript sequence
22: Filtered transcripts(Based machine learning)
1
Select the assembly transcript file in FASTA format
Please provide a value for this option.
Input the peak file in BED format
No bed dataset available.
2
Select the peak region file in BED format
Epoch
50
Learning Rate
0.0001
Initial learning rate
Learning Rate
1e-05
Decayed learning rate
The fixed–length sliding window length
50
The stride
20
The steps length of sliding window
The threshold of filtering HC peak
0.5
Execute
3
Click to run this function
What it does
In this function, All peaks derived from the HC transfrags are designated as positive samples and utilized as input for the weakly supervised learning framework to facilitate model training (Huang et al, 2021; Song et al, 2024).
Inputs
Input the transfrags file: Input the transfrags in FASTA format
Input the peak region file: The enriched peak region matrix in BED format
Parameters
Epoch: The epoch of training model (default: 50)
History
search datasets
Unnamed history
1.21 GB
24
6
29 : bowtie2_readsCoverage_summary.txt (on Ath.input.rep1_1.fastq_clean_reads_R1.fastq and Ath.input.rep1_2.fastq_clean_reads_R2.fastq
28 : bowtie2_alignment_summary.txt (on Ath.input.rep1_1.fastq_clean_reads_R1.fastq and Ath.input.rep1_2.fastq_clean_reads_R2.fastq
27 : bowtie2_alignment.bam (on Ath.input.rep1_1.fastq_clean_reads_R1.fastq and Ath.input.rep1_2.fastq_clean_reads_R2.fastq)
26 : bowtie2_readsCoverage_summary.txt (on Ath.ip.rep1_1.fastq_clean_reads_R1.fastq and Ath.ip.rep1_2.fastq_clean_reads_R2.fastq
25 : bowtie2_alignment_summary.txt (on Ath.ip.rep1_1.fastq_clean_reads_R1.fastq and Ath.ip.rep1_2.fastq_clean_reads_R2.fastq
24 : bowtie2_alignment.bam (on Ath.ip.rep1_1.fastq_clean_reads_R1.fastq and Ath.ip.rep1_2.fastq_clean_reads_R2.fastq)
23 : PSOL Negative Increasement