3000788 Intro to Comp Molec Biol

Week 10: Systems biology and online resources

Fall 2024



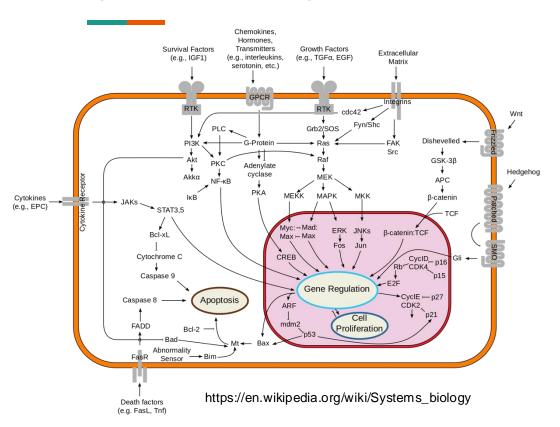
Sira Sriswasdi, PhD

- Research Affairs
- Center of Excellence in Computational Molecular Biology (CMB)
- Center for Artificial Intelligence in Medicine (CU-AIM)

Part 1: Systems biology

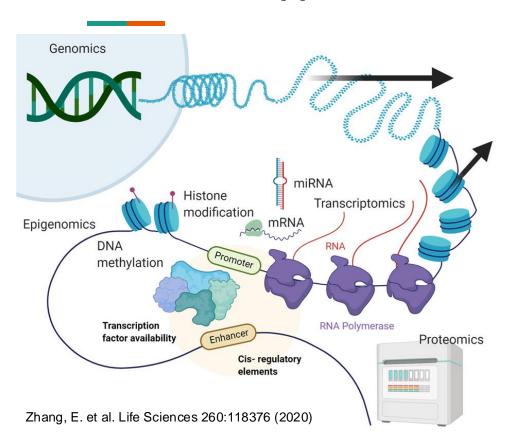
- Biological systems consists of components and interactions
- Gene expression = snapshot of gene activity
- Interaction network = mechanistic diagram
- Systems biology = understanding the dynamics of the systems
 - Multi-omics
 - Model the time-dependent dynamics

Systems biology



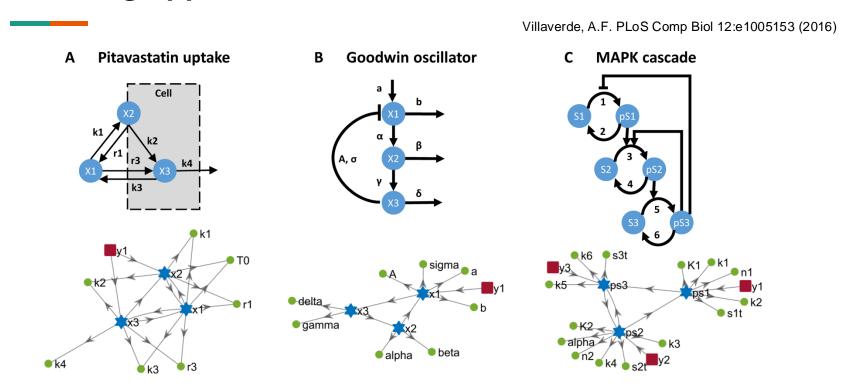
- A (biological) system consists of components (genes) and rules (gene expression regulations) that control its characteristics (phenotypes)
- Systems biology = integration of data and model to fully understand the system

Multi-omics approach



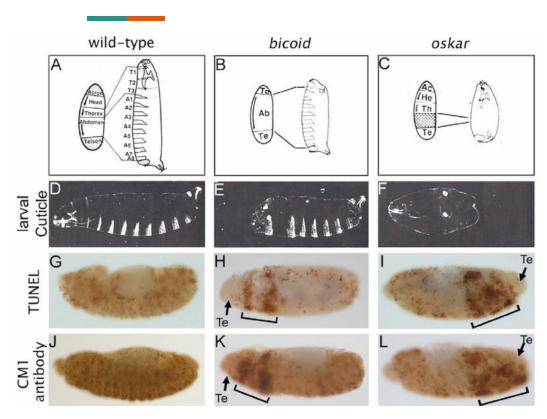
- Genomics → gene state
- Epigenomics → chromatin state
- Transcriptomics \rightarrow gene expression
- Proteomics → protein expression
- Other assays → protein function
- Provide mechanistic understanding

Modeling approach



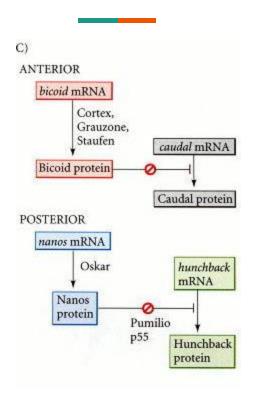
Represent molecular interaction as network of physical-chemical reactions

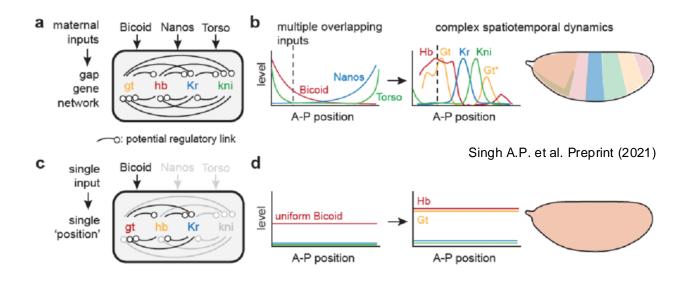
Why is systems biology interesting?



- How does gene and protein know where to activate in the body during development?
- Each cell starts in identical state!

Why is systems biology interesting?





Gene regulatory dynamics lead to specific expression pattern over time

A simple enzymatic reaction

$$E + S \underset{\mathsf{k}_{\mathsf{off}}}{\longleftrightarrow} ES \xrightarrow{\mathsf{k}_{\mathsf{react}}} E + P$$

-
$$\frac{d[S]}{dt}$$
 = Rate of change in $S = -k_{on}[E][S] + k_{off}[ES]$

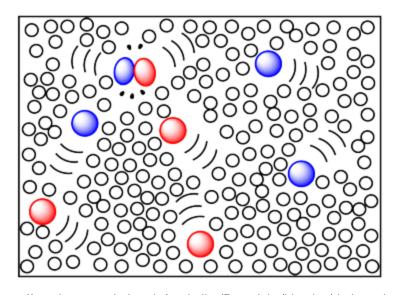
-
$$\frac{d[E]}{dt}$$
 = Rate of change in $E = -k_{on}[E][S] + k_{off}[ES] + k_{react}[ES]$

-
$$\frac{d[P]}{dt}$$
 = Rate of change in $P = \frac{k_{react}}{ES}$

A key mental image

Rate of E + S \rightarrow ES = Rate of E meeting S x Rate of binding

Rate of E + S \rightarrow ES = [E][S] x k



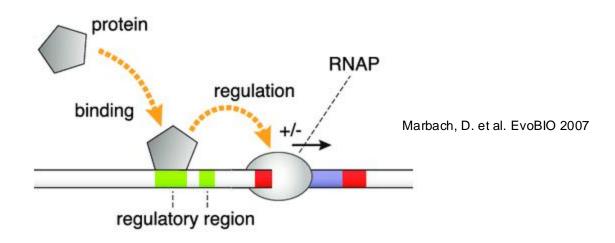
Rate of E meeting S scales linearly with [E] and [S]

Rate of binding is a constant for E and S

https://employees.csbsju.edu/cschaller/Reactivity/kinetics/rkphase.htm

- Molecules must find each other in 3D to bind and interact
- Reaction takes time (and typically energy)

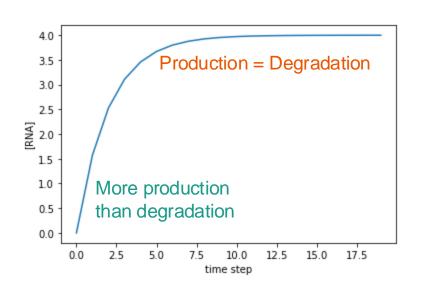
From enzymatic reaction to transcription



- TF + DNA $_{inactive} \leftrightarrow TF$ -DNA
- RNAP + TF-DNA \rightarrow RNAP + TF-DNA + RNA
- RNA \rightarrow degraded RNA

A differential equation for gene expression

 $k_{\text{transcription}}$ $DNA \rightarrow DNA + RNA$ $RNA \rightarrow \emptyset$ $k_{\text{degradation}}$



- $\frac{d[RNA]}{dt}$ = rate of change of RNA = $k_{transcription} k_{degradation}[RNA]$
- What would the graph of [RNA] looks like?

Simulating differential equation

$$- \frac{d[RNA]}{dt} = k_{transcription} - k_{degradation}[RNA]$$

- Start at an initial condition $x(0) = x_0$,
 - Calculate $x(1) = x(0) + x'(0) = x_0 + k_{transcription} k_{degradation} x_0$
 - Calculate $x(2) = x(1) + x'(1) = x(1) + k_{transcription} k_{degradation} x(1)$
 - $= x_0 + 2 \cdot k_{transcription} k_{degradation}(x_0 + x(1))$
 - [RNA] after 2 timesteps = initial [RNA] + 2 units of transcription combined degradation of [RNA] at both timesteps

A toy example

$$-\frac{d[RNA]}{dt} = 4 - 0.2 [RNA]$$

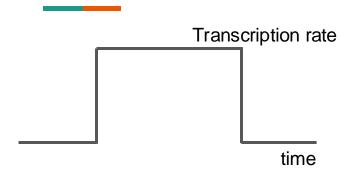
- Start at an initial condition x(0) = 100,
 - -x(1) = 100 + 4 20 = 84
 - -x(2) = 84 + 4 17 = 71
 - -x(3) = 71 + 4 14 = 61
 - x(4) = 61 + 4 12 = 53
 - -x(5) = 53 + 4 10 = 47

Another toy example

$$-\frac{d[RNA]}{dt} = 4 - 0.2 [RNA]$$

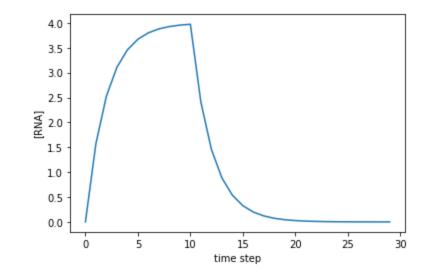
- Start at an initial condition x(0) = 0,
 - -x(1) = 0 + 4 0 = 4
 - -x(2) = 4 + 4 1 = 7
 - -x(3) = 7 + 4 1 = 10
 - -x(4) = 10 + 4 2 = 12
 - -x(5) = 12 + 4 2 = 14

Time-dependent activation

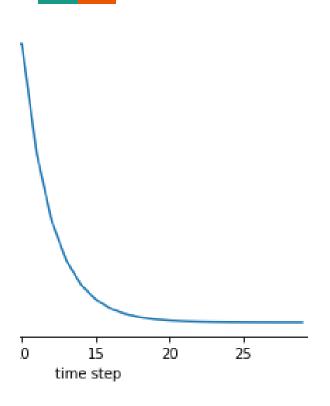


- First half of the dynamics is the same as before
- Second half is when RNA degrades until the [RNA] falls back to the basal level

```
def time_transcription(rna, time, k_trans, k_deg):
    if time < 10:
        return k_trans - k_deg * rna
    else:
        return - k_deg * rna</pre>
```



Exponential decay

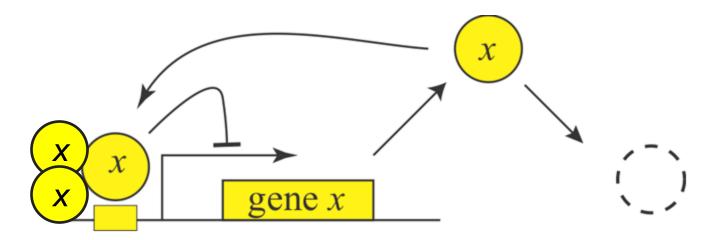


$$- \frac{d[RNA]}{dt} = -k_{\text{degradation}}[RNA]$$

- Fast decay in the beginning because there are a lot of RNA molecules
- Slower decay at the end

$$-\frac{de^{-kt}}{dt} = -ke^{-kt}$$

Negative auto-regulation



http://be150.caltech.edu/2019/handouts/03_small_circuits.html

$$- \frac{d[X]}{dt} = k_{transcription}[DNA_{active}] - k_{degradation}[X]$$

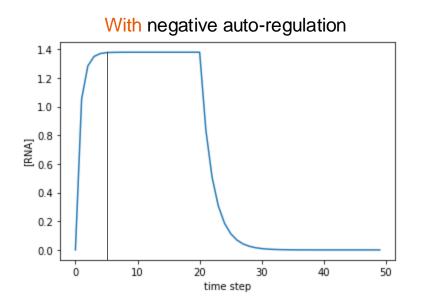
- $[DNA_{active}]$ also depends on [X] through binding dynamics
- Two variables!

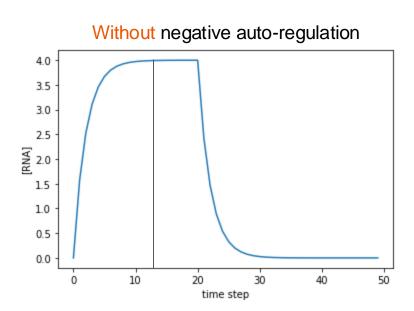
Negative auto-regulation

$$\begin{array}{c} \mathbf{k}_{\mathsf{bind}} \\ \mathbf{n}X + DNA_{active} \leftrightarrow DNA_{inactive} \\ \mathbf{k}_{\mathsf{unbind}} \end{array}$$

- $-\frac{dDNA_{inactive}}{dt} = k_{bind}[X]^{n}[DNA_{active}] k_{unbind}[DNA_{inactive}]$
- At binding equilibrium, fraction of unbound (active) DNA = $\frac{k_{bind}[X]^n}{k_{unbind} + k_{bind}[X]^n}$
- Also known as Hill function

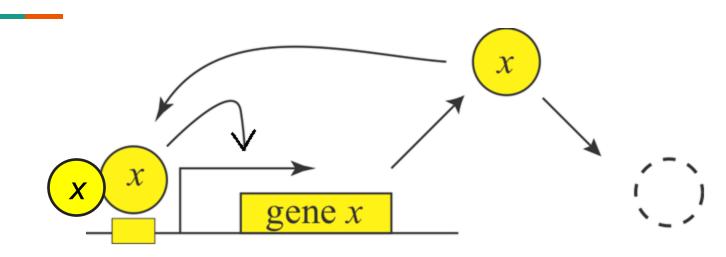
Faster response time with negative auto-regulation





$$\frac{d[X]}{dt} = \frac{k_{transcription}}{1 + (k[X])^n} - k_{degradation}[X]$$

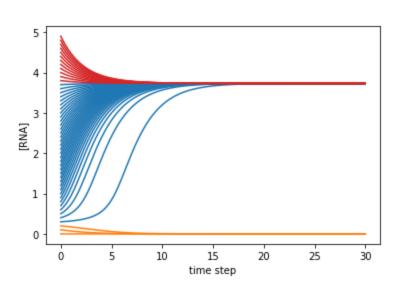
Positive auto-regulation



$$-\frac{d[X]}{dt} = \frac{k_{transcription}(k[X])^2}{1 + (k[X])^2} - k_{degradation}[X]$$

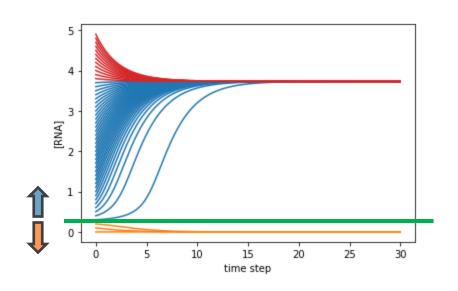
- Can you guess how this system will behave?
 - How many equilibria are there?

Bistability



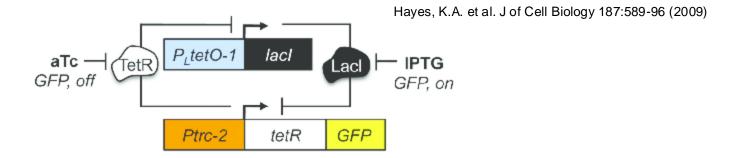
- For **low** [X], the degradation term wins over the transcription and [X] decays down to zero
- For intermediate [X], the transcription term wins over degradation and [X] increases until reaching the other equilibrium
- For high [X], the degradation term wins over the transcription and [X] decays down to the nearest equilibrium

Bistability as memory



- Given a cell in low expression state
- A drug treatment raises the expression level in the cell pass the green threshold
- After stopping the treatment, the cell will "memorize" that it is now in the high expression state

Memory from gene toggle switch



- Two genes repressing each other
- Two equilibria: High Lacl or high GFP
- If GFP is ON, how can we turn it off? Adding aTc or IPTG?
- Once GFP is OFF, can we stop supplying those molecules to the cell?

Two-gene linear system

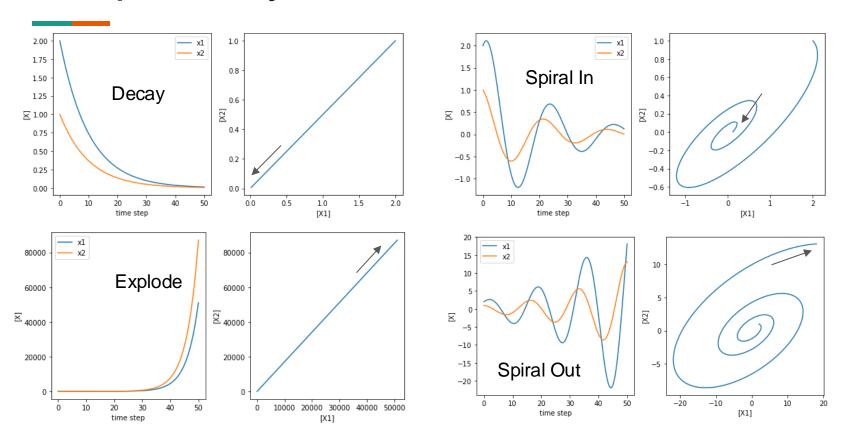
$$\frac{d[X1]}{dt} = k_{1,1}[X1] + k_{1,2}[X2]$$

$$\frac{d[X2]}{dt} = k_{2,1}[X1] + k_{2,2}[X2]$$

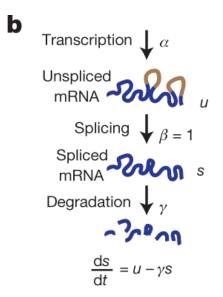
```
1  def two_loci_linear(rna, time, k11, k12, k21, k22):
2     dx1_dt = k11 * rna[0] + k12 * rna[1]
3     dx2_dt = k21 * rna[0] + k22 * rna[1]
4     return [dx1_dt, dx2_dt]
```

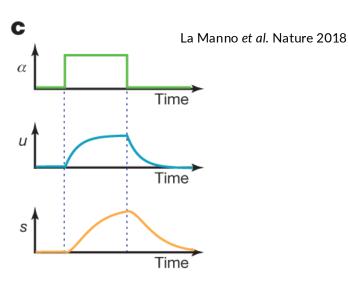
- A simple two-gene system with only linear effects
- Depending on $k_{i,i}$, interesting dynamics can be derived
 - Actually, depending on **Trace** and **Determinant** of the matrix $\{k_{i,j}\}$

Some possible dynamics



RNA velocity model





- Two-step process: From DNA to unspliced RNA to spliced RNA
- $\frac{d[U]}{dt} = \alpha \beta[U], \frac{d[S]}{dt} = \beta[U] \gamma[S]$
- Not the same dynamics as simplifying $\frac{d[S]}{dt} = \alpha \gamma[S]$

Part 2: Useful online resources

- Use public datasets to develop hypothesis
- New findings can still be discovered in published data
- Be careful of hidden confounding and design factors
- Use multiple tools to confirm

Some motivations

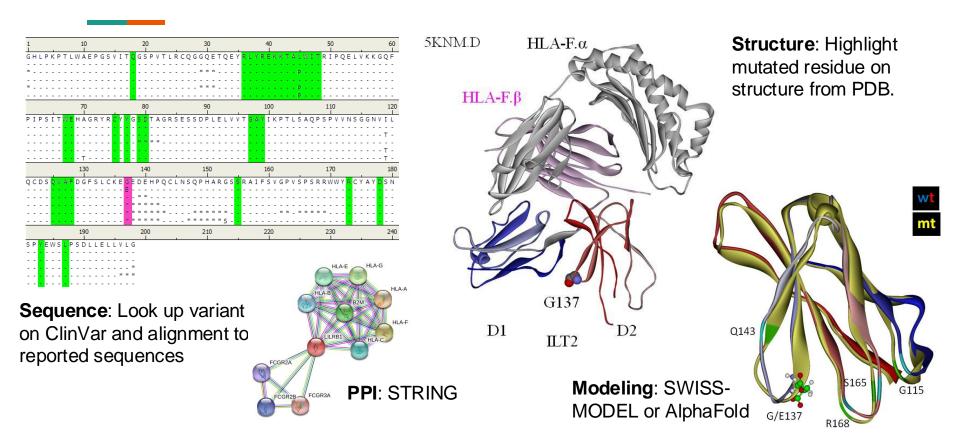
- Quick answers to quick questions
 - Impact of a newly identified mutation?
 - Where does a TF bind on genome?
- Free hypothesis generation
 - What genes are consistently up-regulated in a certain disease?



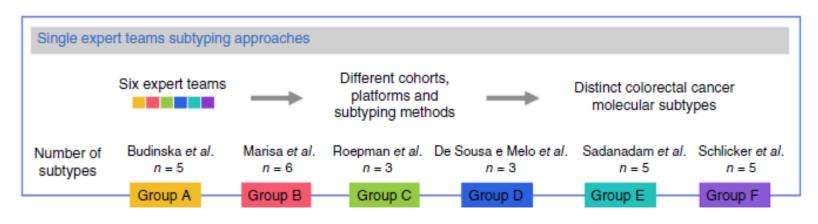
Image from boston.lti.cs.cmu.edu

- Combine data from multiple modalities
 - Omics
 - Clinical
 - Imaging

Example: G137E mutation on ILT2



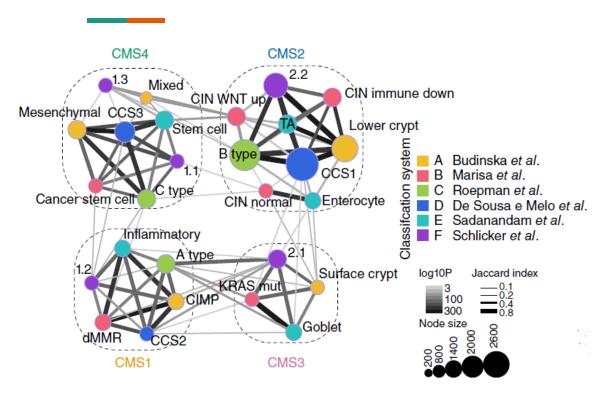
Example: Consensus cancer subtyping



Guinney et al. Nat Medicine 21: 1350-1356 (2015)

- Different clinicians characterize cancer patients differently
- But there should be a common molecular basis!

Example: Consensus cancer subtyping



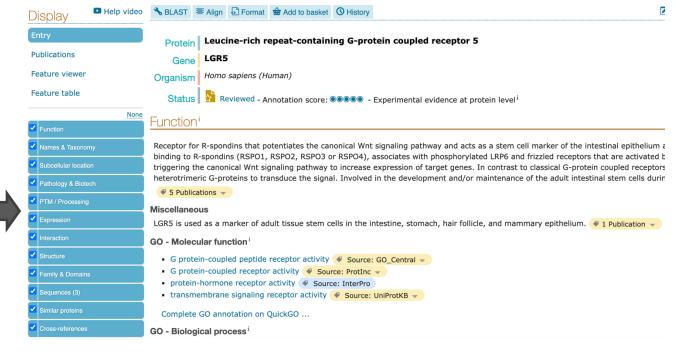
- Compare clinician's decisions on a common group of patients
- Identify 4 major groups:
 Consensus Molecular
 Subtype (CMS)

Guinney et al. Nat Medicine 21: 1350-1356 (2015)

Knowledgebase

- NCBI/GenBank
- Ensembl
- Uniprot
- GeneCard
- ENCODE
- Human Protein Atlas

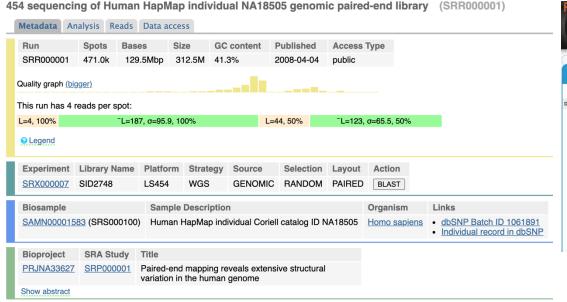
UniProtKB - 075473 (LGR5_HUMAN)



Accession IDs

ID system	Accession
Gene Symbol	LGR5
HGNC gene	4504
Entrez (NCBI)	8549
RefSeq transcript	NM_00367
Ensembl	ENSG00000139292
Uniprot protein	O75473
OMIM	606667

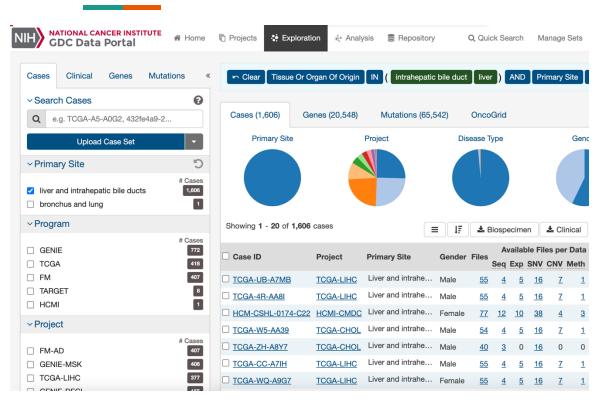
Data repository





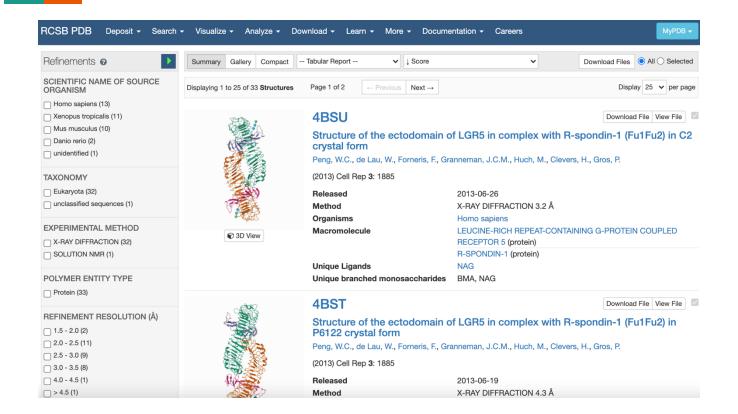
Reanalysis with a common pipeline

Genomic Data Commons



- Cancer multi-omics
 - Exome
 - RNA-seq
 - Methylation
- Clinical and demographic data
- Some with histopathological images and radiographic images

PDB: 3D structures



Gene Expression Omnibus / ArrayExpress



You query contains a term which has too many synonyms and more specific phrases in EFO. The results shown below do not include those expanded terms.

Search results for breast cancer metastasis

1 − 20 of 1,417+ results Sort by: Relevance ▼ ✓ ^

E-MTAB-4801 • 9 January 2019 • 3 links • 3 files

Variation in RNA expression in a panel of 30 breast cancer cell lines

... type comparison design EFO_0001745 cell line cell line BT20 BT474 BT549 ... 27 other values Homo sapiens female mammary gland invasive ductal carcinoma breast ductal adenocarcinoma metaplastic breast carcinoma squamous cell breast carcinoma, acantholytic variant breast adenocarcinoma breast...

E-MTAB-8807 • 4 April 2020 • 1 link • 2 files

Estrogen receptor beta inhibits cholesterol biosynthesis through overexpression of mir-181a-5p

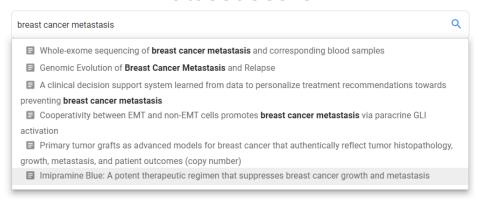
in Triple Negative Breast Cancer

... carcinoma squamous cell breast carcinoma, acantholytic variant not specified invasive breast ductal carcinoma invasive lobular carcinoma adenocarcinoma atypical carcinoma carcinoma ER beta negative ER beta positive HCC1806 null null null epithelial cell squamous cell breast carcinoma, acantholytic...

Search for omics datasets

Google Dataset / FigShare

Dataset Search



1,494,669 results found



Investigating novel mechanisms of metastasis in ...



Contribution with the first "this how contribution of both contributions of the contribution of the contri

Stratification of radiosensitive brain metastases based o...



Identification of Molecular Mediators of Endocrine ...

- Google Dataset compiles links from major public data repositories
- Figshare is a free repository that scientists often use for non-omics data

MSigDB: Curated gene sets

Human MSigDB Collections

(browse 6449 gene sets)





Gene Symbols

JSON bundle

NCBI (Entrez) Gene IDs

The 33196 gene sets in the Human Molecular Signatures Database (MSigDB) are divided into 9 major collections, and several subcollections. See the table below for a brief description of each, and the Human MSigDB Collections: Details and Acknowledgments page for more detailed descriptions. See also the MSigDB Release Notes.

Click on the "browse gene sets" links in the table below to view the gene sets in a collection. Or download the gene sets in a collection by clicking on the links below the "Download Files" headings. For a description of the GMT file format see the Data Formats in the Documentation section. The gene sets can be downloaded as NCBI (Entrez) Gene Identifiers or HUGO (HGNC) Gene Symbols. There are also JSON bundles containing the HUGO (HGNC) Gene Symbols along with some useful metadata. An XML file containing all the Human MSiqDB gene sets is available as well.

H: hallmark gene sets (browse 50 gene sets)	Hallmark gene sets summarize and represent specific well- defined biological states or processes and display coherent expression. These gene sets were generated by a computational methodology based on identifying overlaps between gene sets in	Download GMT Files Gene Symbols NCBI (Entrez) Gene IDs	
	other MSigDB collections and retaining genes that display coordinate expression. details	JSON bundle	
C1: positional gene sets (browse 299 gene sets)	Gene sets corresponding to human chromosome cytogenetic bands. details	Download GMT Files Gene Symbols NCBI (Entrez) Gene IDs	
		JSON bundle	

including online pathway databases and the biomedical literature.

Many sets are also contributed by individual domain experts. The

gene set page for each gene set lists its source. The C2 collection is divided into the following two sub-collections: Chemical and

genetic perturbations (CGP) and Canonical pathways (CP), details

- Function-based / Diseasebased / Publication-based / Genomic location-based
- For gene panel, enrichment test, etc.

miRNA target



Release 7.1: June 2016

Agarwal et al., 2015

Search for predicted microRNA targets in mammals

[Go to TargetScanMouse]
[Go to TargetScanWorm]
[Go to TargetScanFly]
[Go to TargetScanFish]

1.	. Se	lect	а	speci	es	Human	•
----	------	------	---	-------	----	-------	---

AND

2. Enter a human gene symbol (e.g. "Hmga2") LIN28A or an Ensembl gene (ENSG0000149948) or transcript (ENST0000403681) ID

AND/OR

- 3. Do one of the following:
- Select a broadly conserved* microRNA family Broadly conserved microRNA families ▼
- Select a conserved* microRNA family Conserved microRNA families ▼
- Select a poorly conserved but confidently annotated microRNA family Poorly conserved microRNA families *
- Select another miRBase annotation

Other miRBase annotations

Note that most of these families are star miRNAs or RNA fragments misannotated as miRNAs

Enter a microRNA name (e.g. "miR-9-5p")

Submit Reset



Apologies for the longer-than-usual wait. miRBase 21 is now available on the website, and all data available for download on the FTP site. As usual, the release

notes describe the major changes. Of particular note this time, the Genome Reference Consortium have released a new human genome assembly, GRCh38. We

By sam (June 26, 2014)

miRBase: the microRNA database

niRBase 21 finally arrives

Help | FAO

Comments

Citation | Policy

have therefore remapped the human [...]

	miRDB
Target Search	Choose one of the following search options:
Target Mining	Search by miRNA name
Custom Prediction	Human ▼ Go Clear
FuncMir Collection	Search by gene target
Data Download	Human ▼ Gene Symbol ▼ Go Clear
<u>Statistics</u>	miRDB is an online database for miRNA target prediction and functional annotations.

bioinformatics tool, MirTarget, which was developed by analyzing thousands of miRNA-t

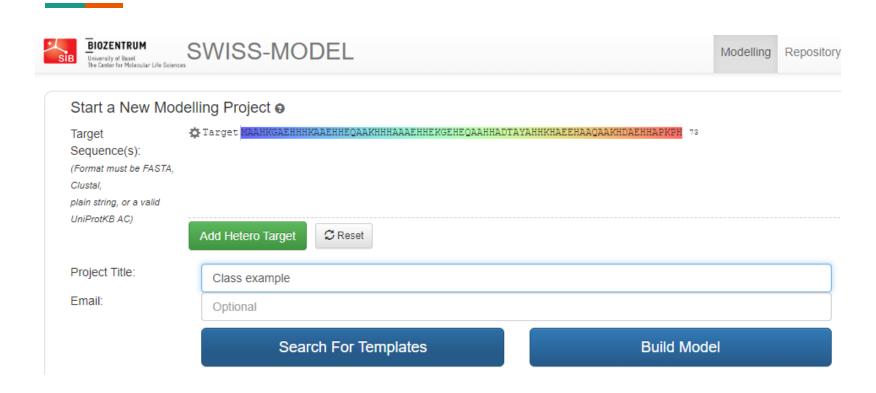
sequencing experiments. Common features associated with miRNA target binding have targets with machine learning methods, miRDB hosts predicted miRNA targets in five si

As a recent update, users may provide their own sequences for customized target pred

computational analyses and literature mining, functionally active miRNAs in humans an

well as associated functional annotations, are presented in the FuncMir Collection in mi

SWISS-MODEL: Homology modeling



AI-assisted protein folding (ColabFold)

- 3. Enter the amino acid sequence(s) to fold
 Enter the amino acid sequence(s) to fold:

 If you enter only a single sequence, the monomer model will be used.
 If you enter multiple sequences, the multimer model will be used.

 sequence_1: "MAAHKGAEHHHKAAEHHEQAAKHHHAAAEHHEKGEHEQA
 sequence_2: "Insert text here
 - https://github.com/sokrypton/ColabFold

5. Run AlphaFold and download prediction Once this cell has been executed, a zip-archive v In case you are having issues with the relaxation violations. run_relax: 🗸 Relaxation is faster with a GPU, but we have four reverting to using without GPU. relax use gpu: Show code

- Accessible, simplified Python interface

GenePattern / Galaxy



Features

Powerful genomics tools in a user-friendly interface



GenePattern provides hundreds of analytical tools for the analysis of gene expression (RNA-seq and microarray), sequence variation and copy number, proteomic, flow cytometry, and network analysis. These tools are all available through a Web interface with no programming experience required.

GenePattern Notebook



The GenePattern Notebook environment extends the Jupyter Notebook system, allowing researchers to create documents that interleave formatted text, graphics and other multimedia, executable code, and GenePattern analyses, creating a single "research narrative" that puts scientific discussion and analyses in the same place.

Blog > GP updates

- GenePattern Coverage and Support December 23, 2021 - January 2, 2022
- · End of Support for GParc
- End of support for modules using the deprecated GenePattern patch mechanism.
- End of support for Cufflinks suite of modules
- End of support for MAGeCK

view more >

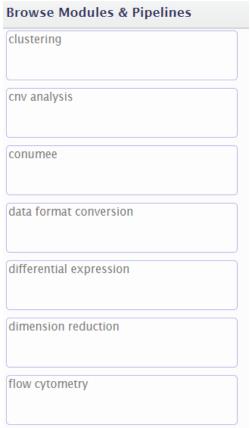
Papers > Related Publications

- Comparability and reproducibility of biomedical data
- ToppCluster: a multiple gene list feature analyzer for

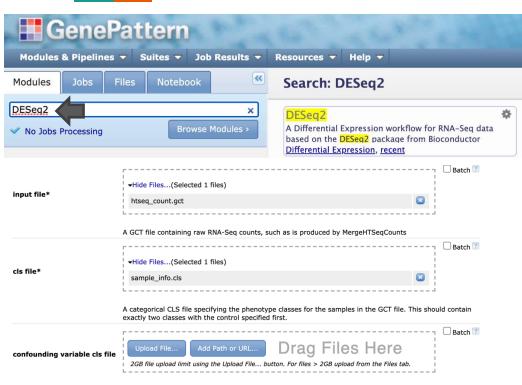
Content



- Modules = standard bioinformatics tools
- Notebook = Python environment (similar to Google Colab) with specific bioinformatics library



DESeq2 on GenePattern



- Input expression data (RNA-seq read count) and sample label
- .gct and .cls are text files

#1.2						
60488	12					
Name	Description	sample1	sample2	sample3	sample4	9
ENSG00000242268.2	ENSG00000242268.2	2	10	0	2	
ENSG00000270112.3	ENSG00000270112.3	8	6	0	0	
ENSG00000167578.15	ENSG00000167578.15	102	633	468	1200	
ENSG00000273842.1	ENSG00000273842.1	0	0	0	0	
ENSG00000078237.5	ENSG00000078237.5	370	364	1220	692	

12 2 1 # alive dead 1 1 0 0 1 0 0 0 1 0 1 1

A categorical CLS file specifying an additional confounding variable, mapped to the input file samples. Use this for a two-factor comparison.

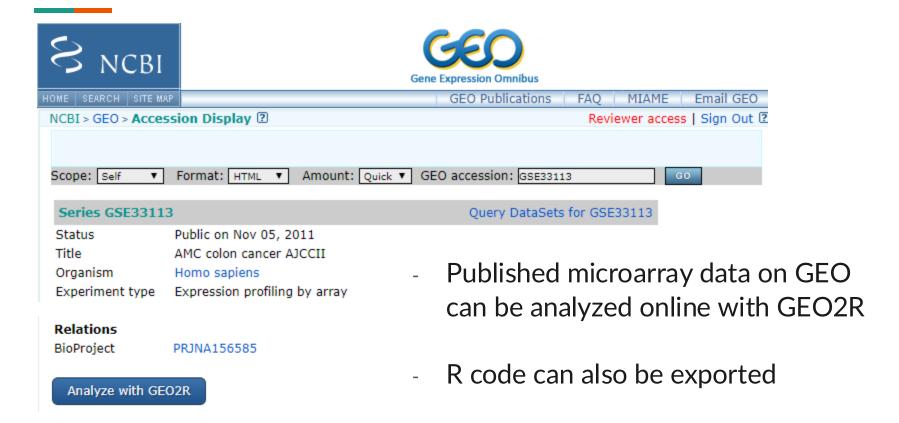
DESeq2 output

471244. DESeq2 📵 Source: GenePattern production (new) submitted: Oct 25 01:06:41 AM, completed: Oct 25 01:10:38 AM, size: 30 MB Show details Comments (0) Tags (0) input.file: expression.gct cls.file: label.cls class_example.Positive.vs.Negative.DESeq2_results_report.txt (670.0 KB) (Last modified: 2022-10-25 01:10:26.0) class_example.Positive.vs.Negative.QC.DispEsts.png (78.0 KB) (Last modified: 2022-10-25 01:10:26.0) class_example.Positive.vs.Negative.QC.MAplot.png (9.0 KB) (Last modified: 2022-10-25 01:10:26.0) ✓ class_example.Positive.vs.Negative.mean_values_by_class.txt (230.0 KB) (Last modified: 2022-10-25 01:10:26.0) class example. Positive. vs. Negative. normalized counts. qct (14.5 MB) (Last modified: 2022-10-25 01:10:26.0) ✓ class example. Positive. vs. Negative. normalized counts.txt (14.5 MB) (Last modified: 2022-10-25 01:10:26.0) el class example. Positive. vs. Negative.... 0 downregulated genes report.txt (3.0 KB) (Last modified: 2022-10-25 01:10:26.0) class_example.Positive.vs.Negative.top20_upregulated_genes_report.txt (3.0 KB) (Last modified: 2022-10-25 01:10:26.0) stderr.txt (1.0 KB) (Last modified: Tue Oct 25 01:10:37 UTC 2022)

stdout.txt (4.0 KB) (Last modified: Tue Oct 25 01:10:37 UTC 2022)

qp execution log.txt (1.0 KB) (Last modified: Tue Oct 25 01:10:38 UTC 2022)

GEO2R: Differential expression



Demo time!

- Let's try some online tools:
 - ColabFold interface for AlphaFold2
 - STRING for viewing protein-protein interaction
 - GEO2R for differential expression analysis of microarray data

Any question?