

Tracking Principles / Practicals

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Abstract

In the lecture, we will overview methods used for tracking in cell and developmental biology. Focus is made on the details of generally used two-step strategy namely particle detection and linking. In practical works, we use several image processing and analysis tools and explore strategies for quantifying the microtubule polarity of EB1 labeled cells within *Drosophila* embryo. Students learn how to measure their movement in image sequences by manual tracking, kymograph, automated tracking and by PIV (Particle Image Velocimetry).

1 Lecture Notes

Time series of digital images, usually called 'a stack', contains temporal dynamics of position and intensity. By analyzing these dynamics, we can extract numerical parameter which then enables us to characterize the biological system. There are three types of dynamics.

1. Position does not change but intensity changes over time.
2. Position changes but the intensity does not change.
3. Both Position and Intensity change over time.

The lecture is about the measurement of type (i), generally called object tracking.

For recent review on object tracking in cell and developmental biology, see articles by Meijering et al. (2012). A bit old review on single particle tracking by Saxton is still an excellent textbook affording rich biophysical insights for analyzing tracking results (Saxton and Jacobson, 1997). For the classic "tracking" mentioned in the talk, or the presumptive fate map, see Vogt (1925). This historical article could be downloaded via Google books. Magnificent movies produced by Keller et. al. using single plane illumination microscopy and extensive image processing could be accessed through the web page of the paper in Science (Keller et al., 2008). Comparison of segmentation strategy in object tracking was published in 2001 by Cheezum et al. (2001) and has been a frequently cited paper in the field of biological object tracking. For overall review on image analysis methods see Hamilton (2009).

1.1 Segmentation techniques

- Manual tracking
ImageJ "manual tracker plugin", we will do this in the practical.
- Manual contour tracing and centroid
- Thresholding

- Local intensity maxima
- Gaussian-fitting
 - Many applications for getting sub-pixel resolution coordinates.
 - You also saw this already in super-resolution microscopy talks by Ricardo and Meilke.
 - Vaccinia Virus tracking example
- Active Contour (SNAKES), Level-set
 - <http://iac1.ece.jhu.edu/projects/gvf/>
 - Quimp11, ImageJ Plugin
<http://www2.warwick.ac.uk/fac/sci/systemsbiology/staff/bretschneider/quimp/>
- Machine Learning
 - There is no tracking package that does tracking using machine learning based segmentation, but it is a strong candidate in coming days.
 - Fiji plugin “Advanced Weka Segmentation”
http://fiji.sc/Advanced_Weka_Segmentation
- Pattern matching (cross-correlation or sum-of-difference technique)
 - 3D tracking of macrophage-like cells.
See Grabher et al. (2007)

1.2 Linking techniques

- Nearest-Neighbor Linking
This is the simplest principle linking an object in one time point to the other.
- Evaluation of Overlaps
- Graph-theory based global optimization
 - Vallotton et al. (2003)
- Designing cost functions for the global optimization.
 - Sbalzarini and Koumoutsakos (2005)
 - Sbalzarini (2006)
- “Tracklets”, a new way of dealing with dividing cells.
 - Jaqaman and Danuser (2009)
 - Bise et al. (2011)

2 Sample Sequences

Get Image Files

If capturing of EB3 image sequences were successful, we use them. Otherwise, we use an example image sequence of EB1 labeled cells. EB1 behaves similar to EB3 and moves along microtubules. A file named **eb18.b.tif** is a sequence taken from single cultured cell labeled with eb1.

Download and save the image files, either the sequence you have taken or the example sequence eb18.b.tif in your local computer. We try several different ways to measure movement of EB3 or EB1 protein by tracking. Before tracking, examine the sequence first using stack related functions.

Open the file in Fiji by

[File > Open...]

Check dimensions of the image by

[Image > Properties...]

...Image stacks are by default taken as a z-series and not t-series. Set Slices to 1, and Frames to appropriate size (number of frames).

Examine the sequence using stack tools.

Explore stack functions.

Start animation, Stop animation, change frame rates, Manipulations... These functions are located under

[Image > Stack >]

A useful plugin for stack

Running Z Projector plugin averages defined number of frames with moving window. This processing is powerful when you want to reduce noise from time-lapse sequences. Drawback is that some details of movement might be attenuated due to averaging. The plugin could be downloaded from

- http://valelab.ucsf.edu/~nico/IJplugins/Running_ZProjector.html

1. Download the plugin from above URL.
2. Install the plugin using [Plugin > Install...]
3. Restart Fiji, just to be sure with the installation.
4. Try running it on EB1 or EB3 sequence.

A plugin for sample images

Download the plugin "Sample Image Loader" linked in the following page and install just like you did with the running Z-projector.

- <http://cmci.embl.de/downloads/sampleimageloader>

3 Manual Tracking

In the simplest case, tracking can be done manually. The user can read out the coordinate position of the target object directly from the imaging software. In ImageJ, user can read out position coordinate indicated in the status bar by placing the cross-hair pointer over the object. Then coordinates can be listed in standard spreadsheet software such as Microsoft Excel or R for further analysis.

An ImageJ plug-in is also freely available to assist such simple way of tracking. An obvious disadvantage of the manual tracking is that the mouse-clicking by the user could be erroneous, as we are still human who gets tired after thousands of clicking. For such errors, measurement errors can be estimated by tracking the same object several times and this error could then be indicated together with the results. Otherwise, automated tracking is more objective, but if you have only limited number of tracks to analyze, manual tracking is a best choice before start to explore complex parameter space of automated tracking setting.

The manual tracking can be assisted by an ImageJ plug-in "Manual Tracking". This plug-in enables the user to record x-y coordinates of the position where the user clicked using mouse in each frame within a stack. The download site has a detailed instruction on how to use.

Download and Installation

Manual Tracking (Manual_Tracking.class) is a plugin bundled with Fiji. If you are using ImageJ, download the plugin file from the link below.

- <http://rsbweb.nih.gov/ij/plugins/track/track.html>

Measurements

1. Your task now is to track the movement of individual EB3 signal using the ManualTracker PlugIn. To ease detection of the signal, enhance contrast and convert the image stack from 16-bit to 8-bit.

[Image > Adjust > Brightness and Contrast]

and then convert to 8-bit

[Image > Type > 8bit]

2. Then
 - (a) Check "centering correction", use Local Maximum.
 - (b) Start manual tracking by clicking "Add track".
 - (c) End tracking by "End Track"
 - (d) Show tracks by "Drawing" Functions"

You could track different particles by repeating steps between 2 and 3. Results window will list the measured positions for particles and step 4 will show a track overlaid image stack.

3. Tracked data can be saved by activating "Result" window and

[File > Save as...]

4. **OPTIONAL:** Copy and paste the result table and plot the track in Excel or R and calculate a histogram of velocity distribution.

4 Kymograph

Kymographs are a two-dimensional time traces, where time t is in Y axis and space along a one-dimensional contour is in X, and the dynamical variable $F(x,t)$ is visualized as an image. Kymographs provide a fast and convenient way to visualize motion and dynamics in microscopy images. We try measuring the speed of EB1 or EB3 movement using Kymographs.

1. Open the image stack.
2. Contrast enhance,
[Image > Adjust > Brightness and Contrast]
and then convert to 8-bit
[Image > Type > 8bit]
3. Do maximum intensity Z-projection
[Image > Stacks > Z-projection].
4. Choose segmented line ROI tool (to do this, right click line-ROI icon in the menu bar. You will see a drop down selection). With projection image generated above, trace one of the tracks in the projection image.

5. Go back to the stack (click the window and activate it - or bring it to the front -),
 [Edit > Selection > Restore Selection]
 Then
 [Image > Stacks > Reslice...]
 Don't change parameters in the dialog window, simply OK. Resulting image is the kymograph.
6. To estimate the velocity of movement from kymograph, you simply need to measure the slope of the diagonal line shown in the kymograph. Since x-axis is distance and y-axis is time, if the starting point of the diagonal line is (x_1, y_1) and the end point of the line is (x_2, y_2) then the *slope = velocity* is

$$velocity = (x_2 - x_1) / (y_2 - y_1)$$

Here is a simple macro to calculate this automatically from straight line selection.

ImageJ macro for measuring kymograph

```
// scale and dt must be manually set
var xyscale=0.207; //um
var dt=0.69; //sec

// sampling ROI coordinates, should be a straight line selection
print("-----");
getLine(x1, y1, x2, y2, lineWidth);
print("start (" + x1 + " , " + y1 + ")    end (" + x2 + " , " + y2 + ") ");

// calculation of speed.
dx=abs(x2-x1);
dy=abs(y2-y1);
dx *= xyscale;
dy *= dt;
velocity= dx/dy;
//output in log window.
print("dx = " + dx + "um    during:" + dy + "sec");
print("Velocity = " + d2s(velocity, 3) + " [um/s]");
```

To use this macro, do

[File > New > Script]
 to launch script editor, then select

[Language > ImageJ Macro]
 in the script editor menu, copy and paste the code above. You could also copy and paste from the following page.

- <http://cmci.embl.de/documents/121005advancedimg>

7. **OPTIONAL:** Collect your data in Excel or R and plot tracks and a histogram of velocity distribution.
8. **OPTIONAL:** For those interested in using kymograph intensively, there is a good plugin that assists you in doing so efficiently.
 - Multiple Kymograph Plugin for ImageJ
<http://www.embl.de/eamnet/html/kymograph.html>
9. **OPTIONAL:** To quantify ambiguous kymograph (meaning hard to draw a line, texture like streaks) use:
 - Kymoquant macro for ImageJ
<http://cmci.embl.de/downloads/kymoquant>

5 Automated Tracking: ParticleTracker Plugin

We use open source software Fiji for particle tracking. Fiji is a distribution of ImageJ, bundled with many useful plugins.

Download ParticleTracker Plugin and Install

The plugin could be down loaded from

- <http://www.mosaic.ethz.ch/Downloads/ParticleTracker>

To install the downloaded plugin, [Plugins > Install...]. You could also directly copy the file to plugins folder under Fiji directory and do [Help > Refresh Menus].

Just in case if the above plugin somehow does not work, try download and use the version linked in the flowing site.

- <http://cmci.embl.de/downloads/particletracker2d>

Start the ParticleTracker plugin

Start the particleTracker by [Plugins > Mosaic > ParticleTracker 2D/3D].

Study Dot Detection Parameter

This tracking tool has two parts. First, all dots in each frame are detected, and then dots in successive frames are linked. The first task then is to determine three parameters for dot detection. There are three parameters.

1. **Radius**
Expected diameter of dot to be detected in pixels.
2. **CutOff**
Cutoff level for the none-particle discrimination criteria, a value for each dot that is based on intensity moment order 0 and 2.
3. **Percentile**
Larger the value, more particles with dark intensity will be detected. Corresponds to the area proportion below intensity histogram in the upper part of the histogram.

Try setting different numbers for these parameters and click "Preview Detected". Red circles appear in the image stack. You could change the frames using the slider below the button.

After some trials, set parameters to what you think is optimum.

Set Linking parameters

Two parameters for linking detected dots should be set.

1. **Link Range**
...could be more than 1, if you want to link dots that disappears and reappears. If not, set the value to 1.
2. **Displacement**
...expected maximum distance that dots could move from one frame to the next. Unit is in pixels.

After parameters are set, click "OK". Tracking starts.

Inspect the Tracking Results

When tracking is done, a new window titled "Results" appears. At the bottom of the window, there are many buttons. Click "Visualize all trajectories", and then a duplicate of the image stack overlaid with trajectories will appear.

Select a region within the stack using rectangular ROI tool and then click "Focus on Area". This will create another image stack, with only that region. Since this image is zoomed, you could carefully check if the tracking was successful or not.

If you think the tracking was not successful, then you should reset all the parameters and do the tracking again.

Export the tracking results

To analyze the results in R, data should be saved as a file. To do so, first click "All Trajectories to Table". Results table will then created. In this results table window, select [File > Save As...] and save the file on your desktop. By default, file type extension is ".xls", excel format, but change this to ".csv". CSV stands for "comma separated file", and this is more classic but general data format which you could easily import in many software including R.

Further analysis

OPTIONAL: Import your results from Excel or R and plot tracks and a histogram of velocity distribution.

6 Particle Image Velocimetry (PIV)

Download and Installation

There is a **PIV** plugin bundled with Fiji, but we use another PIV plugin developed by Qingzong Tseng. You need several files for this plugin to run. Here is the recipe for running the plugin in 64-bit Windows OS.

1. Go to
 - <https://sites.google.com/site/qingzongtseng/piv>and scroll the page down to "Installation" section. There you will find links to **PIV.jar**, **javacv.jar** and **jna.jar** file. Download these three files.
2. Move or copy above three files to the **plugin folder** within Fiji or ImageJ folder.
3. Go to
 - <https://sites.google.com/site/qingzongtseng/template-matching-ij-plugin>(or you could click "template matching and slice alignment" link in the side bar in Zong's page) and scroll down to "Installation" section. There you will find a link to OpenCV library for 64-bit windows. Download the file and unzip it. You will find two files, cv100.dll and cxcore100.dll, then.
4. Move or copy above two files, cv100.dll and cxcore100.dll, to the Fiji or ImageJ folder (**not the plugin folder**).

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