Plotting Chromosight's output

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Chromosight generates tabular text files with loops coordinates and scores. This file can be loaded into your favorite scripting language for visualization. For the purpose of this demonstration, we show how to plot the contact maps with detected coordinates using python, pandas and cooler.

The data shown here was generated with the following commands:

```
chromosight detect data_test/example.cool -m8000 -M50000 -p0.35 detect/example_loops chromosight detect data_test/example.cool --pattern borders detect/example_borders chromosight detect data_test/example.cool --pattern hairpins detect/example_hairpins
```

Which will detect all loops of size 8-50kb in example.cool and filter those with a score above 0.35. The output files will be located in the detect/ folder.

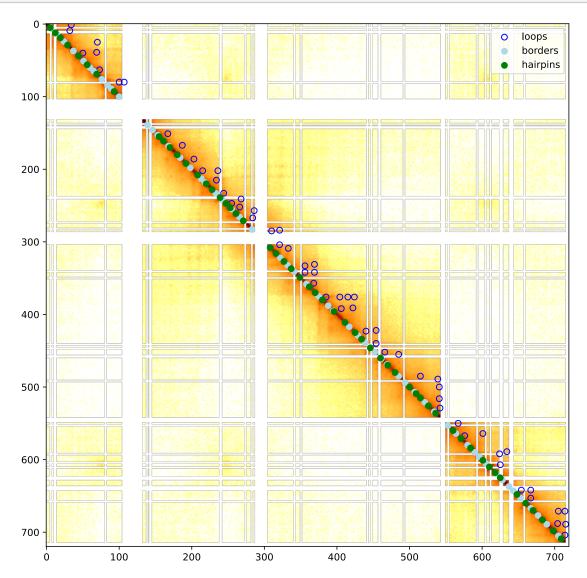
```
[35]: %config InlineBackend.figure_format = 'svg'
import re
import json
import numpy as np
import pandas as pd
import cooler
import matplotlib.pyplot as plt
import chromosight.utils.detection as cud

# Load detected patterns' tables
loops = pd.read_csv('detect/example_loops.tsv', sep='\t')
borders = pd.read_csv('detect/example_borders.tsv', sep='\t')
hairpins = pd.read_csv('detect/example_hairpins.tsv', sep='\t')

# Load Hi-C data in cool format
c = cooler.Cooler("../../data_test/example.cool")
```

View the whole genome matrix

To plot the whole matrix with patterns, the matrix is extracted from the cool file and columns bin1 and bin2 are used. Those columns contain the genome-wide bin number of pattern coordinates, and matches the whole genome matrix. Plotting the whole genome is straightforward, but likely to take too much memory for larger genomes.



View a matrix region

To reduce the amount of memory required, we can define a region of interest. The corresponding matrix region can be fetched from the cool file using cooler, and patterns falling within that region can be filtered using pandas. Since we want to overlay the patterns on top of the region matrix, the bin1 and bin2 columns should be adjusted to be relative to the region's start instead of the genome.

```
[]: def subset_region(df, region):
         Given a pattern dataframe and UCSC region string, retrieve only patterns in_{11}
      \hookrightarrow that region.
         11 11 11
         # Split the region string at each occurrence of - or : (yields 3 elements)
         chrom, start, end = re.split('[-:]', region)
         start, end = int(start), int(end)
         # Only keep patterns on the same chromosome as the region and
         # within the start-end interval
         subset = df.loc[
              (df.chrom1 == chrom) &
              (df.chrom2 == chrom) &
              (df.start1 >= start) &
              (df.start2 >= start) &
              (df.end1 < end) &
              (df.end2 < end),:
         1
         return subset
```

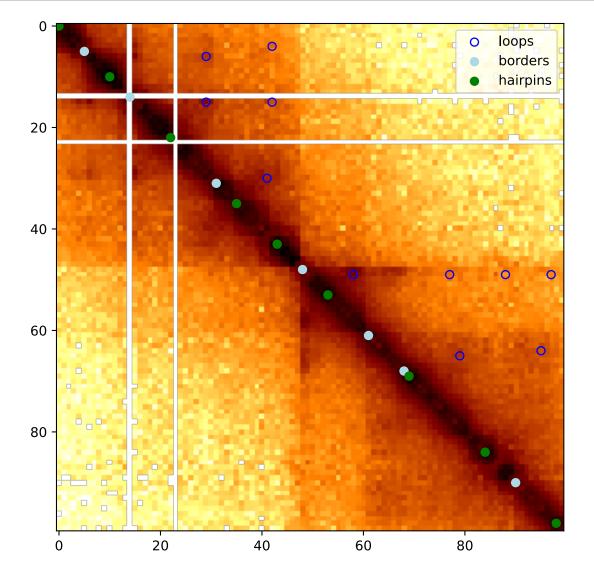
```
[37]: # Select a region of interest
region = 'chr2:200000-300000'
mat = c.matrix(sparse=False, balance=True).fetch(region)

loops_sub = subset_region(loops, region)
borders_sub = subset_region(borders, region)
hairpins_sub = subset_region(hairpins, region)

# Make genome-based bin numbers relative to the region
for df in [loops_sub, borders_sub, hairpins_sub]:
    df.bin1 -= c.extent(region)[0]
    df.bin2 -= c.extent(region)[0]
```

```
[38]: %matplotlib inline
plt.figure(figsize=(7, 7))
plt.imshow(np.log10(mat), cmap='afmhot_r')
plt.scatter(loops_sub.bin2, loops_sub.bin1, edgecolors='blue',__

facecolors='none', label='loops')
plt.scatter(borders_sub.bin2, borders_sub.bin1, c='lightblue', label='borders')
plt.scatter(hairpins_sub.bin2, hairpins_sub.bin1, c='green', label='hairpins')
plt.legend()
plt.show()
```

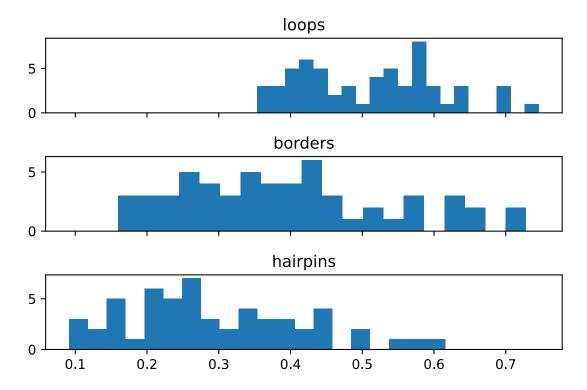


Plot the distribution of scores

Scores of detected patterns are provided as Pearson correlation coefficient with the template and are stored in the 'score' column of the tabular output. Their distribution can be viewed with regular histogram functions. Since we use a threshold for detection (the --pearson option in the command line interface), the score lower end of the distribution will be truncated at this threshold.

Different patterns will have different score distributions and default thresholds.

<Figure size 576x576 with 0 Axes>



Looking at detected patterns

Windows around detected patterns in the processed matrix are stored in the JSON / npy file when running chromosight's detect or quantify commands. These windows are in the same order as the coordinates in the output table.

```
[40]: # Load input json file into a dictionary
loop_wins = json.load(open('detect/example_loops.json', 'r'))
# Note that keys are string, as required by the JSON format,
# so we convert them to int() for convenience
loop_wins = {int(i): np.array(w) for i, w in loop_wins.items()}
# Make an empty 3D array of shape N_coords x height x width
wins = np.zeros((len(loop_wins.items()), *loop_wins[0].shape))
# Fill the 3D array with windows values
for i, w in loop_wins.items(): wins[i] = w
```

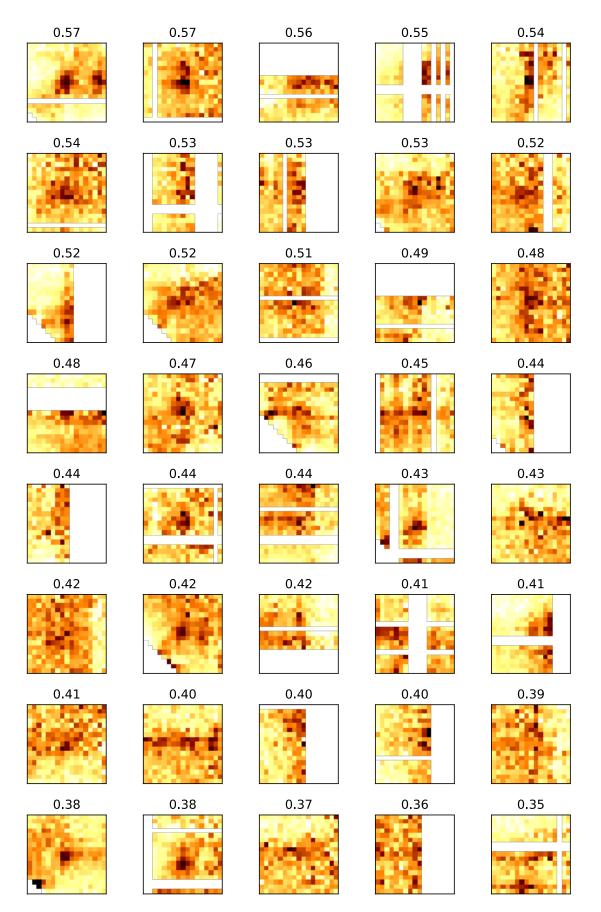
For example, we can plot the best 40 windows around detected loops ordered by score:

```
[41]: %matplotlib inline
plt.figure(figsize=(10, 10))

fig, ax = plt.subplots(8, 5, figsize=(8, 12))

for i, n in enumerate(np.argsort(loops.score)[39::-1]):
    m, s = np.nanmean(loop_wins[n]), np.nanstd(loop_wins[n])
    ax.flat[i].imshow((loop_wins[n] - m) / s, cmap='afmhot_r', vmax=4)
    ax.flat[i].set_title(f'{loops.score[n]:.2f}')
    ax.flat[i].set_xticks([])
    ax.flat[i].set_yticks([])
    plt.tight_layout()
```

<Figure size 720x720 with 0 Axes>



The pileup can also be re-computed from these windows using chromosight's helper function. This is useful to plot the pileup for a subset of the detected patterns, or just to generate the pileup plot with different aesthetics.

```
[42]: %matplotlib inline
  plt.figure(figsize=(4, 4))
  pileup = cud.pileup_patterns(wins)
  plt.imshow(pileup, cmap='coolwarm', vmax=1.8, vmin=0)
```

[42]: <matplotlib.image.AxesImage at 0x7f571dcb6f50>

