Supplementary material for

Estimating the effectiveness of routine asymptomatic PCR testing at different frequencies for the detection of SARS-CoV-2 infections

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## A. Comparison with previous results

We compared our findings for the probability of detection by PCR as a function of time since infection to two existing<sup>1,2</sup> results (Figure S1A). Kucirka et al. fit a polynomial curve to results from 8 studies that estimated PCR sensitivity as a function of time since symptom onset or exposure. Hay & Kennedy-Schaffer fit a model motivated by prior knowledge of viral load dynamics to PCR sensitivity as a function of symptom onset as compiled by Borremans et al.<sup>3</sup>

Our model, fitted to the SAFER study data, found a much higher probability of detecting infections between 1 and 3 days after infection than the model fitted by Kucirka et. al. The peak probability of detection was also later on during the course of infection for Kucirka et. al, who estimated it to be around 8 days after infection compared to 4 days found by our model and Hay & Kennedy-Schaffer et. al. Our estimated probability of detection was also consistently lower than that found by both other models for 7 days after infection onwards.

We summarised the SAFER study data using the median infection times inferred by our model to provide point estimates of the observed probability of detection for each day since infection (shown in the orange points in Figure S1A). We then re-fit the Kucirka model to its original data set extended to include the HCW data from the SAFER study (Figure S1B). Doing this gave estimates of the probability of detection between 1 and 3 days after infection from the Kucirka model that were very similar to those found by our model.

However, the Kucirka model still estimates higher probabilities of detection than our model from 7 days after infection onwards. This seems to be a feature of the SAFER study data rather than a poor model fit to the SAFER data (Figure S1A).

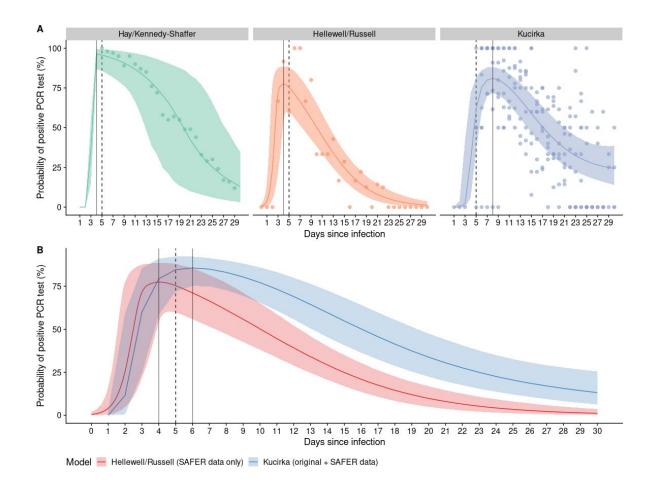


Figure S1: A) Three PCR detection probability curves as a function of the time since infection along with data used to fit the curve. From L - R: (Green) A curve fit to data from Borremans et al (2020) by Hay & Kennedy-Shaffer et al (2020). (Orange) Our curve fit to data on HCWs, the points show the estimated sensitivity of PCR tests at each time point using the median infection time estimated by our model. (Purple) The curve fit Kucirka et al (2020) to data combined from 8 studies. B) A copy of our PCR detection probability curve (same as orange curve in A) is shown in red. The blue curve shows the results of fitting the model developed by Kucirka et al 2020 to their data (purple points in A) combined with our data (orange points in A).

## B. Sensitivity Analysis

It is possible that the fitting procedure that led to our PCR positivity curve over time was heavily influenced by a single individual, given that our curve was jointly fit to the posterior distributions of all individuals (representing the likely time at which they were exposed to SARS-CoV-2). Therefore, to check the influence of each individual on the resulting curve, we performed a leave-one-out sensitivity analysis, whereby we fit the same curve to the 27 possible sets of 26 individuals if one individual is left out in each fit.

The medians of the resulting 27 fits and their corresponding 95% credible intervals are plotted in Figure S2, where it is possible to see that each run is largely in agreement with all of the others. In particular, the 95% credible intervals of all curves are in almost exact agreement. The median of one curve is noticeably lower than the others after the 11-12 day mark. However, on inspection of the 95% credible interval for this curve in comparison to the other 26, it is possible to see that this variation is captured within the 95% credible intervals of the other curves. Specifically, we notice that the 95% credible interval of the curve with a lower median is almost identical to the 95% credible intervals of the other 26 curves.

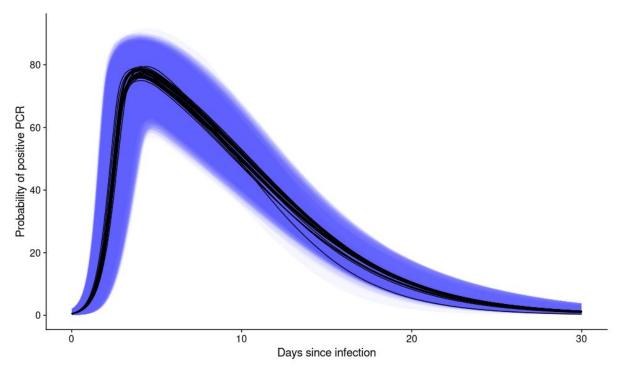


Figure S2: Multiple PCR positivity curves superimposed on top of each other, each curve shows the fitted PCR positivity curve while leaving out data for a different one of the 27 individuals in the data set each time. There is one curve whereby the median posterior probability is around 5% lower from ~12 days after infection onwards if data for an individual is excluded. This suggests that one individual out of the 27 HCWs continued to test positive for a long time after their inferred infection date, which could possibly bias our PCR positivity upwards slightly towards the tail of the distribution.

## C. Routine Asymptomatic Testing Model

To calculate the probability that a symptomatic infection is detected prior to symptom onset, let I be the set of the possible testing times for a given test frequency  $f_x$ , which given explicitly, can be written as

$$I = \{[0, f_x, 2f_x, \ldots], [1, f_x + 1, 2f_x + 1, \ldots], \ldots, [f_x - 1, 2f_x - 1, 3f_x - 1 \ldots]\}$$

The maximum values of  $i \in I$  are set at 30 since testing PCR positive 30 days after infection is unlikely.

For the given testing times  $i \in I$ , if we denote the  $j_{th}$  testing time in i as  $i_j$ , the number of testing times in i as |i|, and d as the delay between test and result, the probability of detecting an infection before symptom onset for testing times i is equal to

$$P_i = \sum_{j=1}^{|i|} g(i_j)p(i_j - d) \prod_{k=j-1}^{0} \left( (1 - g(i_k))(1 - p(i_k - d)) \right)$$

where g(t) is the probability of no onset before time t and p(t) is the probability of a positive test at time t.

Noting that  $|I|=f_x$ , the probability of detecting a symptomatic infection before symptom onset over all possible testing time variations  $i \in I$  is therefore

$$\frac{1}{f_x} \sum_{i \in I} P_i$$

For asymptomatic infections, the value of  $g(t)=1 \ \forall t$  because there will never be an onset time. For detection within seven days we consider

$$I^* = \{[0, f_x, 2f_x, \ldots], [1, f_x+1, 2f_x+1, \ldots], \ldots, [f_x-1, 2f_x-1, 3f_x-1\ldots]\}$$

with values up to 7-d, since a positive test needs to be performed by this point to be returned within 7 days. For the given testing times  $i \in I^*$ , the probability of detecting an asymptomatic infection within 7 days is

$$P_i^* = \sum_{j=1}^{|i|} p(i_j) \prod_{k=j-1}^{0} 1 - p(i_k),$$

and lastly the probability of detecting an asymptomatic infection within 7 days over all testing time variations  $i \in I^*$  is equal to

$$\frac{1}{f_x} \sum_{i \in I^*} P_i^*.$$

#### D. Detection via Lateral Flow test

During 2020, lateral flow tests (LFTs) with a turnaround time of roughly 30 minutes for the detection of SARS-CoV-2 have been developed and evaluated<sup>6</sup>. Such tests typically have a lower mean sensitivity than standard PCR tests. However, the faster turnaround time can aid the logistical challenge posed by rapid large-scale testing.

Throughout our analysis in the main text, a positive PCR test is defined by a cycle threshold (Ct) value of less than or equal to 37. However, given that we have Ct values over time within our dataset, we were also able to redefine a positive test, using a Ct value threshold that reflects the sensitivity of the more recent LFTs, which can generally detect infectiousness but not always infection. Specifically, we assumed a test is positive if the corresponding Ct value is less than or equal to 28, which corresponds to the value below which the majority of samples would be expected to be culture positive<sup>4</sup>.

Using this LFT-like definition for a positive test, we re-fitted our model. The probability of detection over the course of an infection is shown in Figure S3. The uncertainty is wider, given that we now have fewer positive tests to fit to in total, as the threshold is essentially redefining tests that were classified as positive as negative. However, we are still able to obtain a well-identified curve, with a peak probability of detection of 60% on around day 5.

Compared to Figure 3, which showed the probability of detection via PCR over the course of an infection, the probability of detection via LFT has the following characteristics: a less steep increase initially, a much steeper decrease after the peak, a lower peak (~60% instead of 78%) and a negligible probability of detection after around 17-18 days rather than for around 30 days for PCR.

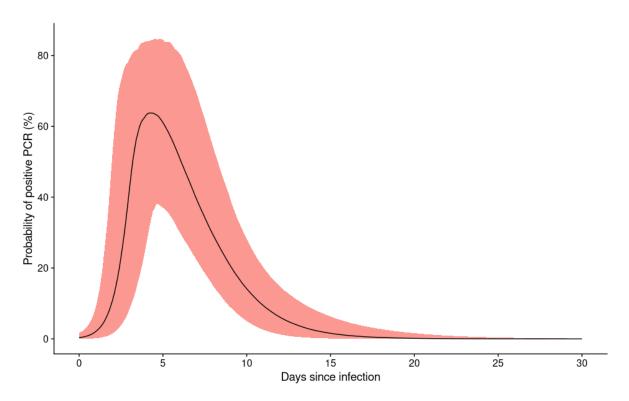


Figure S3: the probability of detection over the course of an infection via lateral flow test. The black filled line represents the median of the posterior distribution of the fitted curve and the coloured region represents the 95% credible interval of the inferred posterior.

## E. Further Methodology

## 1. Empirical distribution of PCR positivity

The grey interval in Figure 3A is calculated from the posterior samples of the likely infection time for each individual ( $T_i$ ). If we let the  $j_{th}$  posterior sample of  $T_i$  be denoted  $T_{ij}$ , then  $d_{ij}$ , which denotes the time from infection until each test  $t_{n,i}$ , performed on individual i, for each sample  $T_{ij}$  is given by

$$d_{ij} = t_{n,i} - T_{ij}$$

Each  $d_{ij}$  is rounded to the nearest discrete day and for each MCMC iteration, j, we calculate the proportion of tests where  $d_{ij}=k$  which were positive for each discrete day k since infection, denoted  $p_j(k)$ . We then calculate the mean and 95% uncertainty intervals of  $p_j(k)$  for each day k over all MCMC samples j. This can be considered a graphical representation of the "data" that the PCR positivity regression is fit to (the precise values rely on the infection time draws at each iteration of the MCMC).

### 2. Sources of uncertainty

We infer likely infection times using a Bayesian inference framework, where we are able to include sources of uncertainty in a statistically robust manner. Such sources of uncertainty include:

- The censored nature of the interval between the *last asymptomatic report* and *first symptomatic report* for all individuals
- The incubation period, which is assumed to be a Gamma distribution parameterised using fitted estimates from Lauer et al. (2020)<sup>5</sup>
- As the data to which we fit the curve to is sampled from the inferred posterior
  distributions corresponding to the likely time at which individuals were infected, the
  95% credible interval of our PCR positivity curve over time includes the uncertainty
  within each of the posterior distributions of likely infection time

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## I. References

- 1. Hay JA, Kennedy-Shaffer L, Kanjilal S, Lipsitch M, Mina MJ. Estimating epidemiologic dynamics from single cross-sectional viral load distributions. medRxiv. 2020 Oct 13;2020.10.08.20204222.
- Kucirka LM, Lauer SA, Laeyendecker O, Boon D, Lessler J. Variation in False-Negative Rate of Reverse Transcriptase Polymerase Chain Reaction—Based SARS-CoV-2 Tests by Time Since Exposure. Ann Intern Med [Internet]. 2020 May 13 [cited 2020 Nov 10]; Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7240870/
- 3. Benny B, Amandine G, Kc P, Van S. Quantifying antibody kinetics and RNA shedding during early-phase SARS-CoV-2 infection. :20.
- 4. Singanayagam A, Patel M, Charlett A, Bernal JL, Saliba V, Ellis J, et al. Duration of infectiousness and correlation with RT-PCR cycle threshold values in cases of COVID-19, England, January to May 2020. Eurosurveillance. 2020 Aug 13;25(32):2001483.
- 5. Lauer SA, Grantz KH, Bi Q, Jones FK, Zheng Q, Meredith HR, et al. The Incubation Period of Coronavirus Disease 2019 (COVID-19) From Publicly Reported Confirmed Cases: Estimation and Application. Ann Intern Med. 2020 May 5;172(9):577–82.
- SARS-CoV-2 Coronavirus Nucleocapsid Antigen-Detecting Half-Strip Lateral Flow Assay Toward the Development of Point of Care Tests Using Commercially Available Reagents | Analytical Chemistry [Internet]. [cited 2020 Dec 1]. Available from: https://pubs.acs.org/doi/abs/10.1021/acs.analchem.0c01975