

- ecodive: Parallel and Memory-Efficient R Package for
- **Ecological Diversity Analysis**
- Daniel P Smith 6, 2, Sara J Javornik Cregeen 6, 2, and Joseph F
- 4 Petrosino (1) 1,2
- 5 1 The Alkek Center for Metagenomics and Microbiome Research, Department of Molecular Virology and
- 6 Microbiology, Baylor College of Medicine, Houston, TX 77030, USA 2 Department of Molecular Virology
- and Microbiology, Baylor College of Medicine, Houston, TX, USA ROR ¶ Corresponding author

DOI: 10.xxxxx/draft

Software

- Review 🗗
- Repository 🗗
- Archive 🗗

Editor: Open Journals ♂

Reviewers:

• @openjournals

Submitted: 01 January 1970 Published: unpublished

License

Authors of papers retain copyrights and release the work under a Creative Commons Attribution 4.Q, International License (CC BY 4.0)0

Summary

Characterizing the composition of biological communities is a fundamental task in ecology, but the calculations involved can be computationally prohibitive. ecodive is an R package that addresses this challenge by providing a highly optimized implementation of common ecological diversity metrics, including alpha-diversity (within-sample richness and evenness) and beta-diversity (between-sample dissimilarity). These metrics can incorporate species counts, relative abundances, and evolutionary relationships, providing a multi-faceted view of ecological structure. By leveraging a compiled C library with pthreads for parallelization, ecodive delivers substantial performance gains in both speed and memory usage, enabling researchers to analyze larger datasets more efficiently.

Statement of Need

The analysis of ecological diversity in large-scale studies is often hampered by the computational demands of calculating metrics across thousands of communities, a common requirement in modern microbiome research. This is particularly true for phylogenetic metrics like Faith's PD (Faith, 1992) and the UniFrac distance family (C. Lozupone & Knight, 2005), which integrate species abundance with evolutionary data from phylogenetic trees. The resulting high demand on processing time and memory can limit the scope and scale of scientific inquiry.

ecodive overcomes these limitations by offering a significantly faster and more memoryefficient solution. This allows researchers to analyze more samples, explore more complex
questions, and obtain more robust insights from their data. By providing a high-performance,
parallelized engine for these calculations, ecodive empowers researchers to push the boundaries
of large-scale ecological analysis.

Comparison to Existing Packages

While numerous R packages can calculate diversity metrics, our comparison focuses on those that provide their own implementations: abdiv (Bittinger, 2020), adiv (Pavoine, 2020), ampvis2 (Andersen et al., 2018), ecodist (Goslee & Urban, 2007), entropart (Marcon & Herault, 2015), GUniFrac (Chen et al., 2023), phyloregion (Daru et al., 2020), phyloseq (McMurdie & Holmes, 2013), picante (Kembel et al., 2010), and vegan (Oksanen et al., 2025). ecodive sets itself apart from these packages through its superior performance. Furthermore, ecodive has zero external R dependencies. This makes it a lightweight, stable, and secure computational backend, minimizing installation conflicts and simplifying long-term maintenance for developers who build upon it.



- 40 Comprehensive benchmarks, conducted using the bench package (Hester & Vaughan, 2025),
- demonstrate these advantages across a range of metrics (Figures 1-3). The complete benchmark
- code and results are available in the package vignette (vignette('benchmark')) and online.

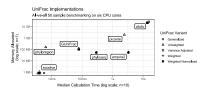


Figure 1: Figure 1: UniFrac benchmarks. ecodive demonstrates substantial performance gains for UniFrac, being 2 to 3,900x faster and using 50 - 32,000x less memory, which helps overcome computational bottlenecks in large-scale analyses.

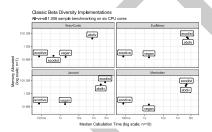


Figure 2: Figure 2: Classic beta diversity benchmarks. ecodive is 6 to 2,300x faster and uses 1 to 1,800x less memory, enabling more efficient analysis of community dissimilarities.

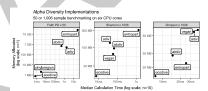


Figure 3: Figure 3: Alpha diversity benchmarks. ecodive is 2 to 43,000x faster and uses 1 to 33,000x less memory, significantly accelerating the analysis of diversity within single samples.

43 Implemented Metrics

- ecodive provides a comprehensive suite of alpha and beta diversity metrics. The current
- 45 implementation includes:

46 Alpha Diversity

- Classic: Shannon Index (Shannon, 1948), Simpson Index (Gini, 1912; Simpson, 1949),
 Inverse Simpson Index (Simpson, 1949), and Chao1 (Chao, 1984).
 - Phylogenetic: Faith's Phylogenetic Diversity (Faith, 1992).

50 Beta Diversity

49

Classic: Bray-Curtis (Bray & Curtis, 1957; Sorenson, 1948), Canberra (Lance & Williams, 1967), Euclidean (Gower & Legendre, 1986; Legendre & Caceres, 2013), Gower (Gower, 1971; Gower & Legendre, 1986), Jaccard (Jaccard, 1908), Kulczynski (Kulczynski, 1927), and Manhattan (Kaufman & Rousseeuw, 1990).



- Phylogenetic: Unweighted UniFrac (C. Lozupone & Knight, 2005), Weighted UniFrac (C. A. Lozupone et al., 2007), Normalized Weighted UniFrac (C. A. Lozupone et al., 2007), Generalized UniFrac (Chen et al., 2012), and Variance Adjusted Weighted UniFrac (Chang et al., 2011).
- For the most up-to-date list and detailed descriptions, please refer to the official ecodive documentation at https://cmmr.github.io/ecodive/reference/index.html.

Example Usage

ecodive is designed for ease of use and integrates seamlessly with existing bioinformatics workflows, such as those using phyloseq objects. For example, calculating weighted UniFrac

64 distances is straightforward:

library(phyloseq)

Acknowledgements

- This study was supported by NIH/NIAD (Grant number U19 Al144297), and Baylor College
- of Medicine and Alkek Foundation Seed. The authors also acknowledge the use of Google's
- Gemini for assistance in refining this manuscript.

References

- Andersen, K. S., Kirkegaard, R. H., Karst, S. M., & Albertsen, M. (2018). ampvis2: An r package to analyse and visualise 16S rRNA amplicon data. bioRxiv. https://doi.org/10. 1101/299537
- Bittinger, K. (2020). Abdiv: Alpha and beta diversity measures. https://doi.org/10.32614/ CRAN.package.abdiv
- Bray, J. R., & Curtis, J. T. (1957). An ordination of the upland forest communities of southern wisconsin. *Ecological Monographs*, 27(4), 325–349. https://doi.org/10.2307/1942268
- Chang, Q., Luan, Y., & Sun, F. (2011). Variance adjusted weighted UniFrac: A powerful beta diversity measure for comparing communities based on phylogeny. *BMC Bioinformatics*, 12(1). https://doi.org/10.1186/1471-2105-12-118
- Chao, A. (1984). Non-parametric estimation of the number of classes in a population. Scandinavian Journal of Statistics, 11, 265–270. https://doi.org/10.2307/4616294
- Chen, J., Bittinger, K., Charlson, E. S., Hoffmann, C., Lewis, J., Wu, G. D., Collman, R. G., Bushman, F. D., & Li, H. (2012). Associating microbiome composition with environmental covariates using generalized UniFrac distances. *Bioinformatics*, 28(16), 2106–2113. https://doi.org/10.1093/bioinformatics/bts342
- Chen, J., Zhang, X., Yang, L., & Zhang, L. (2023). GUniFrac: Generalized UniFrac distances, distance-based multivariate methods and feature-based univariate methods for microbiome data analysis. https://doi.org/10.32614/CRAN.package.GUniFrac



- Daru, B. H., Karunarathne, P., & Schliep, K. (2020). Phyloregion: R package for biogeographic regionalization and macroecology. *Methods in Ecology and Evolution*, *11*, 1483–1491. https://doi.org/10.1111/2041-210X.13478
- Faith, D. P. (1992). Conservation evaluation and phylogenetic diversity. *Biological Conservation*, 61, 1–10. https://doi.org/10.1016/0006-3207(92)91201-3
- Gini, C. (1912). Variabilita e mutabilita. Tipografia di Paolo Cuppini.
- Goslee, S. C., & Urban, D. L. (2007). The ecodist package for dissimilarity-based analysis of ecological data. *Journal of Statistical Software*, 22, 1–19. https://doi.org/10.18637/jss.v022.i07
- Gower, J. (1971). A general coefficient of similarity and some of its properties. *Biometrics*, 27(4), 857–871. https://doi.org/10.2307/2528823
- Gower, J., & Legendre, P. (1986). Metric and euclidean properties of dissimilarity coefficients.

 Journal of Classification, 3, 5–48. https://doi.org/10.1007/BF01896809
- Hester, J., & Vaughan, D. (2025). *Bench: High precision timing of r expressions*. https://doi.org/10.32614/CRAN.package.bench
- Jaccard, P. (1908). Nouvelles recherches sur la distribution florale. *Bulletin de La Societe Vau*doise Des Sciences Naturelles, 44(163), 223–270. https://doi.org/10.5169/seals-268384
- Kaufman, L., & Rousseeuw, P. J. (1990). Finding groups in data: An introduction to cluster analysis. John Wiley & Sons. https://doi.org/10.1002/9780470316801
- Kembel, S. W., Cowan, P. D., Helmus, M. R., Cornwell, W. K., Morlon, H., Ackerly, D. D., Blomberg, S. P., & Webb, C. O. (2010). Picante: R tools for integrating phylogenies and ecology. *Bioinformatics*, 26, 1463–1464. https://doi.org/10.1093/bioinformatics/btq166
- Kulczynski, S. (1927). Die pflanzenassoziationen der pieninen. Bulletin International de l'Académie Polonaise Des Sciences Et Des Lettres, Classe Des Sciences Mathématiques Et Naturelles, Série B: Sciences Naturelles, 57–203.
- Lance, G. N., & Williams, W. T. (1967). A general theory of classificatory sorting strategies II.
 Clustering systems. *The Computer Journal*, 10(3). https://doi.org/10.1093/comjnl/10.3.
- Legendre, P., & Caceres, M. (2013). Beta diversity as the variance of community data:
 Dissimilarity coefficients and partitioning. *Ecology Letters*, 16(8). https://doi.org/10.
 1111/ele.12141
- Lozupone, C. A., Hamady, M., Kelley, S. T., & Knight, R. (2007). Quantitative and qualitative beta diversity measures lead to different insights into factors that structure microbial communities. *Applied and Environmental Microbiology*, 73(5), 1576–1585. https://doi.org/10.1128/aem.01996-06
- Lozupone, C., & Knight, R. (2005). UniFrac: A new phylogenetic method for comparing microbial communities. *Applied and Environmental Microbiology*, 71(12), 8228–8235. https://doi.org/10.1128/AEM.71.12.8228-8235.2005
- Marcon, E., & Herault, B. (2015). Entropart: An r package to measure and partition diversity.

 Journal of Statistical Software, 67(8), 1–26. https://doi.org/10.18637/jss.v067.i08
- McMurdie, P. J., & Holmes, S. (2013). Phyloseq: An r package for reproducible interactive analysis and graphics of microbiome census data. *PloS One*, 8(4), e61217. https://doi.org/10.1371/journal.pone.0061217
- Oksanen, J., Simpson, G. L., Blanchet, F. G., Kindt, R., Legendre, P., Minchin, P. R., O'Hara, R. B., Solymos, P., Stevens, M. H. H., Szoecs, E., Wagner, H., Barbour, M., Bedward, M., Bolker, B., Borcard, D., Carvalho, G., Chirico, M., De Caceres, M., Durand, S., ... Borman,



- T. (2025). Vegan: Community ecology package. https://doi.org/10.32614/CRAN.package. vegan
- Pavoine, S. (2020). Adiv: An r package to analyse biodiversity in ecology. *Methods in Ecology and Evolution*, 11, 1106–1112. https://doi.org/10.1111/2041-210X.13430
- Shannon, C. E. (1948). A mathematical theory of communication. *Bell System Technical Journal*, 27(3), 379–423. https://doi.org/10.1002/j.1538-7305.1948.tb01338.x
- Simpson, E. H. (1949). Measurement of diversity. *Nature*, 163(4148), 688-688. https://doi.org/10.1038/163688a0
- Sorenson, T. (1948). A method of establishing groups of equal amplitude in plant sociology based on similarity of species content. *Kongelige Danske Videnskabernes Selskab*, 5, 1–34.

