

Class 09: Structural Bioinformatics (pt1)

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the PDB database

The main database for structural biology is called the PDB. Let's have a look at what it contains:

Download a CSV file from the PDB site (accessible from "Analyze" > "PDB Statistics" > "by Experimental Method and Molecular Type").

```
stats <- read.csv("Data Export Summary.csv")
stats
```

	Molecular.Type	X.ray	EM	NMR	Integrative	Multiple.methods
1	Protein (only)	176,204	20,299	12,708	342	218
2	Protein/Oligosaccharide	10,279	3,385	34	8	11
3	Protein/NA	9,007	5,897	287	24	7
4	Nucleic acid (only)	3,066	200	1,553	2	15
5	Other	173	13	33	3	0
6	Oligosaccharide (only)	11	0	6	0	1
	Neutron	Other	Total			
1	83	32	209,886			
2	1	0	13,718			
3	0	0	15,222			
4	3	1	4,840			
5	0	0	222			

```
6      0      4      22
```

```
stats$Total
```

```
[1] "209,886" "13,718" "15,222" "4,840" "222" "22"
```

Oh, these are characters not numeric...

```
as.numeric(sub(",", "", stats$Total))
```

```
[1] 209886 13718 15222 4840 222 22
```

```
library(readr)
```

```
stats <- read_csv("Data Export Summary.csv")
stats
```

```
# A tibble: 6 x 9
  `Molecular Type`    `X-ray`    EM    NMR Integrative `Multiple methods` Neutron
  <chr>              <dbl> <dbl> <dbl>          <dbl>          <dbl> <dbl>
1 Protein (only)      176204 20299 12708          342          218    83
2 Protein/Oligosacch~ 10279 3385 34            8           11     1
3 Protein/NA          9007 5897 287           24           7     0
4 Nucleic acid (only) 3066 200 1553           2          15     3
5 Other               173 13 33            3           0     0
6 Oligosaccharide (o~ 11 0 6            0           1     0
# i 2 more variables: Other <dbl>, Total <dbl>
```

```
n.total <- sum(stats$Total)
n.total
```

```
[1] 243910
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy. Give your answer to 2 significant figures

```
n.xray <- sum(stats$`X-ray`)
percent.xray <- n.xray / n.total * 100
percent.xray
```

```
[1] 81.48087
```

There are 81.48 percent Xray structures in the PDB

```
n.em <-sum(stats $ `EM`)  
percent.em <- n.em / n.total * 100  
percent.em
```

```
[1] 12.21516
```

There are 12.22, percent EM structures in the PDB

Q2: What proportion of structures in the PDB are protein?

```
round(stats$Total[1]/n.total * 100, 2)
```

```
[1] 86.05
```

Exploring PDB structures

Package for structural bioinformatics

```
library(bio3d)  
hiv <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
hiv
```

Call: read.pdb(file = "1hsg")

Total Models#: 1

Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [HOH (127), MK1 (1)]

Protein sequence:

PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF

+ attr: atom, xyz, seqres, helix, sheet,
calpha, remark, call

Let's first use the Mol* viewer to explore this structure.



Figure 1: My first view of HIV-Pr

And a view of the ligand (ball and stick) with catalytic ASP 25 amino acids(spacefill) and all the important active site water molecule (spacefill)



PDB object in R

```
head(hiv$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elesy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>

```
3 <NA>      C  <NA>
4 <NA>      O  <NA>
5 <NA>      C  <NA>
6 <NA>      C  <NA>
```

Extract the sequence

```
pdbseq(hiv)
```

```
  1   2   3   4   5   6   7   8   9  10  11  12  13  14  15  16  17  18  19  20
"P" "Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G" "G" "Q" "L" "K"
 21  22  23  24  25  26  27  28  29  30  31  32  33  34  35  36  37  38  39  40
"E" "A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M" "S" "L" "P" "G"
 41  42  43  44  45  46  47  48  49  50  51  52  53  54  55  56  57  58  59  60
"R" "W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V" "R" "Q" "Y" "D"
 61  62  63  64  65  66  67  68  69  70  71  72  73  74  75  76  77  78  79  80
"Q" "I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L" "V" "G" "P" "T"
 81  82  83  84  85  86  87  88  89  90  91  92  93  94  95  96  97  98  99   1
"P" "V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T" "L" "N" "F" "P"
  2   3   4   5   6   7   8   9  10  11  12  13  14  15  16  17  18  19  20  21
"Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G" "G" "Q" "L" "K" "E"
 22  23  24  25  26  27  28  29  30  31  32  33  34  35  36  37  38  39  40  41
"A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M" "S" "L" "P" "G" "R"
 42  43  44  45  46  47  48  49  50  51  52  53  54  55  56  57  58  59  60  61
"W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V" "R" "Q" "Y" "D" "Q"
 62  63  64  65  66  67  68  69  70  71  72  73  74  75  76  77  78  79  80  81
"I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L" "V" "G" "P" "T" "P"
 82  83  84  85  86  87  88  89  90  91  92  93  94  95  96  97  98  99
"V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T" "L" "N" "F"
```

```
chainA_seq <- pdbseq(trim.pdb(hiv, chain = "A"))
```

I can interactively view these PDB objects in R with the new **bio3dview** package. This is not on CRAN yet

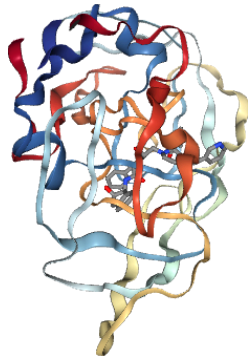
To install this I can setup **pak** package and use it to install **bio3dview** from GitHub. In my console I first run **install.packages("pak")**, then I run **pak::pak("bioboot/bio3dview")** and **install.packages("NGLVieweR")**

```
library(bio3dview)
```

```
view.pdb(hiv)
```

PhantomJS not found. You can install it with `webshot::install_phantomjs()`. If it is installed

file:///private/var/folders/qq/5dd7db0d36v0_h2n3j30cvg80000gn/T/Rtmp768XBi/file100e46e95863,

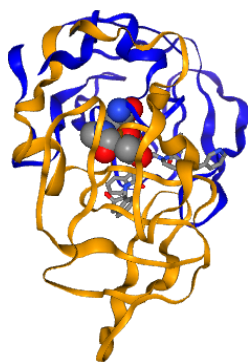


Change some settings

```
sel <- atom.select(hiv, resno=25)
```

```
view.pdb(hiv, highlight = sel, highlight.style= "spacefill", colorScheme = "chain", col = c("l
```

file:///private/var/folders/qq/5dd7db0d36v0_h2n3j30cqh80000gn/T/Rtmp768XBi/file100e6044ef4e,



Predict protein flexibility

We can run a bioinformatics calculation to predict protein dynamics- i.e. functional motions. We will use the `nma()` function:

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file
PDB has ALT records, taking A only, `rm.alt=TRUE`

```
adk
```

Call: `read.pdb(file = "6s36")`

```
Total Models#: 1
  Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)
Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]
```

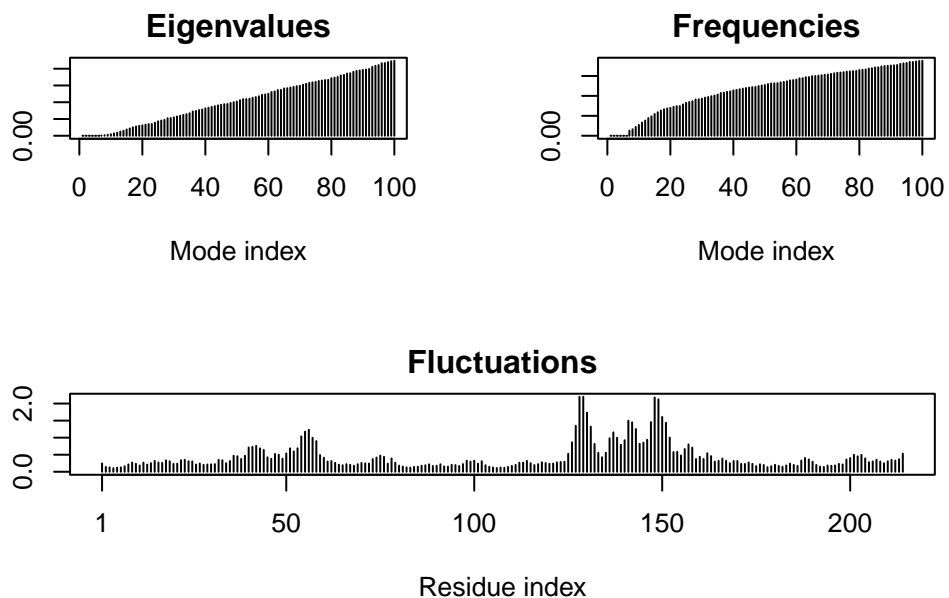
```
Protein sequence:
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV
DELVIALVKERIAQEDCRNGFLLDGFPRTPQADAMKEAGINVDYVLEFDVPDELIVDKI
VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQM TAPLIG
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

```
m <- nma(adk)
```

```
Building Hessian...      Done in 0.02 seconds.
Diagonalizing Hessian... Done in 0.079 seconds.
```

```
plot(m)
```



Generate a “trajectory” of predicted motion

```
mktrj(m, file = "ADK_nma.pdb")
```

```
view.nma(m)
```

```
file:///private/var/folders/qq/5dd7db0d36v0_h2n3j30cqg80000gn/T/Rtmp768XBi/file100e56c2b872
```

