

# Class 09: Structural Bioinformatics (pt1)

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## the PDB database

The main database for structural biology is called the PDB. Let's have a look at what it contains:

Download a CSV file from the PDB site (accessible from “Analyze” > “PDB Statistics” > “by Experimental Method and Molecular Type”.

```
stats <- read.csv("Data Export Summary.csv")
stats
```

	Molecular.Type	X.ray	EM	NMR	Integrative	Multiple.methods
1	Protein (only)	176,204	20,299	12,708	342	218
2	Protein/Oligosaccharide	10,279	3,385	34	8	11
3	Protein/NA	9,007	5,897	287	24	7
4	Nucleic acid (only)	3,066	200	1,553	2	15
5	Other	173	13	33	3	0
6	Oligosaccharide (only)	11	0	6	0	1
	Neutron	Other	Total			
1	83	32	209,886			
2	1	0	13,718			
3	0	0	15,222			
4	3	1	4,840			
5	0	0	222			

```
6      0      4      22
```

```
stats$Total
```

```
[1] "209,886" "13,718"  "15,222"  "4,840"   "222"      "22"
```

Oh, these are characters not numeric...

```
as.numeric(sub(", ", "", stats$Total))
```

```
[1] 209886 13718 15222 4840   222     22
```

```
library(readr)
```

```
stats <- read_csv("Data Export Summary.csv")  
stats
```

```
# A tibble: 6 x 9  
#> #> `Molecular Type`    `X-ray`    EM    NMR Integrative `Multiple methods` Neutron  
#> <chr>          <dbl> <dbl> <dbl>        <dbl>          <dbl> <dbl>  
#> 1 Protein (only) 176204 20299 12708        342          218     83  
#> 2 Protein/Oligosacch~ 10279  3385   34          8           11      1  
#> 3 Protein/NA       9007   5897   287         24            7      0  
#> 4 Nucleic acid (only) 3066    200   1553         2           15      3  
#> 5 Other             173     13    33          3            0      0  
#> 6 Oligosaccharide (o~ 11      0     6          0           1      0  
#> # i 2 more variables: Other <dbl>, Total <dbl>
```

```
n.total <- sum(stats$Total)  
n.total
```

```
[1] 243910
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy. Give your answer to 2 significant figures

```
n.xray <- sum(stats$`X-ray`)  
percent.xray <- n.xray / n.total * 100  
percent.xray
```

```
[1] 81.48087
```

There are 81.48 percent Xray structures in the PDB

```
n.em <- sum(stats $ `EM`)  
percent.em <- n.em / n.total * 100  
percent.em
```

```
[1] 12.21516
```

There are 12.22, percent EM structures in the PDB

Q2: What proportion of structures in the PDB are protein?

```
round(stats$Total[1]/n.total * 100, 2)
```

```
[1] 86.05
```

## Exploring PDB structures

Package for structural bioinformatics

```
library(bio3d)  
  
hiv <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
hiv
```

Call: read.pdb(file = "1hsg")

Total Models#: 1  
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)  
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

```
Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
```

Protein sequence:

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPQMIGGIGGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWPQMIGGIGGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,
       calpha, remark, call
```

Let's first use the Mol\* viewer to explore this structure.



Figure 1: My first view of HIV-Pr

And a view of the ligand (ball and stick) with catalytic ASP 25 amino acids(spacefill) and all the important active site water molecule (spacefill)



## PDB object in R

```
head(hiv$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elesy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>

```

3 <NA>      C  <NA>
4 <NA>      0  <NA>
5 <NA>      C  <NA>
6 <NA>      C  <NA>

```

Extract the sequence

```
pdbseq(hiv)
```

```

 1  2  3  4  5  6  7  8  9  10  11  12  13  14  15  16  17  18  19  20
"P" "Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G" "G" "Q" "L" "K"
 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40
"E" "A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M" "S" "L" "P" "G"
 41 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60
"R" "W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V" "R" "Q" "Y" "D"
 61 62 63 64 65 66 67 68 69 70 71 72 73 74 75 76 77 78 79 80
"Q" "I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L" "V" "G" "P" "T"
 81 82 83 84 85 86 87 88 89 90 91 92 93 94 95 96 97 98 99 1
"P" "V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T" "L" "N" "F" "P"
 2  3  4  5  6  7  8  9  10 11 12 13 14 15 16 17 18 19 20 21
"Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G" "G" "Q" "L" "K" "E"
 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41
"A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M" "S" "L" "P" "G" "R"
 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60 61
"W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V" "R" "Q" "Y" "D" "Q"
 62 63 64 65 66 67 68 69 70 71 72 73 74 75 76 77 78 79 80 81
"I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L" "V" "G" "P" "T" "P"
 82 83 84 85 86 87 88 89 90 91 92 93 94 95 96 97 98 99
"V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T" "L" "N" "F"

```

```
chainA_seq <- pdbseq(trim.pdb(hiv, chain = "A"))
```

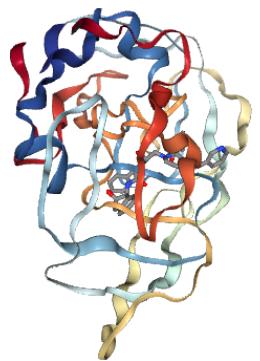
I can interatively view these PDB objects in R with the new **bio3dview** package. This is not on CRAN yet

To install this I can setup **pak** package and use it to install **bio3dview** from GitHub. In my console I first run `install.packages("pak")`, then I run `pak::pak("bioboot/bio3dview")` and `install.packages("NGLVieweR")`

```
library(bio3dview)

view.pdb(hiv)
```

PhantomJS not found. You can install it with `webshot::install_phantomjs()`. If it is installed  
file:///private/var/folders/qq/5dd7db0d36v0\_h2n3j30cqg80000gn/T/Rtmp768XBi/file100e46e95863

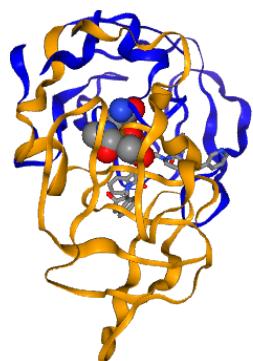


Change some settings

```
sel <- atom.select(hiv, resno=25)
```

```
view.pdb(hiv, highlight = sel,highlight.style= "spacefill", colorScheme = "chain", col = c("1
```

```
file:///private/var/folders/qq/5dd7db0d36v0_h2n3j30cqg80000gn/T/Rtmp768XBi/file100e6044ef4e,
```



## Predict protein flexibility

We can run a bioinformatics calculation to predict protein dynamics- i.e. functional motions.  
We will use the `nma()` function:

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file  
PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

Call: `read.pdb(file = "6s36")`

Total Models#: 1

Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)

Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]

Protein sequence:

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMLRAAVKSGSELGKQAKDIDMAGKLVT  
DELVIALVKERIAQEDCRNGFLLDGFPRTRIPQADAMKEAGINVVDYVLEFDVPDELIVDKI  
VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG  
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

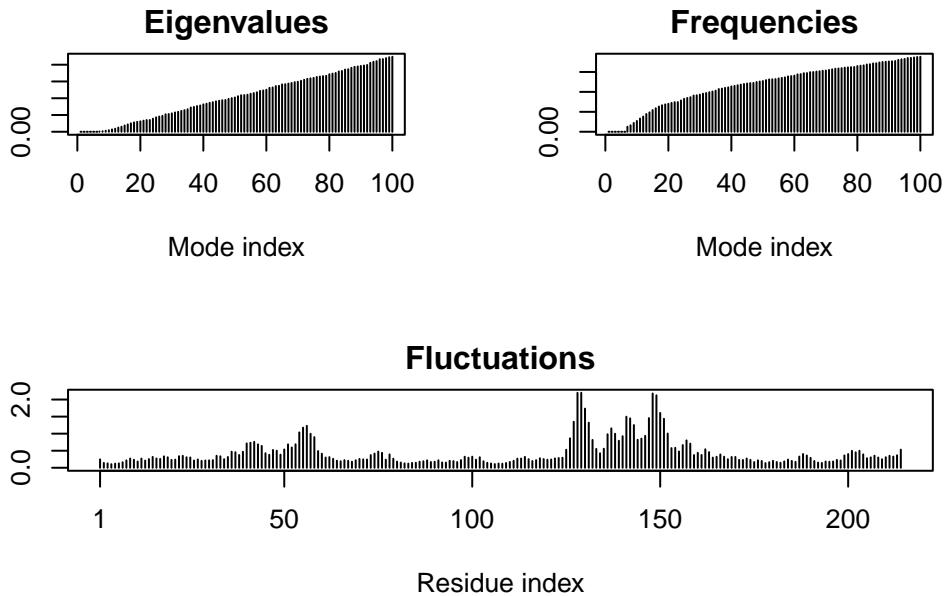
+ attr: atom, xyz, seqres, helix, sheet,  
calpha, remark, call

```
m <- nma(adk)
```

Building Hessian... Done in 0.02 seconds.

Diagonalizing Hessian... Done in 0.079 seconds.

```
plot(m)
```



Generate a “trajectory” of predicted motion

```
mktrj(m, file ="ADK_nma.pdb")
```

```
view.nma(m)
```

```
file:///private/var/folders/qq/5dd7db0d36v0_h2n3j30cqg80000gn/T/Rtmp768XBi/file100e56c2b872
```

