



Bioinformatics

doi.10.1093/bioinformatics/xxxxxx

Advance Access Publication Date: Day Month Year

Application Note

OXFORD

Databases and ontologies

WTFgenes: What's The Function of these genes? Static sites for model-based gene set analysis

lan H. Holmes¹

¹Department of Bioengineering, University of California, Berkeley, CA 94720, USA

Abstract

Motivation. A common technique for interpreting experimentally-identified lists of genes is to look for enrichment of genes associated to particular Gene Ontology terms. The most common technique uses the hypergeometric distribution; more recently, a model-based approach was proposed. These approaches must typically be run using downloaded software, or on a server. Results. We develop a collapsed likelihood for model-based gene set analysis and present WTFgenes, an implementation of both hypergeometric and model-based approaches, that can be published as a static site with computation run in JavaScript on the user's web browser client. Apart from hosting, zero server resources are required: the site can (for example) be served directly from an Amazon S3 bucket. A C++11 implementation yielding identical results runs roughly twice as fast as the JavaScript version. Availability and Implementation. WTFgenes is available from https://github.com/evoldoers/wtfgenes under the BSD3 license. Contact. lan Holmes ihholmes+wtfgenes@gmail.com. Supplementary Information. None.

Introduction

Gene Set Enrichment Analysis (GSEA) is a common technique for finding patterns, such as over-represented ontology terms, in a set of experimentally identified genes (Boyle *et al.*, 2004). The typical approach is a one-tailed Fisher's Exact Test (based on the hypergeometric distribution, which models the number of term-associations if the gene set was chosen by chance), with a suitable correction for multiple hypothesis testing. The GSEA has been implemented many times on various platforms (Robinson *et al.*, 2002; Khatri *et al.*, 2002; Zeeberg *et al.*, 2003; Boyle *et al.*, 2004; Bauer *et al.*, 2008; Jiao *et al.*, 2012; Chen *et al.*, 2013).

A Bayesian alternative to GSEA, which more directly addresses some of the multiple testing issues (for example, by modeling the ways that an observed gene list can be broken down into complementary gene sets), is Model-based Gene Set Analysis (MGSA). In contrast to GSEA, which just rejects a null hypothesis that genes are chosen by chance, the MGSA approach explicitly models the alternative hypothesis that the gene set was generated from a few random ontology terms. This approach was introduced by Lu *et al.* (2008) and developed by Bauer *et al.* (2010), who demonstrated by simulation its increased statistical power when the observed gene set is indeed generated by a small number of terms. It has been implemented in Java (Bauer *et al.*, 2010) and R (Bauer *et al.*, 2011), but remains significantly less well-developed than GSEA.

The graphical model underpinning MGSA is sketched in Figure 1. For each of the m terms there is a boolean random variable T_j ("term j is activated"). For each of the n genes there is a directly-observed

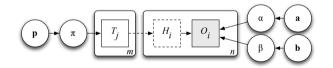


Fig. 1. Model-based explanation of observed genes (O_i) using ontology terms (T_j) , following Bauer et al. (2010). Other variables and hyperparameters are defined in the text. Circular nodes indicate continuous-valued variables or hyperparameters; square nodes indicate discrete-valued (boolean) variables. Dashed lines indicate deterministic relationships; shaded nodes indicate observations. Plates (rounded rectangles) indicate replication.

boolean random variable O_i ("gene i is observed in the gene set"), and one deterministic boolean variable H_i ("gene i is activated") defined by $H_i = 1 - \prod_{j \in G_i} T_j$ where G_i is the set of terms associated with gene i (including directly annotated terms, as well as ancestral terms implied by transitive closure of the directly annotated terms). The probability parameters are π (term activation), α (false positive) and β (false negative), and the respective hyperparameters are $\mathbf{p} = (p_0, p_1)$, $\mathbf{a} = (a_0, a_1)$ and $\mathbf{b} = (b_0, b_1)$. The model is

$$P(T_j = 1|\pi) = \pi$$

$$P(O_i = 1|H_i = 0, \alpha) = \alpha$$

$$P(O_i = 1|H_i = 1, \beta) = 1 - \beta$$

© The Author 2016. Published by Oxford University Press. All rights reserved. For permissions, please e-mail: journals.permissions@oup.com









2 lan H. Holmes

with $\pi \sim \text{Beta}(\mathbf{p})$, $\alpha \sim \text{Beta}(\mathbf{a})$ and $\beta \sim \text{Beta}(\mathbf{b})$. The model of Bauer et al. (2010) is similar but used an ad hoc discretized prior for π , α and β .

Most MGSA and GSEA implementations are designed for desktop use. Several GSEA implementations are designed for the web, such as DAVID-WS (Jiao *et al.*, 2012) and Enrichr (Chen *et al.*, 2013; Gundersen *et al.*, 2015; Kuleshov *et al.*, 2016) which has a rich dynamic web front-end. However, web-facing GSEA implementations generally require a server-hosted back end that executes code. Further, there are no web-based MGSA implementations.

Results

In developing our MGSA sampler, we introduce a collapsed version of the model in Figure 1 by integrating out the probability parameters. Let $c_p = \sum_j^m T_j$ count the number of activated terms, $c_g = \sum_i^n H_i$ the activated genes, $c_a = \sum_i^n O_i (1-H_i)$ the false positives and $c_b = \sum_i^n O_i H_i$ the false negatives. Then

$$P(\mathbf{T}, \mathbf{O}|\mathbf{a}, \mathbf{b}, \mathbf{p}) = Z(c_p; m, \mathbf{p}) Z(c_a; n - c_g, \mathbf{a}) Z(c_b; c_g, \mathbf{b})$$

where

$$Z(k; N, \mathbf{A}) = \binom{N}{k} \frac{B(k + A_0, N - k + A_1)}{B(A_0, A_1)}$$

is the beta-binomial distribution for k successes in N trials with pseudocounts $\mathbf{A} = (A_0, A_1)$, using the beta function

$$B(x,y) = \int_0^1 t^{x-1} (1-t)^{y-1} dt = \frac{\Gamma(x)\Gamma(y)}{\Gamma(x+y)}$$

Integrating out probability parameters improves sampling efficiency and allows for higher-dimensional models where, for example, we observe multiple gene sets and give each term its own probability π_j or each gene its own error rates (α_i,β_i) . Our implementation by default uses uninformative priors with hyperparameters $\mathbf{a}=\mathbf{b}=\mathbf{p}=(1,1)$ but this can be overridden by the user.

The MCMC sampler uses a Metropolis-Hastings kernel where each proposed move perturbs some subset of the term variables. These moves include *flip*, where a single term is toggled; *step*, where any activated term and any one of its unactivated ancestors or descendants are toggled; *jump*, where any activated term and any unactivated term are toggled; and *randomize*, where all term variables are uniformly randomized. The relative rates of these moves can be set by the user.

The sampler of Bauer *et al.* (2010) implemented only the *flip* move. To test the relative efficacy of the newly-introduced moves we measured the autocorrelation of the term variables for one of the GO Project's test sets, containing 17 *S.cerevisiae* mating genes. ¹ The results, shown in Figure 2, led us to set the MCMC defaults such that the *flip*, *step*, and *jump* moves are equiprobable, while *randomize* is disabled.

We have implemented both GSEA (with Bonferroni correction) and MGSA (as described above), in both C++11 and JavaScript. The JavaScript version can be run as a command-line tool using node, or via a web interface in a browser, and includes extensive unit tests. The two implementations use the same random number generator and yield numerically identical results. The C++ version is about twice as fast: a benchmark of MGSA on a late-2014 iMac (4GHz Intel Core i7), using the abovementioned 17 yeast mating genes and the relevant subset of 518 GO terms, run for 1,000 samples per term, took 37.6 seconds of user time for the C++ implementation and 79.8 seconds in JavaScript. By contrast, the GSEA approach is almost instant.

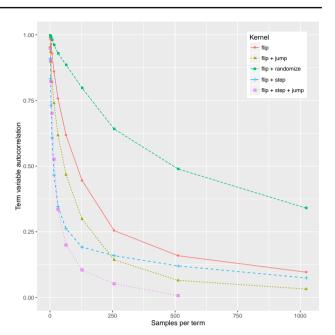


Fig. 2. Autocorrelation of term variables, as a function of the number of MCMC samples, for several MCMC kernels on a set of 17 S.cerevisiae mating genes. A rapidly-decaying curve indicates an efficiently-mixing kernel. The kernel incorporating flip, step and jump moves (defined in the text) mixes most efficiently.

Our JavaScript version of MGSA, when used as a web application, includes visual feedback on the log-likelihood during a sampling run. It can be deployed as a "static site", i.e. consisting only of static files (HTML, CSS, JSON, and JavaScript) that can be hosted via a minimal web server with no need for dynamic code execution. This has considerable advantages: static web hosting is generally much cheaper, and far more secure, than running server-hosted web applications.

Discussion

JavaScript genome browsers such as JBrowse (Buels *et al.*, 2016) are consistent with a broader web trend of producing static sites where possible. We have implemented such a static site approach for ontological term analysis of gene sets that offers both Bayesian (MGSA) and frequentist (GSEA) approaches. It is not intended as a direct competitor to existing GSEA services such as DAVID-WS or Enrichr, but rather as a lightweight static site-generator that allows comparison of MGSA and GSEA approaches.

The model-based approach of MGSA is versatile: it can readily be extended to applications that are structured temporally (Hejblum *et al.*, 2015), spatially (Lin *et al.*, 2015), or by genomic region (McLean *et al.*, 2010); that use domain-specific biological knowledge (Szczurek and Beerenwinkel, 2014); or that incorporate additional lines of evidence such as quantitative data (Kalaitzis and Lawrence, 2011). We hope our development of a collapsed likelihood for MGSA, and evaluation of different MCMC kernels, will assist these efforts.

Funding

IHH was partially supported by NHGRI grant R01-HG004483.



¹ Gene IDs: STE2, STE3, STE5, GPA1, SST2, STE11, STE50, STE20, STE4, STE18, FUS3, KSS1, PTP2, MSG5, DIG1, DIG2, STE12.





WTFgenes 3

References

- Bauer, S., Grossmann, S., Vingron, M., and Robinson, P. N. (2008). Ontologizer 2.0–a multifunctional tool for GO term enrichment analysis and data exploration. *Bioinformatics*, 24(14), 1650–1651.
- Bauer, S., Gagneur, J., and Robinson, P. N. (2010). GOing Bayesian: model-based gene set analysis of genome-scale data. *Nucleic Acids Res.*, **38**(11), 3523–3532.
- Bauer, S., Robinson, P. N., and Gagneur, J. (2011). Model-based gene set analysis for Bioconductor. *Bioinformatics*, 27(13), 1882–1883.
- Boyle, E. I., Weng, S., Gollub, J., Jin, H., Botstein, D., Cherry, J. M., and Sherlock, G. (2004). GO::TermFinder-open source software for accessing Gene Ontology information and finding significantly enriched Gene Ontology terms associated with a list of genes. *Bioinformatics*, 20(18), 3710–3715.
- Buels, R., Yao, E., Diesh, C. M., Hayes, R. D., Munoz-Torres, M., Helt, G., Goodstein, D. M., Elsik, C. G., Lewis, S. E., Stein, L., and Holmes, I. H. (2016). JBrowse: a dynamic web platform for genome visualization and analysis. *Genome Biol.*, 17, 66.
- Chen, E. Y., Tan, C. M., Kou, Y., Duan, Q., Wang, Z., Meirelles, G. V., Clark, N. R., and Ma'ayan, A. (2013). Enrichr: interactive and collaborative HTML5 gene list enrichment analysis tool. *BMC Bioinformatics*, 14, 128.
- Gundersen, G. W., Jones, M. R., Rouillard, A. D., Kou, Y., Monteiro, C. D., Feldmann, A. S., Hu, K. S., and Ma'ayan, A. (2015). GEO2Enrichr: browser extension and server app to extract gene sets from GEO and analyze them for biological functions. *Bioinformatics*, **31**(18), 3060–3062
- Hejblum, B. P., Skinner, J., and Thiebaut, R. (2015). Time-Course Gene Set Analysis for Longitudinal Gene Expression Data. *PLoS Comput. Biol.*, 11(6), e1004310.
- Jiao, X., Sherman, B. T., Huang, d. a. W., Stephens, R., Baseler, M. W., Lane, H. C., and Lempicki, R. A. (2012). DAVID-WS: a stateful web service to facilitate gene/protein list analysis. *Bioinformatics*, 28(13),

- 1805-1806.
- Kalaitzis, A. A. and Lawrence, N. D. (2011). A simple approach to ranking differentially expressed gene expression time courses through Gaussian process regression. BMC Bioinformatics, 12, 180.
- Khatri, P., Draghici, S., Ostermeier, G. C., and Krawetz, S. A. (2002). Profiling gene expression using onto-express. *Genomics*, 79(2), 266–270.
- Kuleshov, M. V., Jones, M. R., Rouillard, A. D., Fernandez, N. F., Duan, Q., Wang, Z., Koplev, S., Jenkins, S. L., Jagodnik, K. M., Lachmann, A., McDermott, M. G., Monteiro, C. D., Gundersen, G. W., and Ma'ayan, A. (2016). Enrichr: a comprehensive gene set enrichment analysis web server 2016 update. *Nucleic Acids Res.*, 44(W1), W90–97.
- Lin, Z., Sanders, S. J., Li, M., Sestan, N., State, M. W., and Zhao, H. (2015). A Markov random field-based approach to characterizing human brain development using spatial-temporal transcriptome data. *Ann Appl Stat*, **9**(1), 429–451.
- Lu, Y., Rosenfeld, R., Simon, I., Nau, G. J., and Bar-Joseph, Z. (2008).
 A probabilistic generative model for GO enrichment analysis. *Nucleic Acids Res.*, 36(17), e109.
- McLean, C. Y., Bristor, D., Hiller, M., Clarke, S. L., Schaar, B. T., Lowe, C. B., Wenger, A. M., and Bejerano, G. (2010). GREAT improves functional interpretation of cis-regulatory regions. *Nat. Biotechnol.*, 28(5), 495–501.
- Robinson, M. D., Grigull, J., Mohammad, N., and Hughes, T. R. (2002).
 FunSpec: a web-based cluster interpreter for yeast. *BMC Bioinformatics*,
 3. 35.
- Szczurek, E. and Beerenwinkel, N. (2014). Modeling mutual exclusivity of cancer mutations. *PLoS Comput. Biol.*, 10(3), e1003503.
- Zeeberg, B. R., Feng, W., Wang, G., Wang, M. D., Fojo, A. T., Sunshine, M., Narasimhan, S., Kane, D. W., Reinhold, W. C., Lababidi, S., Bussey, K. J., Riss, J., Barrett, J. C., and Weinstein, J. N. (2003). GoMiner: a resource for biological interpretation of genomic and proteomic data. *Genome Biol.*, **4**(4), R28.



