



# PO: Precision Oncology Course

## Alignments



# Alignments

What is an alignment?

**Sequence alignments** are a way of arranging the sequences of DNA, RNA or proteins in order to **identify regions of similarity** that may be a consequence of functional, structural, or evolutionary relationships between the sequences.

```
ACGTCCTTGACTGG - TTAAAATAC
AC - TCTTGACTGGATTAAACATAC
```

# Alignments

## Elements of an alignment

Alignment seeks to **reduce gaps and mismatches** and **maximize matches**.

```
ACGTTTGCAGTAAATGCGGACTGA - T
ACGTTGTGCAGTAAATGCGGGA -- GACT
```

↓  
mismatch  
(SNVs)

↓  
match

↓ ↓  
gap  
(indels)

# Alignments

## Elements of an alignment

In the construction, each of these components has a penalty value associated. For gaps there is a penalty value for opening the gap and another for extending it.

ACGTTTGCAGTAAATGCGGA<sup>CT</sup>GAT  
ACGTTGTGCAGTAAATGCGGA<sup>-</sup>GACT

→ 1 gap 3 mismatches

ACGTTTGCAGTAAATGCGGA<sup>CT</sup>GAT  
ACGTTGTGCAGTAAATGCGGA<sup>--</sup>GACT

→ 1 extended gap 1 mismatch

ACGTTTGCAGTAAATGCGGA<sup>CT</sup>GA<sup>-</sup>T  
ACGTTGTGCAGTAAATGCGGA<sup>--</sup>GACT

→ 2 gaps

# Alignments

## Objectives

The comparison between sequences in sequence alignment allows to:

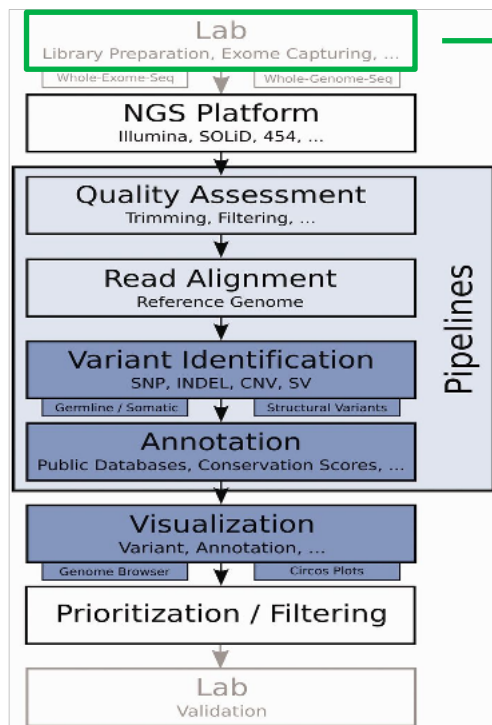
1. Find homologous **positions**
2. Determine the **homology** degree
3. Identify **functional domains**
4. **Compare** the gene with its product
5. Identify **differences**



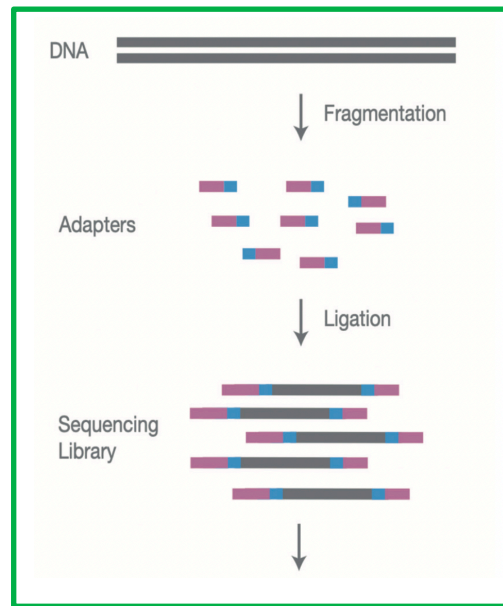
Objective of Variant Calling in Next  
Generation Sequencing (NGS)

# Next Generation Sequencing (NGS)

## Workflow

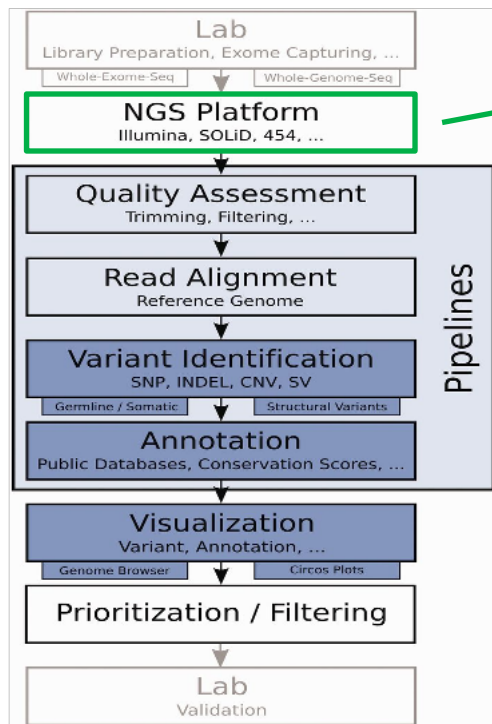


**Library Preparation:**  
DNA is fragmented into small pieces to form a **library**.

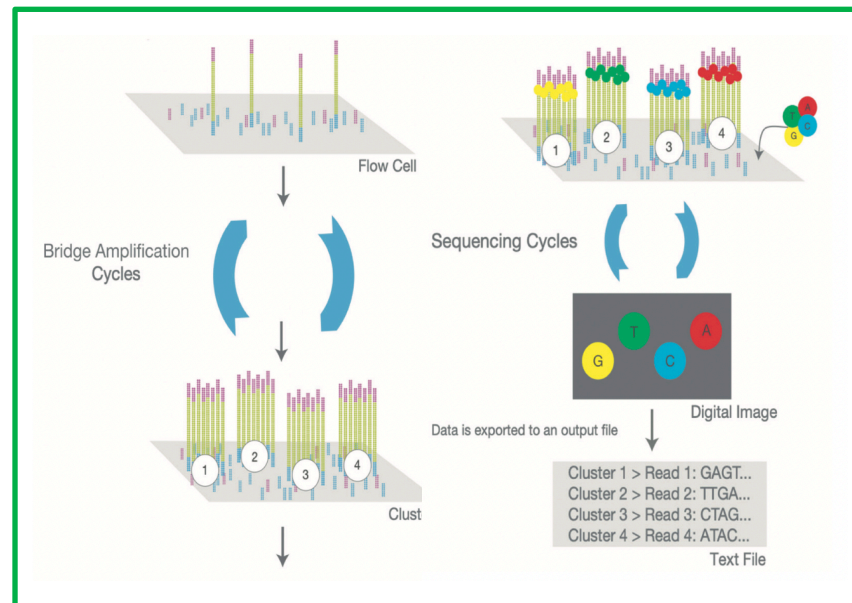


# Next Generation Sequencing (NGS)

## Workflow

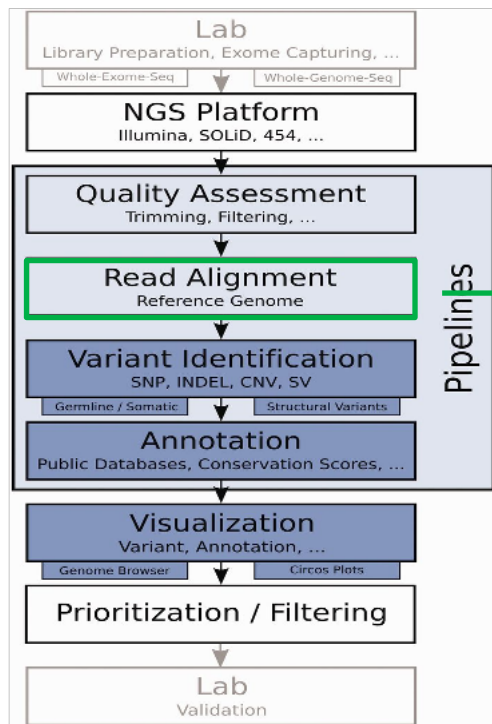


**Sequencing:**  
Fragments are sequenced and stored as **reads** in text files.



# Next Generation Sequencing (NGS)

## Workflow



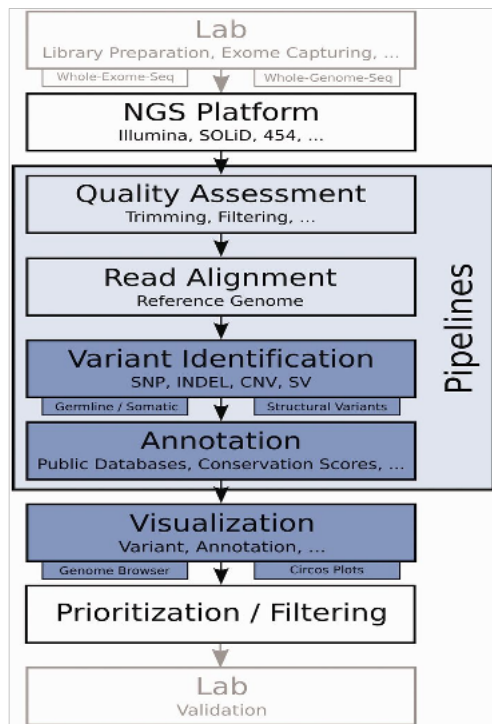
### Alignment:

Align the **reads** against the **reference genome** in order to find out where do they come from.



# Next Generation Sequencing (NGS)

## Workflow



Video: <https://youtu.be/fCd6B5HRaZ8>

# Short Read Aligners (SRA)

## Challenges

Need to **align billions of reads** to a very **large reference genome**: SRA must be **extraordinarily efficient algorithms**

- Speed
- Memory use

As we need to align short reads, **a read may align in multiple positions**

- Either report multiple positions
- Or pick heuristically one of them

**Different NGS technologies** have different error profiles to take into account:

- **Roche 454**: Insertions or deletions in homopolymer runs
- **Illumina**: Increasing likelihood of sequence errors towards the end of the read

# Alignment

## Reads

Short reads are stored in **text files** with the extension **FASTQ**.

Identifier	●	@SRR566546.970 HWUSI-EAS1673_11067_FC7070M:4:1:2299:1109 length=50
Sequence	●	TTGCCTGCCTATCATTTTAGTGCCTGTGAGGTGGAGATGTGAGGATCAGT
'+' sign	●	+
Quality scores	●	hhhhhhhhhhghghghhhhhfhhhhhfffffe'ee['X]b[d[ed'[Y[~Y
Identifier	●	@SRR566546.971 HWUSI-EAS1673_11067_FC7070M:4:1:2374:1108 length=50
Sequence	●	GATTTGTATGAAAGTATACAACTAAACTGCAGGTGGATCAGAGTAAGTC
'+' sign	●	+
Quality scores	●	hhhhgfhhcghghggfcffdhfehhhhcehdchhdhahehffffde'bVd

Hosseini, M.; Pratas, D.; Pinho, A.J. A Survey on Data Compression Methods for Biological Sequences. *Information* **2016**, 7, 56. doi: 10.3390/info7040056

# Alignment

## Reference genome

The reference genome is stored in another **text file** with the extension **FASTA**.

The diagram illustrates the FASTA format with three entries. Each entry consists of a header line starting with a greater-than sign (>) and a sequence line. The labels 'Header' and 'Sequence' are placed to the left of each line, with red lines connecting them to the corresponding lines in the FASTA entries. The first entry has a header '>VIT\_201s0011g03530.1' and a sequence 'AATTAAGCATAAAATACTCACTCTTACCCCCTTATTTTCTTATCTCTCATCACTTTTGGTGCGAAGGACCATGAGAACAAGCTGCAATGGGTGTAGGGTTCTTCGCAAGGCATGCAGCCAAGACTGCATCA'. The second entry has a header '>VIT\_201s0011g03540.1' and a sequence 'CAGGTAGCGTGAAGTTAAACCCTAGCGCTTTAGACAAACAGCTGTAGTCACCGCCCACAAACACCAGCCTCTGAGACACCACCTCAAACCTTTCCACTTAAATACACATCCCTCACACCCTTTTCAATTC'. The third entry has a header '>VIT\_201s0011g03550.1' and a sequence 'CATGCAAAGCTGAACGCGATGCTGTGATTGGTGGTAAGTGGTAGTTGAGTAAATTTGACAGTGAA GCCGAAATGGTAAAAGACTAAGGCTAGAAGTAGAATACCACTGTTCTTCTCATCACGTGGGCCCA'.

```
>VIT_201s0011g03530.1
AATTAAGCATAAAATACTCACTCTTACCCCCTTATTTTCTTATCTCTCATCACTTTTGGTGCGAAG
GACCATGAGAACAAGCTGCAATGGGTGTAGGGTTCTTCGCAAGGCATGCAGCCAAGACTGCATCA

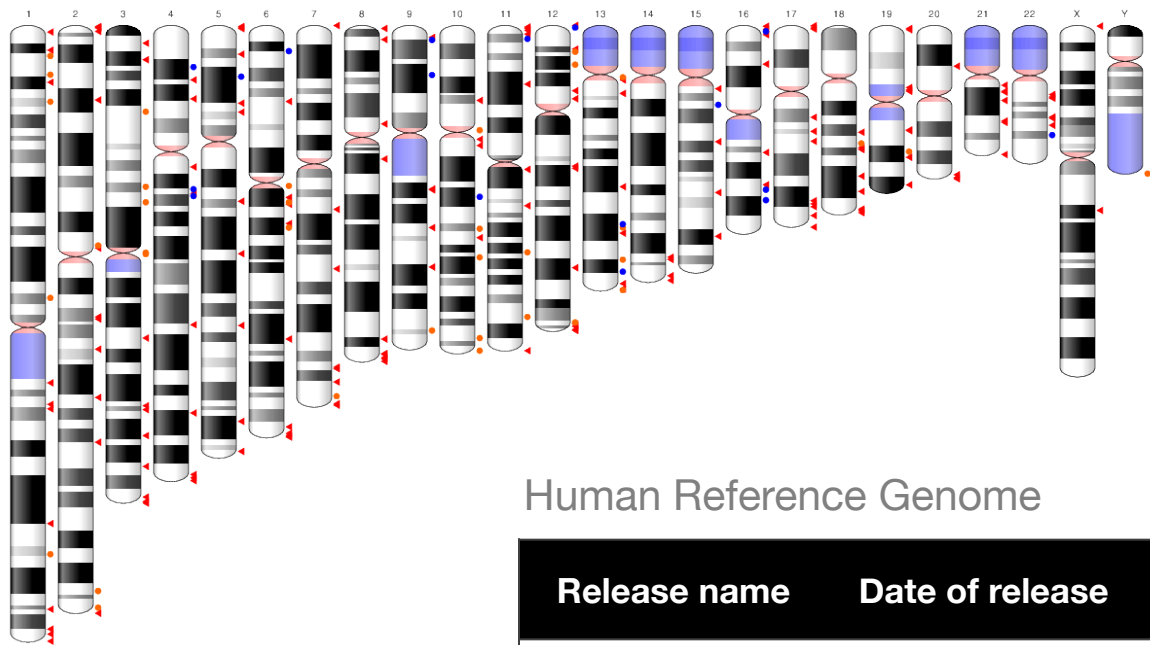
>VIT_201s0011g03540.1
CAGGTAGCGTGAAGTTAAACCCTAGCGCTTTAGACAAACAGCTGTAGTCACCGCCCACAAACACC
AGCCTCTGAGACACCACCTCAAACCTTTCCACTTAAATACACATCCCTCACACCCTTTTCAATTC

>VIT_201s0011g03550.1
CATGCAAAGCTGAACGCGATGCTGTGATTGGTGGTAAGTGGTAGTTGAGTAAATTTGACAGTGAA
GCCGAAATGGTAAAAGACTAAGGCTAGAAGTAGAATACCACTGTTCTTCTCATCACGTGGGCCCA
```

Hosseini, M.; Pratas, D.; Pinho, A.J. A Survey on Data Compression Methods for Biological Sequences. *Information* **2016**, 7, 56. doi: 10.3390/info7040056

# Alignment

## Reference genome



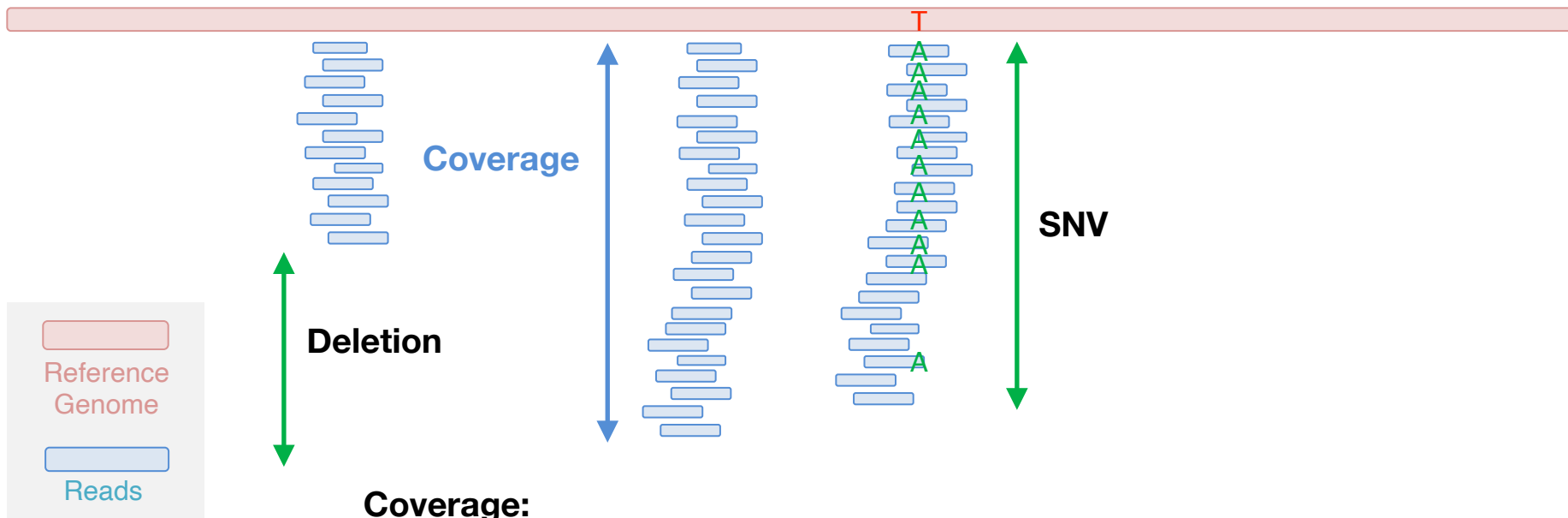
- ◄ Region containing alternate loci
- Region containing fix patches
- Region containing novel patches

## Human Reference Genome

Release name	Date of release	Equivalent UCSC version	Base Pairs
GRCh38	Dec 2013	hg38	3,609,003,417
GRCh37	Feb 2009	hg19	3,326,743,047

# Alignment

Reads are mapped against the reference genome in order to identify differences



**Coverage:**

Number of reads that align to the same region (60x, 100x,...)

# Alignment

## Indexing

In order to optimize the alignment process, these algorithms use a strategy called **indexing**. **As in books**, indexing allows to organize information in a more easier and faster way to search.

## Index

### A

Additive color model, 3

### B

Binding, 20

Bitmap image

defined, 9

resolution of, 11

tonal range in, 11

Bleed, checking, 46

Blue line, 42

### C

Chroma, 2

CMS. See Color management

system

CMY color model, 4

Color

characteristics of, 2

checking definitions, 46

Color proof

checking, 50

contract, 40, 50

separation-based, 40

Color separations. See

Separations

Color space, 4

Color value, 2

Commercial printing

inking, 18

offsetting, 19

platemaking, 17

press check, 40–43, 51

terminology, 5–8

types of, 16–??

wetting, 18

Continuous-tone art

defined, 5

Contract proof, 40, 50

Creep, 21

### G

Gamut, color, 4

GCR (gray-component

replacement), 7

Gravure, 22

Gray, shades of, 13

### H

Halftone cell, 13

Halftone dot, 5, 13

Halftone frequency, 12

Halftone screen

defined, 5

moiré patterns, 9, 15

process colors, 6

Hand-off, 37

creating report for, 43

organizing files for, 46

High-fidelity color, 15

Hue, 2

### Position N

### Position 2

CTGC CGTA AACT AATG

### Position 1

ACTG CCGT AAAC TAAT

ACTG \*\*\*\* AAAC \*\*\*\*

\*\*\*\* CCGT \*\*\*\* TAAT

ACTG \*\*\*\* \*\*\*\* TAAT

\*\*\*\* \*\*\*\* AAAC TAAT

ACTG CCGT \*\*\*\* \*\*\*\*

\*\*\*\* CCGT AAAC \*\*\*\*

# Alignment

## Elements to consider

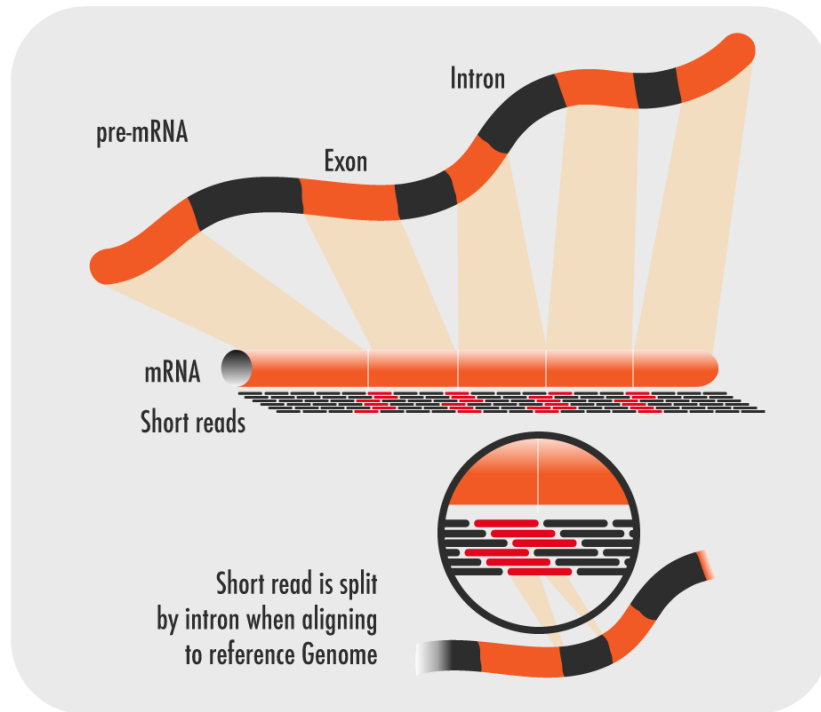
- Read type: DNA, RNA, etc.
- Read length
- Paired-end or single-end
- Computational requirements (number of processors, memory)
- Number of mismatches (limitation in allowed differences)
- Sequencing errors (can be platform dependent)



# Alignment

## Elements to consider

- Read type: DNA, RNA, etc.



Mackenzie RJ. RNA-seq: Basics, Applications and Protocol. *Technology Networks Genomic Research*, **2018**. <https://www.technologynetworks.com/genomics/articles/rna-seq-basics-applications-and-protocol-299461>

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# Alignment

Elements to consider

- Read length

The longer the reads, the higher the likelihood of sequencing errors towards the end.

# Alignment

## Elements to consider

- Read type: DNA, RNA, etc.
- Read length
- Paired-end or single-end
- Computational requirements (number of processors, memory)
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# Alignment

## Elements to consider

- Paired-end or single-end

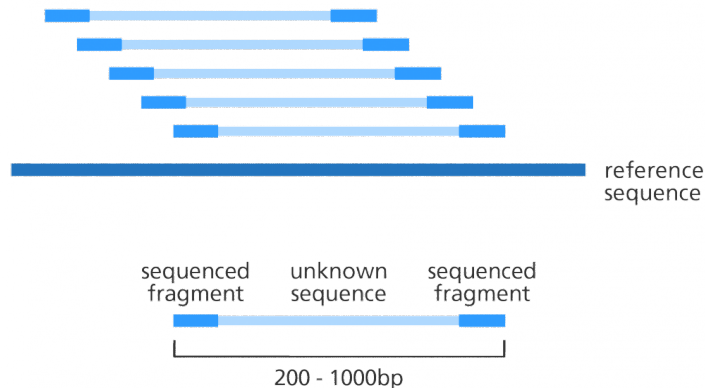
In a **paired-end** experiment, **both ends** of the fragment are sequenced.

The **distance between** the each paired read is **known (insert size)**, thus the aligners can **map** the reads **more precisely**. Important for correctly mapping **repetitive regions** of the genome.

Single-end reads



Paired-end reads



# Alignment

## Elements to consider

- Read type: DNA, RNA, etc.
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# Alignment

## Errors and biases

- Sequencing errors:
  - Increases mismatches
  - Higher at the end of the reads
  - Technology-dependent
- Different regions in the DNA sequence cause aligning biases:
  - Repetitive regions:
    - Found in different locations
    - Place of sequencing errors
    - Place of real mutations and structural variants
  - Difficulties in the alignment of indels (gaps)

# Alignment

## Solutions

Post-alignment Quality Control and local realignment of indels.

**More about indel realignment:** <https://qcb.ucla.edu/wp-content/uploads/sites/14/2016/03/GATKwr12-3-IndelRealignment.pdf>



# Thanks!



*cnio* stop cancer