

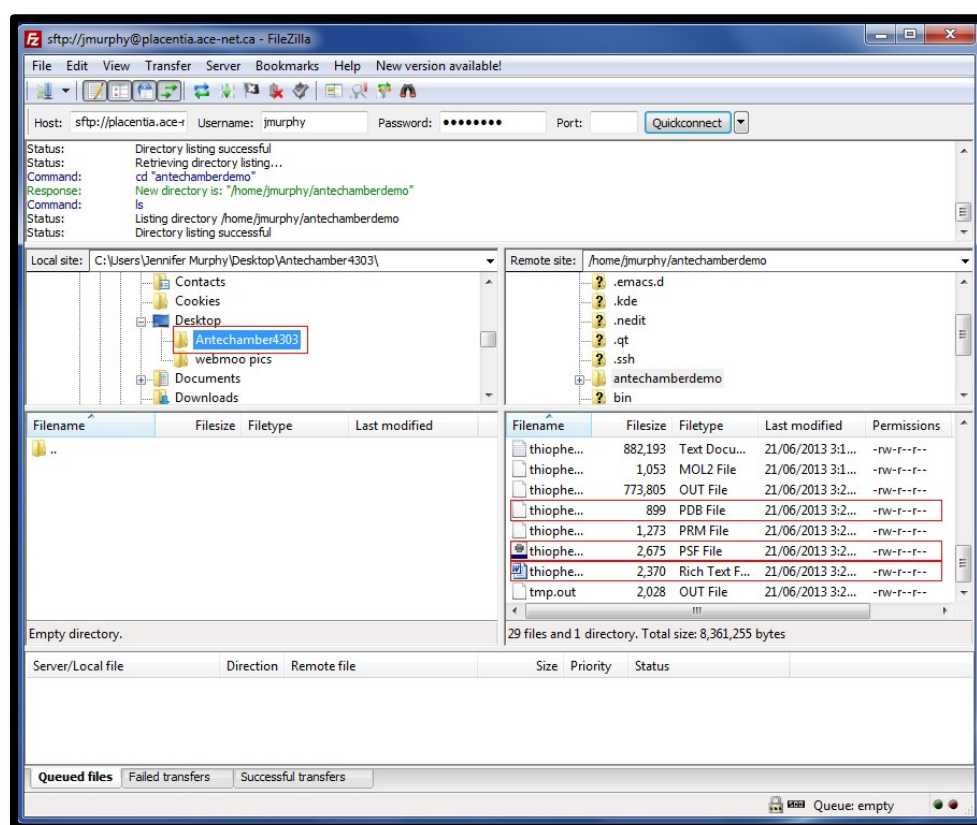
CHEM 4305 Term Project:

Simulating Molecular Liquids

Part 2: Building a Molecular Liquid and Equilibrating the System

18. Get your .psf, .pdb and .rtf back onto the desktop using Filezilla.

This is similar to Step 9. Open **Filezilla** and make a folder on your desktop to transfer the files to.



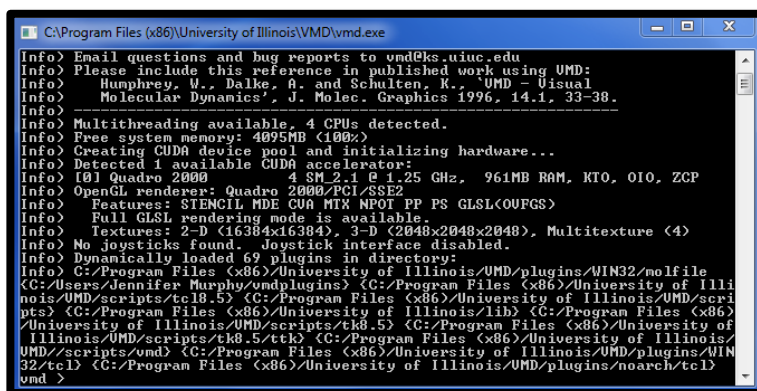
19. Download the liquefy.tcl plugin

Enter the URL: <http://www.chem.mun.ca/homes/cnrhome/liquefy.tcl> into your web browser. Save the file to your computer.

19. Open VMD from the desktop

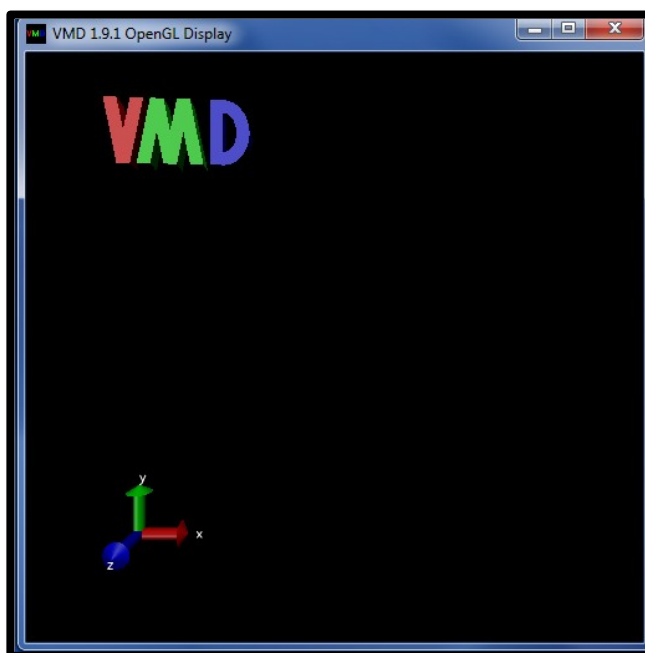
Find the program “VMD” in your Program Files directory under “University of Illinois.” There should be three windows that come up, shown below.

Command
Window

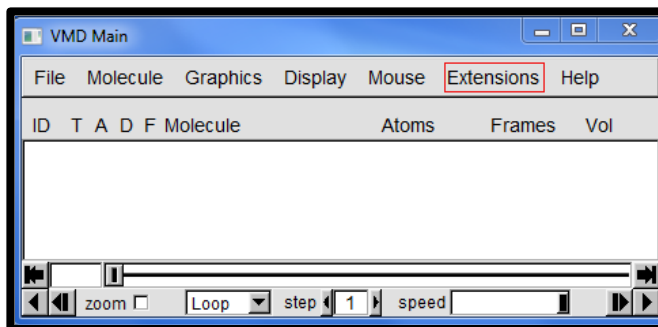


```
C:\Program Files (x86)\University of Illinois\VMD\vmd.exe
Info> Email questions and bug reports to vmd@ks.uiuc.edu
Info> Please include this reference in published work using VMD:
Info> Humphrey, W., Dalke, A. and Schulten, K., 'VMD - Visual
Info> Molecular Dynamics', J. Molec. Graphics 1996, 14.1, 33-38.
Info>
Info> Multithreading available, 4 CPUs detected.
Info> Free system memory: 4095MB (100%)
Info> Creating CUDA device pool and initializing hardware...
Info> Detected 1 available CUDA accelerator:
Info> [0] Quadro 2000 4 SM 2.1 @ 1.25 GHz, 961MB RAM, KTO, OIO, ZCP
Info> OpenGL renderer: Quadro 2000/PCI/SSE2
Info> Features: STENCIL MDE CUA MTX NPOT PP PS GLSL<OUFFGS>
Info> Full GLSL rendering mode is available.
Info> Textures: 2-D (16384x16384), 3-D (2048x2048x2048), Multitexture (4)
Info> No joysticks found. Joystick interface disabled.
Info> Dynamically loaded 69 plugins in directory:
Info> C:/Program Files (x86)/University of Illinois/VMD/plugins/VIN32/molfile
Info> C:/Users/Jennifer Murphy/vmdplugins C:/Program Files (x86)/University of Illi
nois/VMD/scripts/tcl8.5 C:/Program Files (x86)/University of Illinois/VMD/scri
pts C:/Program Files (x86)/University of Illinois/lib C:/Program Files (x86)
/University of Illinois/VMD/scripts/tk8.5 C:/Program Files (x86)/University of
Illinois/VMD/scripts/tk8.5/ttk C:/Program Files (x86)/University of Illinois/
VMD/scripts/vmd C:/Program Files (x86)/University of Illinois/VMD/plugins/VIN
32/tcl C:/Program Files (x86)/University of Illinois/VMD/plugins/noarch/tcl
vmd >
```

Graphics
Window



Program
Window

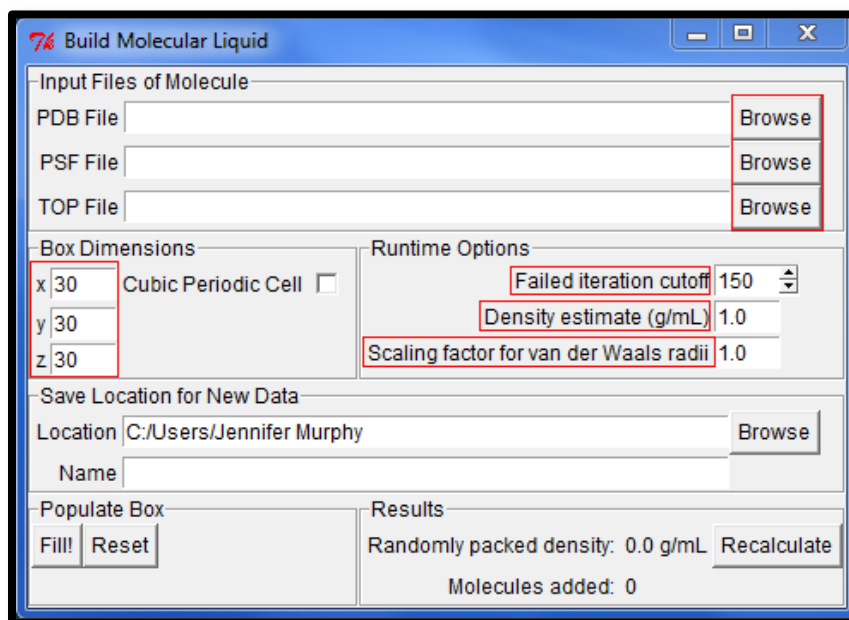


20. Execute the Liquefy Script

In the command window, type the following commands:

```
vmd > source Downloads/liquefy.tcl  
vmd > Liquefy::liquefy_gui
```

A new window will appear. You will use this extension to build a unit cell of your liquid.



Browse to put in the **.pdb**, **.psf** and **.rtf** files that you have transferred over from placentia in the “Input Files of Molecule” section. For your project, change x, y, and z lengths under “Box Dimensions” to 40 Å × 40 Å × 40 Å. Enter the density of your liquid in the “Density estimate” box. Give your liquid an appropriate name and be sure you save it to the same folder as your other files. When you are sure everything is put in the plugin correctly, click **fill** to construct the unit cell of randomly oriented molecules. Click the “Fill!” button to generate a unit cell for your simulation.

You may need to make several attempts to generate a unit cell with the correct density. You can increase the number of iterations up to a maximum of 1000 and the scaling for the van der Waals radii can be decreased to a minimum of 0.75 if your density is too low. It is OK if your density is too low – the box size will decrease during the simulation to achieve the optimal density.

76 Build Molecular Liquid

Input Files of Molecule

PDB File

PSF File

TOP File

Box Dimensions

x Cubic Periodic Cell ☒

y

z

Runtime Options

Failed iteration cutoff

Density estimate (g/mL)

Scaling factor for van der Waals radii

Save Location for New Data

Location

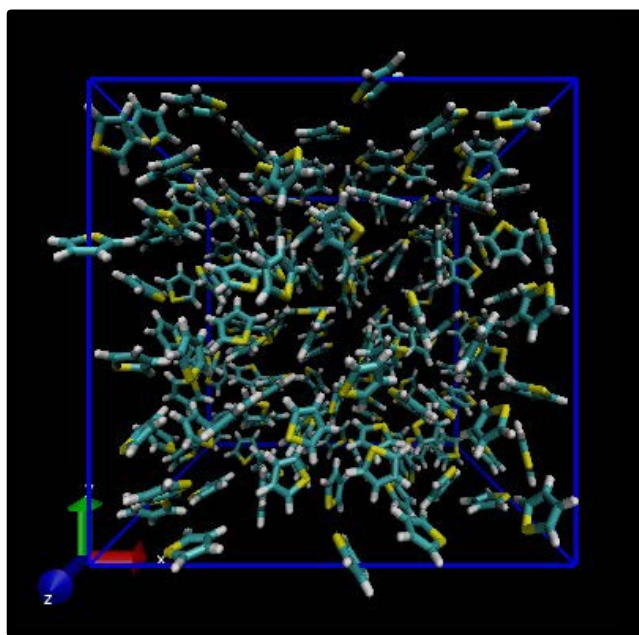
Name

Populate Box

Results

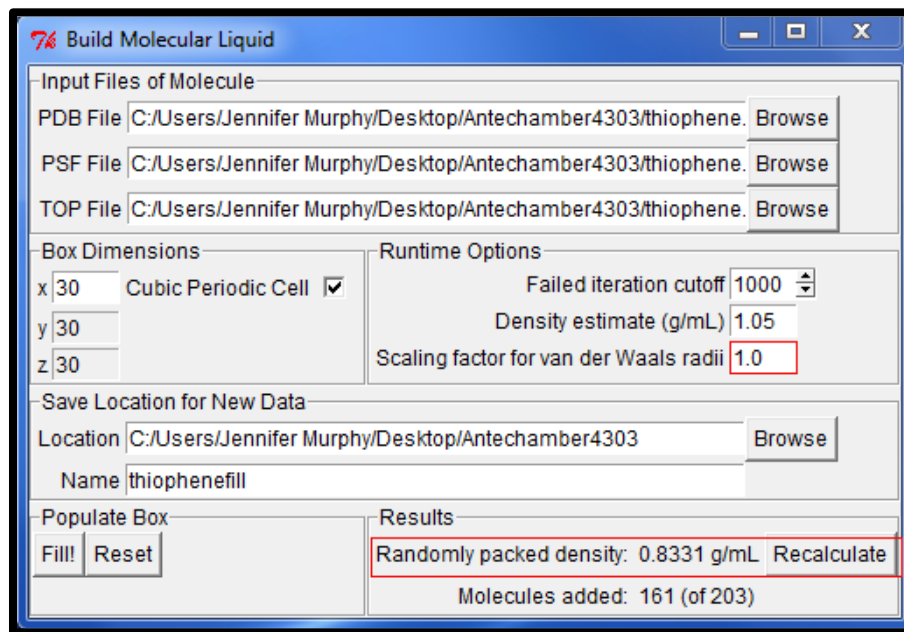
Randomly packed density: 0.0 g/mL

Molecules added: 0



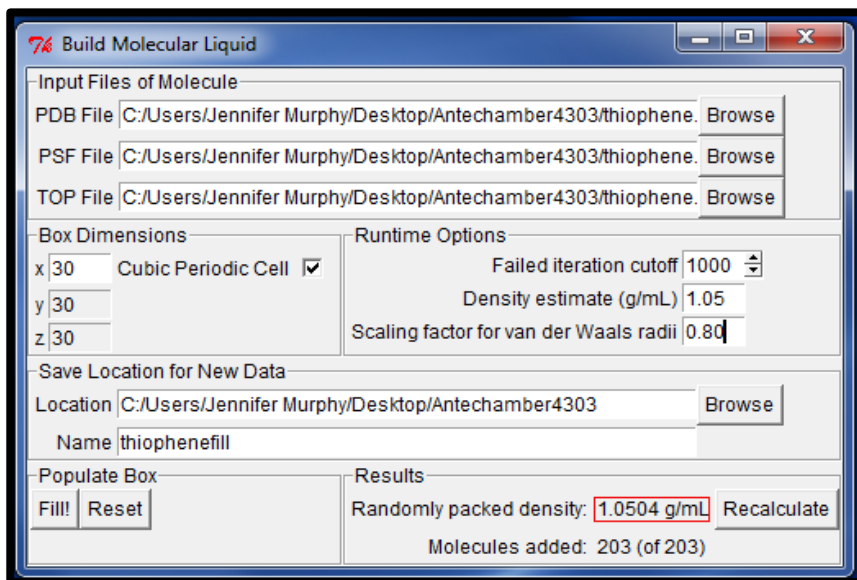
21. Check your density value

Your box should be filled with the molecule like the picture shown above. Look at the main VMD window to see what the density of the liquid is. It's OK if your density differs somewhat from the experimental density; the equilibration simulation will correct this.



As you can see, the **density** is only 0.8331 g/mL. Now we can play around with the scaling factor for van der Waals radii to get the density approximation closer to the experimental value. Also, only 161 molecules out of a possible 203 based on the dimensions of the box were used, this should be higher. You can start by changing the scaling factor to 0.80 and filling the box again.

In this case, the density after altering the scaling factor works out to **1.0504 g/mL** and **all 203 molecules are used**. This may not be true for your liquid and you may have to play around with the scaling factor before you get a density that is close to the experimental value.



22. Copy the sample NAMD configuration file to your directory on placencia

Edit the NAMD file so that it matches your molecule.

```
$ cp /home/crowley/chm4305/moleculefill.namd .
$ nano moleculefill.namd
```

Your input file will look like the following. You should edit the lines like “moleculefile.pdb” to reflect the names of the PSF and PDB files generated using Liquefy and the prm file generated using antechamber.

```
# input
coordinates          moleculefill.pdb
extendedSystem       moleculefill.xsc
structure            moleculefill.psf
parameters           molecule.prm
paratypecharm        on

# output
set output           moleculefill
outputname            $output
dcdfile              ${output}.dcd
xstFile              ${output}.xst
dcdfreq              10000
xstFreq              10000
binaryoutput         no

binaryrestart         no
outputEnergies        100
restartfreq           1000

fixedAtoms           off

# Basic dynamics
exclude              scaled1-4
1-4scaling            1
COMmotion            no
dielectric            1.0

# Simulation space partitioning
switching             on
switchdist            9
cutoff                10
pairlistdist          12

# Multiple timestepping
firsttimestep         0
timestep              1
```

```

stepspercycle      20
nonbondedFreq      2
fullElectFrequency  4

# Temperature control

set temperature      298
temperature          $temperature; # initial temperature

# Langevin Dynamics
langevin            on; # do langevin dynamics
langevinDamping     1; # damping coefficient (gamma) of 1/ps
langevinTemp        $temperature; # bath temperature
langevinHydrogen    no; # don't couple langevin bath to hydrogens
seed                12345

# Pressure control
langevinPiston      on
langevinPistonTarget 1.01325; # in bar -> 1.01325 bar = 1 atm
langevinPistonPeriod 2000
langevinPistonDecay  100
langevinPistonTemp   $temperature
useFlexibleCell      no
useGroupPressure      no
fixedAtomsForces      off

# PBC
wrapAll              on
PME                  yes
PMEGridSpacing       1.0

# Scripting
minimize             1000
reinitvels           $temperature
run                  1000000

```

23. Submit the NAMD simulation to the queue on placencia

```
$ nano script-2.sh

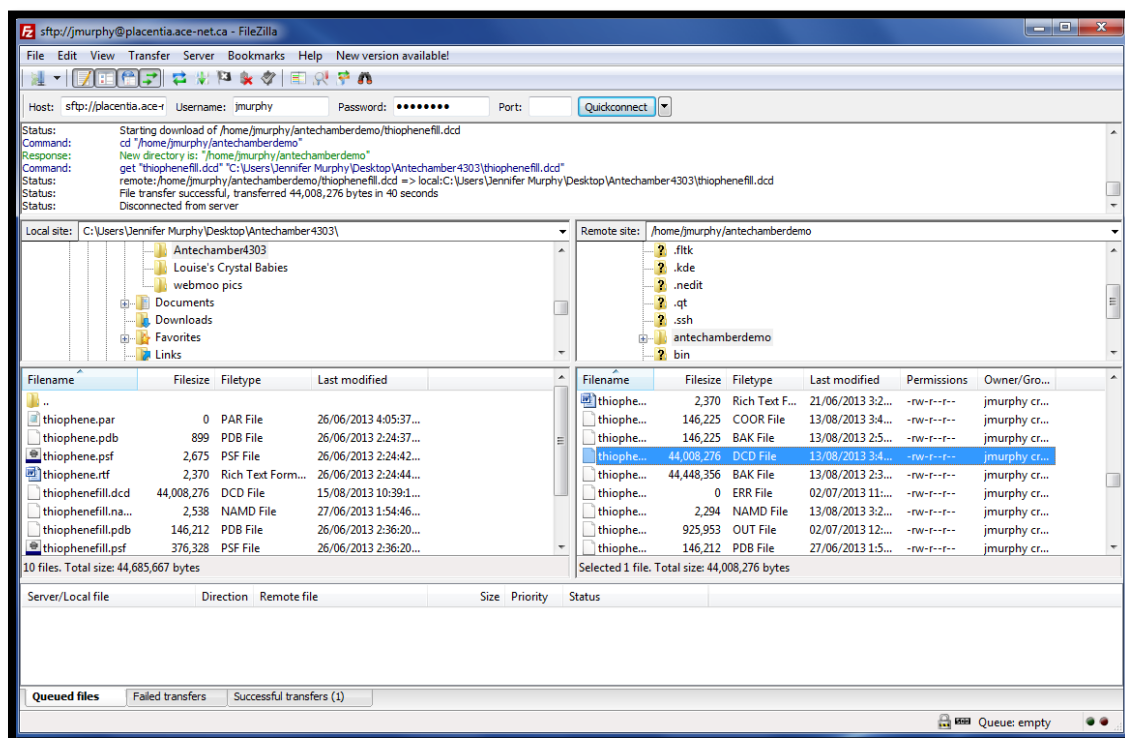
#!/bin/bash
#$ -l h_rt=24:00:00
#$ -cwd

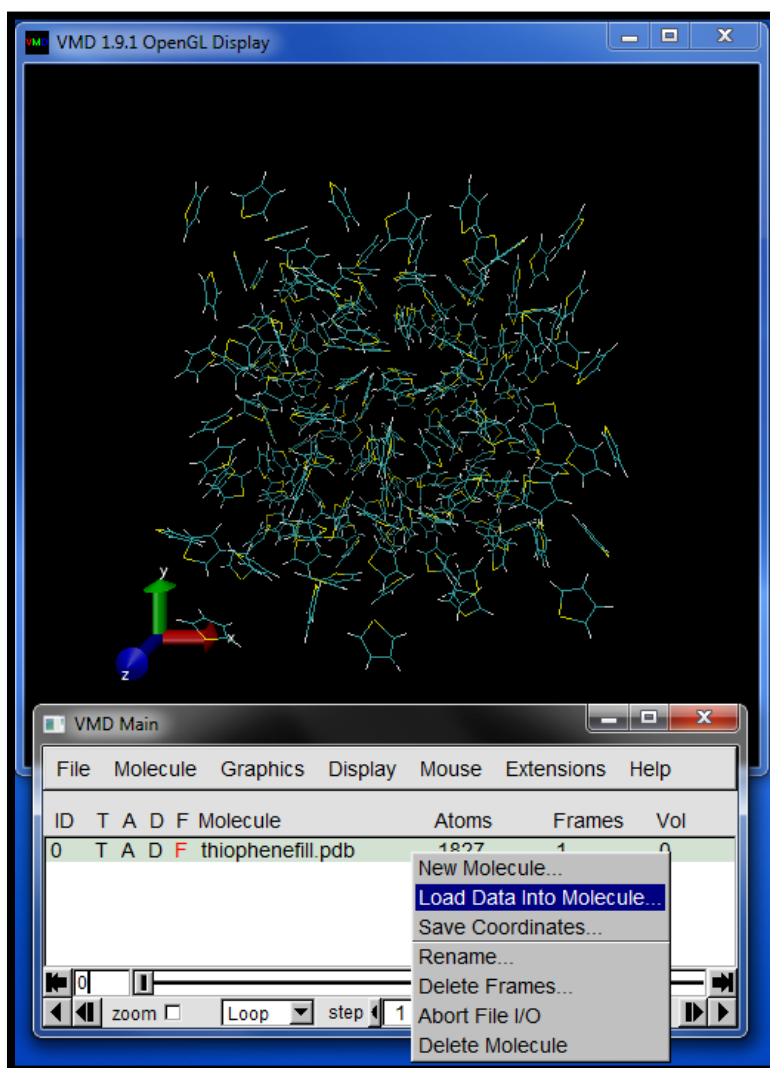
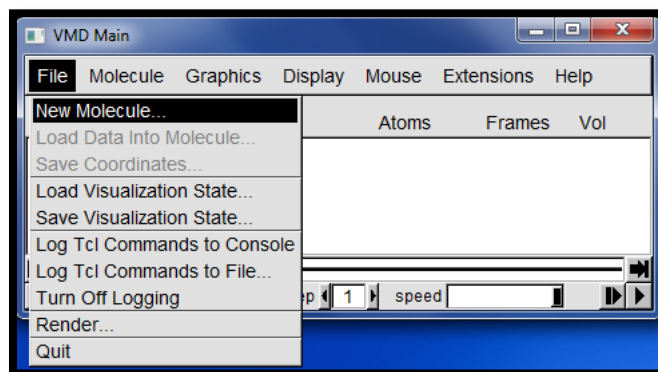
module load namd
namd2 moleculefill.namd 1> moleculefill.out 2> moleculefill.err

$ qsub script-2.sh
```

24. Lights, camera, action!

It is time to check out your movie! The trajectory of your MD simulation will be saved to a DCD file in this directory. To view the trajectory, use filezilla to transfer your .dcd file to your desktop. Open vmd and load your moleculefill.psf. Then, right click on the moleculefill.psf file so that the whole line turns green and a drop down menu comes down. Select 'load data onto molecule' and browse for the thiophenefill.dcd file, click 'load'.





25. Extracting energies/volumes and making graphs in gnuplot.

Use the following commands to extract the potential and total energy from your .namd output file.

```
$ grep "^ENERGY": moleculefill.out > energy_eq.txt
```

You need to look at your .namd output files to see what columns the potential and total energy are in. The potential energy is in 14 and total energy is in 12. An example, from the end of an output file is shown below. The timestep is in column 2.

ETITLE:	TS	BOND	ANGLE	DIHED	IMPRP	
ELECT	VDW					
BOUNDARY	MISC	KINETIC		TOTAL	TEMP	
POTENTIAL	TOTAL3	TEMP				
AVG	PRESSURE	GPRESSURE	VOLUME	PRESSAVG	GPRESSAVG	
ENERGY:	100000	2418.5540	435.0343	304.9260	35.2647	-
1108.3803	-330.4578					
0.0000	0.0000	1580.1563		3335.0973	290.1554	
1754.9410	3354.8077	291.0				
053	1278.5353	751.4656	27477.3849	15.7136	3.1915	

Write a script for your graphs, copy and paste the one below.

```
$ nano liquidenergygraphs_eq
```

```
# Gnuplot script file for plotting data in file "energy_eq.txt"
# This file is called liquidenergygraphs_eq
set terminal png
set output "liquidenergygraphs_eq.png"
set autoscale # scale axes automatically
unset log # remove any log-scaling
unset label # remove any previous labels
set xtic auto # set xtics automatically
set ytic auto # set ytics automatically
set title "Equilibration energies for liquid thiophene"
set xlabel "Timestep"
set ylabel "Energy (kJ)"
set key auto
set xr [0.0:100000]
set yr [-60000:60000]
plot "energy_eq.txt" using 2:12 title 'Total' with linespoints , \
      "energy_eq.txt" using 2:14 title 'Potential' with points
```

We are using GNUPLOT to make our graphs. For more information about GNUPLOT, check out this website: <http://people.duke.edu/~hpgavin/gnuplot.html>

```
$ module load gnuplot
$ gnuplot

G N U P L O T
Version 4.6 patchlevel 0      last modified 2012-03-04
Build System: Linux x86_64

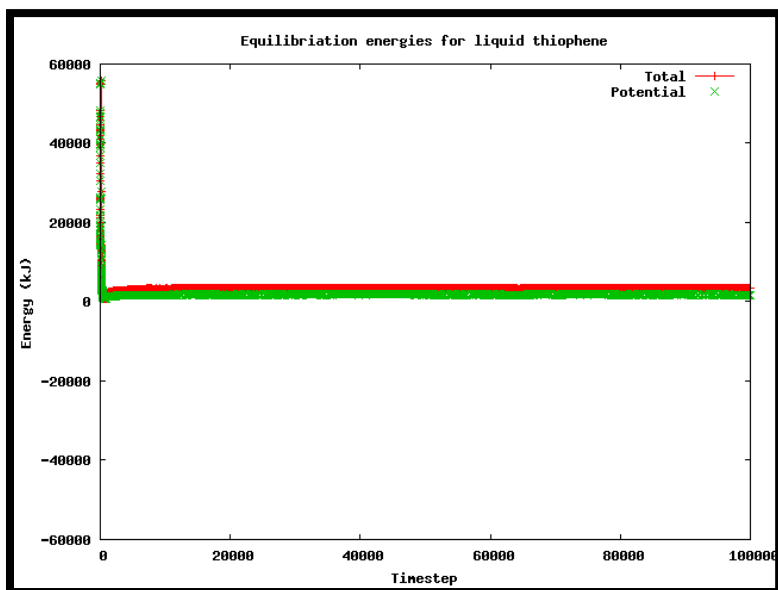
Copyright (C) 1986-1993, 1998, 2004, 2007-2012
Thomas Williams, Colin Kelley and many others

gnuplot home:      http://www.gnuplot.info
faq, bugs, etc:    type "help FAQ"
immediate help:    type "help"    (plot window: hit 'h')

gnuplot> load 'liquidenergygraphs_eq'
```

To get your graph, you must transfer your `liquidenergygraphs_eq.png` file back to your PC using Filezilla.

You may need to adjust the scale of the plot to a value that's appropriate for your data. You can adjust the scale by changing the `"set yr [-60000:60000]"` line in the gnuplot input file.



You should also plot of volume to make sure density of the simulation has reached equilibrium. Note: Volume is in column 19.

```

$ nano liquidvol_eq

# Gnuplot script file for plotting data in file "energy_eq.txt"
#      This file is called   thiophenevol_eq
set terminal png
set output "liquidvol_eq.png"
set      autoscale           # scale axes automatically
unset log                    # remove any log-scaling
unset label                  # remove any previous labels
set xtic auto                # set xtics automatically
set ytic auto                # set ytics automatically
set title   "Equilibration   volume for liquid thiophene"
set xlabel "Timestep"
set ylabel "Volume)"
set key auto
set xr [0.0:100000]
set yr [0:35000]
plot      "energy_eq.txt" using 2:19 title 'Volume' with linespoints

$ gnuplot

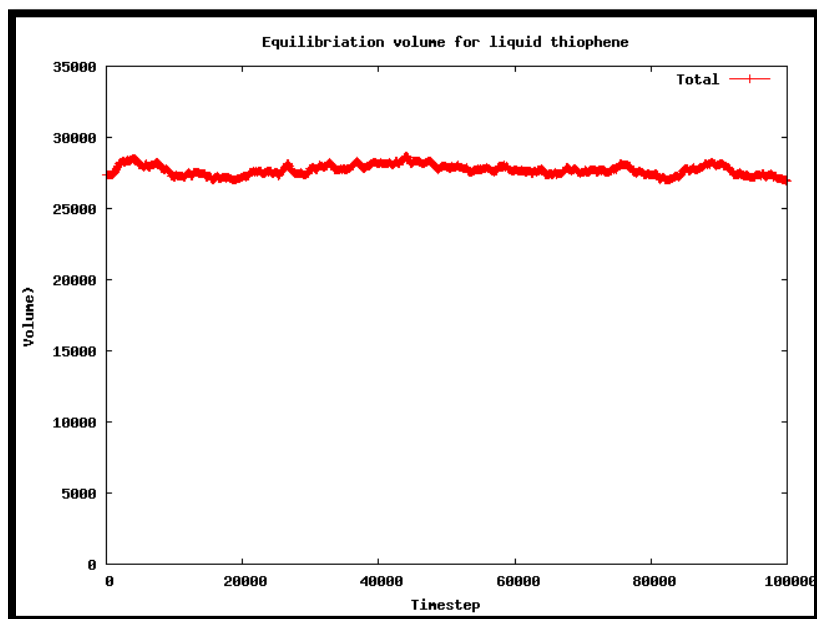
G N U P L O T
Version 4.6 patchlevel 0      last modified 2012-03-04
Build System: Linux x86_64

Copyright (C) 1986-1993, 1998, 2004, 2007-2012
Thomas Williams, Colin Kelley and many others

gnuplot home:      http://www.gnuplot.info
faq, bugs, etc:    type "help FAQ"
immediate help:    type "help"   (plot window: hit 'h')

gnuplot> load 'liquidvol_eq'

```



Stage 2 Complete

Download the initial PDB generated by Liquefy and the final coordinates from the NAMD simulation (moleculefill.coor), and the PNG files of the plots of the cell volume and potential energy to show convergence of this simulation.