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# SCANPY: large-scale single-cell gene expression data analysis

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## Abstract

SCANPY is a scalable toolkit for analyzing single-cell gene expression data. It includes methods for preprocessing, visualization, clustering, pseudotime and trajectory inference, differential expression testing, and simulation of gene regulatory networks. Its Python-based implementation efficiently deals with data sets of more than one million cells (<https://github.com/theislab/Scanpy>). Along with SCANPY, we present ANNDATA, a generic class for handling annotated data matrices (<https://github.com/theislab/anndata>).

**Keywords:** Single-cell transcriptomics, Machine learning, Scalability, Graph analysis, Clustering, Pseudotemporal ordering, Trajectory inference, Differential expression testing, Visualization, Bioinformatics

## Background

Simple integrated analysis work flows for single-cell transcriptomic data [1] have been enabled by frameworks such as SEURAT [2], MONOCLE [3], SCDE/PAGODA [4], MAST [5], CELL RANGER [6], SCATER [7], and SCRAN [8]. However, these frameworks do not scale to the increasingly available large data sets with up to and more than one million cells. Here, we present a framework that overcomes this limitation and provides similar analysis possibilities. Moreover, in contrast to the existing R-based frameworks, SCANPY's Python-based implementation is easy to interface with advanced machine-learning packages, such as TENSORFLOW [9].

## Results

**SCANPY integrates canonical analysis methods in a scalable way**

SCANPY integrates the analysis possibilities of established R-based frameworks and provides them in a scalable and modular form. Specifically, SCANPY provides preprocessing comparable to SEURAT [10] and CELL RANGER [6], visualization through TSNE [11, 12], graph-drawing [13–15] and diffusion maps [11, 16, 17], clustering similar

to PHENOGRAPH [18–20], identification of marker genes for clusters via differential expression tests and pseudotemporal ordering via diffusion pseudotime [21], which compares favorably [22] with MONOCLE 2 [22], and WISHBONE [23] (Fig. 1a).

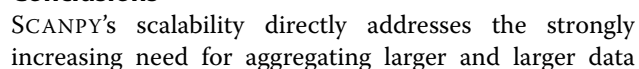
**SCANPY is benchmarked in comparisons with established packages**

In a detailed clustering tutorial of 2700 peripheral blood mononuclear cells (PBMCs), adapted from one of SEURAT's tutorials ([http://satijalab.org/seurat/pbmc3k\\_tutorial.html](http://satijalab.org/seurat/pbmc3k_tutorial.html)) [2], all steps starting from raw count data to the identification of cell types are carried out, providing speedups between 5 and 90 times in each step ([https://github.com/theislab/scanpy\\_usage/tree/master/170505\\_seurat](https://github.com/theislab/scanpy_usage/tree/master/170505_seurat)). Benchmarking against the more run-time optimized CELL RANGER R kit [6], we demonstrate a speedup of 5 to 16 times for a data set of 68,579 PBMCs (Fig. 1a,b, [https://github.com/theislab/scanpy\\_usage/tree/master/170503\\_zheng17](https://github.com/theislab/scanpy_usage/tree/master/170503_zheng17)) [6]. Moreover, we demonstrate the feasibility of analyzing 1.3 million cells without subsampling in a few hours of computing time on eight cores of a small computing server (Fig. 1c, [https://github.com/theislab/scanpy\\_usage/tree/master/170522\\_visualizing\\_one\\_million\\_cells](https://github.com/theislab/scanpy_usage/tree/master/170522_visualizing_one_million_cells)). Thus, SCANPY provides tools with speedups that enable an analysis of data sets with more than one million cells and an interactive analysis with run times of the order of seconds for about 100,000 cells.

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sets [30] across different experimental setups, for example within challenges such as the Human Cell Atlas [31]. Moreover, being implemented in a highly modular fashion, SCANPY can be easily developed further and maintained by a community. The transfer of the results obtained with different tools used within the community is simple, as SCANPY's data storage formats and objects are language independent and cross-platform. SCANPY integrates well into the existing Python ecosystem, in which no comparable toolkit yet exists.

During the revision of this article, the **loom file** format (<https://github.com/linnarsson-lab/loompy>) was proposed for HDF5-based storage of annotated data. Within a joint effort of facilitating data exchange across different labs, ANNDATA now supports importing and exporting to loom (<https://github.com/linnarsson-lab/loompy>). In this context, we acknowledge the discussions with S. Linnarsson, which motivated us to extend ANNDATA's previously static to a dynamic HDF5 backing. Just before submission of this manuscript, a C++ library that provides simple interfacing of HDF5-backed matrices in R was made available as a preprint [32].

## Methods

### SCANPY's technological foundations

SCANPY's core relies on NUMPY [33], SCIPY [34], MATPLOTLIB [35], PANDAS [36], and H5PY [37]. Parts of the toolkit rely on SCIKIT-LEARN [27], STATSMODELS [38], SEABORN [39], NETWORKX [28], IGRAPH [14], the TSNE package of [40], and the Louvain clustering package of [41]. The ANNDATA class—available within the package ANNDATA—relies only on NUMPY, SCIPY, PANDAS, and H5PY.

SCANPY's Python-based implementation allows easy interfacing to advanced machine-learning packages such as TENSORFLOW [9] for deep learning [42], LIMIX for linear mixed models [43], and GPY/GPFLOW for Gaussian processes [44, 45]. However, we note that the Python ecosystem comes with less possibilities for classical statistical analyses compared to R.

### Comparison with existing Python packages for single-cell analysis

Aside from the highly popular sclvm (<https://github.com/PMBio/scLVM>) [46, 47], which uses Gaussian process latent variable models for inferring hidden sources of variation, there are, among others, the visualization frameworks FASTPROJECT (<https://github.com/YosefLab/FastProject>) [48], ACCENSE (<http://www.cellaccense.com/>) [49], and SPRING (<https://github.com/AllonKleinLab/SPRING>) [15]—the latter uses the JavaScript package (<http://d3js.org>) D3.js for the actual visualization and Python only for

preprocessing—the trajectory inference tool SCIMITAR (<https://github.com/dimenwarper/scimitar>), the clustering tool PHENOGGRAPH (<https://github.com/jacoblevine/PhenoGraph>) [19], the single-cell experiment design tool MIMOSCA (<https://github.com/asncd/MIMOSCA>) [50], UMIS (<https://github.com/vals/umis>) for handling raw read data [51], the tree-inference tool ECLAIR (<https://github.com/GGiecold/ECLAIR>) [52], and the framework FLOTILLA (<https://github.com/yeolab/flotilla>), which comes with modules for simple visualization, simple clustering, and differential expression testing. Hence, only the latter provides a data analysis framework that solves more than one specific task. In contrast to SCANPY, however, FLOTILLA is neither targeted at single-cell nor at large-scale data and does not provide any graph-based methods, which are the core of SCANPY. Also, FLOTILLA is built around a complicated class STUDY, which contains data, tools, and plotting functions. SCANPY, by contrast, is built around a simple HDF5-backed class ANNDATA, which makes SCANPY both scalable and extendable (law of Demeter).

## Availability and requirements

SCANPY's and ANNDATA's open-source code are maintained on GITHUB (<https://github.com/theislab/scanpy>, <https://github.com/theislab/anndata>) and published under the BSD3 license.

SCANPY and ANNDATA are released via the Python packaging index: <https://pypi.python.org/pypi/scanpy> and <https://pypi.python.org/pypi/anndata>.

Demonstrations and benchmarks discussed in the main text are all stored at [https://github.com/theislab/scanpy\\_usage](https://github.com/theislab/scanpy_usage) and summarized here:

- Analyzing 68,579 PBMCs (Fig. 1) in a comparison with the CELL RANGER R kit [6]: [https://github.com/theislab/scanpy\\_usage/tree/master/170503\\_zheng17](https://github.com/theislab/scanpy_usage/tree/master/170503_zheng17).
- Clustering and identifying cell types, adapted from and benchmarked with [http://satijalab.org/seurat/pbmc3k\\_tutorial.html](http://satijalab.org/seurat/pbmc3k_tutorial.html) and one of SEURAT's tutorials [2]: [https://github.com/theislab/scanpy\\_usage/tree/master/170505\\_seurat](https://github.com/theislab/scanpy_usage/tree/master/170505_seurat).
- Visualizing and clustering 1.3 million cells (Fig. 1c): [https://github.com/theislab/scanpy\\_usage/tree/master/170522\\_visualizing\\_one\\_million\\_cells](https://github.com/theislab/scanpy_usage/tree/master/170522_visualizing_one_million_cells).
- Reconstructing branching processes via diffusion pseudotime [21]: [https://github.com/theislab/scanpy\\_usage/tree/master/170502\\_haghverdi16](https://github.com/theislab/scanpy_usage/tree/master/170502_haghverdi16).
- Simulating single cells using gene regulatory networks [24]: [https://github.com/theislab/scanpy\\_usage/tree/master/170430\\_krumsiek11](https://github.com/theislab/scanpy_usage/tree/master/170430_krumsiek11).
- Analyzing deep-learning results for single-cell images [25]: [https://github.com/theislab/scanpy\\_usage/tree/master/170529\\_images](https://github.com/theislab/scanpy_usage/tree/master/170529_images).

The data sets used in demonstrations and benchmarks are three data sets from 10x Genomics.

Programming language: Python

Operating system: Linux, Mac OS and Windows

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#### Authors' contributions

FAW conceived the project and developed the software. PA co-developed the software, mainly in regard to architecture and maintainability. FJT supervised the project and helped interpret and present the results. FAW wrote the manuscript with the help of PA and FJT. All authors read and approved the final manuscript.

#### Ethics approval and consent to participate

Ethics approval was not applicable for this study.

#### Competing interests

None of the authors declare competing interests.

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