MTGOsc: MTGO analysis on single cells

Nelson Nazzicari, Simone Marini, Danila Vella 2019-03-08

Contents

5 43161

	Example data: bladder basal epithelial cells
E	xample data: bladder basal epithelial cells
	Γ GOsc comes with some example data from the Mouse Atlas in the form of a (simplified for storage reasons) urat object. Let's start with loading Seurat and MTGO:
	brary(Seurat) brary(MTGOsc)
Lo	ading MTGOsc also loads three objects: bladder, markers, and mouse.pathways.
	his is a (simplified) Seurat object int(bladder)
	An object of class seurat in project Bladder_rm.batch_dge.txt 269 genes across 1028 samples.
	his come from applying Seurat function FindAllMarkers() to the bladder dataset ad(markers)
## ## ## ## ## ## ## ## ## ## ##	krt15 1.523860e-202 1.959457 0.994 0.269 2.070317e-198 igfbp2 3.880320e-196 1.857702 1.000 0.322 5.271803e-192 trf 2.219344e-186 1.747788 0.896 0.189 3.015200e-182 krt5 1.329655e-180 1.730088 0.976 0.276 1.806470e-176 gsdmc2 1.235398e-172 1.711701 0.908 0.208 1.678412e-168 gsto1 2.506894e-172 1.629429 0.985 0.299 3.405866e-168
	his is a simple gene -> pathway dictionary ad(mouse.pathways)
## ##	gene pathway 1 43160 Metabolism 2 43160 Biological oxidations 3 43160 Phase I - Functionalization of compounds 4 43161 Metabolism

Biological oxidations

```
## 6 43161 Phase I - Functionalization of compounds
```

Bladder object contains three clusters:

Calling MTGOsc

For this tutorial we'll focus on Basal epitelial cells cluster, which is of intermediate size. We create two "selector" arrays, one for cells and one for genes.

```
cells.selected = bladder@ident == 'Basal epithelial cell(Bladder)'
genes.selected = subset(markers, cluster == 'Basal epithelial cell(Bladder)')$gene
my.bladder = bladder
my.bladder@data = my.bladder@data[genes.selected, cells.selected]
```

MTGOsc needs a folder where to save temporary files and final results.

```
root = tempdir() #change this to your preferred local path
dir.create(root, recursive = TRUE, showWarnings = FALSE)
```

We are now ready to start with MTGOsc:

Q: 0.4237832874196492

```
#building a genes-pathways dictionary
dict = write.dictionary(genes=mouse.pathways$gene, terms = mouse.pathways$pathway, outfolder = root)
#computign gene coexpression (default function is 'cor')
coexp = write.coexpressionMatrix(geneExpression = my.bladder, outfolder = root)
#thinning coexpression network via scale criterion
edges = write.edges(coexpression = coexp, outfolder = root, keep.weights = FALSE, fun = scale_free_thre
## gamma: 2.05513085766315 (target:2)
## threshold: 0.3
## network edges: 33
#writing a parameter file, useful for MGTO
write.paramFile(outfolder = root)
#actual call to MTGO
call.MTGO(outfolder = root, verbose = TRUE)
## =====> MTGO executed with the following OUTPUT:
## /tmp/RtmpMasKnT/
## Q: -0.028007346189164332
## QGO: 0.0
## Cluster Mean Density: 0.07261904761904762
## Q: 0.4109274563820004
## QGO: 0.65
## Cluster Mean Density: 0.19977106227106228
```

At this point networks are saved on disk, in "Network" subfolder, in the form of two html files. In the first one (ClusterNetwork.html) each functional module is collapsed to a single node, the second one (FullNetwork.html) where each gene is represented and functional modules are color coded.

Gene module enrichment on Reactome

Here we look for Reactome pathway enrichment of the genes constituting the thinned network. This procedure is complementary to the exctraction of Reactome pathways by MTGO-SC.

```
# load libraries for gene enrichment on Reactome (those are on Bioconductor, not CRAN)
library(ReactomePA)
library(clusterProfiler)
library(org.Mm.eg.db)
#a support function to take care of gene upper/lower case convention
firstup = function(x) {
  x = tolower(x)
  substr(x, 1, 1) = toupper(substr(x, 1, 1))
  return(x)
}
#the list of all genes involved in the cluster
genes = unique(c(as.character(edges$gene1), as.character(edges$gene2)))
#correct casing of gene names
genes = firstup(genes)
#translating gene names to ENTREZID via org.Mm.eg.db database
genes = bitr(genes, fromType="SYMBOL", toType="ENTREZID", OrgDb="org.Mm.eg.db")
## Warning in bitr(genes, fromType = "SYMBOL", toType = "ENTREZID", OrgDb =
## "org.Mm.eg.db"): 9.09% of input gene IDs are fail to map...
#the actual enrichment
enriched = enrichPathway(gene=genes$ENTREZID, pvalueCutoff=0.05, readable=T, organism = "mouse", pAdjus
```