

Supplementary file 6: Analysis of the TCGA GBM microarray dataset

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We read the matrix and perform normalization.

```
library(cola)

m = read.table("https://jokergoo.github.io/cola_examples/TCGA_GBM/unifiedScaled.txt",
  header = TRUE, row.names = 1, check.names = FALSE)
m = as.matrix(m)

subtype = read.table("https://jokergoo.github.io/cola_examples/TCGA_GBM/TCGA_unified_CORE_ClaNC840.txt",
  sep = "\t", header = TRUE, check.names = FALSE, stringsAsFactors = FALSE)
subtype = structure(unlist(subtype[1, -(1:2)]), names = colnames(subtype)[-(1:2)])
subtype_col = structure(seq_len(4), names = unique(subtype))

m = m[, names(subtype)]
m = adjust_matrix(m)

library(preprocessCore)
cn = colnames(m)
rn = rownames(m)
m = normalize.quantiles(m)
colnames(m) = cn
rownames(m) = rn
```

First we apply standard consensus partitioning analysis with “ATC” as the top-value method and “skmeans” as partitioning method.

```
set.seed(123)
res = consensus_partition(m, top_value_method = "ATC", partition_method = "skmeans",
  cores = 4, anno = subtype, anno_col = subtype_col)
```

In the following plot, **cola** suggests 5 as the best number of subgroups, but we select 4 as the best k because it gives more stable classification.

```
suggest_best_k(res)
```

```
## The best k suggested by this function might not reflect the real
## subgroups in the data (especially when you expect a large best k). It
## is recommended to directly look at the plots from
## select_partition_number() or other related plotting functions.
```

```
## [1] 5
## attr(,"optional")
## [1] 2 3 4
```

```
select_partition_number(res)
```

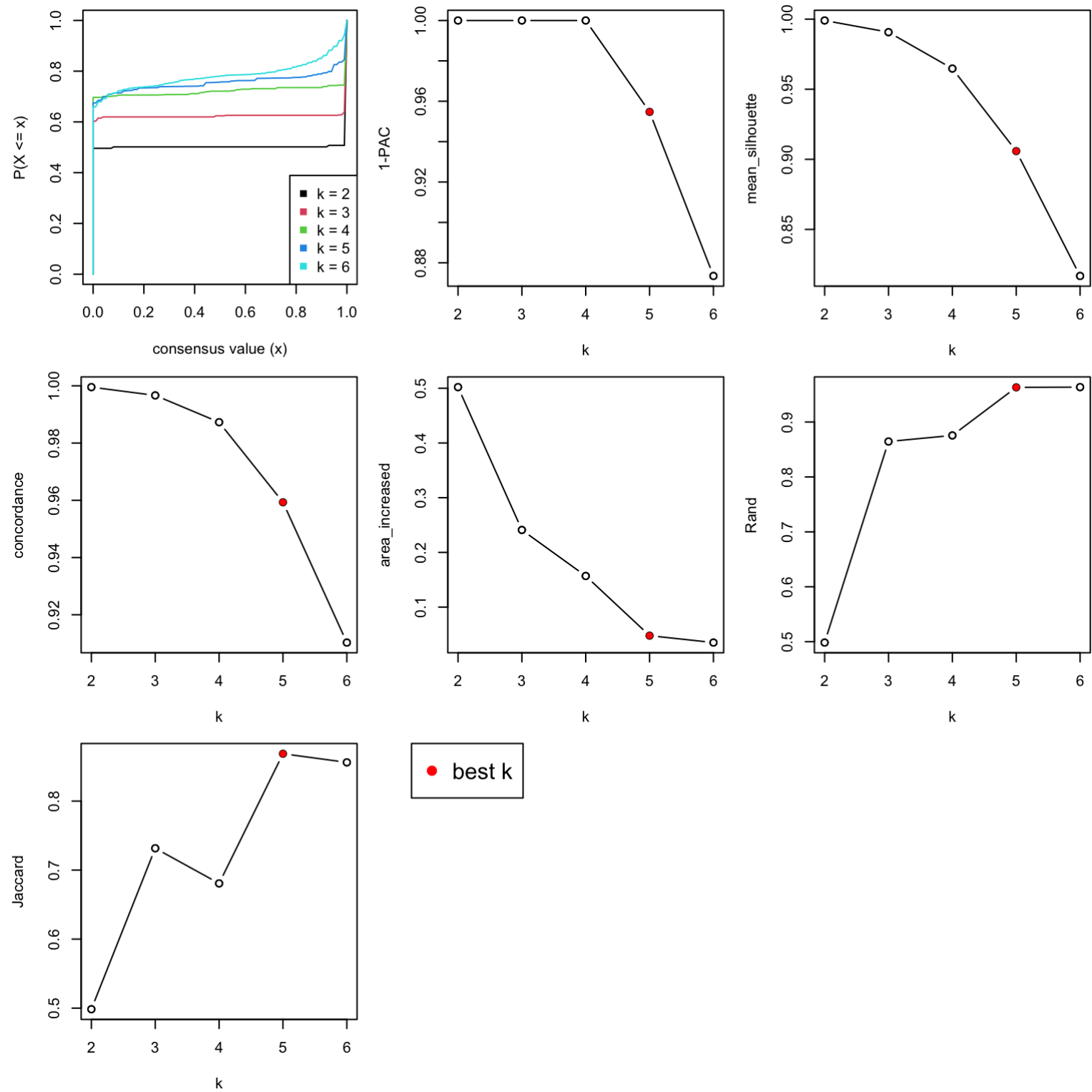


Figure S6.1. Select the best number of groups.

The signature heatmap with 4 subgroups.

```
get_signatures(res, k = 4)
```

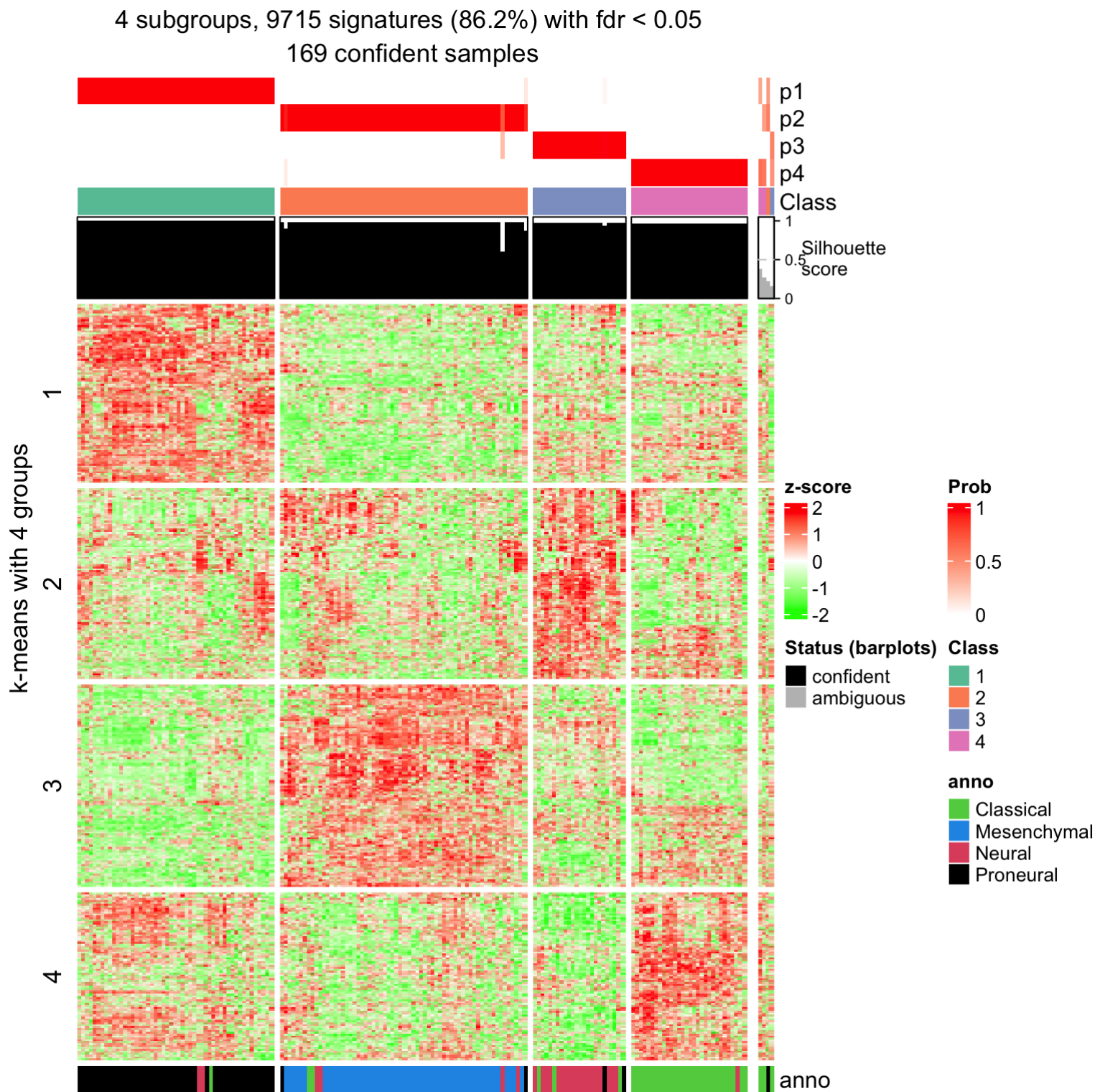


Figure S6.2. Signature heatmap of CP classification with 4 subgroups.

Next we apply hierarchical consensus partitioning (HCP) on the same matrix:

```
set.seed(123)
rh = hierarchical_partition(m, cores = 4, anno = subtype, anno_col = subtype_col)
```

The subgroup hierarchy:

```
collect_classes(rh)
```

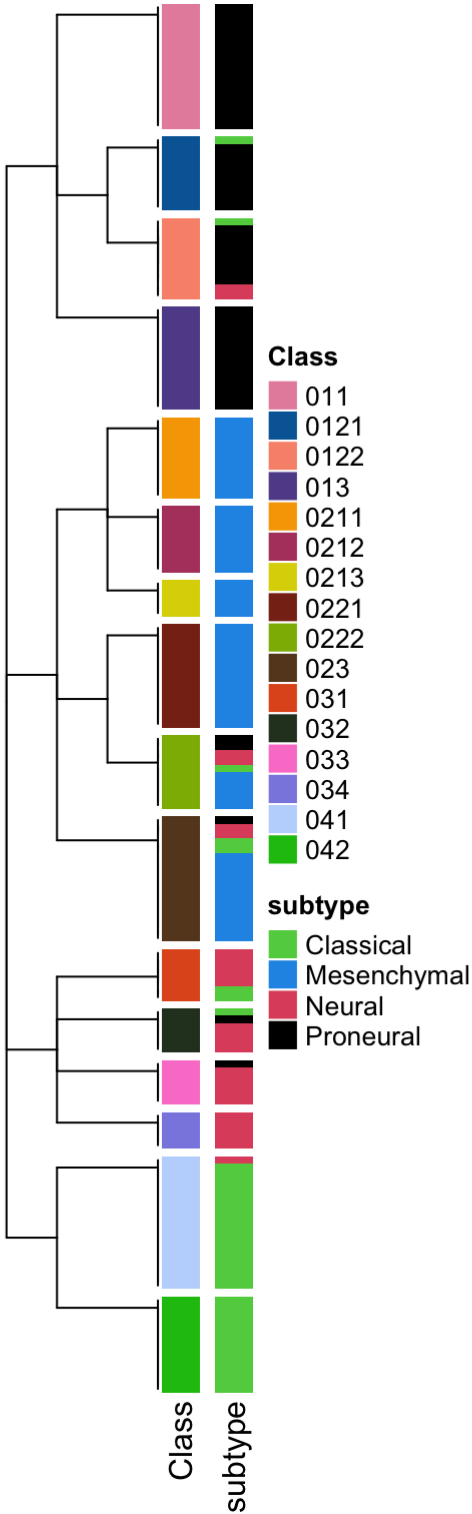


Figure S6.3. Subgroup hierarchy under HCP.

And the signature heatmap under HCP classification:

```
get_signatures(rh)
```

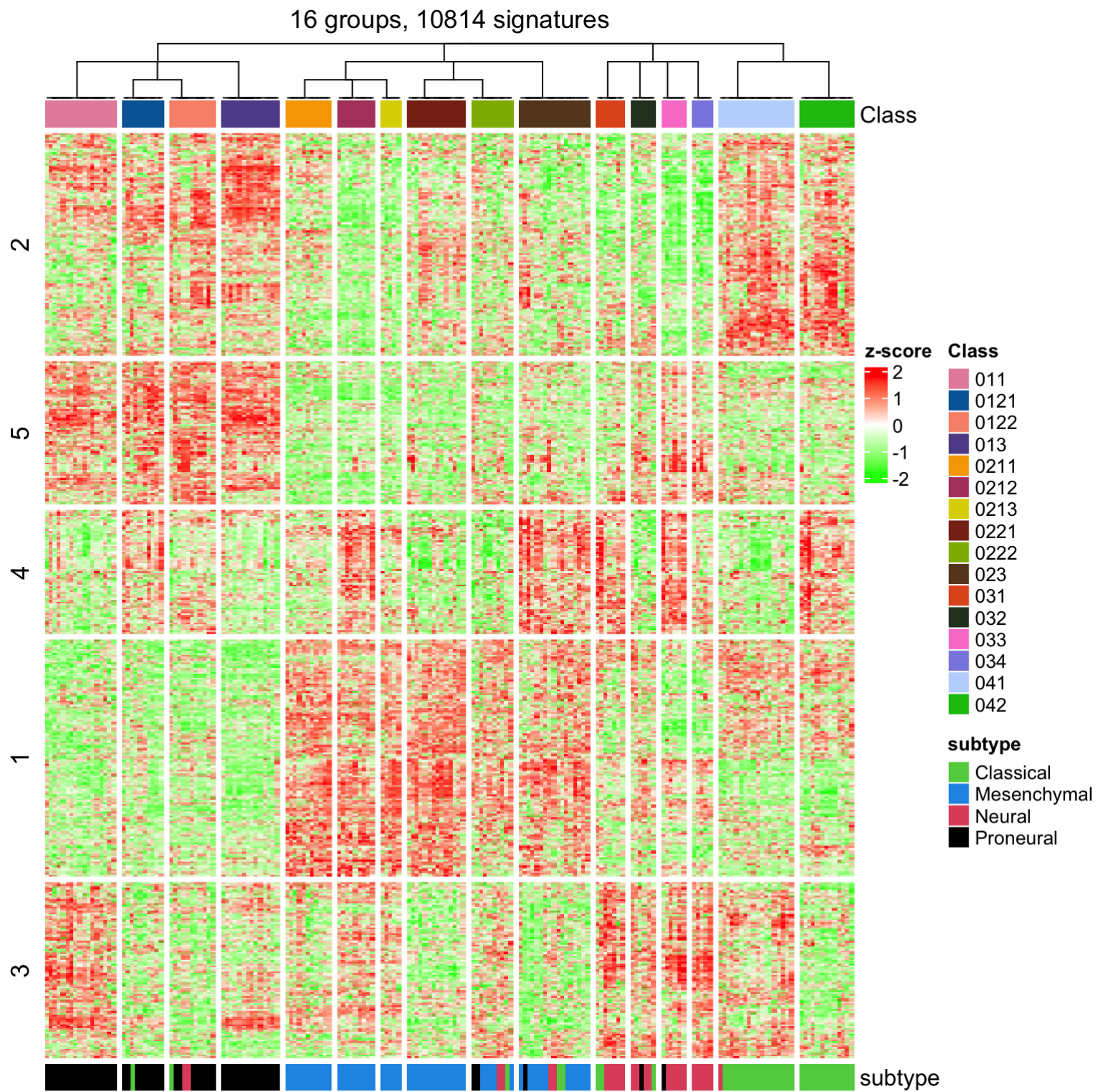


Figure S6.4. Signature heatmap under HCP classification.

The statistics on each node:

```
df = node_info(rh)
df
```

##	id	best_method	depth	best_k	n_columns	n_signatures	p_signatures	is_leaf
## 1	0	ATC:skmeans	1	4	173	9686	0.8596787077	FALSE
## 2	01	ATC:skmeans	2	3	52	4051	0.3595455756	FALSE
## 3	011	ATC:skmeans	3	2	17	55	0.0048815124	TRUE
## 4	012	ATC:skmeans	3	2	21	625	0.0554717316	FALSE
## 5	0121	not applied	4	NA	10	NA	NA	TRUE
## 6	0122	not applied	4	NA	11	NA	NA	TRUE
## 7	013	ATC:skmeans	3	2	14	8	0.0007100382	TRUE
## 8	02	ATC:skmeans	2	3	66	4781	0.4243365581	FALSE
## 9	021	ATC:skmeans	3	3	25	806	0.0715363451	FALSE
## 10	0211	not applied	4	NA	11	NA	NA	TRUE
## 11	0212	not applied	4	NA	9	NA	NA	TRUE
## 12	0213	not applied	4	NA	5	NA	NA	TRUE
## 13	022	ATC:skmeans	3	2	24	666	0.0591106772	FALSE
## 14	0221	ATC:skmeans	4	2	14	30	0.0026626431	TRUE
## 15	0222	not applied	4	NA	10	NA	NA	TRUE
## 16	023	ATC:skmeans	3	2	17	227	0.0201473329	TRUE
## 17	03	ATC:skmeans	2	4	24	1376	0.1221265643	FALSE
## 18	031	not applied	3	NA	7	NA	NA	TRUE
## 19	032	not applied	3	NA	6	NA	NA	TRUE
## 20	033	not applied	3	NA	6	NA	NA	TRUE
## 21	034	not applied	3	NA	5	NA	NA	TRUE
## 22	04	ATC:skmeans	2	2	31	1954	0.1734268217	FALSE
## 23	041	ATC:skmeans	3	2	18	257	0.0228099760	TRUE
## 24	042	ATC:skmeans	3	2	13	175	0.0155320848	TRUE

And the statistics on non-leaf nodes:

```
df[!df$is_leaf, ]
```

##	id	best_method	depth	best_k	n_columns	n_signatures	p_signatures	is_leaf
## 1	0	ATC:skmeans	1	4	173	9686	0.85967871	FALSE
## 2	01	ATC:skmeans	2	3	52	4051	0.35954558	FALSE
## 4	012	ATC:skmeans	3	2	21	625	0.05547173	FALSE
## 8	02	ATC:skmeans	2	3	66	4781	0.42433656	FALSE
## 9	021	ATC:skmeans	3	3	25	806	0.07153635	FALSE
## 13	022	ATC:skmeans	3	2	24	666	0.05911068	FALSE
## 17	03	ATC:skmeans	2	4	24	1376	0.12212656	FALSE
## 22	04	ATC:skmeans	2	2	31	1954	0.17342682	FALSE

Session Info

sessionInfo()

```
## R version 4.1.0 (2021-05-18)
## Platform: x86_64-apple-darwin17.0 (64-bit)
## Running under: macOS Big Sur 10.16
##
## Matrix products: default
## BLAS: /Library/Frameworks/R.framework/Versions/4.1/Resources/lib/libRblas.dylib
## LAPACK: /Library/Frameworks/R.framework/Versions/4.1/Resources/lib/libRlapack.dylib
##
## locale:
## [1] C/UTF-8/C/C/C/C
##
## attached base packages:
## [1] stats      graphics  grDevices  utils      datasets  methods   base
##
## other attached packages:
## [1] knitr_1.33          rmarkdown_2.9      preprocessCore_1.54.0
## [4] RColorBrewer_1.1-2  cola_1.9.4         BiocManager_1.30.16
## [7] colorout_1.2-2
##
## loaded via a namespace (and not attached):
## [1] colorspace_2.0-2      rjson_0.2.20        ellipsis_0.3.2
## [4] mclust_5.4.7          circlize_0.4.13     markdown_1.1
## [7] XVector_0.32.0        GlobalOptions_0.1.2 clue_0.3-59
## [10] rstudioapi_0.13       bit64_4.0.5         AnnotationDbi_1.54.1
## [13] Polychrome_1.2.6      fansi_0.5.0         xml2_1.3.2
## [16] codetools_0.2-18      splines_4.1.0       doParallel_1.0.16
## [19] cachem_1.0.5          impute_1.66.0       jsonlite_1.7.2
## [22] Cairo_1.5-12.2        annotate_1.70.0      cluster_2.1.2
## [25] png_0.1-7             data.tree_1.0.0     compiler_4.1.0
## [28] httr_1.4.2            assertthat_0.2.1    Matrix_1.3-4
## [31] fastmap_1.1.0         htmltools_0.5.1.1   tools_4.1.0
## [34] gtable_0.3.0          glue_1.4.2          GenomeInfoDbData_1.2.6
## [37] dplyr_1.0.7           Rcpp_1.0.7          slam_0.1-48
## [40] Biobase_2.52.0        eulerr_6.1.0        jquerylib_0.1.4
## [43] vctrs_0.3.8           Biostrings_2.60.1   iterators_1.0.13
## [46] xfun_0.24             stringr_1.4.0       lifecycle_1.0.0
## [49] irlba_2.3.3           XML_3.99-0.6        dendextend_1.15.1
## [52] zlibbioc_1.38.0       scales_1.1.1        microbenchmark_1.4-7
## [55] parallel_4.1.0        ComplexHeatmap_2.9.3 yaml_2.2.1
## [58] memoise_2.0.0         gridExtra_2.3        ggplot2_3.3.5
## [61] sass_0.4.0            stringi_1.6.2        RSQLite_2.2.7
## [64] highr_0.9             genefilter_1.74.0    S4Vectors_0.30.0
## [67] foreach_1.5.1         BiocGenerics_0.38.0 shape_1.4.6
## [70] GenomeInfoDb_1.28.0   rlang_0.4.11        pkgconfig_2.0.3
## [73] matrixStats_0.59.0    bitops_1.0-7        evaluate_0.14
## [76] lattice_0.20-44       purrr_0.3.4         bit_4.0.4
## [79] tidysselect_1.1.1     magrittr_2.0.1      R6_2.5.0
## [82] IRanges_2.26.0        magick_2.7.2         generics_0.1.0
## [85] DBI_1.1.1             pillar_1.6.1         survival_3.2-11
## [88] KEGGREST_1.32.0       scatterplot3d_0.3-41 RCurl_1.98-1.3
## [91] tibble_3.1.2          crayon_1.4.1         utf8_1.2.1
## [94] skmeans_0.2-13        viridis_0.6.1        GetoptLong_1.0.3
## [97] grid_4.1.0            blob_1.2.1           digest_0.6.27
## [100] xtable_1.8-4          brew_1.0-6           stats4_4.1.0
## [103] munsell_0.5.0         viridisLite_0.4.0    bslib_0.2.5.1
```