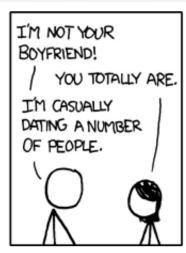
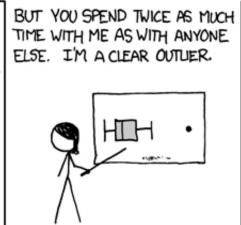
# Binary Response Variables, Random vs Fixed Effects, and Outliers Biology 683

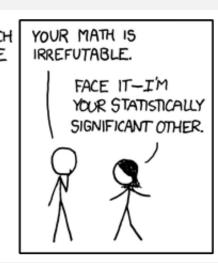
Lecture GLM2

Heath Blackmon

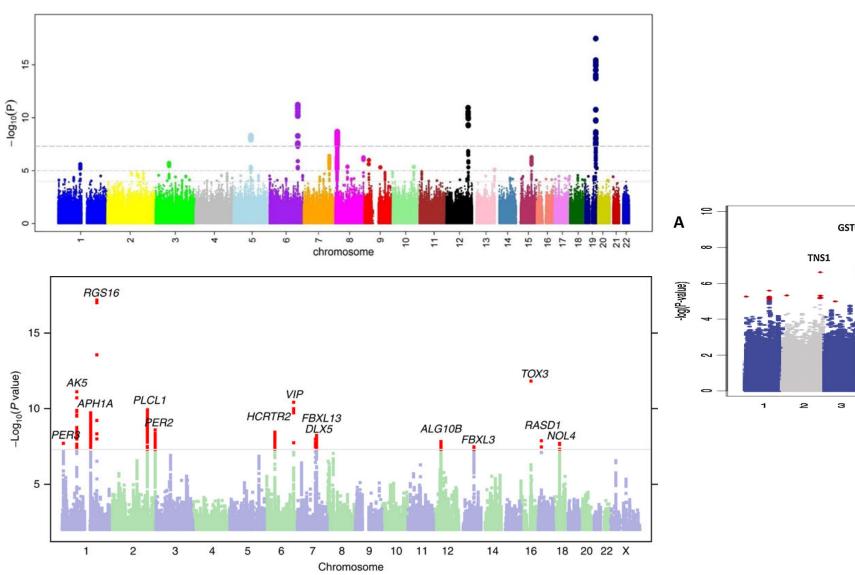


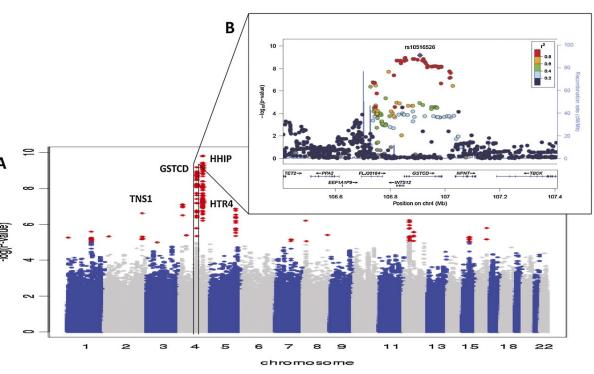






## **GWAS**





#### **GWAS**

#### **GWAS:** Genome wide association study.

The goal of GWAS is to determine what genes have alleles that are responsible for a trait of interest. The trait can be any measurable trait in any organism that you wish to study. For instance, a disease in humans, an economically important trait of a crop or domestic animal, an adaptation like a certain color pattern in birds, etc.

## **GWAS – Discrete condition (often disease)**



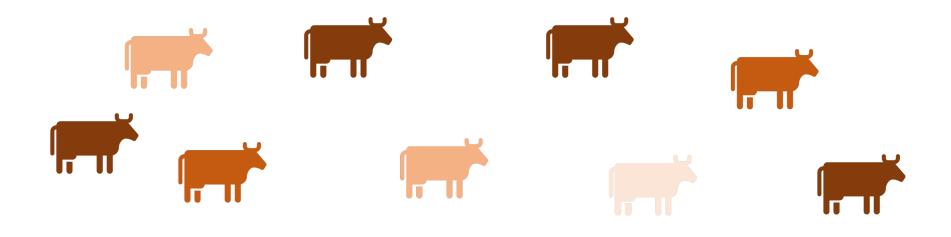
>case 1

CATACTACTGAACGTTTGCTCCTGCtactatctctctctctctctctctctctctctctctCATGC >case 2

CATACTACTGAACGTTTGCTCCTGCtactatctctctctctctctctctctctctctctctctCATGC >control 1

CATACTACTGAACGTTTGCTCCTGCtactatctctctctctctctctctctctctctctctCATGC >control 3

#### **GWAS – Continuous trait**



>sample 1

 ${\tt CATACTACTGAACGTTTGCTCCTGCtactatctctctctctctctctctctctctctctctct}$ 

>sample 2

>sample 3

 ${\tt CATACTACTGAACGTTTGCTCCTGCtactatctctctctctctctctctctctctctctctct}$ 

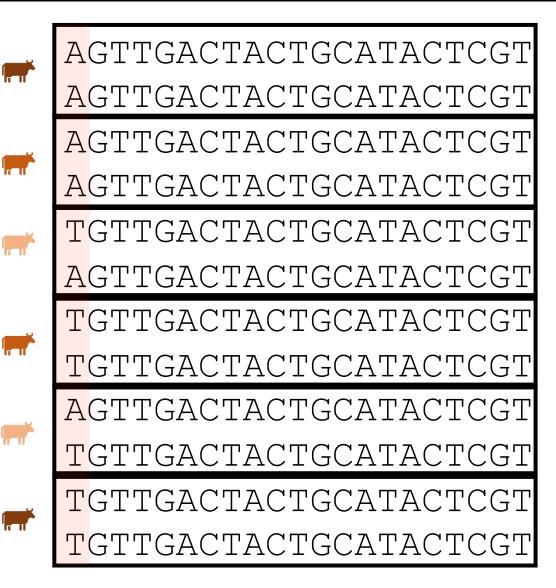
>sample 4

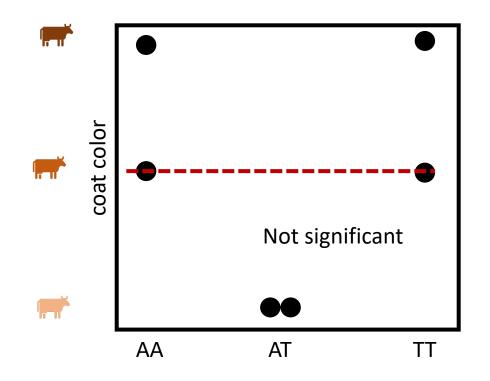
>sample 5

CATACTACTACTGAACGTTTGCTCCTGCtactatctctctctctctctctctctctctctctctctCATGC

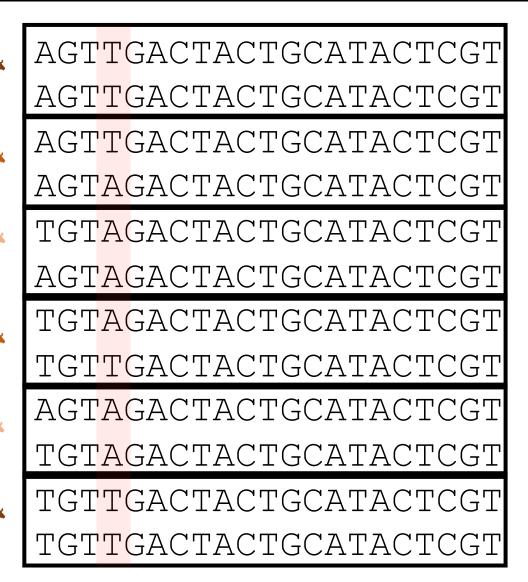
>sample 6

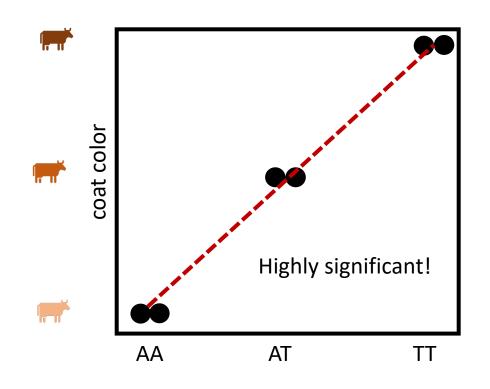
#### **GWAS – Continuous trait**





#### **GWAS – Continuous trait**





#### **GWAS - Problems**

What is the problem with doing this across the whole genome?

#### Multiple tests lead to more false positives!

- 1) require a higher level of significance 5x10<sup>-8</sup>
- 2) only look at the very most significant
- 3) lots of more complicated approaches too!

What is one of the most basic requirements of almost all statistical tests?

#### Tests normally assume independence of the data points!

- 1) Samples from a population will be related to each other due to ancestry (trees!)
- DNA sequencing is not done equally in all groups of people (western samples are usually over represented

## **GWAS - Problems**

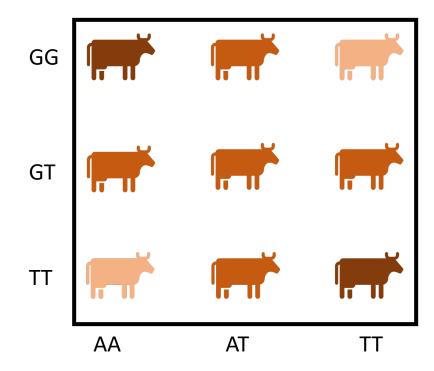
## What is epistasis?

It is the case where the impact of a genotype at one locus depend on the genotype at another locus!

To find this type of effect you would need to look at all pairs of genotypes. How many tests would we need to do then?

About 6 orders of magnitude more tests would be required.

Humans have around 4,000,000 sites that are variable like this which equates to 7,999,998,000,000



## **GWAS - Problems**

What about the environment?

Many diseases have a strong environmental component (heart disease, diabetes, cancer, etc.)

If these are left out of the study often what is discovered is actually genetic variation that happens to coincide with environmental factors?

If a disease is more common in Europeans than Africans or Asians but it is because of a lifestyle characteristic any genetic variation that is common in Europeans but rare in Africans and Asians could appear associated with the disease.

## **Applications and benefits of GWAS**

Widely used in agriculture and domestication. For instance, you can do a GWAS on wild strains of rice (which have lots of variation in things we care about like grain size, growing time, etc.) This GWAS can tell you what variants at what locations in the genome should be introgressed into domestic varieties in hopes of introducing favorable traits.

Widely used in medicine to identify the genes responsible for disease. This is often the first step necessary in being able to create an animal model for a disease that will then allow researchers to study the disease and develop pharmaceutical interventions.



Many diseases have multiple different underlying genetic causes and treatments that are available may only work on some forms of the diseases. Thanks to GWAS studies we now know what these different causal genes are. Now patients can be genotyped at these causative loci and medication can be tailored to their version of the disease. (precision or personalized medicine)

Applied in a more limited fashion to inform patients of risk that they will develop more severe versions of a disease. For instance depending on your genotype at a gene you may choose a more aggressive form of treatment.

## **Complex Disease / Complex Phenotypes**

Many diseases are what we call complex diseases. There is no one gene responsible for the disease. Instead the disease can manifest due to variations in 10 or 100s of different genes in the genome acting in concert with the environment. GWAS is less insightful (though still important) for diseases like this.

Schizophrenia: Not really a clearly delineated disease like say COVID, type 1 diabetes, or cystic fibrosis. Instead it is a constellation of symptoms that individuals exhibit to varying degrees.

From studies of multiple generations of families we know that 20-30% of risk is inherited (genetic). The other 70-80% of your risk of developing the disease is environmental and is poorly understood. Massive GWAS studies with 1000s of individuals have studied the genetic component. These studies have identified more than 100 different loci in the genome that seem to have some predictive power. However, these variations are associated with an increase in risk and there are many people who carry many alleles that increase risk but that never develop the disease.

## Mixed models

Mixed models are models that include fixed and and random effects.

Fixed effects can be repeated by other researchers. These are the variables that you are interested in studying.

Random effects are usually nuisance parameters. These are variables that other researchers cannot replicate and you are not interested in inferring anything about them.

## Mixed models

Fixed effects are the variables whose impact we wish to determine

- Characteristics of the media or habitat
- Experimental treatments
- Age groups
- Time points
- Mutant genotypes

Conclusions that you reach are only applicable to the groups or treatments you include in the study

# Example of fixed effects

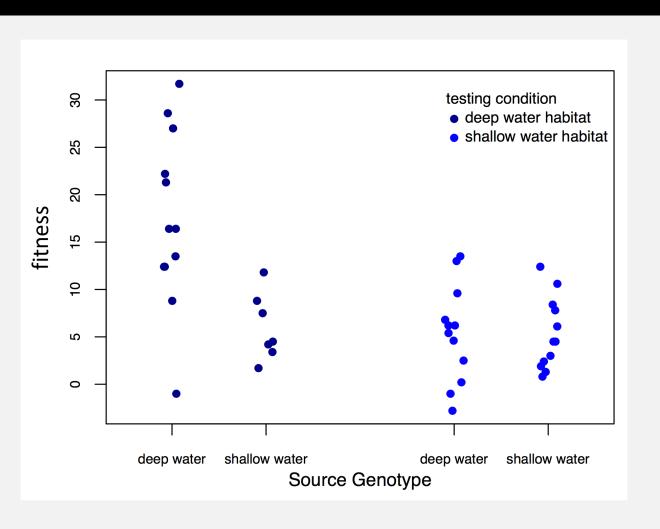
Reciprocal relocation experiment to investigate how genotype and habitat interact to determine the fitness of stickleback fish (Rundle 2002).

<b>C</b>	1 1. * 1 . 1
Source	habitat

		Shallow	Deep
Test habitat	Shallow	12 fish	11 fish
	Deep	7 fish	11 fish



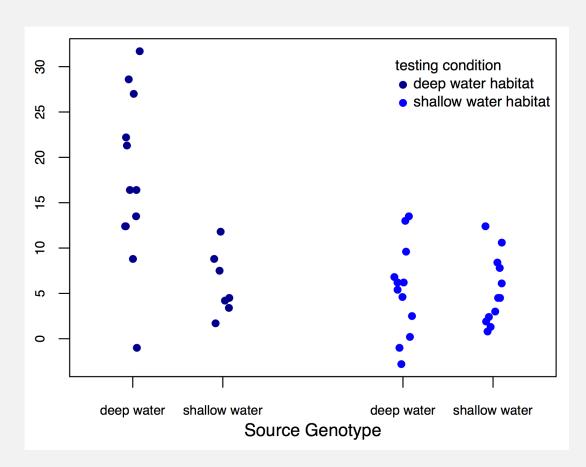
# Example of fixed effects (two factor ANOVA)



```
> anova(lm(fitness ~ genotype * test.habitat))
Analysis of Variance Table
Response: fitness
                       Df Sum Sq Mean Sq F value
                                                      Pr(>F)
                           363.49 363.49 9.7045 0.0034403 **
genotype
                          550.55 550.55 14.6986 0.0004485 ***
test.habitat
genotype:test.habitat 1
                           333.58 333.58 8.9059 0.0048864 **
Residuals
                       39 1460.77
                                    37,46
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' '1
> summary(lm(fitness ~ genotype * test.habitat))
Call:
lm(formula = fitness ~ genotype * test.habitat)
Residuals:
    Min
                  Median
-18.4750 -3.6917
                           3.4583 14.2250
                 -0.8083
Coefficients:
                                 Estimate Std. Error t value Pr(>|t|)
(Intercept)
                                   17.475
                                                      9.891 3.49e-12 ***
                                  -11.489
genotypeshallow
                                               2.911 -3.947 0.000321 ***
test.habitatshallow
                                  -12.125
                                              2.499 -4.853 1.99e-05 ***
genotypeshallow:test.habitatshallow 11.448
                                              3.836
                                                      2.984 0.004886 **
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' '1
Residual standard error: 6.12 on 39 degrees of freedom
Multiple R-squared: 0.4606,
                             Adjusted R-squared: 0.4192
F-statistic: 11.1 on 3 and 39 DF, p-value: 2.093e-05
```

## Interpreting Coefficients

```
> summary(lm(fitness ~ genotype * test.habitat))
Call:
lm(formula = fitness ~ genotype * test.habitat)
Residuals:
    Min
                   Median
                                        Max
-18.4750 -3.6917 -0.8083
                            3.4583 14.2250
Coefficients:
                                   Estimate Std. Error t value Pr(>|t|)
                                     17.475
                                                        9.891 3.49e-12 ***
(Intercept)
                                                1.767
genotypeshallow
                                    -11.489
                                                2.911 -3.947 0.000321
test.habitatshallow
                                    -12.125
                                                2.499 -4.853 1.99e-05 ***
genotypeshallow:test.habitatshallow 11.448
                                                3.836
                                                        2.984 0.004886 **
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' '1
Residual standard error: 6.12 on 39 degrees of freedom
                              Adjusted R-squared: 0.4192
Multiple R-squared: 0.4606,
F-statistic: 11.1 on 3 and 39 DF, p-value: 2.093e-05
```



## What is a random effect

These are randomly sampled categories of a variable that represent groups of individual measurements. Usually random effects are not repeatable.

- Study sites
- Environmental chambers
- Families made up of siblings
- Measurements within individuals

Conclusions that you reach are applicable only to the sample being studied.

## What is a random effect

#### Sometimes random effects are a nuisance

- Field sites
- Environmental chambers
- Field plots
- Repeated measures

Occasionally random effects are of great interest

- Families Heritability
- Individuals Breeding value

# Example of random effects

Impact of selective regime on horn size.

Measure both the left and right horn in 25 beetles from two different selective regimes.

Horn size	Beetle	Selective regime	
256	1	High	
276	1	High —	1950
321	2	High	126
321	2	High —	
423	3	Low	124
401	3	Low —	
381	4	Low	126
409	4	Low —	



# Example of random effects

Identifying the predictors for the presence or absence of Chrysina beetles.

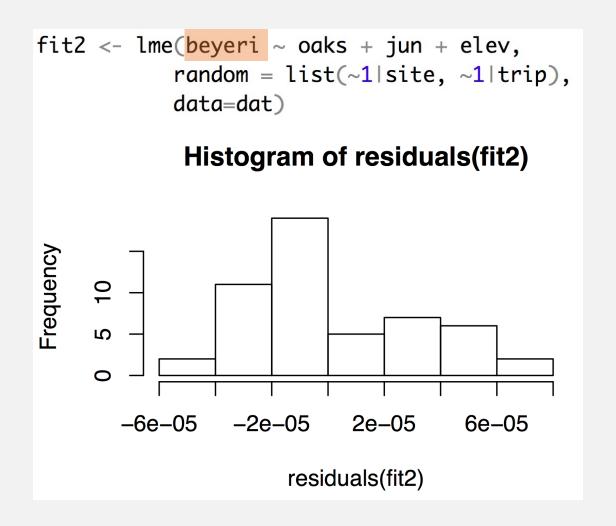
Number collected	oak	juniper	site	date	trip
8	1	0	21	210	Α
2	1	1	13	210	Α
1	0	1	31	211	Α
5	0	1	15	212	Α
4	1	1	21	242	В
6	1	0	13	242	В
0	1	1	31	245	В
7	1	1	15	245	В

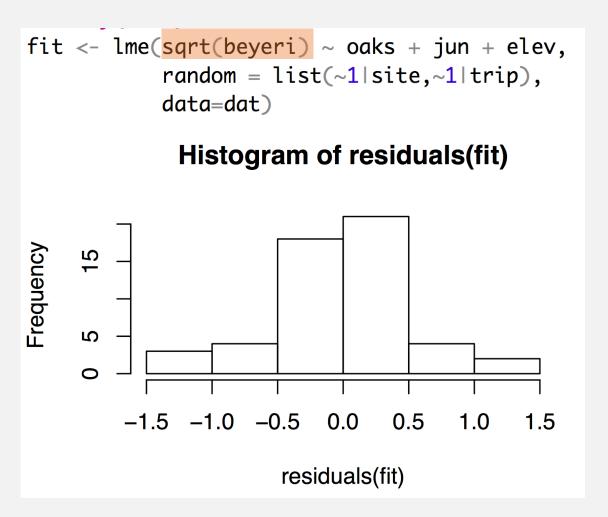


# Implementing a mixed effects model

Mixed effect models can be fit using the LME function from the package nlme.

## Implementing a mixed effects model





# Implementing a mixed effects model

## Mixed effect models can be fit using the LME function from the package nlme.

```
fit <- lme(sqrt(beyeri) ~ oaks + jun + elev,
             random = list(~1|site,~1|trip),
             data=dat)
> summarv(fit)
Linear mixed-effects model fit by REML
Data: dat
      AIC
                     logLik
               BIC
 153.6247 166.7231 -69.81233
Random effects:
Formula: ~1 | site
        (Intercept)
StdDev: 2.272341e-05
Formula: ~1 | trip %in% site
       (Intercept) Residual
StdDev: 0.8169537 0.00264004
Fixed effects: sqrt(beyeri) ~ oaks + jun + elev
               Value Std.Error DF t-value p-value
(Intercept) -1.6788177 1.5125428 26 -1.109931 0.2772
            0.9707025 0.2943403 22 3.297892 0.0033
           -0.0860503 0.2460159 22 -0.349775 0.7298
jun
            0.0012513 0.0008072 22 1.550167 0.1354
elev
Correlation:
    (Intr) oaks jun
oaks 0.530
    0.197 -0.097
elev -0.993 -0.584 -0.250
```

```
fit <- lme(sqrt(beyeri) ~ oaks + elev,
            random = list(~1|site,~1|trip),
            data=dat)
> summarv(fit)
Linear mixed-effects model fit by REML
 Data: dat
       AIC
               BIC logLik
 150.7722 162.1231 -69.3861
Random effects:
 Formula: ~1 | site
         (Intercept)
StdDev: 2.191491e-05
 Formula: ~1 | trip %in% site
        (Intercept)
                      Residual
StdDev: 0.8096043 0.002598713
Fixed effects: sqrt(beyeri) ~ oaks + elev
                Value Std.Error DF t-value p-value
(Intercept) -1.5744919 1.4695032 26 -1.071445 0.2938
            0.9607576 0.2903283 23 3.309211 0.0031
oaks
            0.0011807 0.0007746 23 1.524404 0.1410
elev
 Correlation:
     (Intr) oaks
oaks 0.563
elev -0.994 -0.631
```

```
fit <- lme(sqrt(beyeri) ~ oaks,
            random = list(~1|site,~1|trip),
            data=dat)
> summary(fit)
Linear mixed-effects model fit by REML
 Data: dat
       AIC
                      loaLik
                BIC
 138.5902 148.1504 -64.29512
Random effects:
 Formula: ~1 | site
        (Intercept)
StdDev: 2.309943e-05
 Formula: ~1 | trip %in% site
       (Intercept)
                      Residual
StdDev: 0.8202517 0.002667513
Fixed effects: sqrt(beyeri) ~ oaks
               Value Std.Error DF t-value p-value
(Intercept) 0.6514133 0.1674341 26 3.890566
                                             6e-04
            1.2400606 0.2281742 24 5.434710
oaks
                                             0e+00
 Correlation:
     (Intr)
oaks -0.734
```

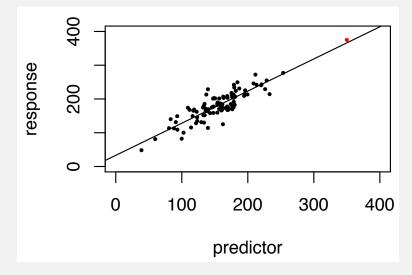
## Considerations for models with random effects

- Most software will assume that all factors are fixed unless you specify them as mixed.
- Designating factors as random effects takes extra work.
- The lm function treats all predictors as fixed effects.
- Treating random effects as fixed effects is fundamentally wrong.

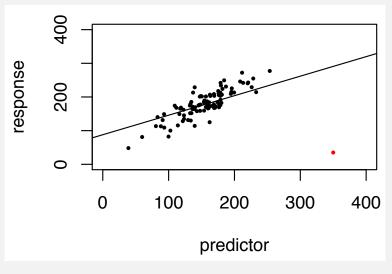
# 4 Types of outliers

1) Obviously erroneously recorded or measured data

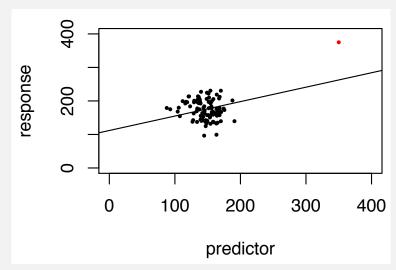
2) Extreme data point that doesn't impact statistic of interest but does impact p-value.



3) Extreme data point that impacts statistic of interest.



4) Extreme data point that creates significance.



## Links

MCMCMglmm: Fit mixed models with phylogenetic or pedigree information in a Bayesian framework.

Outlier Package: Apply outlier tests to identify possible outlier datapoints - I don't recommend this.

If time permits: repeated median regression (Theil-Sen Estimator)

- simulate x and y with medium strength relationship and add one outlier
- standard regression record Beta
- record slope between all pairs and find median slope
- repeat standard regression without outlier