ANOVA Analysis of Variance

Stats and Genetics/Evolution

The Correlation between relatives on the supposition of Mendelian Inheritance

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Communicated by Professor J. ARTHUR THOMSON

With Four Figures in Text

(MS. received 15 June 1918. Read 8 July 1918. Issued separately 1 October 1918)

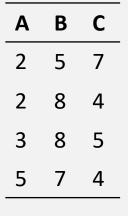
ON THE "PROBABLE ERROR" OF A COEFFICIENT OF CORRELATION DEDUCED FROM A SMALL SAMPLE

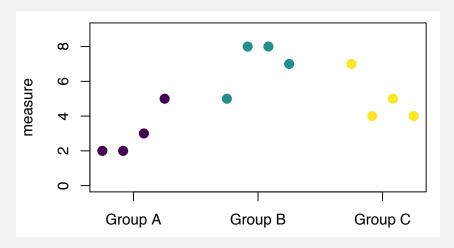
Fisher 1921

Author's Note (CMS 1.2a)

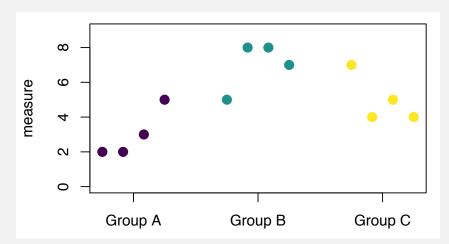
This is the second of three papers dealing with the sampling errors of correlation coefficients covering the cases (i) "The frequency distribution of the values of the correlation coefficient in samples from an indefinitely large population," *Biometrika*, Vol. 10, pp. 507-521, 1915.

- Used to compare the means among more than two groups
- If you are comparing three groups, for instance, you cannot just do three pair-wise t-tests — this approach would cause too many false positives
- ANOVA takes into account the fact that you are comparing multiple groups and controls the false positive rate.



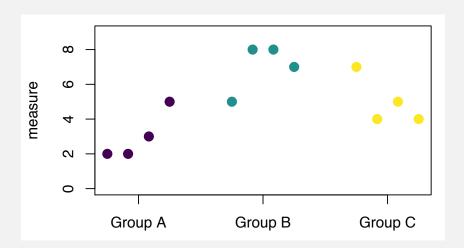






$$f \ statistic = \frac{\frac{\sum_{j=1}^{p} n_{j}(\bar{x}_{j} - \bar{x})^{2}}{df_{ssb}}}{\frac{\sum_{j=1}^{p} \sum_{i=1}^{n_{j}} (x_{ij} - \bar{x}_{j})^{2}}{df_{ssw}}}$$

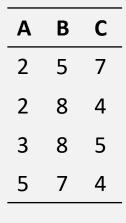
В	С
5	7
8	4
8	5
7	4
	5 8 8

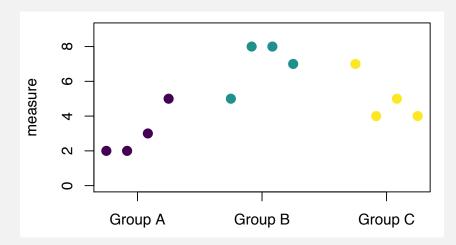


$$f \ statistic = \frac{\sum_{j=1}^{p} n_j (\bar{x}_j - \bar{x})^2}{\frac{df_{SSb}}{\sum_{j=1}^{p} \sum_{i=1}^{n_j} (x_{ij} - \bar{x}_j)^2}} \quad \text{number of groups -1}$$

$$number \ of \ sample - number \ of \ groups$$

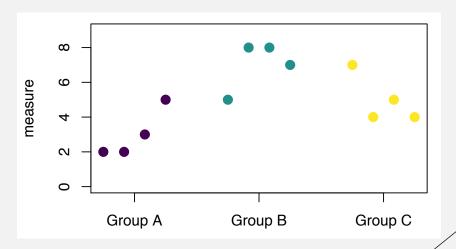
$$df_{SSW}$$

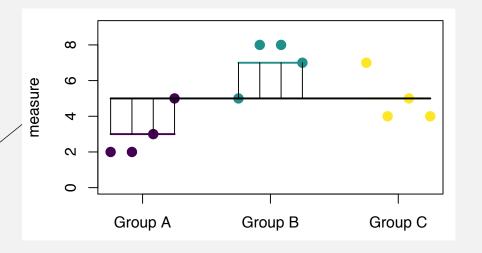




$$f \ statistic = \frac{\frac{\sum_{j=1}^{p} n_j (\bar{x}_j - \bar{x})^2}{df_{ssb}}}{\frac{\sum_{j=1}^{p} \sum_{i=1}^{n_j} (x_{ij} - \bar{x}_j)^2}{df_{ssw}}}$$

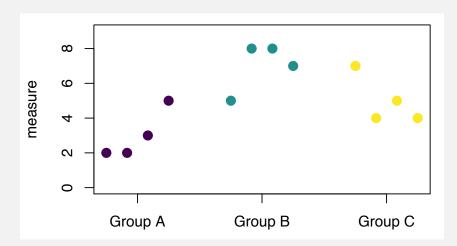


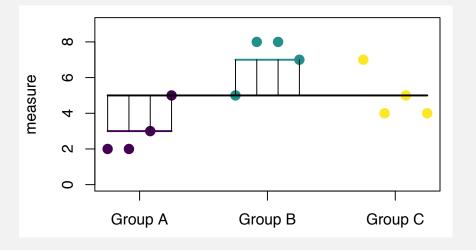




$$f \ statistic = \frac{\sum_{j=1}^{p} n_{j}(\bar{x}_{j} - \bar{x})^{2}}{\frac{\sum_{j=1}^{p} \sum_{i=1}^{n_{j}} (x_{ij} - \bar{x}_{j})^{2}}{df_{ssw}}}$$

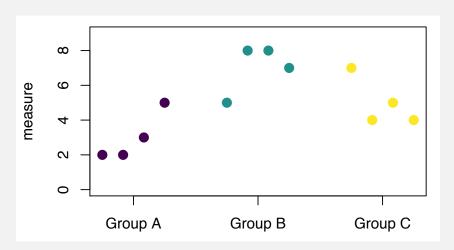
Α	В	С
2	5	7
2	8	4
3	8	5
5	7	4

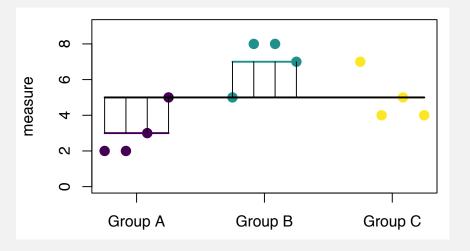


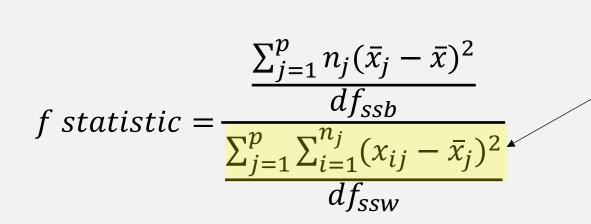


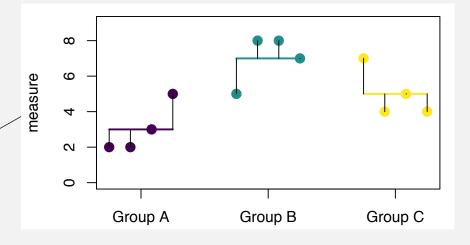
$$f \ statistic = \frac{\frac{\sum_{j=1}^{p} n_j (\bar{x}_j - \bar{x})^2}{df_{ssb}}}{\frac{\sum_{j=1}^{p} \sum_{i=1}^{n_j} (x_{ij} - \bar{x}_j)^2}{df_{ssw}}}$$



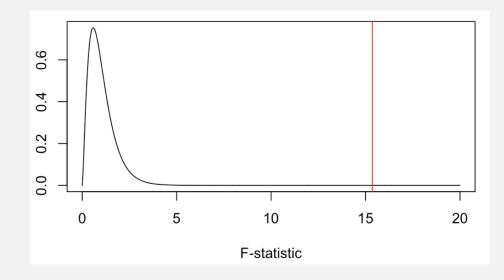








Running ANOVA in R

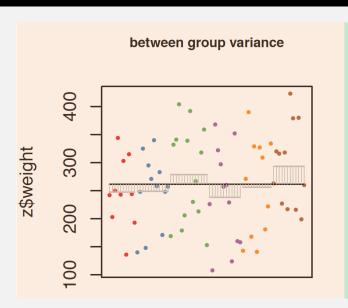


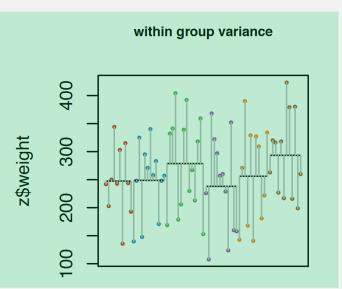
This significant result tells us that at least one of the groups of chickens have significantly different mean weights than at least one other groups. (significant ANOVA result allows us to reject the null that they are all the same)

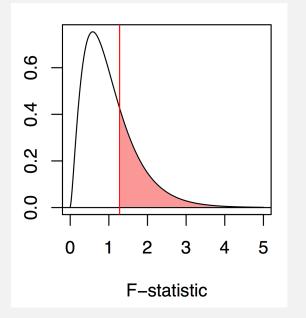
$$f \ statistic = \frac{\sum_{j=1}^{p} n_j (\bar{x}_j - \bar{x})^2}{df_{ssb}}$$

$$\frac{\sum_{j=1}^{p} \sum_{i=1}^{n_j} (x_{ij} - \bar{x}_j)^2}{df_{ssw}}$$

Running ANOVA in R







Go To R

Post-hoc tests

If your ANOVA is significant, you may be interested in discovering which groups are different from one another

A variety of post-hoc comparisons of the means can be used

Fisher's LSD

Least conservative test, basically uses t-tests to compare the means

Scheffe's method

Performs all comparisons simultaneously, but has relatively low power

Tukey-Kramer method

A pair-wise method, like a t-test, but corrected for multiple comparisons

Post-hoc tests

```
> data("chickwts")
> fit <- aov(weight~feed, data=chickwts)
> TukeyHSD(fit)
  Tukey multiple comparisons of means
   95% family-wise confidence level
Fit: aov(formula = weight ~ feed, data = chickwts)
$feed
                           diff
                                        lwr
                                                          p adj
                                                  upr
horsebean-casein
                    -163.383333 -232.346876 -94.41979 0.0000000
linseed-casein
                    -104.833333 -170.587491 -39.07918 0.0002100
meatmeal-casein
                     -46.674242 -113.906207 20.55772 0.3324584
soybean-casein
                     -77.154762 -140.517054 -13.79247 0.0083653
sunflower-casein
                       5.333333
                                 -60.420825 71.08749 0.9998902
linseed-horsebean
                      58.550000
                                 -10.413543 127.51354 0.1413329
meatmeal-horsebean
                     116.709091
                                  46.335105 187.08308 0.0001062
soybean-horsebean
                      86.228571
                                  19.541684 152.91546 0.0042167
sunflower-horsebean
                    168.716667
                                  99.753124 237.68021 0.0000000
meatmeal-linseed
                      58.159091
                                  -9.072873 125.39106 0.1276965
soybean-linseed
                      27.678571
                                 -35.683721 91.04086 0.7932853
sunflower-linseed
                     110.166667
                                  44.412509 175.92082 0.0000884
soybean-meatmeal
                     -30.480519
                                 -95.375109 34.41407 0.7391356
sunflower-meatmeal
                      52.007576
                                 -15.224388 119.23954 0.2206962
```

82.488095

19.125803 145.85039 0.0038845

sunflower-soybean

anova and aov functions will both perform an ANOVA but the results are stored slightly differently. For this posthoc test we want the aov format

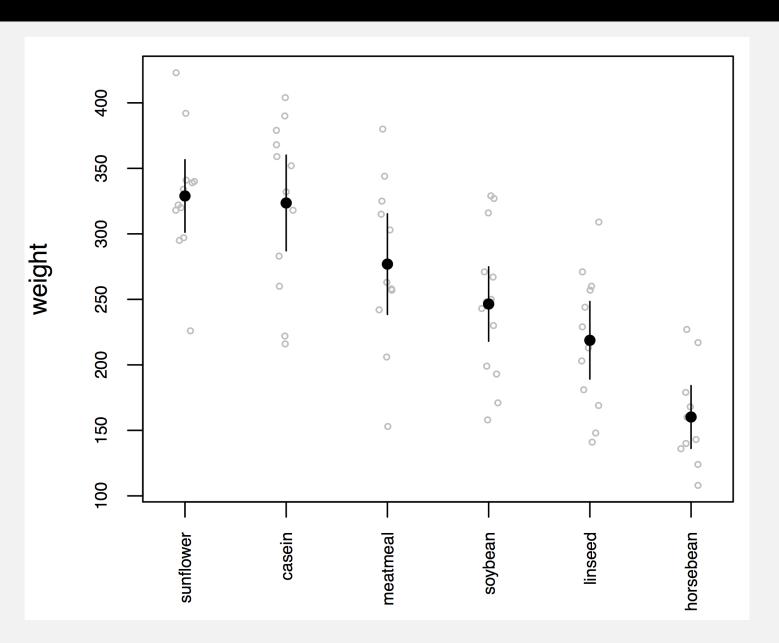
Interpreting post-hoc tests

```
> data("chickwts")
> fit <- aov(weight~feed, data=chickwts)</pre>
> TukeyHSD(fit)
 Tukey multiple comparisons of means
   95% family-wise confidence level
Fit: aov(formula = weight ~ feed, data = chickwts)
$feed
                           diff
                                        lwr
                                                           p adj
                                                   upr
                    -163.383333 -232.346876 -94.41979 0.0000000
horsebean-casein
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soybean-casein
                     -77.154762 -140.517054 -13.79247 0.0083653
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linseed-horsebean
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meatmeal-horsebean
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soybean-horsebean
                      86.228571
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                                  19.125803 145.85039 0.0038845
sunflower-soybean
                      82.488095
```

When we examine all the significantly different ones we can draw several conclusions:

- Chicks fed casein are significantly heavier than those fed horsebean, linseed, and soybean.
- Chicks fed horsebean are significantly lighter than those fed meatmeal, soybean, and sunflower.
- 3) Chicks fed sunflower are significantly heavier than those fed linseed or soybean.

Plotting this kind of data



Our results from the ANOVA and Tukey match up pretty well with our rules of thumb about 95% CI overlaps

Example of code

```
#sets the order of the treatments
chickwts$feed <- factor(chickwts$feed,</pre>
                         levels=c("sunflower", "casein",
                                   "meatmeal", "soybean",
                                   "linseed", "horsebean"))
stripchart(weight ~ feed, data=chickwts,
           method = "jitter", vertical = TRUE, cex.axis = .7,
           col = "qray", pch = 1, cex = .5, las = 3)
#Add error bars:
#First calculate means and SDs
meanShift <- tapply(chickwts$weight, chickwts$feed, mean)</pre>
sdevShift <- tapply(chickwts$weight, chickwts$feed, sd)</pre>
n <- tapply(chickwts$weight, chickwts$feed, length)</pre>
feed_table <- data.frame(mean = meanShift,
                          std.dev = sdevShift, n = n)
#Now add the SEM for each group:
seShift <- 1.96 * sdevShift / sqrt(n)
segments(1:6, meanShift - seShift,
         1:6, meanShift + seShift)
points(meanShift \sim c(1:6), pch = 16)
```

Assumptions of the ANOVA

- The variable is normally distributed within each group
- The variance is the same in the different groups
- The design is balanced you have the same sample size for each group
- But... ANOVA is fairly robust to violations of these assumptions

A Non-Parametric Alternative

- Kruskal-Wallis Test
- Based on ranks
- The multiple-group version of the Mann-Whitney U-test

R-implementation:

> kruskal.test(weight ~ feed, data = chickwts)

Kruskal-Wallis rank sum test

data: weight by feed

Kruskal-Wallis chi-squared = 37.343, df = 5, p-value = 5.113e-07

p-value suggests this test has lower power than ANOVA

A Non-Parametric post-hoc

 Dunn's test – is the non-parametric equivalent of the Tukey

Not in base R need to install: install.packages("dunn.test", dependencies=TRUE)

```
> dunn.test(chickwts$weight, g=chickwts$feed,
            altp = T, method = "bonferroni")
  Kruskal-Wallis rank sum test
data: x and group
Kruskal-Wallis chi-squared = 37.3427, df = 5, p-value = 0
                           Comparison of x by group
                                 (Bonferroni)
Col Mean-I
Row Mean 1
                        horsebea
               casein
                                    linseed
                                              meatmeal
                                                          soybean
horsebea l
             4.813069
              0.0000*
             3.308292 -1.658736
 linseed
              0.0141*
                          1.0000
             1.415755 -3.364059
meatmeal
                                  -1.819817
               1.0000
                         0.0115*
                                     1.0000
 soybean
             2,499922 -2,602093
                                  -0.933255
                                              0.974144
               0.1863
                          0.1390
                                     1.0000
                                                1.0000
sunflowe
            -0.182969
                       -4.987524
                                  -3.491262 -1.594703
                                                        -2.689798
               1.0000
                         0.0000*
                                    0.0072*
                                                1.0000
                                                           0.1072
alpha = 0.05
Reject Ho if p <= alpha
```

ANOVA Summary

- ANOVA is the foundation of essentially all tests comparing multiple means
- Don't make it too complicated the null hypothesis is simple: they are all the same.
- Post-hoc tests are important for determining which means are the source of a significant ANOVA.
- You can only justify a post-hoc test if the ANOVA is significant in the first place.
- Before applying ANOVA, check that your data fit the assumptions
 (consider transforming the data lots of times this will be based on your
 biological knowledge because you will have insufficient data to say much
 about the observed distribution)

ANOVA Practice Problems

- Use the chick weights dataset included in R data("chickwts"). Reduce the data down to just soybean and sunflower. Run an ANOVA and determine whether these foods lead to significant differences in weight.
- Use the offspring.csv file from the course website and determine whether XY and ZW systems have different numbers of male offspring.

Hypothesis testing has limits

Often times we want to say more than something has an effect. We want to understand exactly how a predictor variable impacts a response variable.

Often times we have complex relationships where several variables (continuous and discrete) impact our response variable and we need to understand how all of these things work together to determine our observations.

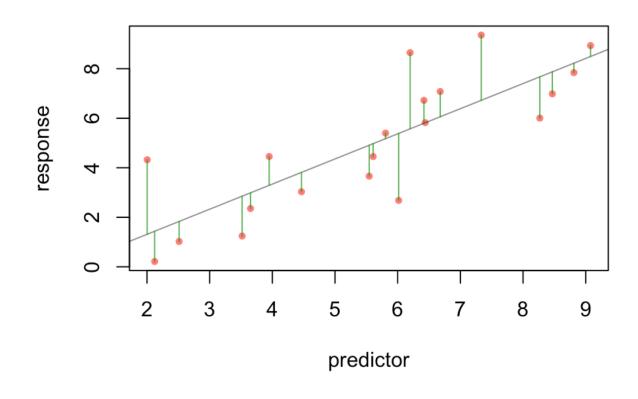
The solution is to build a model that can describe the way that different variables make contributions to an observation.

Regression in R

1) With linear regression we find the linear equation that best predicts the values of Y based on the values of X.

$$2) y = bx + a$$

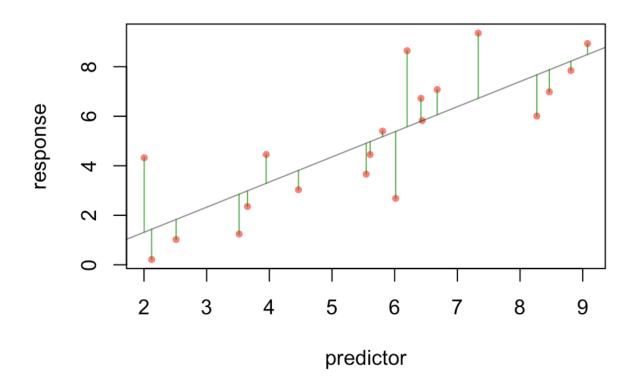
3) Least-squares regression minimizes the squared deviations of the data points from that line.



Example of regression

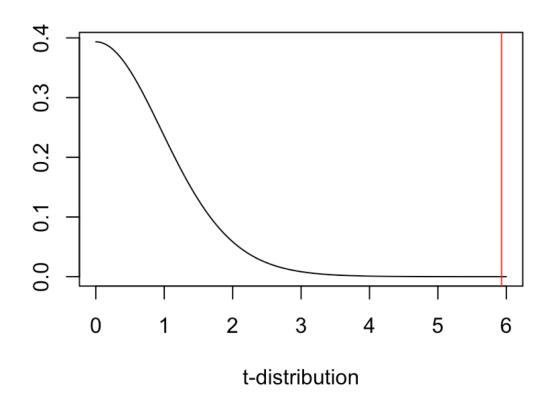
```
set.seed(3)
                                        y = bx + a
x \leftarrow runif(min = 1, max = 10, 20)
y \leftarrow rnorm(20, mean = x, sd = 2)
fit.xy \leftarrow lm(y \sim x)
summary(fit.xy)
Call:
lm(formula = y \sim x)
Residuals:
            1Q Median
   Min
                             3Q
                                    Max
-2.7060 -0.9742 -0.4539 0.9479 3.0728
Coefficients:
            Estimate Std. Error t value Pr(>|t|)
(Intercept) -0.7173
                        1.0302
                                -0.696
                                          0.495
                                 5.943 1.27e-05 ***
             1.0150
                        0.1708
Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' '1
Residual standard error: 1.625 on 18 degrees of freedom
Multiple R-squared: 0.6624, Adjusted R-squared: 0.6437
```

F-statistic: 35.32 on 1 and 18 DF, p-value: 1.267e-05



Example of regression

```
set.seed(3)
                                        y = bx + a
x < -runif(min = 1, max = 10, 20)
y \leftarrow rnorm(20, mean = x, sd = 2)
fit.xy \leftarrow lm(y \sim x)
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Call:
lm(formula = y \sim x)
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Example of regression

```
set.seed(3)
x <- runif(min = 1, max = 10, 20)
y <- rnorm(20, mean = x, sd = 2)
fit.xy <- lm(y ~ x)
summary(fit.xy)</pre>
```

This can help to justify the biological importance assuming you have a regression that is significant. It is the proportion of total variance explained by the regression.

$lm(formula = y \sim x)$

Call:

Residuals:

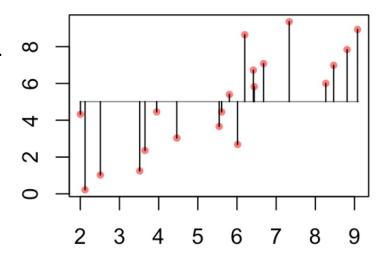
Min 1Q Median 3Q Max -2.7060 -0.9742 -0.4539 0.9479 3.0728

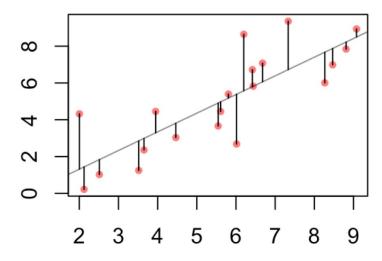
Coefficients:

Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' '1

Residual standard error: 1.625 on 18 degrees of freedom Multiple R-squared: 0.6624, Adjusted R-squared: 0.6437

F-statistic: 35.32 on 1 and 18 DF, p-value: 1.267e-05





Multiple vs Adjusted R-squared

```
Call:
lm(formula = y \sim x)
Residuals:
   Min
           10 Median 30
                                 Max
-2.7060 -0.9742 -0.4539 0.9479 3.0728
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
(Intercept) -0.7173 1.0302 -0.696 0.495
       1.0150 0.1708 5.943 1.27e-05 ***
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' '1
Residual standard error: 1.625 on 18 degrees of freedom
Multiple R-squared: 0.6624, Adjusted R-squared: 0.6437
F-statistic: 35.32 on 1 and 18 DF, p-value: 1.267e-05
```

Adjusted R-squared penalizes for additional parameters

Linear regression uses

- Depict the relationship between two variables in an eye-catching fashion
- Test the null hypothesis of no association between two variables
 - The test is whether or not the slope is zero
- Predict the average value of variable Y for a group of individuals with a given value of variable X
 - variation around the line can make it very difficult to predict a value for a given individual with much confidence
 - Predictions outside of the range of observed data is generally discouraged
- Used both for experimental and observational studies

What are Residuals

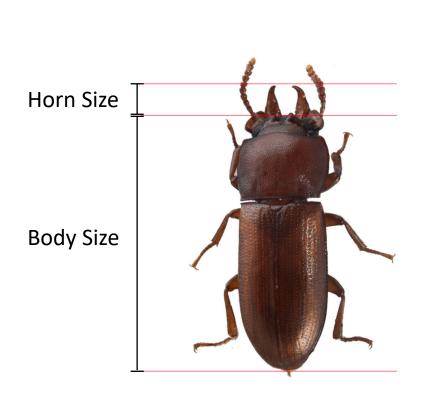
In general, the residual is the individual's departure from the value predicted by the model

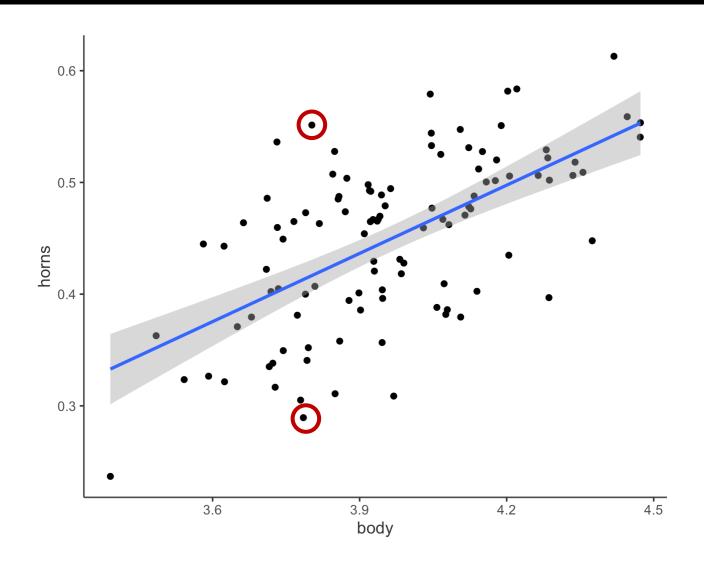
In this case the model is simple – the linear regression – but residuals also exist for more complex models

For a model that fits better, the residuals will be smaller on average

Residuals can be of interest in their own right, because they represent values that have been *corrected* for relationships that might be obscuring a pattern.

What are Residuals





Making that plot

```
ggtheme <- theme_bw() + theme(panel.grid.major = element_blank(),
                              panel.grid.minor = element_blank(),
                              panel.background = element_blank(),
                              panel.border=element_blank(),
                              axis.line = element_line(colour="grey30"),
                              axis.title = element_text(colour="grey20"),
                              axis.text = (element_text(colour="grey30")),
                              legend.title = element_text(colour="grey20"),
                              legend.text = element_text(colour="grey30"))
dat <- read.csv("gnatocerus.csv")</pre>
ggplot(data = dat, aes(x=body, y=horns)) +
  geom_point() + ggtheme +
  geom_smooth(method='lm')
```

Assumptions of Linear Regression

- The true relationship must be linear
- At each value of X, the distribution of Y is normal (i.e., the residuals are normal)
- The variance in Y is independent of the value of X
- Note that there are no assumptions about the distribution of X

Common Problems

Outliers

- Regression is extremely sensitive to outliers
- The line will be drawn to outliers, especially along the x-axis
- Consider performing the regression with and without outliers
- Non-linearity
 - Best way to notice is by visually inspecting the plot and the line fit
 - Try a transformation to get linearity [often a log transformation]
- Non-normality of residuals
 - Can be detected from a residual plot
 - Possibly solved with a transformation
- Unequal variance
 - Usually visible from a scatterplot or from a residual plot

Moving past simple models

- The reason ANOVA is so widely used is that it provides a framework to simultaneously test the effects of multiple factors
- ANOVA also makes it possible to detect interactions among the factors
- ANOVA is a special case of a general linear model
- Linear regression is a special case of a general linear model

GLM and LM function in R

- The GLM and LM function in R takes equations that can be described with the following operators
- + +X include this variable
- : X:Z include the interaction between these variables
- * X*Y include these variables and the interactions between them
- $(X + Z + W)^3$ include these variables and all interactions up to three way

R versus the math implied

glm(y ~ X + W)
$$y_i = \beta_0 + \beta_1 X_i + \beta_2 W_i + \epsilon_i$$

glm(y ~ X * W)
$$y_i = \beta_0 + \beta_1 X_i + \beta_2 W_i + \beta_3 X_i W_i + \epsilon_i$$

Go to R

Betta models