



Chromosome number evolution in skippers (Lepidoptera, Hesperiidae)

Vladimir A. Lukhtanov^{1,2}

1 Department of Entomology, Faculty of Biology, St. Petersburg State University, Universitetskaya nab. 7/9, 199034 St. Petersburg, Russia **2** Department of Karyosystematics, Zoological Institute of Russian Academy of Science, Universitetskaya nab. 1, 199034 St. Petersburg, Russia

Corresponding author: Vladimir A. Lukhtanov (lukhtanov@mail.ru)

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Abstract

Lepidoptera (butterflies and moths), as many other groups of animals and plants, simultaneously represent preservation of ancestral karyotype in the majority of families with a high degree of chromosome number instability in numerous independently evolved phylogenetic lineages. However, the pattern and trends of karyotype evolution in some Lepidoptera families are poorly studied. Here I provide a survey of chromosome numbers in skippers (family Hesperiidae) based on intensive search and analysis of published data. I demonstrate that the majority of skippers preserve the haploid chromosome number n=31 that seems to be an ancestral number for the Hesperiidae and the order Lepidoptera at whole. However, in the tribe Baorini the derived number n=16 is the most typical state which can be used as a (syn)apomorphic character in further phylogenetic investigations. Several groups of skippers display extreme chromosome number variations on within-species (e.g. the representatives of the genus Carcharodus Hübner, [1819]) and between-species (e.g. the genus Agathymus Freeman, 1959) levels. Thus, these groups can be used as model systems for future analysis of the phenomenon of chromosome instability. Interspecific chromosomal differences are also shown to be useful for discovering and describing new cryptic species of Hesperiidae representing in such a way a powerful tool in biodiversity research. Generally, the skipper butterflies promise to be an exciting group that will significantly contribute to the growing knowledge of patterns and processes of chromosome evolution.

Keywords

Lepidoptera, Hesperiidae, karyotype evolution, chromosome number, cryptic species, phylogeny, chromosomal conservatism, chromosomal instability

Introduction

The main karyotypic features of organisms, particularly the number of chromosomes, tend to be stable within species (White 1973, King 1993). New chromosomal rearrangements usually originate as heterozygotes and are often – although not always (Lukhtanov et al. 2011) – associated with heterozygote disadvantage. The spread of such rearrangements to fixation within a large population has low probability (King 1993). Therefore, many organisms are characterized by chromosomal conservatism, a situation in which all closely related taxa demonstrate the same chromosome number.

In contrast to chromosomal conservatism, chromosomal instability characterizes situations where multiple closely related taxa (populations, subspecies and/or species) belonging to a single phylogenetic lineage differ drastically from each other by major chromosomal rearrangements, sometimes resulting in high variability in chromosome number.

Both phenomena - chromosomal conservatism and chromosomal instability are clearly expressed in insects of the order Lepidoptera (butterflies and moths). The modal haploid number of chromosomes (n) of n = 31 or n = 30 (Suomalainen 1969, Lukhtanov 2000) is preserved in the majority of lepidopteran families (Robinson 1971). At the same time, numerous cases of chromosomal instability have been discovered in the butterfly families, e.g. in Papilionidae (Emmel et al. 1995), Pieridae (Lukhtanov 1991, Lukhtanov et al. 2011, Dinca et al. 2011), Nymphalidae (Brown et al. 1992, 2004, 2007a, 2007b) and Riodinidae (Brown et al. 2012). This phenomenon was analyzed in more detail in the family Lycaenidae (Kandul et al. 2004, 2007, Lukhtanov et al. 2005, 2006, 2008, Vershinina and Lukhtanov 2010, 2013, Vila et al. 2010, Talavera et al. 2013, Przybyłowicz et al. 2014).

Skippers (the family Hesperiidae) are studied to a lesser extent with the respect of karyotype evolution than the other butterfly families mentioned above (but see: Emmel and Trew 1973, Saura et al. 2013). This family includes about 4000 species under 567 genera and is a globally distributed group found in all continents except Antarctica (Warren et al. 2008). The tribal level classification of skippers, based on combined analysis of molecular and morphological data, was recently elaborated by Warren and colleagues (Warren et al. 2008, 2009).

Here I provide a first world-wide survey of chromosome numbers in skippers based on intensive search and analysis of published data.

Results

The results of literature search are presented in the Table below. It includes all the discovered chromosome counts except n=13 for *Ochlodes venatus* (Bremer et Grey, 1853), noted by Bigger (1960) as "*Augiades venata*". The name *Ochlodes venatus* was long used for the *Ochlodes* species of Europe, but it actually refers to its Far Eastern sister species, and the European taxon is now called *O. sylvanus* (Esper, 1777) (ICZN 2000). Both European and Far Eastern species have the same chromosome number n=29 (Federley

1938, Lorković 1941, Abe et al. 2006), not n=13 as indicated by Bigger (1960). Thus, the species name used by Bigger (1960) was probably misidentification.

The classification of skippers accepted in this paper follows Warren and colleagues (Warren et al. 2008, 2009).

Discussion

Modal chromosomal numbers

The table gives the chromosome numbers of 205 species of skippers, i.e. about 5% of the species of the world fauna. This number is not enough to infer any final statements about peculiarities of chromosome numbers distribution within the Hesperiidae. However, several tentative conclusions can be made. The haploid chromosome number n=31 was found in 50 studied species of skippers and, thus, it is a clear modal number for the family at whole. Interestingly, n=31 was found in representatives of all investigated subfamilies, except for Heteropterinae. However, in the last subfamily only one species was karyologically studied until now, and discovery of n=31 in Heteropterinae is not excluded in future. The next most common numbers are n=29 (43 species), n=30 (33 species) and n=28 (13 species).

Subfamilies Coeliadinae and Eudaminae have a sharp peak at n=31. In the subfamily Trapezitinae n=31 was also found (only one species studied).

Within the subfamily Pyrginae, the modal number n=31 is found in the tribe Erynnini. The tribe Pyrrhopygini is characterized by the most common n=28. The modal number in the tribe Tagiadini is n=30. The tribe Carcharodini has peaks at n=30 and n=31. In the tribe Pyrgini, n=29, n=30 and n=31 were found as the most common numbers.

In the family Heteropterinae n=29 was found (only one species studied).

Within the subfamily Hesperiinae, the tribes Taractrocerini, Thymelicini, Calpodini, Moncini and Hesperiini are characterized by the most common n=29. Very variable chromosome numbers (from n=5 to n=50) were found in the tribe Aeromachini. It is difficult to infer the modal number for the last tribe. However, it should be noted that one species, *Thoressa varia*, has n=31 as the majority of other skippers. The tribe Baorini (subfamily Hesperiinae) has a clear peak at n=16, so it is exceptional with respect to the modal number of chromosomes.

The overall evidence indicates that chromosome numbers of Coeliadinae, Eudaminae, Trapezitinae, Pyrginae and Hesperiinae conform to the lepidopteran modal of n=31 (Robinson 1971). This number seems to be an ancestral one for the Hesperiidae as for the order Lepidoptera at whole (Suomalainen 1969, Lukhtanov 2000). This modal number (or its deviation to n=30, n=29 and 28) were preserved in the majority of skippers. However, in the tribe Baorini the number n=16 was evolved and, thus, represents a derived trait which can be used as a (syn)apomorphic character in further phylogenetic studies of the family Hesperiidae.

Table 1. Chromosome number of skippers (Lepidoptera, Hesperiidae) of the world fauna (Us are univalents; 2n is diploid chromosome number).

Years of the species descriptions are given square brackets in cases where they were not stated in the original sources but were inferred from reliable external evidence.

#	Species	Haploid chromosome number	Country	Reference
Subfa	mily Coeliadinae			1
1	Bibasis aquilina (Speyer, 1879)	29	Japan	Maeki 1953
	B. a. chrysaeglia (Butler, 1881)	31 (2n=62)	Japan	Abe et al. 2006
2	B. jaina formosana Fruhstorfer, 1911	31	Taiwan	Maeki and Ae 1968b
3	Choaspes benjaminii (Guérin-Méneville, 1843)	31	Japan	Maeki 1953
	Ch. b. japonica (Murray, 1875)	31	Japan	Saitoh et al. 1978
4	Coeliades anchises jucunda (Butler, 1881)	30	Oman	Saitoh 1982
5	C. ernesti (Grandidier, 1867)	31	Madagascar	de Lesse 1972
6	C. fervida (Butler, 1880)	23	Madagascar	de Lesse 1972
7	C. forestan arbogastes (Guenee, 1863)	31	Madagascar	de Lesse 1972
8	C. ramanatek (Boisduval, 1833)	31	Madagascar	de Lesse 1972
Subfa	mily Euschemoninae no chromosomal	data available		
Subfa	mily Eudaminae			
9	Achalarus casica (Herrich-Schäffer, 1869)	29	USA (Texas)	Emmel and Trew 1973
10	A. lyciades (Geyer, 1832)	31	USA (Connecticut)	Maeki 1961
11	A. toxeus (Plötz, 1882)	16	Mexico	Maeki and Remington 1960
12	Astraptes anaphus (Godman et Salvin, 1896)	31	Bolivia	de Lesse 1967a
13	A. fulgerator (Walch, 1775)	31	Peru	Kumagai et al. 2010
14	A. naxos (Hewitson, 1867)	31	Brazil	Saura et al. 2013
15	A. phalaecus (Godman et Salvin, 1893)	25	Guatemala	de Lesse 1967a
16	A. longipennis (Plötz, 1882)	31	Costa Rica	Kumagai et al. 2010
		31	Peru	Kumagai et al. 2010
		31	Brazil	Kumagai et al. 2010
17	Autochton sp.	20, 21	Brazil	Kumagai et al. 2010
18	Chioides albofasciatus (Hewitson, 1867)	31	Mexico	de Lesse 1970a
	Ch. albofasciatus (Hewitson, 1867) (as Ch. catillus)	31	Mexico	Maeki and Remington 1960
	Ch. albofasciatus (Hewitson, 1867)	31	USA (Texas)	Emmel and Trew 1973
19	Entheus priassus pralina Evans, 1952	22	Brazil	Saura et al. 2013
20	Epargyreus barisses (Hewitson, 1874)	31	Argentina	de Lesse 1967
21	E. clarus (Cramer, 1775)	31	USA (Florida)	Maeki 1961
22	E. clavicornis tenda Evans, 1955	ca 29-30	Guatemala	de Lesse 1970a
23	Oechydrus chersis (Herrich-Schäffer, 1869)	31	Bolivia	de Lesse 1967a

#	Species	Haploid chromosome number	Country	Reference
24	Phocides polybius phanias (Burmeister, 1880)	16	Brazil	Saura et al. 2013
25	Tarsoctenus praecia plutia (Hewitson, 1857)	15	Brazil	Saura et al. 2013
26	Thorybes pylades pylades (Scudder, 1870)	31	USA (Connecticut)	Maeki 1961
27	Udranomia spitzi (Hayward, 1942)	29	Brazil	de Lesse and Brown 1971
28	Urbanus dorantes dorantes (Stoll, 1790)	31	Mexico	de Lesse 1970a
29	U. doryssus doryssus (Swainson, 1831)	14	Costa Rica	Kumagai et al. 2010
30	Urbanus proteus (Linnaeus, 1758)	31	Bolivia	de Lesse 1967a
		31	Mexico	de Lesse 1970a
		31	USA (Florida)	Maeki 1961
31	U. simplicius (Stoll, 1790)	31	Argentina	de Lesse 1967a
32	U. teleus (Hübner, 1821)	31	Argentina	de Lesse 1967a
Subfar	mily Pyrginae			
	Pyrrhopygini			
33	Elbella lamprus (Hopffer, 1874)	40	Brazil	de Lesse 1970a
34	(?) Jemadia sp.	32(?)	Brazil	Saura et al. 2013
35	Mimoniades montana J. Zikán, 1938	27	Brazil	Saura et al. 2013
36	M. nurscia (Swainson, 1821)	28	Ecuador	de Lesse 1967a
	M. n. malis (Godman et Salvin, 1879)	28	Colombia	Saura et al. 2013
37	Mimoniades sp.	21	Colombia	Saura et al. 2013
38	Mimoniades sp.	28	Colombia	Saura et al. 2013
39	M. versicolor (Latreille, [1824])	28	Brazil	de Lesse and Brown 1971
40	Pyrrhopyge charybdis Westwood, 1852	14(?)	Brazil	Saura et al. 2013
41	P. pelota Plötz, 1879	28	Argentina	de Lesse 1967a
42	Pyrrhopyge sp.	15	Brazil	Saura et al. 2013
43	Sarbia sp.	30	Brazil	Saura et al. 2013
Tribe '	Tagiadini	-		
44	Daimio tethys (Ménétriés, 1857)	30	Japan	Maeki 1953, Maeki and Makino 1953
45	D. t. moorei Mabille, 1876	30	Taiwan	Maeki and Ae 1968b
46	Eagris lucetia (Hewitson, 1876)	30	Uganda	de Lesse 1968
47	E. sabadius astoria Holland, 1896	30	Kenya	de Lesse 1968
48	Eretis lugens (Rogenhofer, 1891	28	Kenya	de Lesse 1968
	Celaenorrhinini			
49	Sarangesa phidyle (Walker, 1870)	29	Senegal	de Lesse and Condamin 1962
Tribe	Carcharodini		·	<u> </u>
50	Carcharodus alceae (Esper, [1780])	31	Croatia	Lorkovic 1941
51	C. boeticus Reverdin, 1913	43-47	Spain	de Lesse 1960
	C. boeticus Reverdin, 1913	40–52	France	de Lesse 1960
	C. boeticus Reverdin, 1913	38–46	Italy	de Lesse 1960
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		Haploid	_	
#	Species	chromosome	Country	Reference
52	C. dravira (Moore, 1874)	37–48 (with Us)	Iran	de Lesse 1960
53	C. flocciferus (Zeller, 1847)	32–41 (with Us)	France (Cauterets)	de Lesse 1960
54	C. flocciferus (Zeller, 1847)	42–58 (with Us)	Italy	de Lesse 1960
55	C. lavatherae (Esper, [1783])	30	France (Salau, Ariege)	de Lesse 1960
56	C. orientalis Reverdin, 1913	31–32	Lebanon	de Lesse 1960
		30	Turkey (Van)	de Lesse 1960
		30–37 (with Us)	Turkey (Amasya)	de Lesse 1960
57	C. stauderi ambiguus Verity, 1925	30	Lebanon	de Lesse 1960
		30	Turkey	de Lesse 1960
58	Hesperopsis alpheus (W. H. Edwards, 1876) (as Pholisora)	34	USA (Texas)	Emmel and Trew 1973
59	Muschampia nomas (Lederer, 1855)	30	Lebanon	de Lesse 1960
60	M. proteides (Wagner, 1929)	30	Lebanon	Larsen 1975
61	M. proto (Ochsenheimer, 1808)	30	Spain	de Lesse 1960
		30	Lebanon	Larsen 1975
62	Pholisora catullus (Fabricius, 1793)	29	?USA	Lorkovic in Robinson 1971
63	Spialia orbifer (Hübner, [1823])	30	Croatia	Lorkovic 1941
		31	Turkey	de Lesse 1960
64	S. phlomidis (Herrich-Schäffer, [1845])	31	Turkey	de Lesse 1960
65	S. sertorius (Hoffmannsegg, 1804)	31	Slovenia	Lorkovic 1941
Tribe	Erynnini			
66	Chiomara asychis georgina (Reakirt, 1868)	31	Mexico	de Lesse 1970a
	Ch. asychis georgina (Reakirt, 1868)	32	USA (Texas)	Emmel and Trew 1973
67	Chiomara sp.	31	Trinidad	Wesley and Emmel 1975
68	Ebrietas anacreon (Staudinger, 1876)	31	Argentina	de Lesse 1967a
69	E. osyris (Staudinger, 1876)	31	Argentina	de Lesse 1967a
70	Erynnis baptisiae (W. Forbes, 1936)	31	USA (Connecticut)	Maeki 1961
71	E. funeralis (Scudder et Burgess, 1870)	31	Argentina	de Lesse 1967a
72	E. horatius (Scudder et Burgess, 1870)	31	USA (Florida)	Maeki 1961
73	E. icelus (Scudder et Burgess, 1870)	30	USA (Connecticut)	Maeki 1961
74	E. juvenalis juvenalis (Fabricius, 1793)	30	USA (Connecticut)	Maeki 1961
75	E. lucilius (Scudder et Burgess, 1870)	31	USA (Connecticut)	Maeki and Remington 1960a
76	E. marloyi (Boisduval, [1834])	31	Lebanon	de Lesse 1960
77	E. montanus (Bremer, 1861)	31 (2n=62)	Japan	Abe et al. 2006
	E. montanus (Bremer, 1861)	31	Japan	Maeki 1953
	-			

#	Species	Haploid chromosome number	Country	Reference
78	E. persius (Scudder, 1863)	31	USA (Connecticut)	Maeki 1961
79	E. tages (Linnaeus, 1758)	31	Croatia	Lorkovic 1941
		31	France	de Lesse 1960
		31	England	Bigger 1960
80	E. tristis tatius (W. H. Edwards, 1883)	31	USA (Texas)	Emmel and Trew 1973
81	Gesta gesta (Herrich-Schäffer, 1863)	32	Tobago	Wesley and Emmel 1975
82	Grais stigmaticus (Mabille, 1883)	31	Mexico	Maeki and Remington 1960a
83	Theagenes albiplaga (C. Felder et R. Felder, 1867)	31	Bolivia	de Lesse 1967a
Tribe A	Achlyodidini			
84	Achlyodes pallida (R. Felder, 1869) (as A. selva)	15	Bolivia	de Lesse 1967a
		15	Mexico	de Lesse 1970a
85	Zera zera zera (Butler, 1870)	34	Brazil	de Lesse and Brown 1971
Tribe 1	Pyrgini			
86	Anisochoria sublimbata Mabille, 1883	31	Argentina	de Lesse 1967a
87	Antigonus erosus (Hübner, [1812])	31	Mexico	de Lesse 1970a
88	A. liborius Plötz, 1884	31	Argentina	de Lesse 1967a
89	Celotes nessus (W. H. Edwards, 1877)	14, 13	USA (Texas)	Emmel and Trew 1973
90	Heliopetes arsalte (Linnaeus, 1758)	30	Bolivia	de Lesse 1967a
	H. arsalte (Linnaeus, 1758)	30	Mexico	de Lesse 1970a
91	H. laviana (Hewitson, 1868)	29	USA (Texas)	Emmel and Trew 1973
92	H. macaira (Reakirt, [1867])	29	USA (Texas)	Emmel and Trew 1973
93	H. omrina (Butler, 1870)	30	Argentina	de Lesse 1967a
94	Heliopyrgus americanus (Blanchard, 1852)	30	Chile	de Lesse 1967a
95	Paches loxus (Westwood, [1852])	31	Guatemala	de Lesse 1970a
96	<i>Pyrgus aladaghensis</i> De Prins et van der Poorten, 1995	ca 18–21	Turkey	Lukhtanov and Kandul 1995 (in Hesselbarth et al. 1995)
97	P. albescens Plötz, 1884	30 (2n=60)	USA (Texas)	Goodpasture 1976
	P. albescens Plötz, 1884	28	USA (Texas)	Emmel and Trew 1973
98	P. alveus (Hübner, [1803])	24	Finland	Federley 1938
		24	Croatia	Lorkovic 1941
		24	Turkey	Lukhtanov and Kandul 1995 (in Hesselbarth et al. 1995)
99	P. bellieri (Oberthür, 1910)	27	France	de Lesse 1960
100	P. bocchoris (Hewitson, 1874)	30	Argentina	de Lesse 1967a
101	P. bolkariensis De Prins et van der Poorten, 1995	30	Turkey	Lukhtanov and Kandul 1995 (in Hesselbarth et al. 1995)
102	P. cacaliae (Rambur, 1839)	30	Italy	de Lesse 1960
103	P. carlinae (Rambur, [1839])	30	Italy	de Lesse 1960
104	P. carthami (Hübner, [1813])	29	Italy	de Lesse 1960
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#	Species	Haploid chromosome number	Country	Reference
105	P. cirsii (Rambur, [1839])	30	France (Peyreleau, Aveyron)	de Lesse 1960
106	P. fides Hayward, 1940	30	Chile	de Lesse 1967a
107	P. maculates (Bremer et Grey, 1852)	31 (2n=62)	Japan	Abe et al. 2006
108	P. malvae (Linnaeus, 1758)	31	Finland	Federley 1938
		33	England	Bigger 1960
109	P. oileus (Linnaeus, 1767)	30 (2n=60)	USA (Texas)	Goodpasture 1976
		32	USA (Texas)	Emmel and Trew 1973
110	P. onopordi (Rambur, [1839])	30	France	Lorkovic 1941
111	P. serratulae (Rambur, [1839])	30	France	Lorkovic 1941
112	Trina geometrina geometrina (C. Felder et R. Felder, 1867)	31	Brazil	de Lesse and Brown 1971
Subfan	nily Heteropterinae			
113	Butleria quilla Evans, 1939	29	Chile	de Lesse 1967a
Subfan	nily Trapezitinae			
114	Trapezites eliena Hewitson, 1868	31	Australia	Maeki and Ogata 1971
	nily Hesperiinae	31	Australia	Iviacki alid Ogađa 17/1
	Aeromachini			
		2.6	Motor	E 10/0
115	Aegiale hesperiaris (Walker, 1856)	24	Mexico	Freeman 1969
116	Agathymus alliae (Stallings et Turner, 1957)	38	USA (Arizona)	Freeman 1969
117	A. aryxna (Dyar, 1905)	5	Mexico	Freeman 1969
118	A. baueri (Stallings et Turner, 1954)	15	USA (Arizona)	Freeman 1969
119	A. chisosensis (Freeman, 1952)	18	USA (Texas)	Freeman 1969
120	A. estelleae valverdiensis Freeman, 1966	9	USA (Texas)	Freeman 1969
	A. e. estelleae (Stallings et Turner, 1958)	9	Mexico	Freeman 1969
121	A. freemani Stallings, Turner et Stallings, 1960	15	USA (Arizona)	Freeman 1969
122	A. gilberti Freeman, 1964	21	USA (Texas)	Freeman 1969
123	A. mariae chinatiensis Freeman, 1964	22	USA (Texas)	Freeman 1969
	A. mariae lajitaensis Freeman, 1964	22	USA (Texas)	Freeman 1969
	A. mariae mariae (Barnes et Benjamin, 1924)	22	USA or Mexico	Freeman 1969
	A. mariae rindgei Freeman, 1964	22	USA (Texas)	Freeman 1969
124	A. micheneri Stallings, Turner et Stallings, 1961	20	Mexico	Freeman 1969
125	A. neumoegeni florenceae (Stallings et Turner, 1957)	10	USA (Texas)	Freeman 1969
	A. neumoegeni macalpinei (Freeman, 1955)	10	USA (Texas)	Freeman 1969
126	A. polingi (Skinner, 1905)	10	USA (Arizona)	Freeman 1969
127	A. remingtoni (Stallings et Turner, 1958)	9	Mexico	Freeman 1969

#	Species	Haploid chromosome number	Country	Reference
128	Alera vulpina (C. Felder et R. Felder, 1867)	ca27	Ecuador	de Lesse 1967a
129	Ankola fan (Holland, 1844)	10	Uganda	De Lesse 1968
130	Arotis derasa (Herrich-Schäffer, 1870) (as Euphyes)	28	Brazil	de Lesse and Brown 1971
131	Erionota thrax thrax (Linnaeus, 1767)	29	Malaysia	Saitoh and Kumagai 1974
132	Euphyes leptosema Mabille, 1891	ca28	Argentina	de Lesse 1967a
133	Megathymus coloradensis coloradensis Riley, 1877	27	USA	Freeman 1969
134	M. coloradensis kendalli Freeman, 1965	27	USA (South central Texas)	Freeman 1969
	M. coloradensis louiseae Freeman, 1963	27	USA (Western Texas)	Freeman 1969
	M. coloradensis navajo Skinner, 1911	27	USA	Freeman 1969
	M. coloradensis reinthali Freeman, 1963	27	USA (Texas)	Freeman 1969
	M. coloradensis reubeni Stallings, Turner et Stallings, 1963	27	USA (Texas)	Freeman 1969
	M. coloradensis stallingsi Freeman, 1943	27	USA	Freeman 1969
	M. coloradensis wilsonorum Stallings et Turner, 1958	27	?Mexico	Freeman 1969
135	M. violae Stallings et Turner, 1956	27	USA	Maeki 1961, Freeman 1969
136	M. yuccae buchholzi Freeman, 1952	26	USA (Florida)	Freeman 1969
137	Pardaleodes incerta (Snellen, 1872)	17	Uganda	de Lesse 1968
138	Stallingsia maculosus (Freeman, 1955)	50	USA (Texas)	Maeki 1961, Freeman 1969
139	Suastus gremius (Fabricius, 1798)	23	Taiwan	Maeki and Ae 1968b
140	Thoressa varia (Murray, 1875)	31 (2n=62)	Japan	Abe et al. 2006
141	T. varia (Murray, 1875)	31	Japan	Maeki 1953
Tribe B	Baorini			
142	Gegenes gambica (Mabille, 1878)	41	Yemen	Saitoh 1984
		41	Turkey	de Lesse 1960
		41	Lebanon	Larsen 1982
143	Gegenes nostrodamus (Fabricius, 1793)	15	Egypt	Larsen 1982
		15	Israel	Saitoh 1979, Larsen 1982
144	Gegenes pumilio (Hoffmansegg, 1804)	24	France	de Lesse 1960
		24	Alger	de Lesse 1967b
145	Parnara guttata (Bremer et Grey, 1852)	16	Japan	Maeki 1953, Maeki and Makino 1953
	1/2-2/	16	China	Saitoh and Abe 1981
146	Pelopidas conjucta conjucta (Herrich-Schäffer, 1869)	16	Hong Kong	Maeki and Ae 1968a
147	P. jansonis (Butler, 1878)	16 (2n=32)	Japan	Abe et al. 2006

#	Species	Haploid chromosome number	Country	Reference
148	P. mathias (Fabricius, 1798)	16	Japan	Maeki and Remington 1960
149	P. thrax (Hübner, [1821])	16	Lebanon	Larsen 1975
150	Polytremis lubricans (Herrich-Schäffer, 1869)	16	Taiwan	Maeki and Ae 1968b
151	P. pellucida (Murray, 1875)	16, 17, 18 (2n=32, 33)	Japan	Abe et al. 2006
		16	Japan	Maeki and Remington 1960
152	Zenonia zeno (Trimen, 1864)	16	Uganda	de Lesse 1968
Tribe T	aractrocerini			
153	Ocybadistes walkeri sothis Waterhouse, 1933	28	Australia	Maeki and Ogata 1971
154	Potanthus flavus (Murray, 1875)	29 (2n=58)	Japan	Abe et al. 2006
155	Telicota ancilla horisha Evans, 1934	29	Taiwan	Maeki and Ae 1968b
156	Telicota colon stinga Evans, 1949	29	Japan (Okinava)	Abe et al. 2006
157	<i>T. ohara formosana</i> Fruhstorfer, 1911	29 (2n=58)	Taiwan	Abe et al. 2006
Tribe T	Thymelicini			
158	Copaeodes minima (W.H. Edwards, 1870)	29	USA (Florida)	Maeki 1961
159	Thymelicus sylvestris (Poda, 1761)	27	England	Bigger 1960
160	Th. sylvaticus (Bremer, 1861)	10 (2n=20)	Japan	Abe et al. 2006
161	Th. acteon (Rottemburg, 1775)	28	Spain	de Lesse 1970c
162	Th. hyrax (Lederer, 1861)	29	Lebanon	Larsen 1975
163	Th. leoninus (Butler, 1878)	9 (2n=18)	Japan	Abe et al. 2006
164	Th. lineola (Ochsenheimer, 1808)	29	Finland	Federley 1938
		29	Lebanon	Larsen 1975
Tribe C	Calpodini			
165	Ebusus ebusus (Cramer, [1780])	29	Mexico	de Lesse 1970a
166	Lychnuchus celsus (Fabricius, 1793)	30	Brazil	de Lesse and Brown 1971
167	Panoquina hecebolus (Scudder, 1872)	29	USA (Texas)	Emmel and Trew 1973
168	Panoquina ocola (W. H. Edwards, 1863)	29	USA (Texas)	Emmel and Trew 1973
169	P. panoquin (Scudder, 1863)	29	USA (Florida)	Maeki 1961
170	P. panoquinoides (Skinner, 1891)	29	USA (Texas)	Emmel and Trew 1973
Tribe A	Anthoptini no chromosomal data avail	able		
Tribe N	Moncini			
171	Amblyscirtes aenus W.H. Edwards, 1878	28, 29	USA (Texas)	Emmel and Trew 1973
172	A. cassus W. H. Edwards, 1883	29	USA (Texas)	Emmel and Trew 1973
173	A. celia (Skinner, 1895)	29	USA (Texas)	Emmel and Trew 1973
174	A. phylace W.H. Edwards, 1878	29	USA (Texas)	Emmel and Trew 1973
175	A. texanae Bell, 1927	29	USA (Texas)	Emmel and Trew 1973
176	A. vialis (W. H. Edwards, 1862)	29	USA (Connecticut)	Maeki 1961
177	Cymaenes sp.	31	Tobago	Wesley and Emmel 1975

#	Species	Haploid chromosome number	Country	Reference
178	Enosis immaculata immaculata (Hewitson, 1868)	29	Ecuador	Kumagai et al. 2010
179	Lerema accius (Smith, 1797)	29 (2n=58)	USA (Texas)	Goodpasture 1976
		29	USA (Texas)	Emmel and Trew 1973
180	Moeris vopiscus (Herrich-Schäffer, 1869)	27	Peru	Kumagai et al. 2010
181	Nastra lherminier (Latreille, [1824])	30	USA (Connecticut)	Maeki 1961
182	Thargella caura (Plötz, 1882)	25	Brazil	de Lesse and Brown 1971
183	Vettius coryna (Hewitson, [1866])	31, ca32	Ecuador	de Lesse 1967a
184	V. phyllus prona Evans, 1955	26	Brazil	de Lesse and Brown 1971
185	V. triangularis (Hübner, [1831])	26	Brazil	Kumagai et al. 2010
Tribe H	lesperiini			
186	Asbolis capucinus (Lucas, 1857)	48	USA (Florida)	Maeki 1961
187	Cynea iquita (Bell, 1941)	29	Argentina	de Lesse 1967a
188	Hesperia comma (Linnaeus, 1758)	28	Italy	de Lesse 1970c
		28	Lebanon	Larsen 1975
189	H. florinda Butler, 1878	28 (2n=56)	Japan	Abe et al. 2006
190	Hylephila fasciolata (Blanchard, 1852)	29	Argentina	de Lesse 1967a
191	H. phyleus (Drury, 1773)	29	Argentia	de Lesse 1967a
		29	USA (Florida)	Maeki 1961
192	H. signata (Blanchard, 1852)	29	Chile	de Lesse 1967a
193	Ochlodes ochraceus (Bremer, 1861)	29 (2n=58)	Japan	Abe et al. 2006
		24	Japan	Maeki and Remington 1960
194	O. sylvanoides (Boisduval, 1852)	29	USA	Maeki 1961
195	O. sylvanus (Esper, 1777)	29	Finland	Federley 1938
		29	Croatia	Lorkovic 1941
196	O. venatus (Bremer et Grey, 1853) (as sylvanus Esper, 1777)	29 (2n=58)	Japan	Abe et al. 2006
197	Oligoria maculata (W. H. Edwards, 1865)	29	USA (Florida)	Maeki 1961
198	Poanes hobomok hobomok (Harris, 1862)	29	?USA	Lorkovic in Robinson 1971
199	P. taxiles (W. H. Edwards, 1881)	29	USA	Maeki 1961
200	P. zabulon (Boisduval et Le Conte, [1837]) (as Polites zabulon)	29	USA (Connecticut)	Maeki 1961
201	Polites themistocles (Latreille, [1824])	29	USA (Florida)	Maeki 1961
202	P. vibex catilina (Plötz, 1886)	29	Argentina	de Lesse 1967a
	P. vibex praeceps (Scudder, 1872)	27	USA (Texas)	Emmel and Trew 1973
	P. vibex vibex (Geyer, 1832)	29	USA (Florida)	Maeki 1961
203	Wallengrenia egeremet (Scudder, 1863)	28	USA (Texas)	Emmel and Trew 1973
204	W. otho curassavica (Snellen, 1887)	28-30	USA (Texas)	Emmel and Trew 1973
205	W. premnas (Wallengren, 1860)	27	Argentina	de Lesse 1967

Between- and within-species variations in chromosome number

Several groups of skippers display extreme chromosome number variations at the with-in-species level (Table). The most extreme variations in number of chromosome elements were observed in first meiotic metaphase of *Carcharodus boeticus*, *C. dravira* and *C. flocciferus* (Table, de Lesse 1960). The nature of these variations remains unknown, and there are two plausible explanations for this phenomenon. First, this variation can be explained by presence of so-called B-chromosomes (=additional chromosomes, =supernumerary chromosomes) (de Lesse 1960). B-chromosomes consist mainly of repetitive DNA and can sometimes accumulate through processes of mitotic or meiotic drive (Jones et al. 2008). B-chromosomes can be distinguished from normal A-chromosomes because they are usually smaller and can be seen as additional chromosomes present in only some of the individuals in a population (Camacho et al. 2000, Jones et al. 2008).

Second, this kind of variation can be caused by violations in meiotic chromosome pairing resulting in appearance of univalents (instead of bivalents) in meiotic prophase (Lorković 1990). This type of variation was studied in detail by Maeki and Ae (1979) in butterfly genus *Papilio* and is expected if regular or irregular interspecific mating occurs in nature. Anyway, the nature of intraspecific variations observed in *Carcharodus* is different from that discovered in the Wood White butterfly *Leptidea sinapis* (Linnaeus, 1758) (Pieridae). In the last species the compared range of intraspecific variation in chromosome number (from n=28 to n=53) was caused by multiple chromosome fusions/fissions accumulated within the species (Lukhtanov et al. 2011, Dinca et al. 2011).

Between-species variation exists in numerous genera of skippers (Table 1) and is especially expressed in the Nearctic genus *Agathymus* Freeman, 1959, in which the range of haploid numbers was discovered from n = 5 in *A. aryxna* to n=38 in *A. alliae* (Freeman 1969). This range is comparable of even exceeds the range found in chromosomally diverse genera from other butterfly families (Lorković 1990, Lukhtanov et al. 2005, Talavera et al. 2013). Thus, the genera of Hesperiidae can be used as model systems for future analysis of the phenomenon of chromosome instability.

Detecting cryptic species using analysis of chromosomal differences

Recent years karyological data have been widely used in studies of butterfly taxonomy and in biodiversity research as main or additional chracters for detecting cryptic species (e.g. Dinca et al. 2011) and for synonymizing biological entities that were incorrectly described as distinct species (e.g. Vila et al. 2010). The family Hesperiidae is not excluded in this respect. In the genus *Gegenes* Hübner, [1819], two cryptic species *G. pumilio* (n=24) and *G. gambica* (n=41) were discovered through extensive chromosome analysis of different populations (de Lesse 1960, 1967b, Larsen 1982, Saitoh 1984).

In the genus *Pyrgus* Hübner, [1819], our unpublished chromosome data (see Table) were used to recognize and then to describe two morphologically similar species, *P. bolkariensis* and *P. aladaghensis* (De Prins and van der Poorten 1995).

Thus, interspecific chromosomal differences are useful for discovering and describing new cryptic species of Hesperiidae representing in such a way a powerful tool in biodiversity research.

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