

Group 2 : SpikeVar & TykeVar Simulation of Mosaic Variants in Sequencing Data

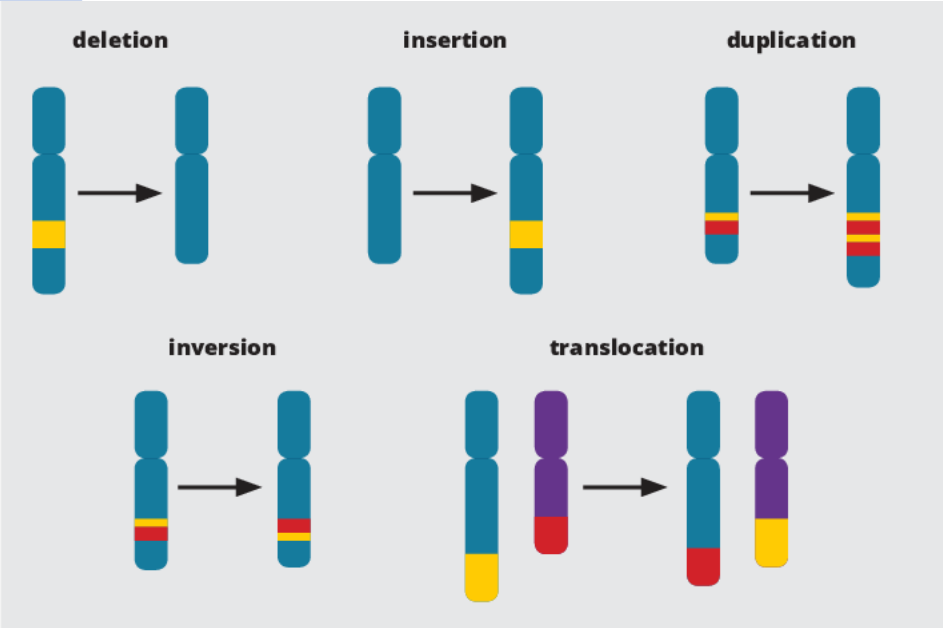
Farhang Jaryani,

Fritz Sedlazeck, Lab

Human Genome Sequencing Center

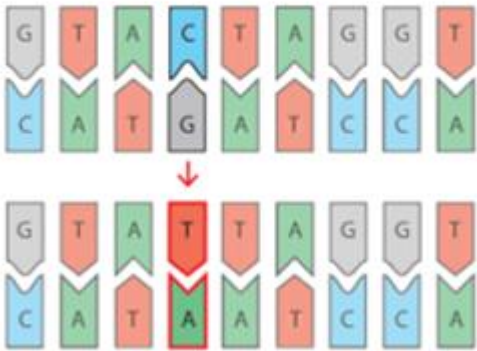
Baylor college of Medicine

Structural Variants



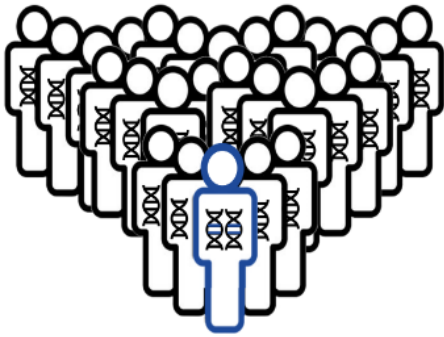
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Deletion	1	3		
Inversion	1	3	2	
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Tandem Duplication	1	1	2	3
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Mobile Element Insertion	1	2	Mobile Element	3
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SNV



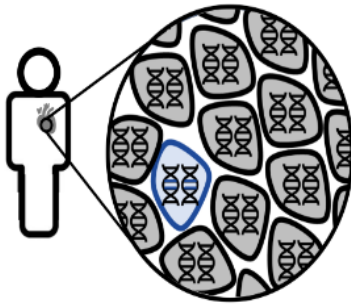
Mosaic Variants

Population of individuals



Rare variants

Population of cells



Mosaic variants

Mosaic Variants:

- ✓ Potentially associated with disease
- ✓ Mixed in sequence data
- ✓ Requires mosaic variant callers to detect



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Disease-associated mosaic variation in clinical exome sequencing: a two-year pediatric tertiary care experience

Cecelia R. Miller, Kristy Lee, Ruthann B. Pfau, Shalini C. Reshmi, Donald J. Corsmeier, Sayaka Hashimoto, Ashita Dave-Wala, Vijayakumar Jayaraman, Daniel Koboldt, Theodora Matthews, Danielle Mouhlas, Maggie Stein, Aimee McKinney, Tom Grossman, Benjamin J. Kelly, Peter White, Vincent Magrini, Richard K. Wilson, Elaine R. Mardis, Catherine E. Cottrell

Comprehensive Cancer Center, Comprehensive Cancer Center - Innate Immunity, Pathology, Pediatrics, Center for Clinical and Translational Science, Comprehensive Cancer Center - Molecular Biology and Cancer Genetics, Comprehensive Cancer Center - Experimental Therapeutics

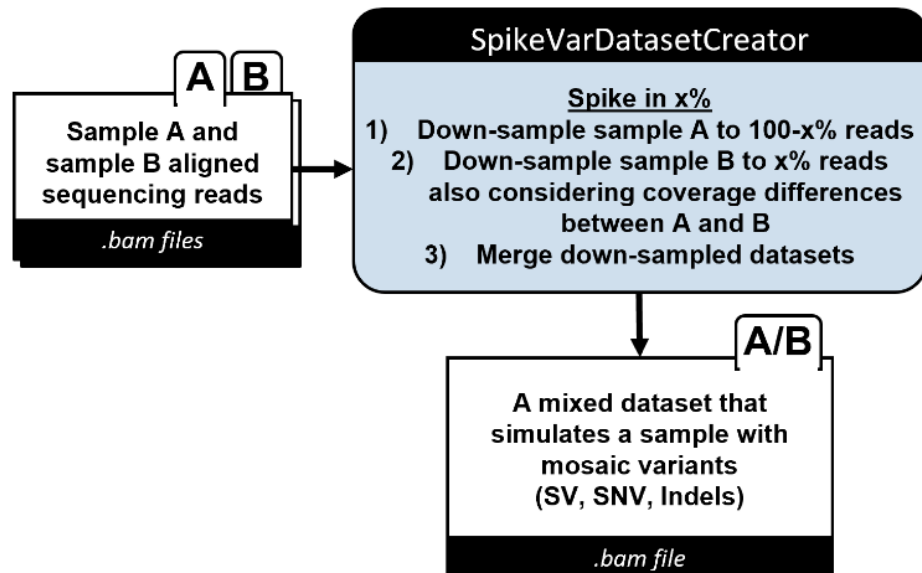
Research output: Contribution to journal › Article › peer-review

Disease-associated mosaic variation in clinical exome sequencing: a two-year pediatric tertiary care experience. Cold Spring Harb Mol Case Stud. 2020;6(3). Epub 20200612. doi: 10.1101/mcs.a005231. PubMed PMID: 32371413; PMCID: PMC7304353.

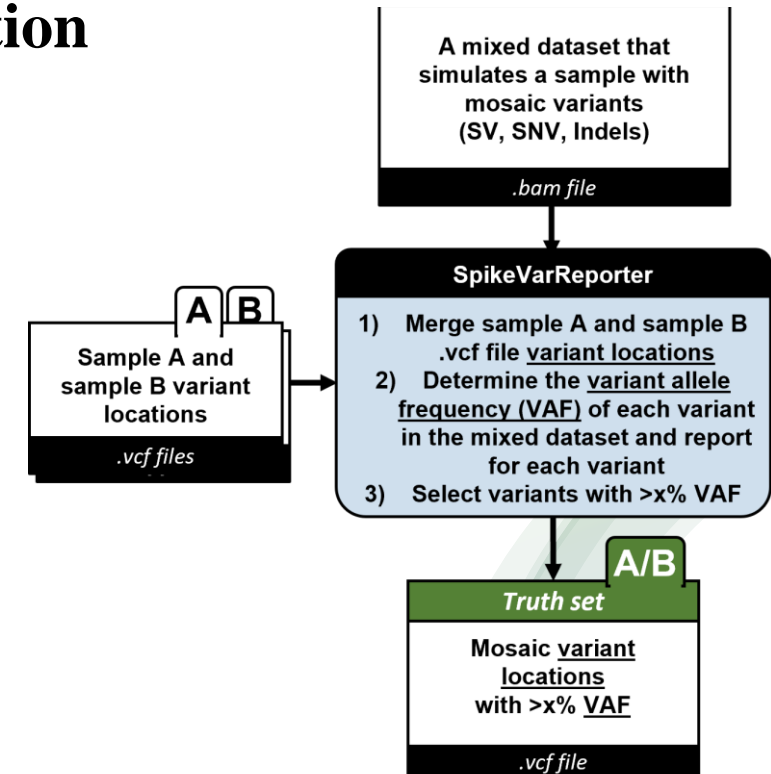
Spike in Known Exogenous Variants

❖ *Introducing User-Defined Spiking in Sequencing Data*

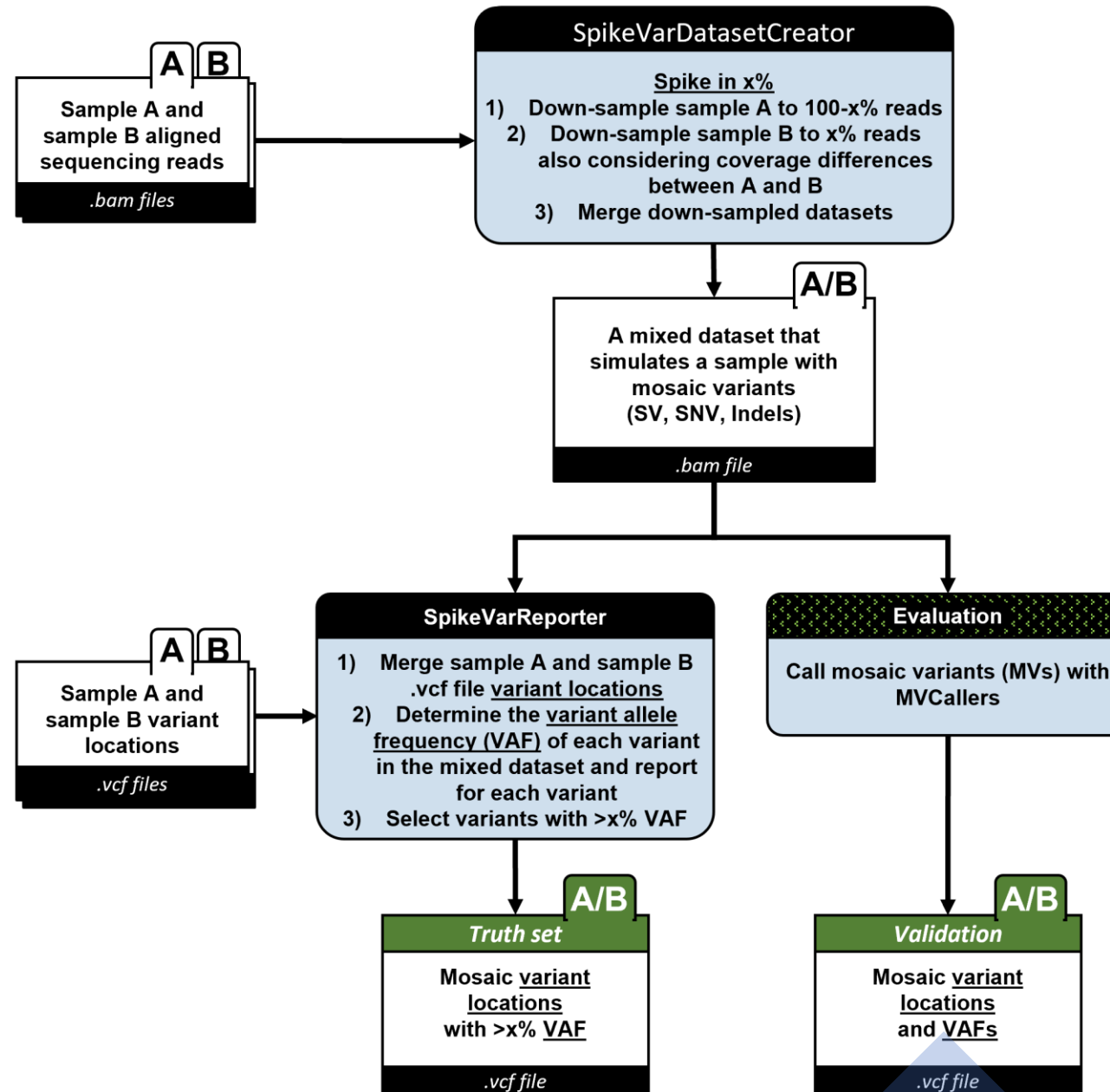
1) SpikeVar DatabaseCreator – Generate Spiked-in Dataset



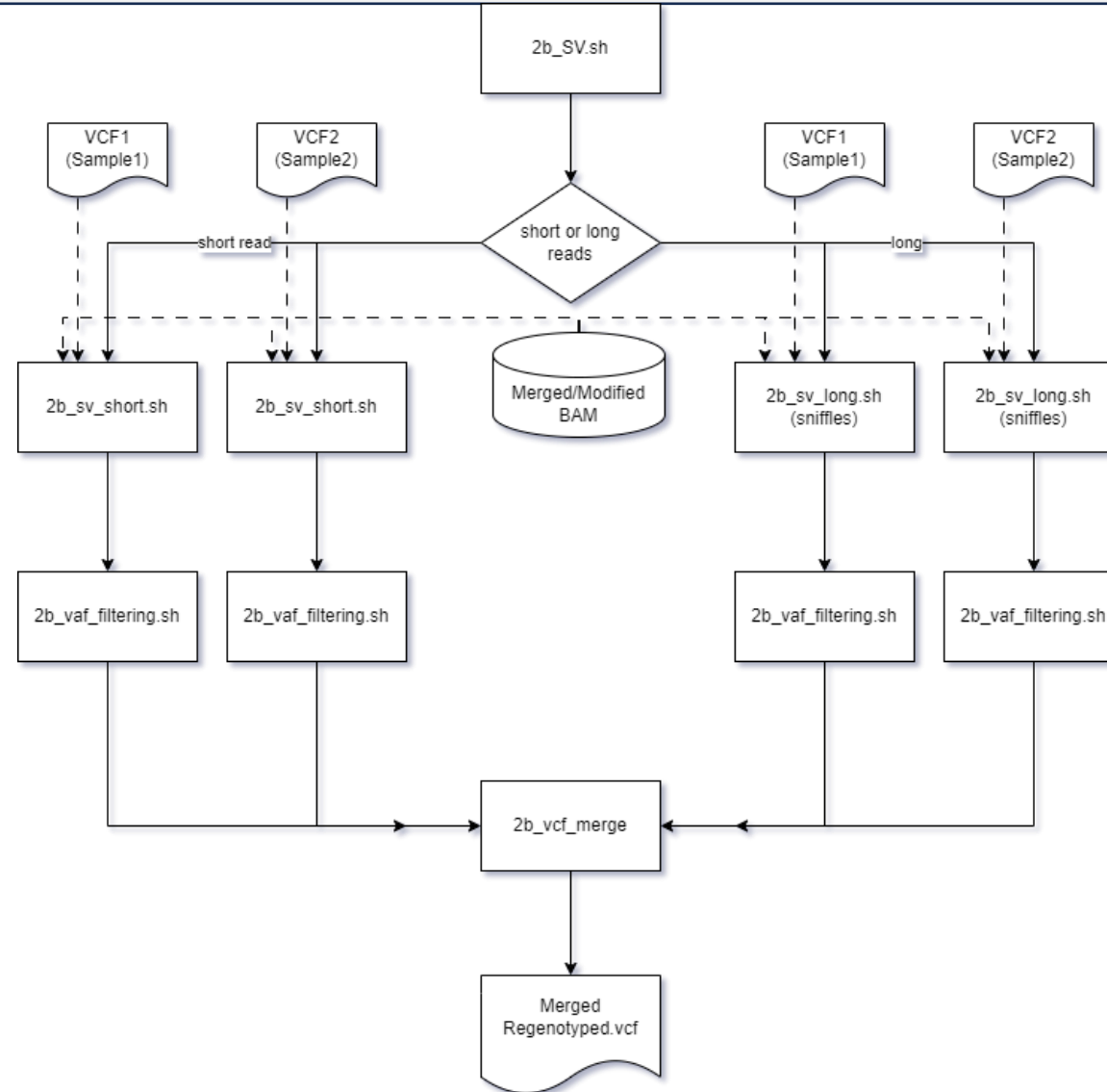
2) SpikeVarReporter – Filter Reads After Variant Allele Frequency Recalculation



Spike in Known Exogenous Variants

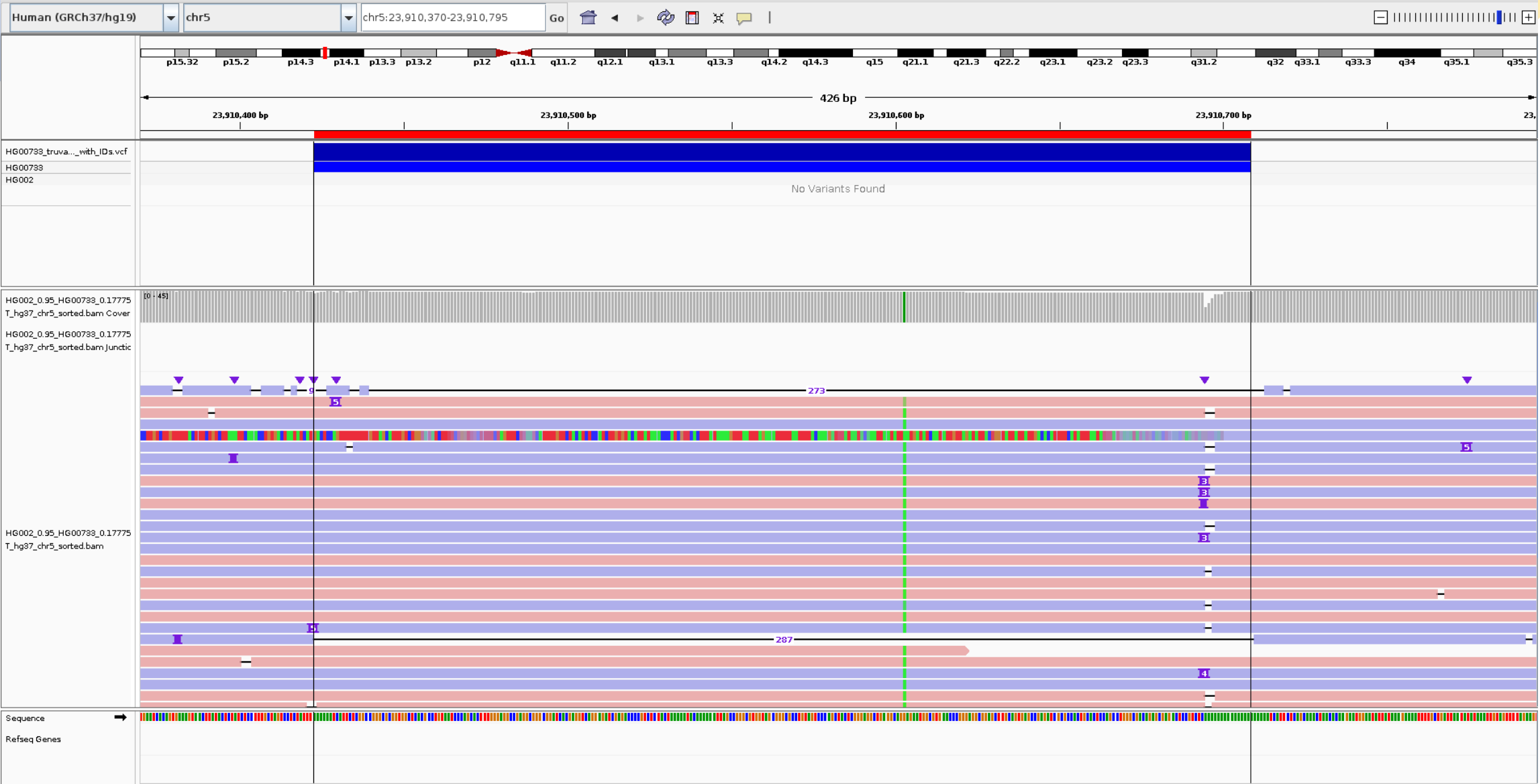


Spike in Known Exogenous Variants



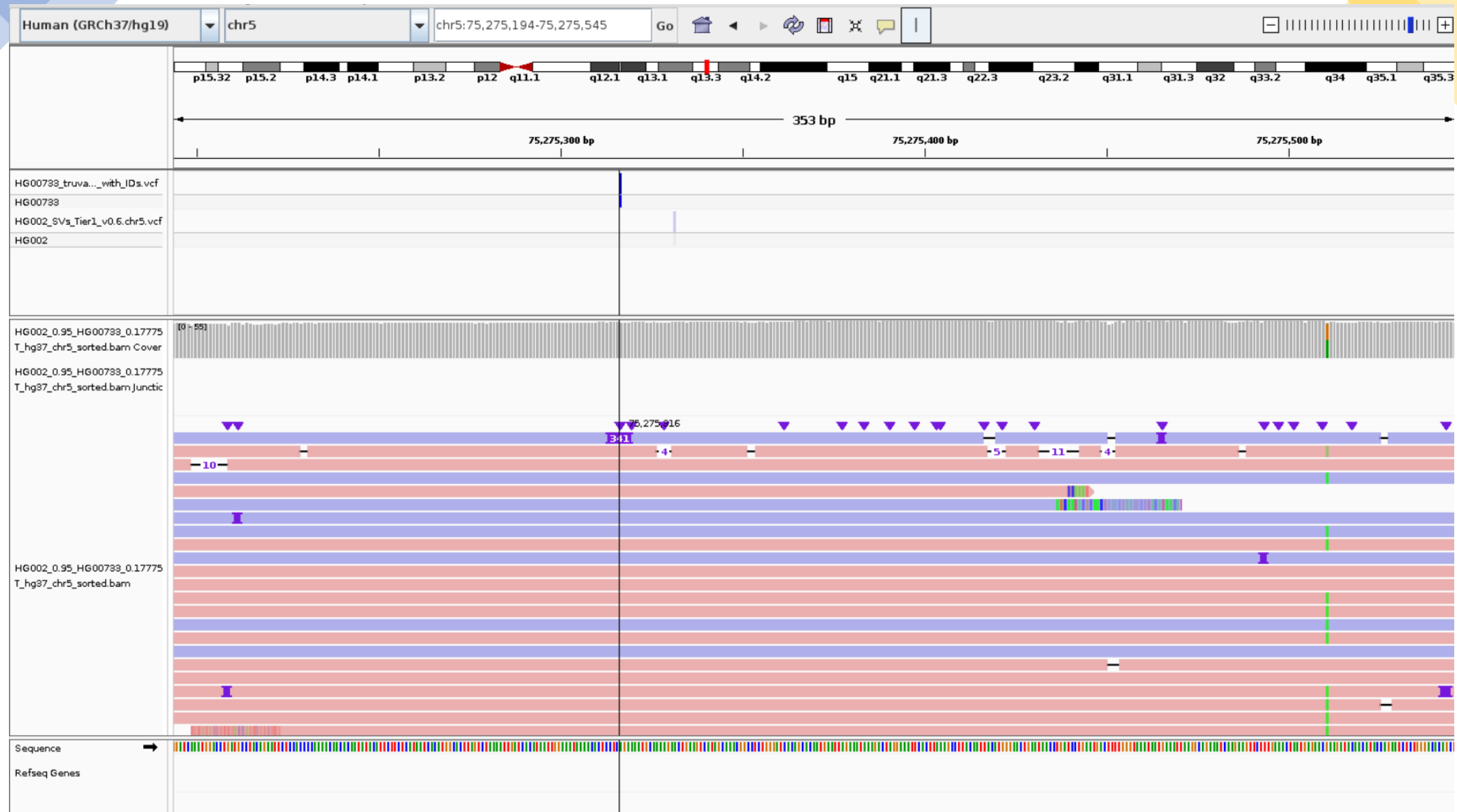
Flowchart of re-genototyping both short and long reads SVs based on the modified generated bam file

Spike in Known Exogenous Variants



Example of a spiked in deletion.

Spike in Known Exogenous Variants



Example of a spiked in insertion.

Track in Your *Key Endogenous Variants*

- ❖ creates a list of random mutations
- ❖ modifies a fraction of existing reads to match the user-defined MV frequency

1) TykeVarSimulator – Generate Simulated VCF

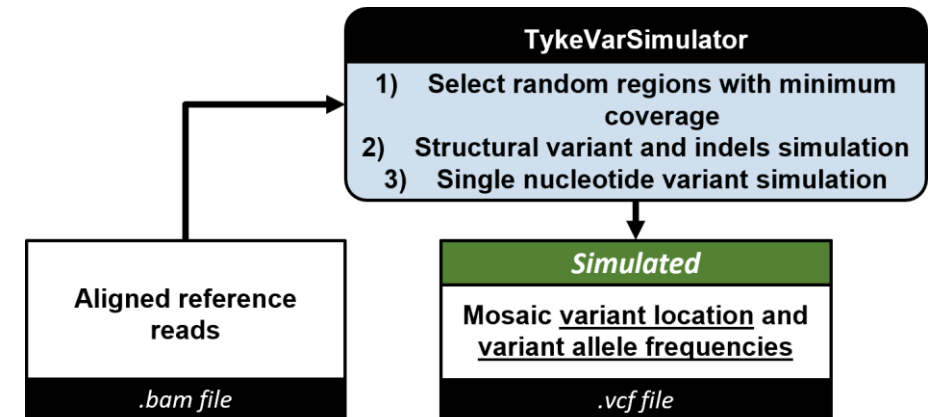
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chr22 32562272 HackIns1 N
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chr22 42134824 HackIns2 N
CCCCAGTGCTCAATCCATTCTCGTTTTTTGGTGGACCTCGGCCTTCGAGTAGGCTTTGATGTACGGTCTTCTCAGTTATAGATGCCTCGTGAGGCCCGCCGTTT>

chr22 43938337 HackIns3 N
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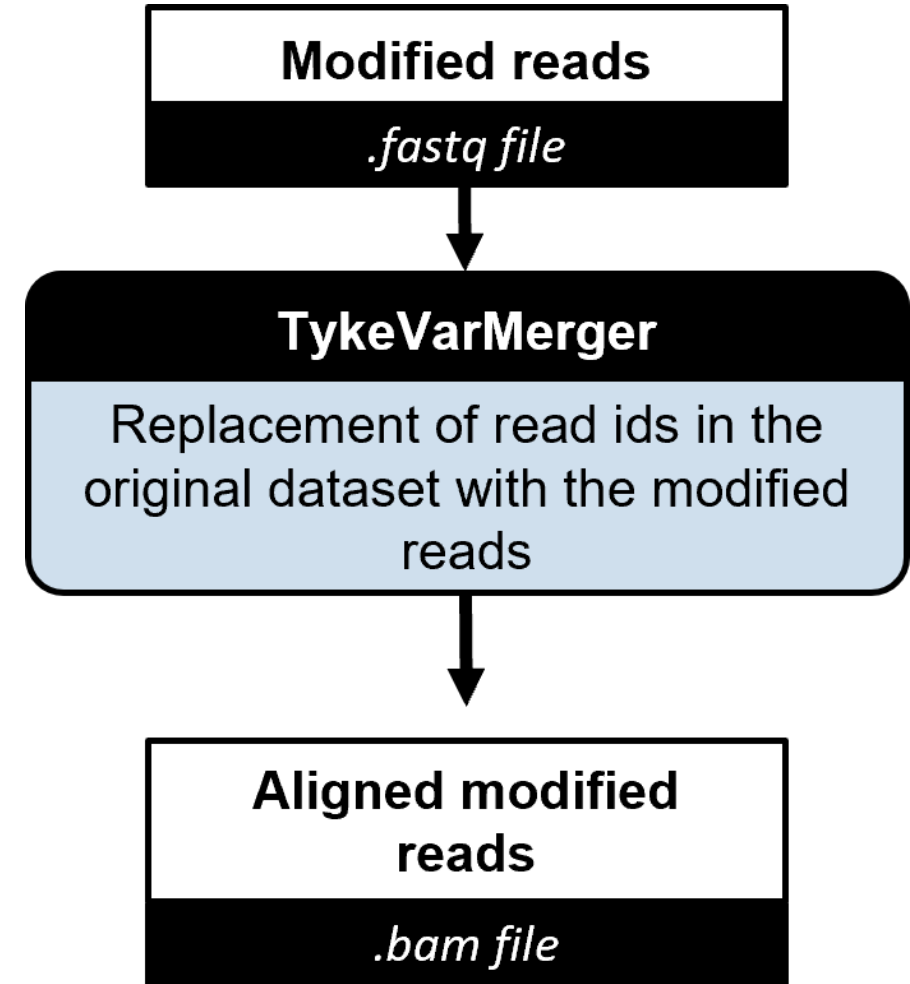
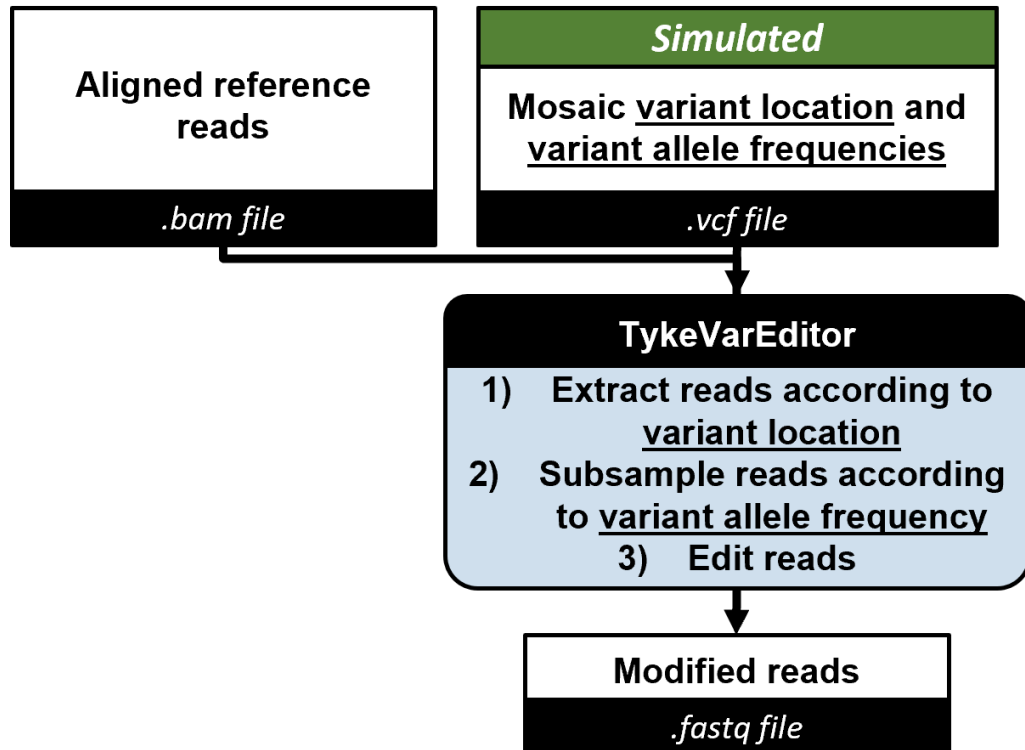
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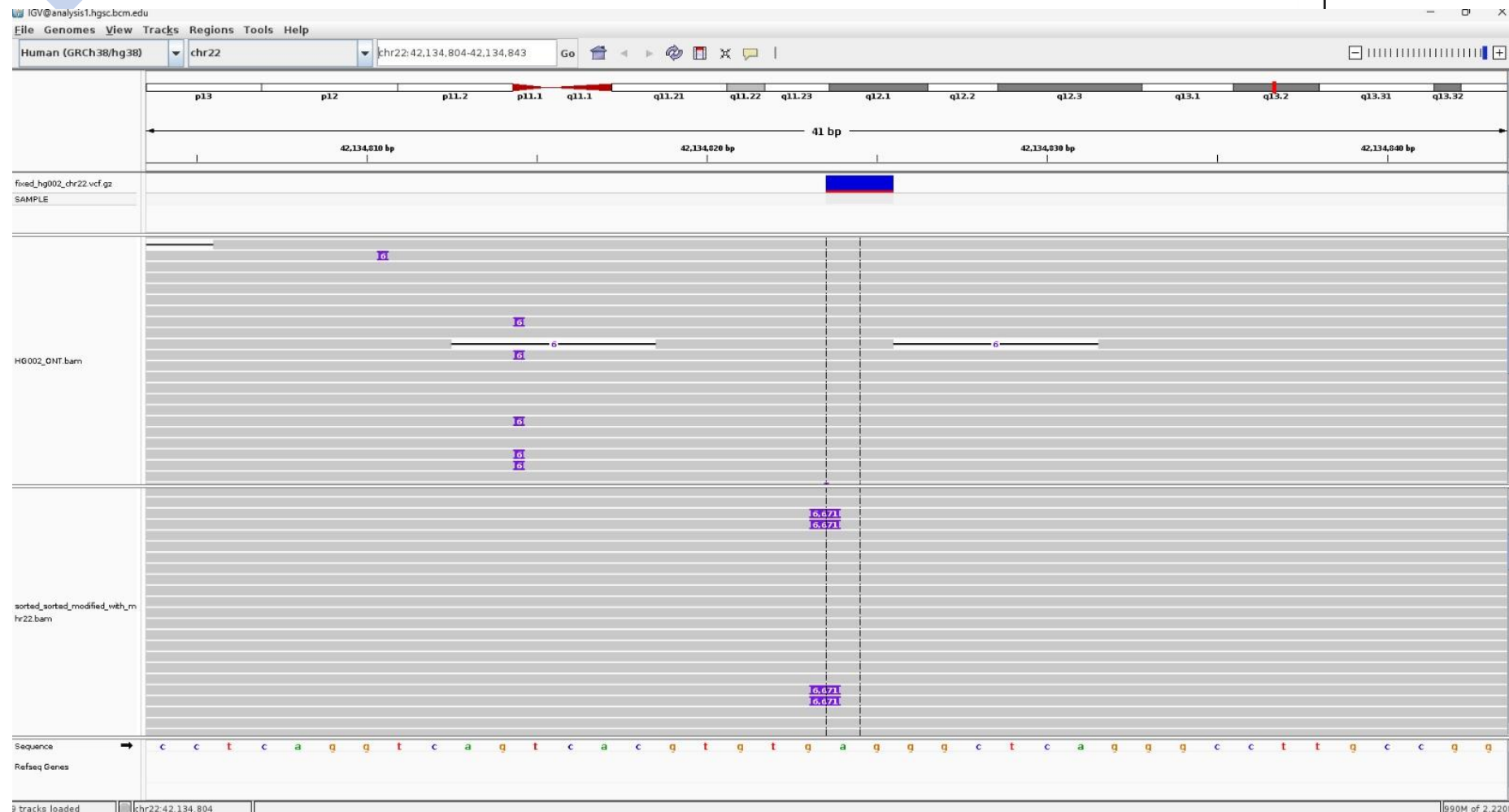
Track in Your *Key Endogenous Variants*

3) TykeVarMerger – Re-Align Modified Reads and Merge Them

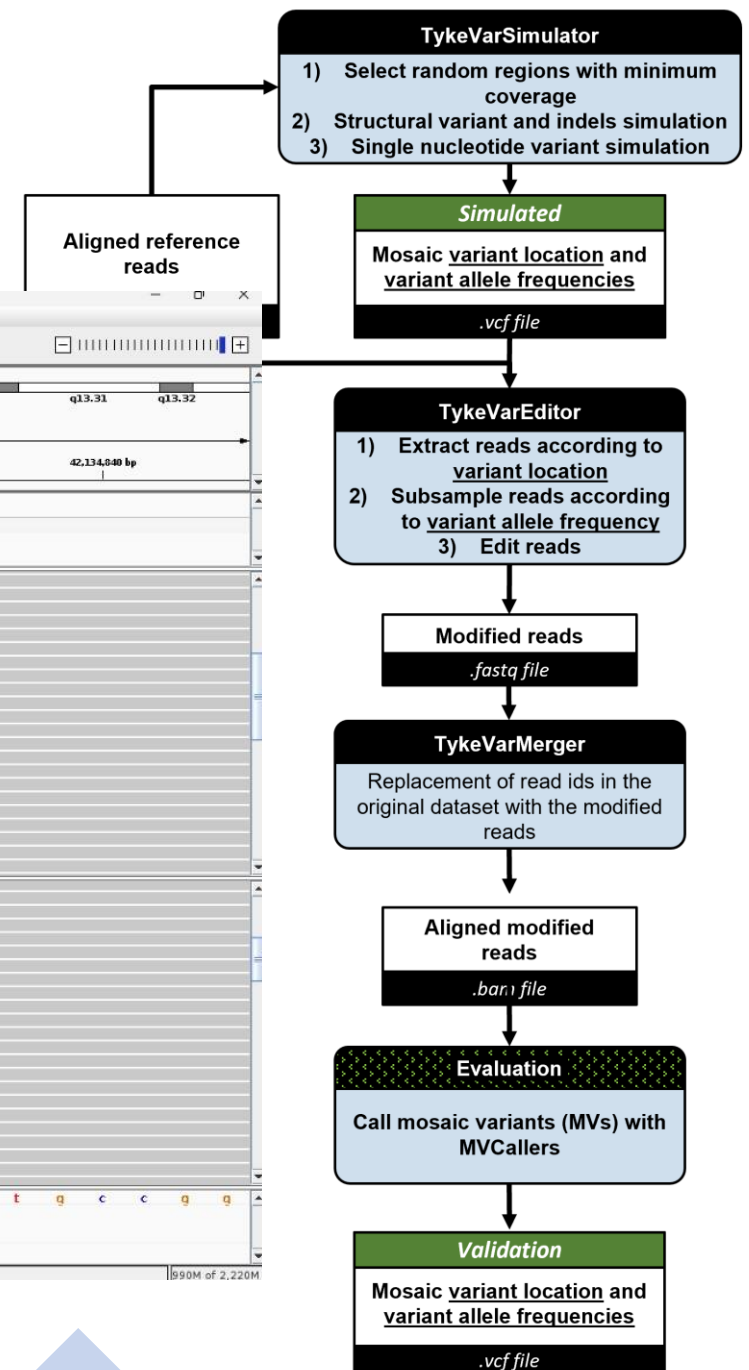
2) TykeVarEditor – Generate Edited Reads Based on Simulated VCF



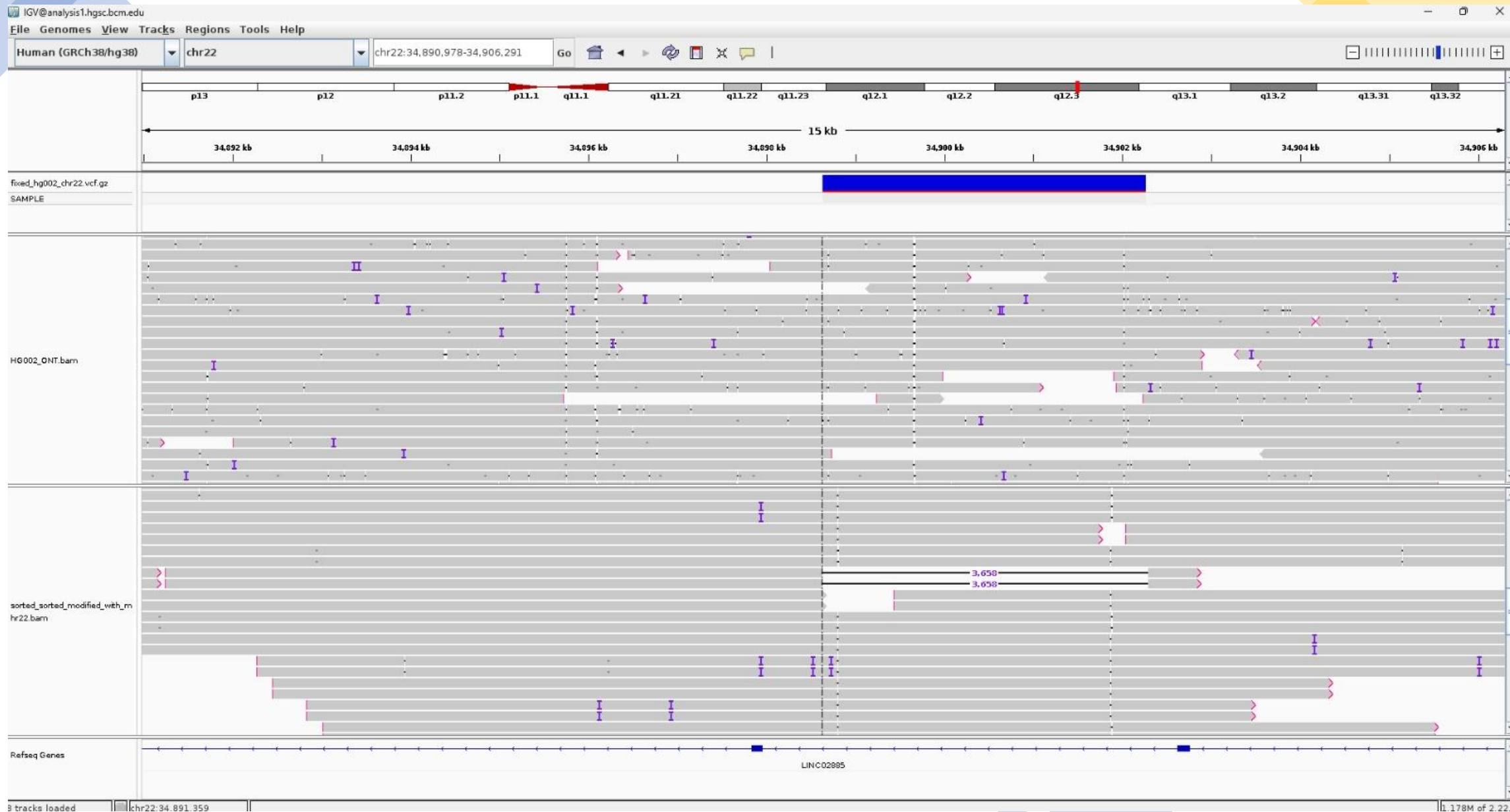
TykeVar – Creation of Sequencing Data With a Subset of Modified Reads



A mosaic SV insertion.



Track in Your *Key Endogenous Variants*



Track in Your Key Endogenous Variants

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chr22 11914735 HackDel12 N <DEL> 60 PASS AF=0.16;END=11924236;PRECISE;SVLEN=-9501;SVTYPE=DEL GT:GQ 0/0:60
chr22 16315573 HackIns19 N TTAATGCACTTACCTCCCAGAGATTGAGGTATTTTACGCTAGGCCGCGATTGCACGTTGATCTGGCGCCGCCCTTGGAAACATTCCTTTCCATTATCG
chr22 16445910 HackIns29 N TCGCTCTACTCGTCAGATGCTCTACGAGTAACATATCGCGCATCGTCTTACGTTAGAAAGTACAGAGACGTGGATTACGCTCGGCCCATGTTCTATCT
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chr22 17737821 HackIns4 N GGAGTGCACGTGACGCGCACTACGCTTACTAGCGTAATGCCGCGCATCGCACGAACGCCAAGGTACAAAACTGTTATATTCTACGCACTGTAAAAATGGGCAS
chr22 19815816 HackDel4 N <DEL> 60 PASS AF=0.13;END=19817115;PRECISE;SVLEN=-1299;SVTYPE=DEL GT:GQ 0/0:60
chr22 20461918 HackIns9 N CCACGCTGATCCTGTGTGCCGCGCATGTGCCCGCTTTGCTGCAGACCTCGAAAAAGGAA ##INFO=<ID=SUPPORT_INLINE,Number=1,Type=Integer,Description="Number of reads supporting an INS/DEL SV (non-split events only)">
chr22 20604273 HackDel10 N <DEL> 60 PASS AF=0.17;END=20607490;PRECISE;SVLEN=- ##INFO=<ID=SUPPORT_LONG,Number=1,Type=Integer,Description="Number of soft-clipped reads putatively supporting the long insertion SV">
chr22 21329741 HackIns18 N GAAGTTATACCACAACGTATCGTCAGACCATATAGGCTTCTCAAGAGCTCACTCT ##INFO=<ID=END,Number=1,Type=Integer,Description="End position of structural variation">
chr22 21641996 HackIns21 N GTTGGCGCTAATCGTCTATTGAAGGCCGTGCGGAGATTACCCCGGTTCCGCAACGT ##INFO=<ID=STDEV_POS,Number=1,Type=Float,Description="Standard deviation of structural variation start position">
chr22 22551854 HackIns28 N GCTCAATTGTATCGTTGCCGACGCCATCATTAAATTTCCCGAGAAGATGGAGTTT ##INFO=<ID=STDEV_LEN,Number=1,Type=Float,Description="Standard deviation of structural variation length">
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chr22 30857173 HackIns20 N CCCGAAATGCCGAAATAATCGTCTTTGAGGTGCATACAAGTCTTGGCACCAGTGCCT ##INFO=<ID=PHASE,Number=.,Type=String,Description="Phasing information derived from supporting reads, represented as list of: HAPLOTYPE,PHASES"
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chr22 32022097 HackIns14 N GGTGTTTAATAAGCATCAACTCGTTTTGAGTGTCTGCGGTGGGATGGATGGGACAA chr22 11895609 Sniffles2.INS.36S0 N GTGAAACCCTGTCTCTACTAAAAATACAAAAAATAGCTGGACATGGTAGCACATGCCTGTAATCCTAGTACTTGGAGGCTGAGC
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chr22 32562272 HackIns1 N GAGTATGTCGTTAAATCTGAACATGGGTGGTGGGTCAGTACGTACCACTGCCAACCCA chr22 16315572 Sniffles2.INS.7ES0 N CTTAATGCACTTACCTCCCAGAGATTGTAGGTATTTTACGCTAGGCCGCGATTGCACGTTGATCTGGCGCCGCCCTTGGAAACA
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chr22 chr22 16437276 Sniffles2.INS.D3S0 N AAAAAATTTGCGAGTGCTCAATGGTCCCCAGGCTGGAGTGCAGTGGCGTGATCTCAGCTCGCTACAACCCCCACCTCCACGCCGCTC
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chr22 16477550 Sniffles2.INS.E1S0 N ATCGCTCTACTCGTCAGATGCTCCTACGAGTAACATATCGCGCATCGTCTTACGGTTAGAAGATCACAGAGACGTGGATTACAGCTC
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chr22 17747883 Sniffles2.INS.FBS0 N CCGAGTGCACGTGACGCGCACTACGCTTACTAGCGTAATGCCGCGCATCGCACGAACGCCAAGGTACAAAACTGTTATATTCTAC
```

Ground Truth VCF

Sniffles Mosaic Results

Acknowledgement



Sedlazeck lab

- Fritz J Sedlazeck
- Luis Paulin
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- Ryan Cheng
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- Farhang Jaryani

