# Reliable identification and quantification of neural cells in microscopic images of neurospheres

RUB



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### Abstract

Neurosphere cultures consisting of primary human neural stem / progenitor cells (hNPC) are used for studying the effects of substances on early neurodevelopmental processes in vitro. Differentiating hNPCs migrate and differentiate into radial glia, neurons, astrocytes, and oligodendrocytes (oligos) upon plating on a suitable extracellular matrix and thus model processes of early neural development. In order to characterize alterations in hNPC development, it is thus an essential task to reliably identify the cell type of each migrated cell in the migration area of a neurosphere. To this end, we introduce and validate a deep learning approach for identifying and quantifying cell types in microscopic images of differentiated hNPC. As we demonstrate, our approach performs with high accuracy and is robust against typical potential confounders. We demonstrate that our deep learning approach reproduces the dose responses of well-established developmental neurotoxic (DNT) compounds and controls, indicating its potential in medium or high throughput in vitro screening studies. Hence, our approach can be used for studying compound effects on neural differentiation processes in an automated and unbiased process.

### Data & Methods

			ı					_
Plate	Stainings		DNT		#Wells distributed			N
Ind.	Neurons	Oligos	Compound ID	Effects	Train	Val	Test	•
<i>I</i> 1	✓		3	Both	36	4	0	
11	✓		11		36	4	0	
11	✓		11		36	4	0	
11	✓	<b>✓</b>	3	Both	36	4	0	
11	✓	<b>√</b>	13	Oligos	36	3	0	
11	✓	✓	4		36	4	0	
11	✓	✓	1	Both	36	3	0	•
<i>l</i> 2		✓	1	Both	36	3	0	•
<i>l</i> 3		✓	9	Both	36	4	0	
11		✓	13	Oligos	36	4	0	
11		✓	6	Both	36	3	0	
<i>l</i> 2		✓	6	Both	36	3	0	
<i>l</i> 2		✓	12	Oligos	36	4	0	a
14		<b>√</b>	2	Oligos	36	4	0	
14		✓	5	Both	36	4	0	
11	<b>√</b>	✓	8	Neurons	36	4	1	    a
<i>l</i> 2	✓	✓	7	Neurons	36	4	1	
11	✓	✓	10	Oligos	36	4	1	Δ
14	<b>√</b>	<b>✓</b>	11		36	4	1	

Detailed Assay & Staining protocol published in [2].

#### Neurosphere data distribution:

- Can model DNT in vitro
- 19 Plates with 40 Neurospheres each
- One sphere per well
- Grown from hNPCs for 7-8 weeks
- Antibody stained, differentiated cells
- 8 different compound concentrations
- 5 replicates each

		Neuron Set	Oligo Set				
	Plates	11	16				
	Total Nuclei	222,423	309,753				
Label	Training	28,548	19,486				
annotated	Validation	3,866	2,443				
	Test	1,114	718				
Label not	Training	158,932	248,908				
annotated	Validation	20,132	27,971				
	Test	9,831	10,227				
Annatations O Data imbalance							

#### Annotations & Data imbalance:

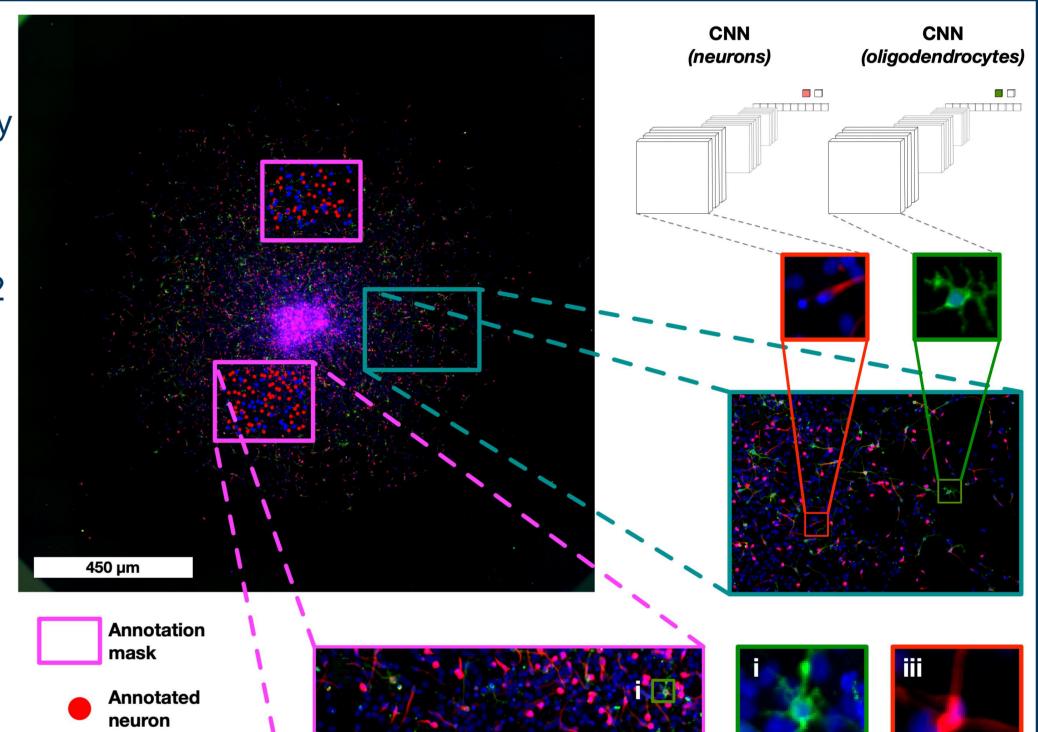
- Roughly 1 out 5 nuclei is a neuron
- Roughly 1 out of 12 nuclei is an oligo

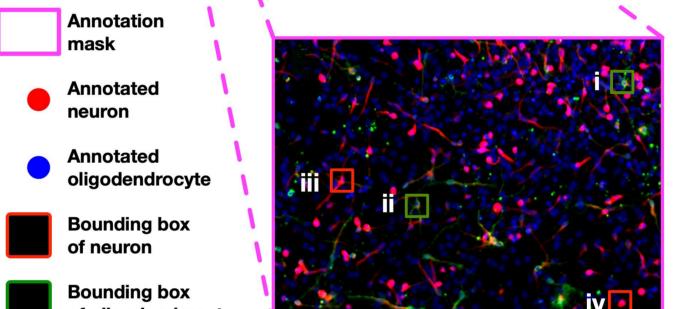
#### **Pre-Processing**

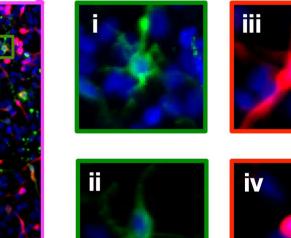
- Capture whole well images via fluorescence microscopy
- Image resolution  $4858\times4858 \ \mu m \rightarrow$ 5520×5520 px
- Annotate every nucleus in 2 ROIs (purple) per neurosphere
- Capture 64×64 px 8-Bit RGB images centered on every nucleus
- Train on ROIs to predict sphere migration area

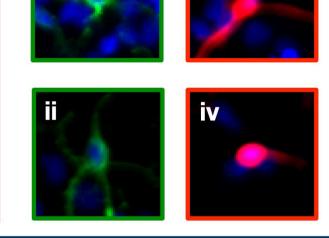
#### The Model

- Modified VGG architecture
- One model for Neurons and Oligodendrocytes each
- Trained with 100 batches
- Up to 5000 epochs & early stopping









# Results & Discussion

### **Internal Validation: External Validation:** Individual heterogeneity: Neurons Normalization approaches: Neurons No normalization (0.831) Individual I1 (0.796) Flat normalization (0.818) Individual I2 (0.705) Per tile, separate channels (0.780) Individual I4 (0.805) Individuals $I\{1,2,4\}$ (0.74) Per well, across channels (0.826) Individual heterogeneity: Oligos Normalization approaches: Oligos No normalization (0.876) Flat normalization (0.876) Per tile, separate channels (0.867) Individual I4 (0.8) Per tile, across channels (0.404) Individuals I {1,2,4} (0.841) Precision-Recall Curves show annotation strength. Legend

entry values within brackets show the respective area

under curves.

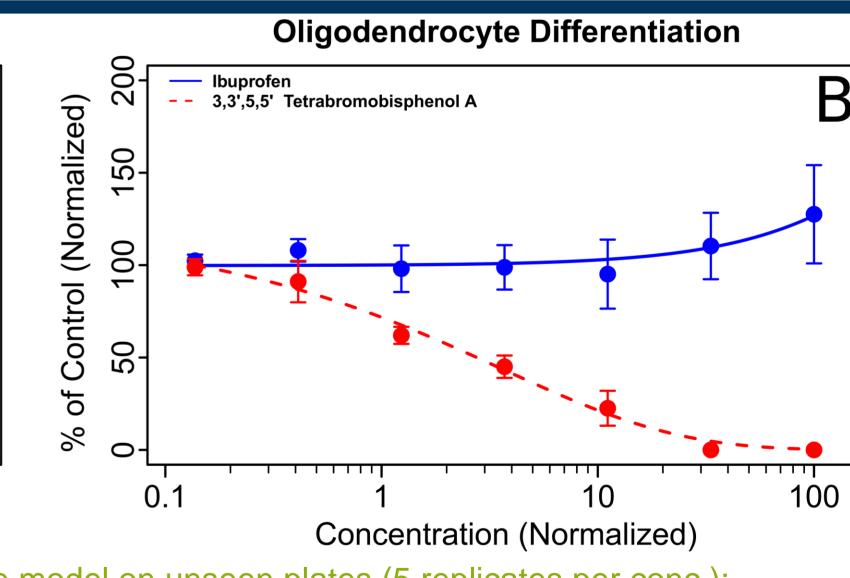
**Individual Heterogeneity** Accounts for ind. morphology.

- New model trained using only plates from 11
- Validate the model performance on the other individuals
- Individually
- Pooled

#### **Normalisations**

- Accounts for staining & lighting.
- No Normalization: RGB
- Values unchanged Flat Normalization: RGB
- Values normalized to 8 bit • Per tile, separate channels: Each channel in
- each tile is min-max normalized independently
- Per tile, across channels: Each channel in each tile is min-max normalized on the same min/max
- Per well, across channels: Each channel in each tile is min-max normalized with min/max from the same well

# **Neuronal Differentiation** Methylmercury(II) chloride 50 00 Concentration (Normalized)



Concentration Response Curves - Validating the model on unseen plates (5 replicates per conc.):

- *Ibuprofen* is a known control, concentration normalized to 20.0 μM
- Methylmercury(II) chloride is a known neuronal DNT, concentration normalized to 2.22 μM
- 3,3',5,5' Tetrabromobisphenol A is a known oligo DNT, concentration normalized to 20.0 μM

- Robust neuron & oligodendrocyte
- Captures dose response relations
- Many confounders accounted for
- Model performance suitable for toxicology
- Regulators (US-EPA, EFSA European
- classifications

- Food Safety Authority) showing interest in assay and endpoints

## **Future Work**

- Extended classifications for Radial Glia Cells and
- Weakly supervised learning to infer morphological

# References & Acknowledgements

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- Masjosthusmann, S., Blum, J., Bartmann, K., Dolde, X., Holzer, A. K., Stürzl, L. C., ... & Fritsche, E. (2020). Establishment of an a priori protocol for the implementation and interpretation of an in-vitro testing battery for the assessment of developmental neurotoxicity. EFSA Supporting Publications, 17(10), 1938E.
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- Astrocytes
- endpoints

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