

Christopher Olsen

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PROFILE SUMMARY

Cell and biochemical assay Research Associate with 5 years of experience in assay development and screening. Versatile skillset with hands-on experience in bench automation, Python coding and primary cell culture in a fast-paced biotech environment.

WORK & LEADERSHIP EXPERIENCE

Blueprint Medicines

Boston, Massachusetts

Contract Research Associate, Biology

August 2024 – Nov 2024

- Expanded pre-clinical lead small molecule testing into dozens of novel immortal cell lines in multiple assays including, proliferation, alphaLISA, immunoblot, and imaging in support of oncology programs.
- Established automation protocols for immunoblot sample processing, dramatically increasing throughput.
- Transitioned key flow cytometry experiments from tube to plate format increasing sample throughput.
- Designed and implemented automation workflows utilizing Biomek i7 liquid handler.

Cellarity

Boston, Massachusetts

Research Associate II, in vitro Pharmacology

June 2022 – Nov 2023

- Developed and ran cell culture assays for small molecule screening in lead optimization campaigns in the hematology disease area and myeloid cell lineages including hands on hematopoietic differentiation.
- Built assay informatics dashboards enabling consistent and regular data analysis for multiple assay readouts including, alphaLISA, TR-FRET, Flow Cytometry, Nanostring, Western Blot, and qPCR.
- Served as point person for troubleshooting and training on multiple instruments for automation including Formulatrix Mantis, FAST, Integra Viaflow 384, and Agilent Bravo
- Established standardized methodologies for evaluating dose response curve fits across projects at all stages of R&D and standardized western blot procedure.

Boston University Hereditary Fructose Intolerance Laboratory

Boston, Massachusetts

Undergraduate Research Associate, The Tolan Lab

September 2020 – May 2022

- Awarded Corporate Sponsorship from New England Bio labs for 2021 summer research project titled: “KHK inhibition as a potential therapeutic target for type 2 diabetes mellitus.”
- Performed Cell Culture in the lab on experiments focusing on recovering insulin sensitivity in HepG2 Cells and worked on protein purification in *E. coli*.
- Managed rodent colony for investigations concerning the efficacy of KHK inhibitors in treating established Insulin Resistance and IP dosing for test compounds.
- Implemented various spectrophotometric assays for measurement of fat accumulation and collagen content in mouse liver tissue in profiling NAFLD in AldolaseB -/- and investigated with western blot.

Symbrosia Inc

Kailua Kona, Hawaii

Remote Research Intern

June 2020 – August 2020

- Performed background research on the Bromoform producing seaweed *Asparagopsis Taxiformis*.
- Presented research recommendations which assisted in selective breeding grant proposal and acquisition.

EDUCATION

Boston University

Boston, Massachusetts

Bachelor of Arts in Biochemistry & Molecular Biology

May 2022

SKILLS, ACTIVITIES & INTERESTS

Skills: Cell Culture, qPCR, Immunoblotting, ELISAs, Reporter Assays, NGS Sample Preparation, MSD, Transfection, NanoString, CRISPR, Mouse dosing via IP dosing, oral gavage, subq dosing, tail vein injection, and tissue extraction, HPLC, SQL, Python, R, Julia, Benchling, Dotmatics, NCBI BLAST Graph Pad Prism, FlowJo.

Activities & Interests: Served as supervisor at Agganis Arena overseeing operations during BU Men's Hockey Games. Served as Treasurer of BU Gaming Club increased revenues by 10%.