

*Mathematical modeling of cancer-related  
molecular mechanisms:*

*cell fate decisions, cancer biology,  
discrete modeling and BiNoM software*

Andrei Zinovyev

U900 Institut Curie/INSERM/Ecole de Mines

*“Computational Systems Biology of Cancer”*

<http://sysbio.curie.fr>

# Computational Systems Biology of Cancer group at Institut Curie, Paris <http://sysbio.curie.fr>



**Emmanuel Barillot, PhD**

Director of the U900 Institut Curie/INSERM/Ecole de Mines ParisTech

Data integration, Systems Biology of Cancer, Dynamics of network motifs



**Laurence Calzone, PhD**

INVADE and PIC SysBio projects

Mathematical modeling of cell signalling in cancer



**David Cohen, PhD**

Atlas of Cancer Cell Signalling

Network building and analysis



**Loredana Martignetti, PhD**

ASSET project

Regulatory network analysis



**Liron Kuperstein, PhD**

ACSN, INVADE and PIC SysBio projects

Systems biology of cancer



**Eric Bonnet, PhD**

BiNoM project

Integrative computational biology



**Bruno Tesson, PhD**

Curie-Servier alliance on basal breast cancer

Cancer bioinformatics



**Valentina Basso, PhD**

Next-generation sequencing

Genomic sequence analysis



**Seriem Sefta**

Retinoblastoma project

Cancer bioinformatics



**Luca Garavito, PhD**

ACSN project, meta-analysis of cancer omics data

Modeling and data analysis for signalling networks



**Antonio Cappuccio, PhD**

'What's in TSLP' project

Systems Immunology



**Paola Vera-Licona, PhD**

Curie-Servier alliance on basal breast cancer

Mathematical modeling of biological networks



**Daniel Rovera, volunteer**

BiNoM project

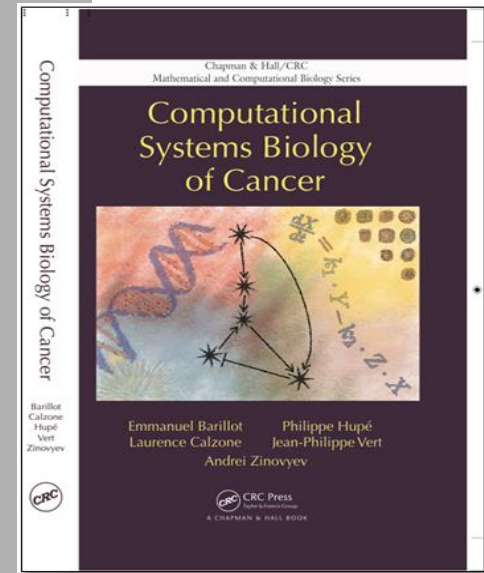
Methods for network analysis



**Andrei Zinovyev, PhD**


Scientific coordinator of the Systems Biology team

Systems Biology of Cancer, Complexity and Model reduction



<http://www.cancer-systems-biology.net/>

# Plan of the presentation

- Recent developments using <sup>n</sup>
  - NaviCell
  - Atlas of Cancer Signaling Networks
  - Creating CellDesigner files
- What can be done for cancer biology, using discrete modeling
  - Cell fate decision model
  - OCSANA (minimal cut sets)
  - Computing phenotype probabilities
  - Model reduction
  - Dosage response curves
  - Sensitivity analysis
  - MaBOSS: discrete states, continuous time

# Biological Network Manager

Part of Curie platform for network systems biology of cancer

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Java library + Cytoscape 2.\* plugin



Manipulating standards

(SBML, BioPAX, CellDesigner, and others)

- importing/exporting/creating
- conversion between formats
- BioPAX 3.0 editor
- merging/decomposing/transforming
- annotating

Network analysis (graph theory, semantics-based)

Analysis of data using biological networks

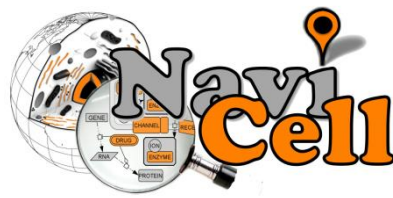
<http://binom.curie.fr/>

Bonnet E., Calzone L., Rovera D., Stoll G., Barillot E. and Zinovyev A..

**BiNoM 2.0, a Cytoscape plugin for accessing and analyzing pathways using standard systems biology formats.** 2013, *BMC Systems Biology*, 7:18.

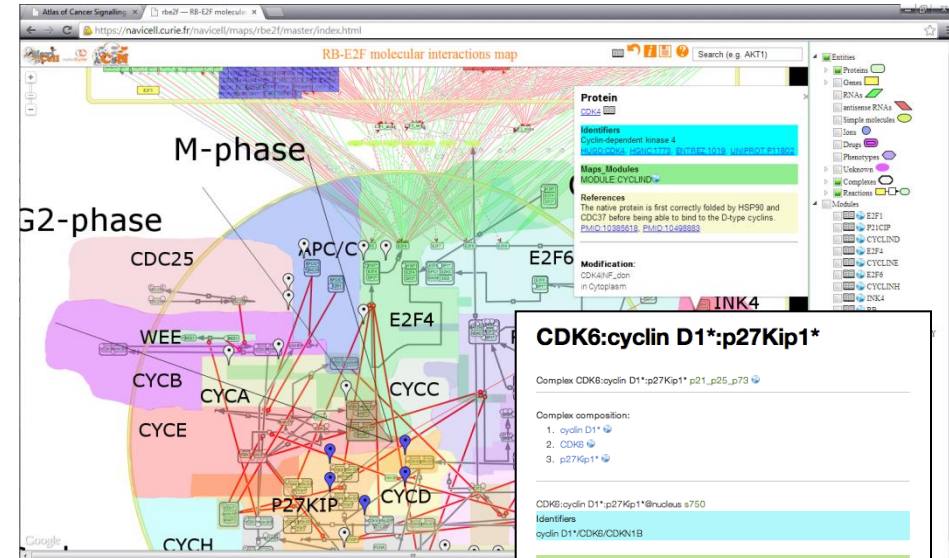
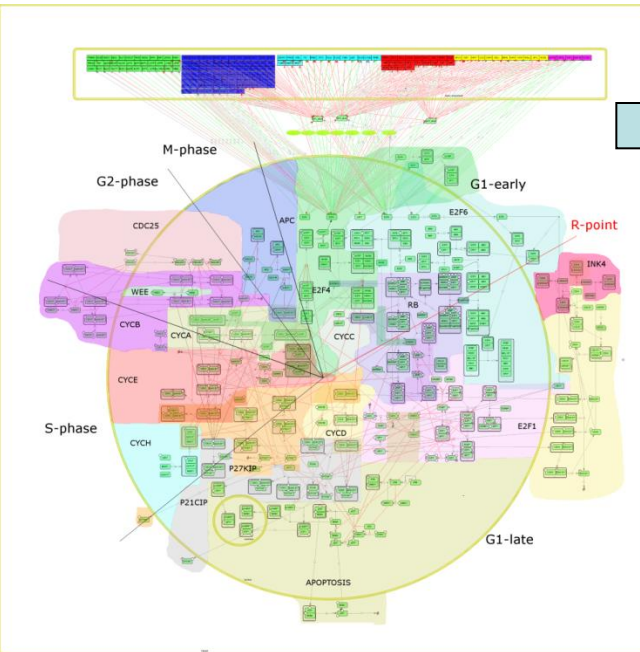
Zinovyev A., Viara E., Calzone L., Barillot E.

**BiNoM: a Cytoscape plugin for manipulating and analyzing biological networks.** 2008. *Bioinformatics* 24(6):876-877



# factory tool

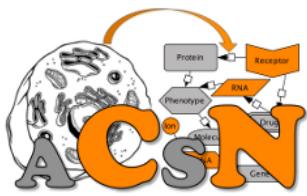
## CellDesigner map



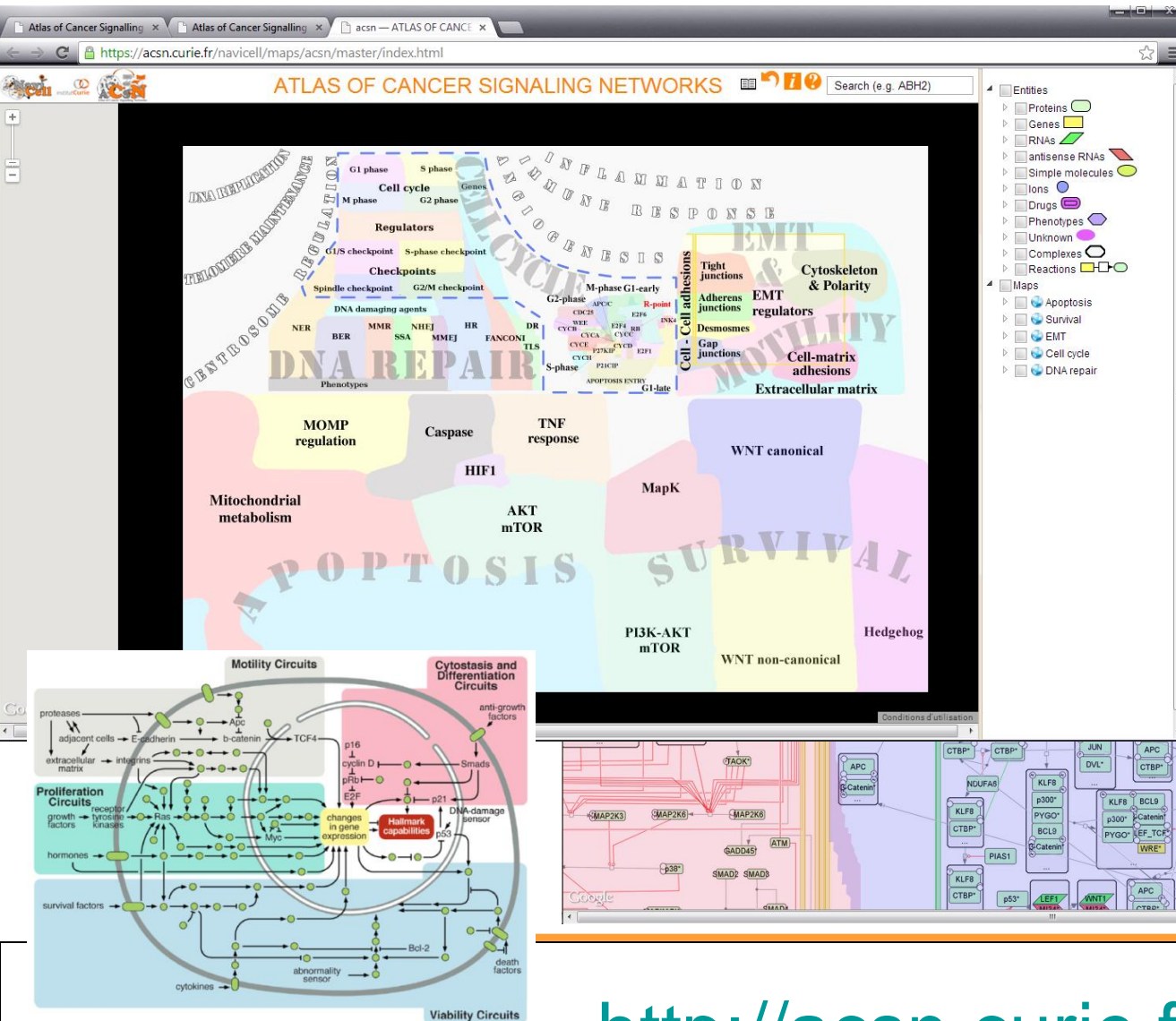
## Google Maps+ WordPress blog

<http://navicell.curie.fr>

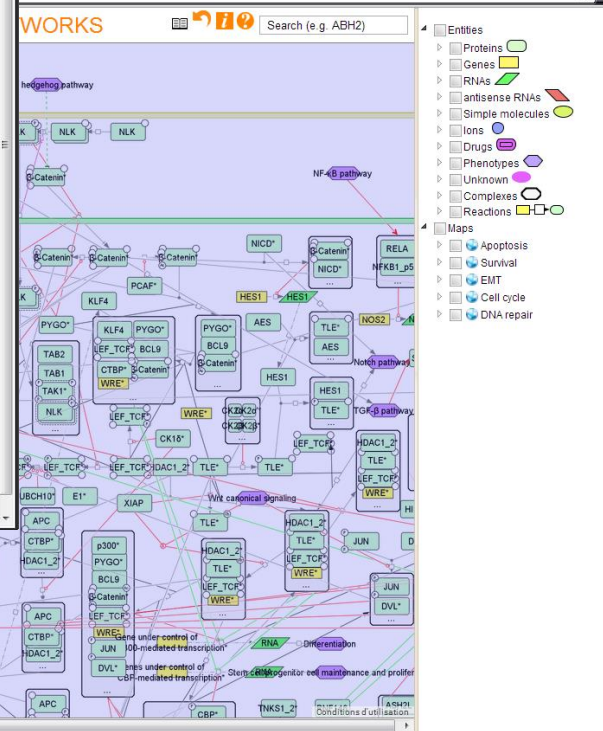




# Atlas of Cancer Signaling Networks



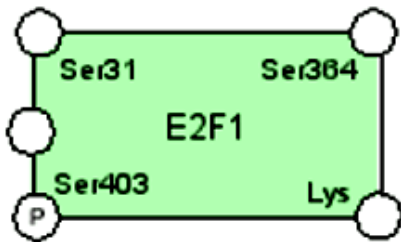
**Inna Kuperstein**  
**At COMBINE,**  
**Thursday, 19<sup>th</sup>**  
**morning session**



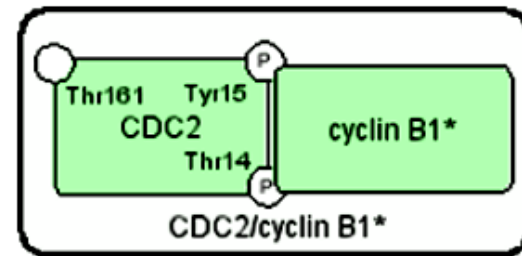
<http://acsn.curie.fr>

# Naming conventions in BiNoM

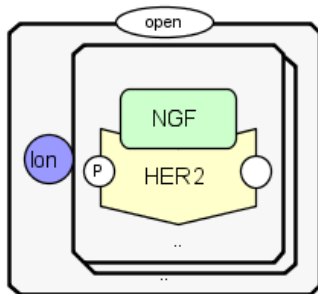
Entity1\_name|Modification1|Modification2|...: Entity2\_name|Modifications...[\_active|\_hmN]@compartment



E2F1|Ser403\_pho@nucleus



CDC2|Tyr15\_pho|Thr14\_pho:cyclinB1\*@cytoplasm

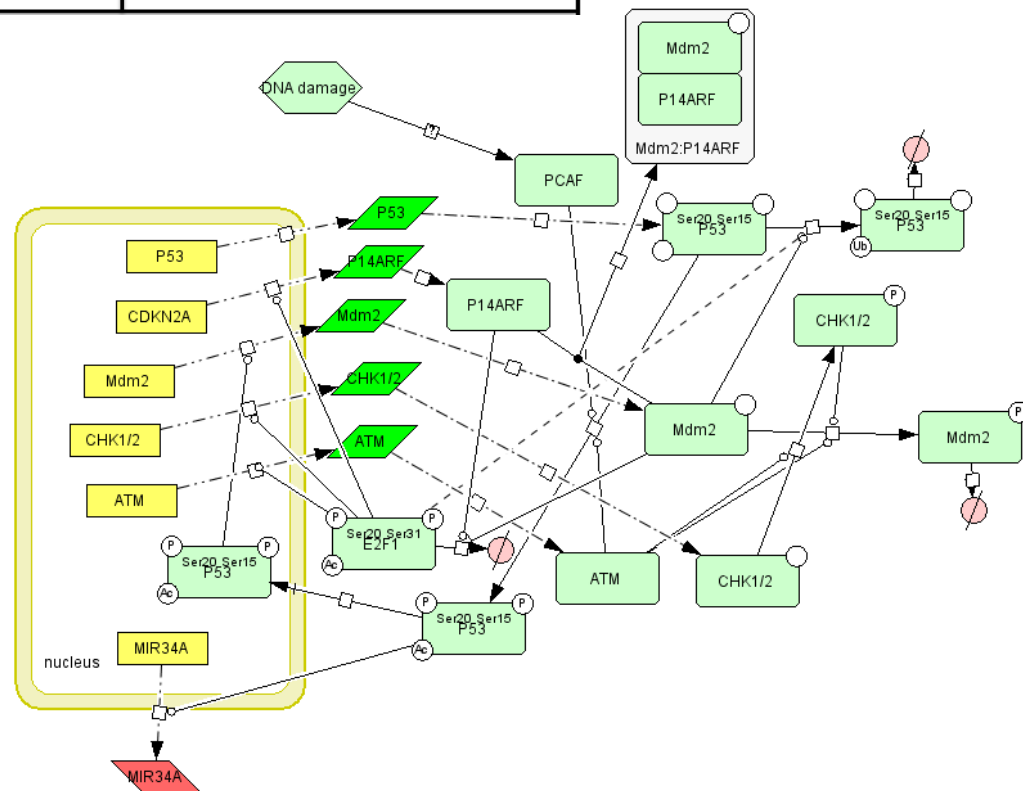


(lon:(HER2|pho|:NGF)|hm2)|open@plasma\_membrane

# Creating CellDesigner files from simple textual description (BiNoM reaction format)

Reactions	Regulators
<ul style="list-style-type: none"> <li>-&gt; STATE_TRANSITION</li> <li>-&gt; KNOWN_TRANSITION_OMMITED</li> <li>-?&gt; UNKNKNOWN_TRANSITION</li> <li>-+&gt; POSITIVE_INFLUENCE</li> <li>- &gt; NEGATIVE_INFLUENCE</li> <li>-/&gt; TRANSPORT</li> <li>-..&gt; TRANSCRIPTION</li> <li>-&gt; TRANSLATION</li> <li>-:&gt; HETERODIMER_ASSOCIATION</li> <li>-=&gt; DISSOCIATION</li> </ul>	<ul style="list-style-type: none"> <li>- CATALYSIS</li> <li>- UNKNOWN_CATALYSIS</li> <li>-  INHIBITION</li> <li>-  UNKNOWN_INHIBITION</li> <li>-* MODULATION</li> <li>-) PHYSICAL_STIMULATION</li> </ul>
	Entities
	<ul style="list-style-type: none"> <li><b>g</b>MDM2 gene of MDM2</li> <li><b>r</b>TP53 RNA of TP53</li> <li><b>ar</b>MIR200 antisense RNA of MIR200</li> </ul>

P14ARF+Mdm2 -> Mdm2:P14ARF  
 rP14ARF -> P14ARF  
 gCDKN2A@nucleus-E2F1|Ser31\_pho|Ser20\_pho|ace -> rP14ARF  
 gMIR34A@nucleus-P53|Ser15\_pho|Ser20\_pho|ace|active-..>arMIR34A  
 E2F1|Ser31\_pho|Ser20\_pho|ace-P14ARF-|Mdm2->null  
 Mdm2|pho->null  
 rCHK1/2->CHK1/2  
 gCHK1/2@nucleus-E2F1|Ser31\_pho|Ser20\_pho|ace-..>rCHK1/2  
 rATM->ATM  
 gATM@nucleus-E2F1|Ser31\_pho|Ser20\_pho|ace-..>rATM  
 DNA\_damage-?>PCAF  
 P53-|E2F1|Ser31\_pho|Ser20\_pho|ace-Mdm2 -> P53|ubi|active  
 CHK1/2 -ATM -> CHK1/2|pho|active  
 rP53 -> P53  
 gP53@nucleus -..> rP53  
 P53|ubi|active -> null  
 rMdm2 -> Mdm2  
 gMdm2@nucleus -P53|Ser15\_pho|Ser20\_pho|ace|active@nucleus -..> rMdm2  
 P53|Ser15\_pho|Ser20\_pho|ace|active -/>  
 P53|Ser15\_pho|Ser20\_pho|ace|active@nucleus  
 P53-PCAF -ATM -> P53|Ser15\_pho|Ser20\_pho|ace|active  
 Mdm2-CHK1/2|pho|active-ATM -> Mdm2|pho



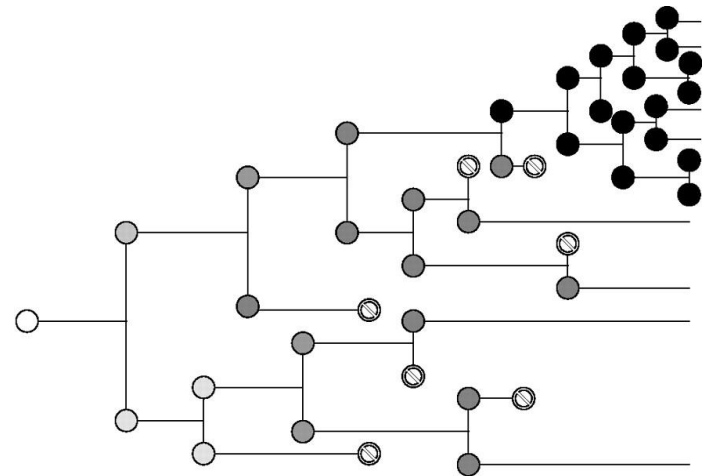
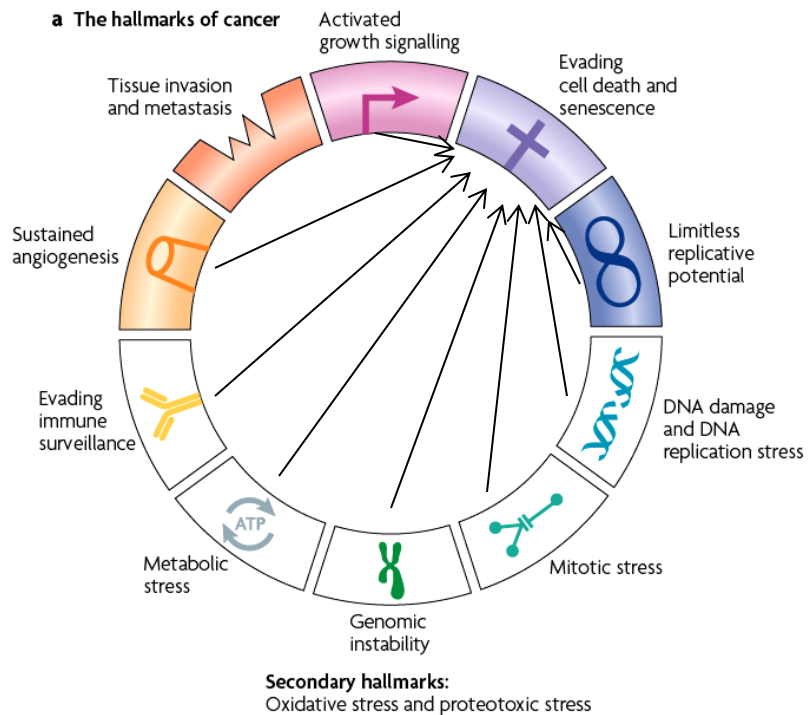


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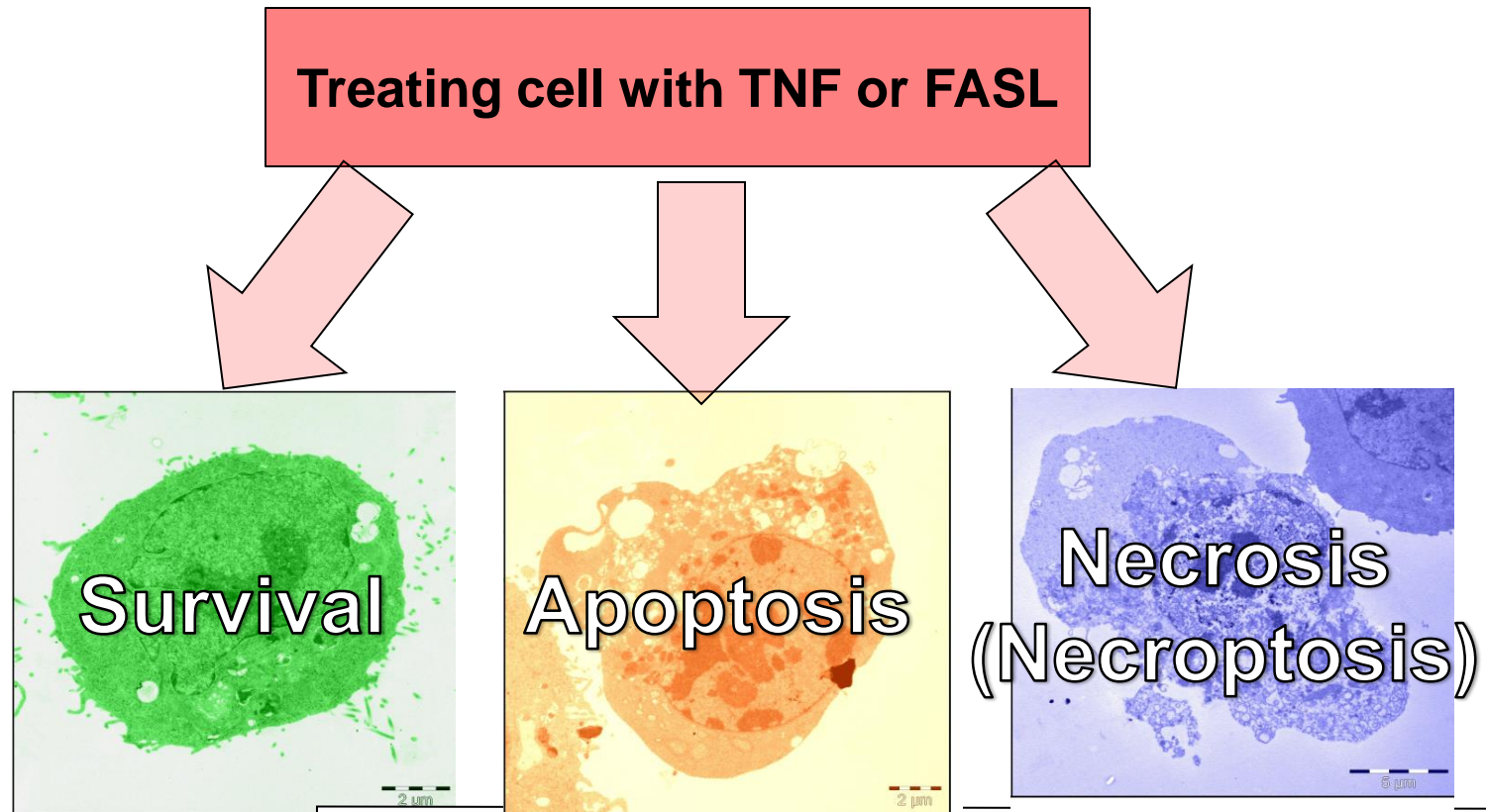
# Cell fate decisions in cell death machinery

Illustrating discrete modeling toolbox

# Cell life/death decisions in cancer



# “Nature integrates empirically”: Apoptosis vs Necrosis vs Survival



OPEN ACCESS Freely available online

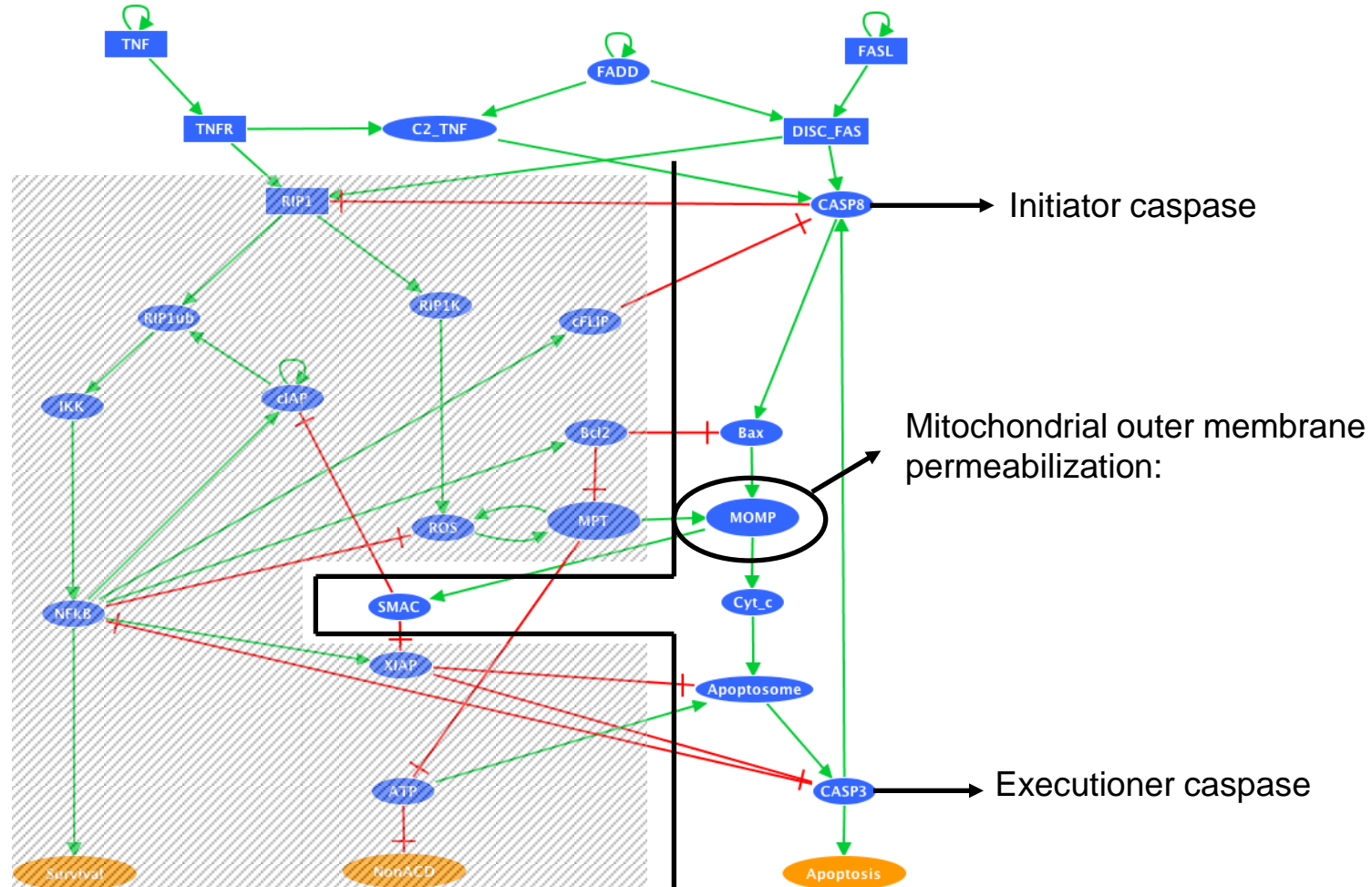
PLoS COMPUTATIONAL BIOLOGY

## Mathematical Modelling of Cell-Fate Decision in Response to Death Receptor Engagement

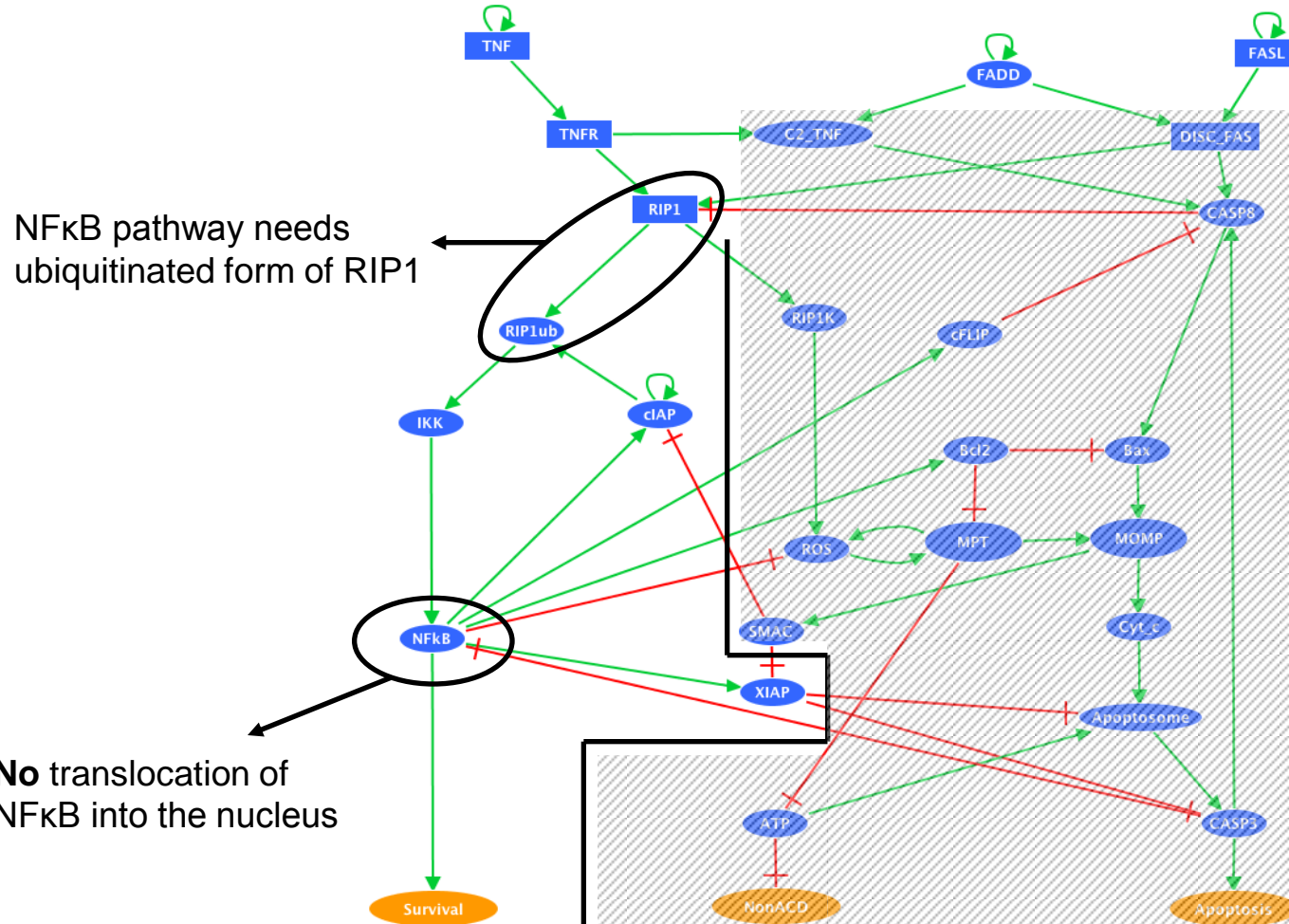
Laurence Calzone<sup>1,2,3\*</sup>, Laurent Tournier<sup>1,2,3</sup>, Simon Fourquet<sup>1,2,3</sup>, Denis Thieffry<sup>4,5</sup>, Boris Zhivotovsky<sup>6</sup>,  
Emmanuel Barillot<sup>1,2,3†</sup>, Andrei Zinovyev<sup>1,2,3†</sup>

<sup>1</sup> Institut Curie, Paris, France, <sup>2</sup> Ecole des Mines ParisTech, Paris, France, <sup>3</sup> INSERM U900, Paris, France, <sup>4</sup> TAGC – INSERM U928 & Université de la Méditerranée, Marseille, France, <sup>5</sup> CONTRAINTE Project, INRIA Paris-Rocquencourt, France, <sup>6</sup> Karolinska Institutet, Stockholm, Sweden

# APOPTOSIS

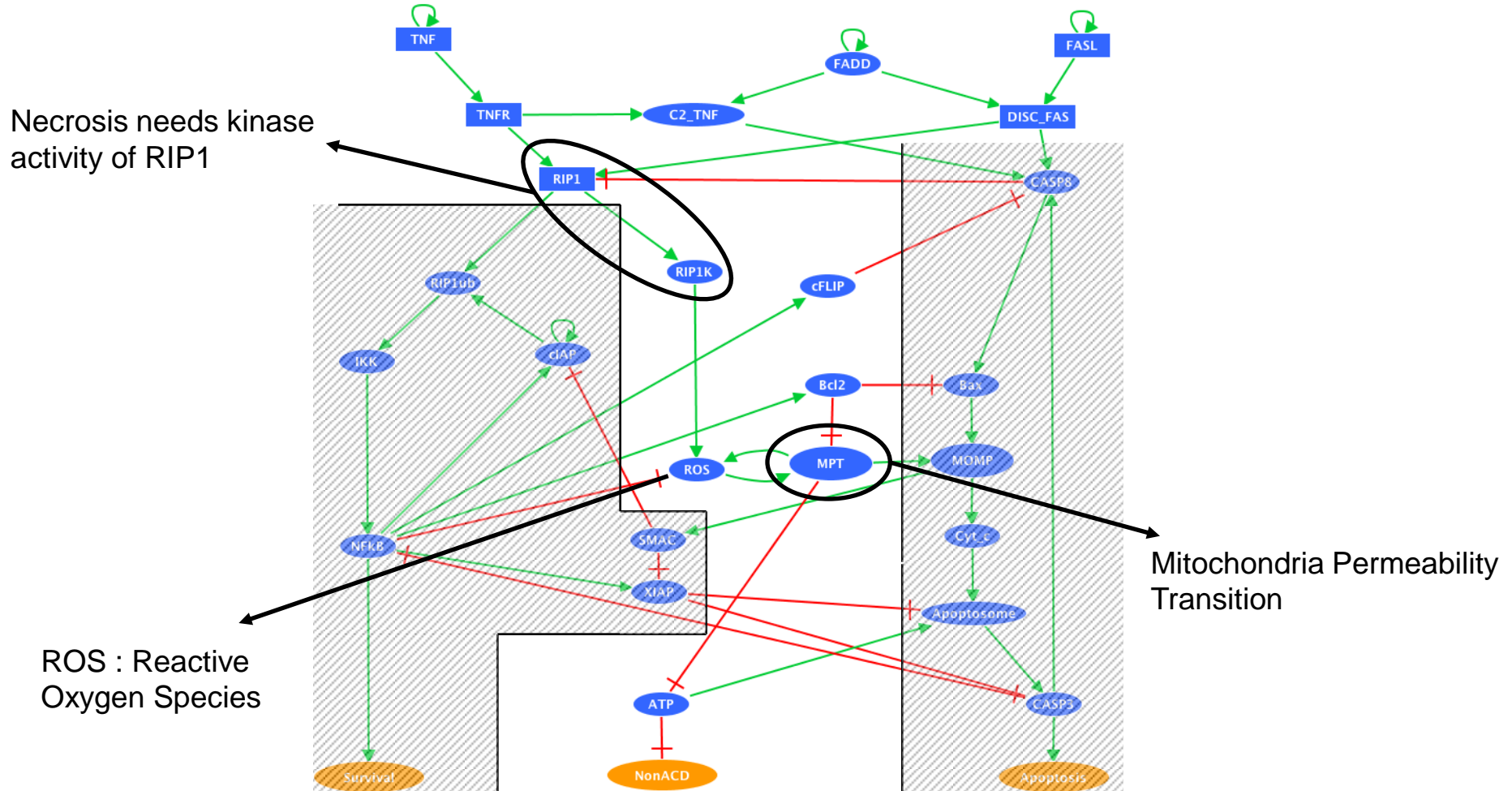


## NFκB pathway



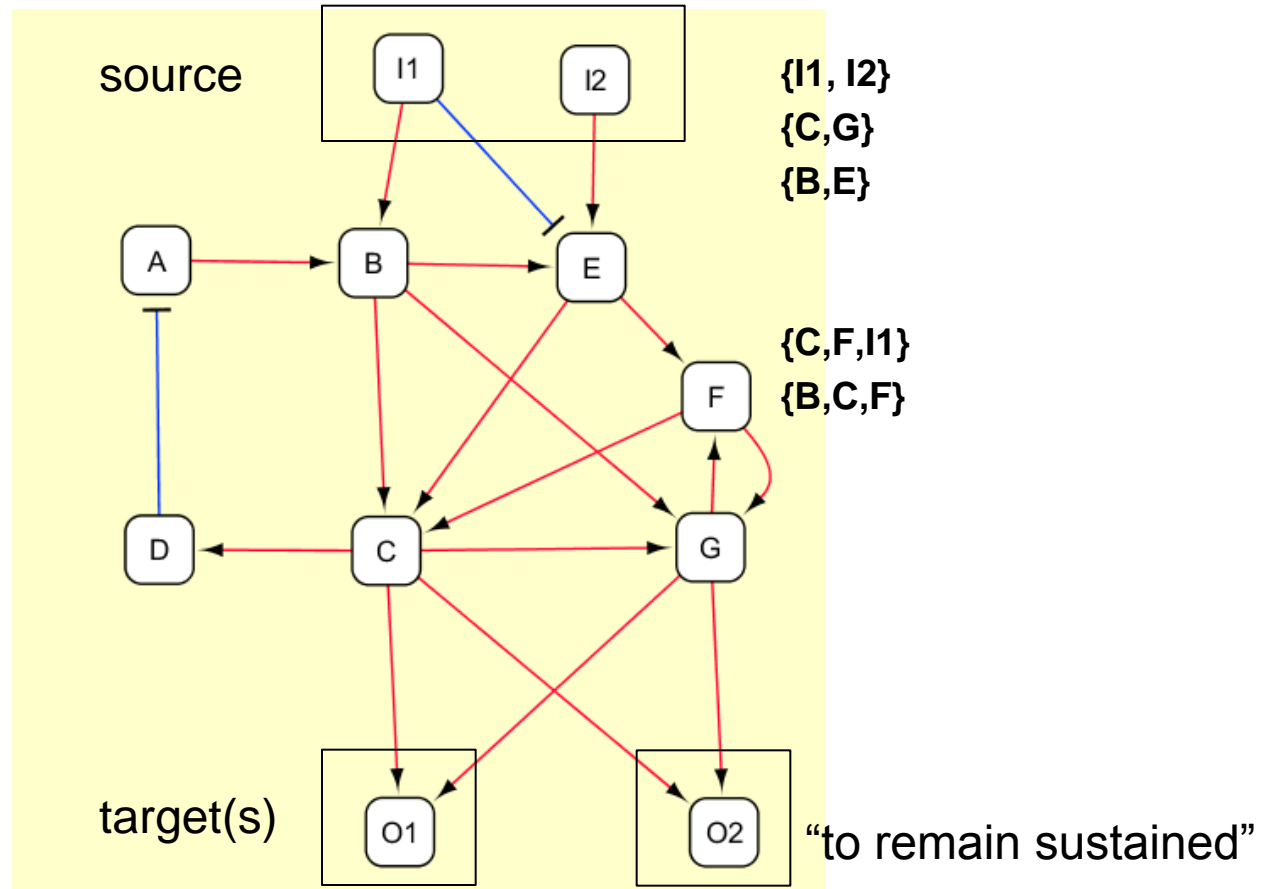


# NECROSIS





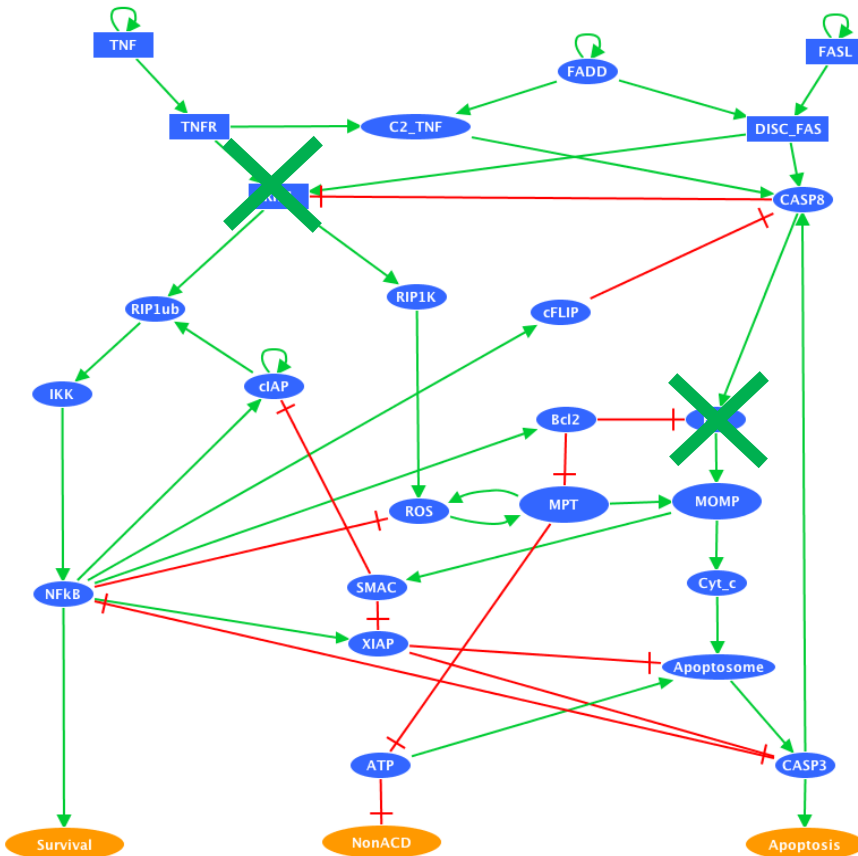
# OCSANA: solving the minimal cut set problem



Vera-Licona, P., et al. OCSANA: optimal combinations of interventions from network analysis. (2013). *Bioinformatics* **15**:29(12).

<http://bioinfo-out.curie.fr/projects/ocsana>

# Cell fate decision in cancer (survival vs apoptosis/necrosis)



## Problem #1 (how): (cancer treatment strategy)

Minimal intervention disabling survival with least effect on cell de

## Problem #2 (cancer treatment strategy)

Minimal intervention disabling cell de survival

Type of Path Analysis	Number of Elementary Paths Nodes	Number and Sizes of CIs	Smallest CIs	Highest Scored CIs (according to OCSANA's score)	Top 5 Most Frequently Present Nodes in all ISs and Number of Occurrences
Shortest Paths	6 elementary paths 20 elementary nodes	Total 130 CIs: 9 CIs of size 2 92 CIs of size 3 24 CIs of size 4	[CASP3 , ATP] [CASP3 , MPT] [CASP3 , ROS] [CASP3 , RIP1K]	[CASP3 , ATP]	XIAP 30 ROS 29 RIP1K 26 MPT 29 ATP 29
Optimal and Sub-optimal Shortest Paths	14 elementary paths 22 elementary nodes	Total 140 CIs: 10 CIs of size 2 59 CIs of size 3 69 CIs of size 4 2 CIs of size 5	[CASP3 , ATP] [CASP3 , RIP1] [CASP8 , RIP1] [BAX , RIP1]	[CASP8 , ATP , RIP1ub]	ROS 38 XIAP 35 NFkB 34 IKK 34 RIP1ub 34
All Non Self-intersecting Paths	124 elementary paths 24 elementary nodes	Total 134 CIs: 9 CIs of size 2 53 CIs of size 3 68 CIs of size 4 4 CIs of size 6	[CASP3 , ATP] [CASP3 , MPT] [MOMP , RIP1] [CASP8 , RIP1]	[ATP , NFkB , MOMP]	ROS 38 XIAP 37 NFkB 33 IKK 33 RIP1ub 33

# Boolean modeling

## Assign logic to nodes

Example of CASP8

**CASP8 = 0** when

DISC-Fas=0 and DISC-TNF=0 and CASP3=0  
(equivalent to no external signals from death receptors  
and no intracellular problems)

cFLIP=1

(equivalent to inhibition by the NFkB pathway)

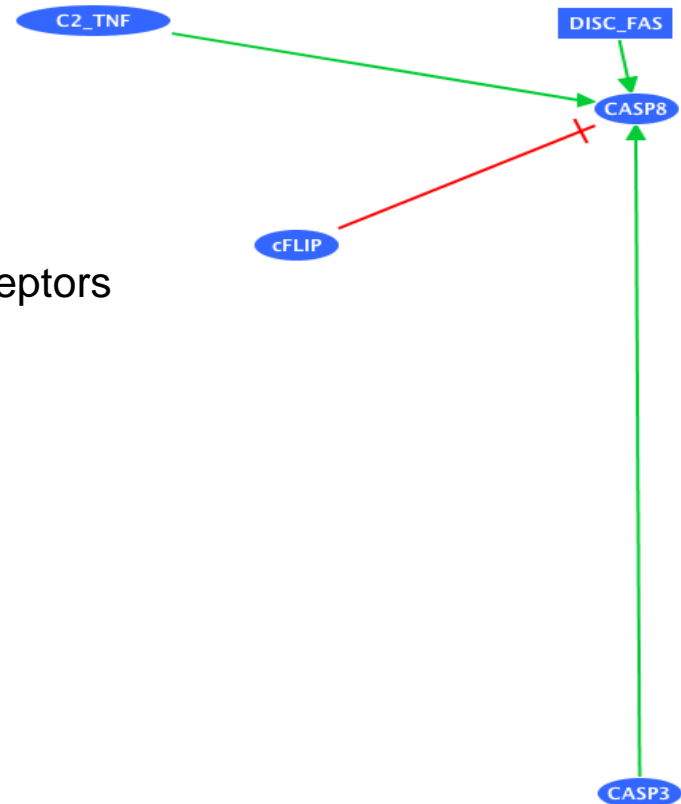
**CASP8 = 1** when

DISC-Fas=1 or/and DISC-TNF=1  
(equivalent to signal from death receptors)

CASP3=1

(amplification signal, feedback activation)

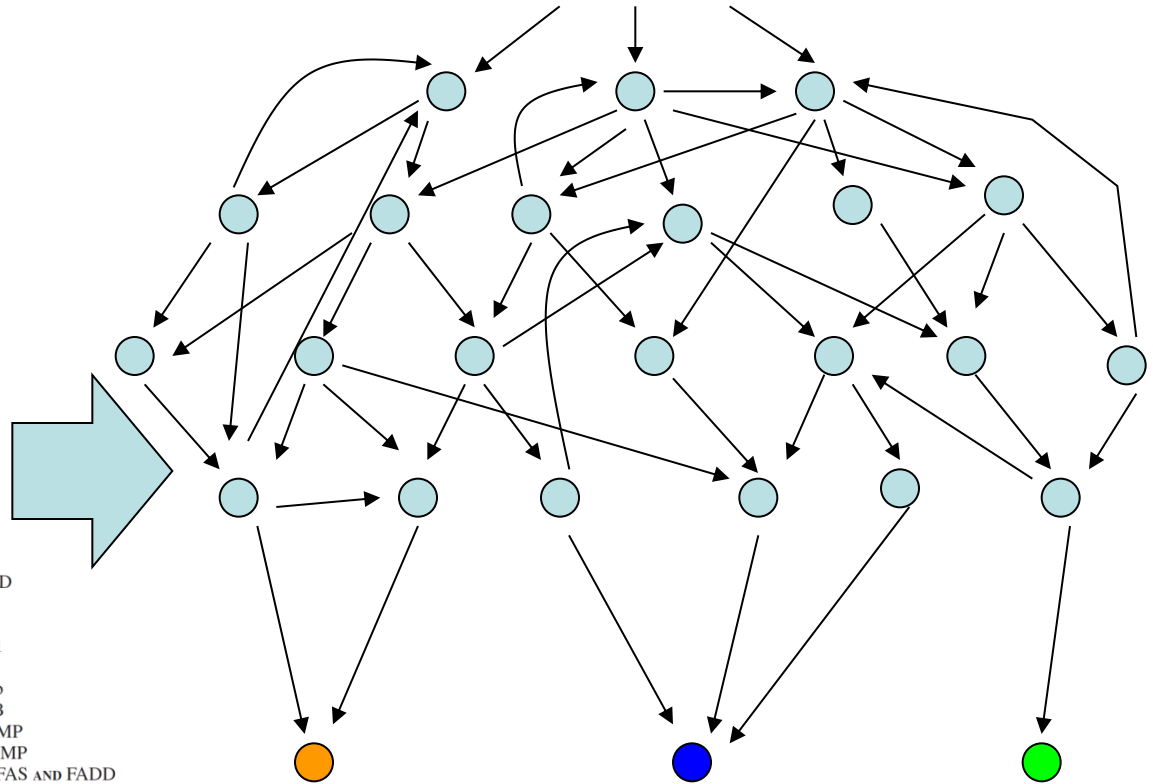
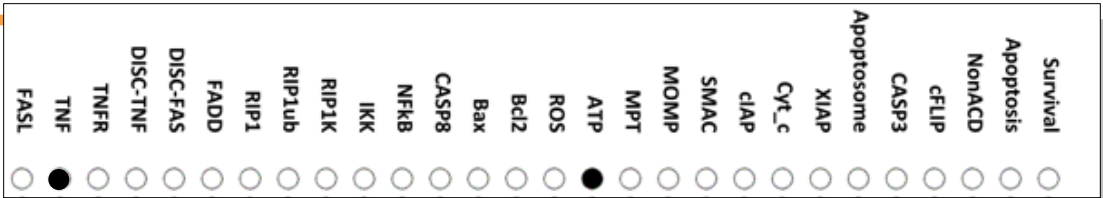
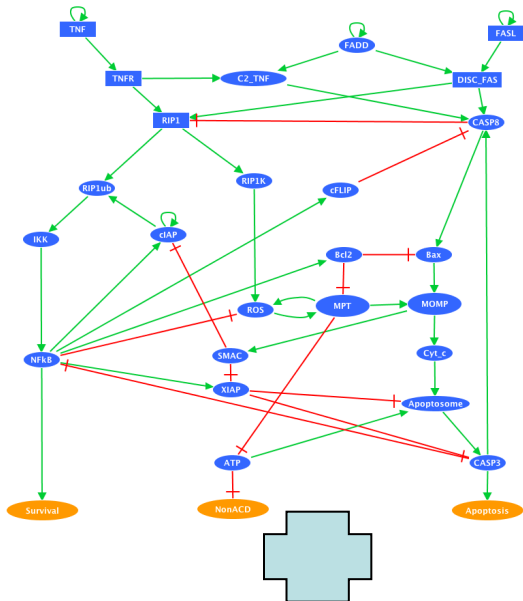
AND no cFLIP





# Asynchronous state transition graph

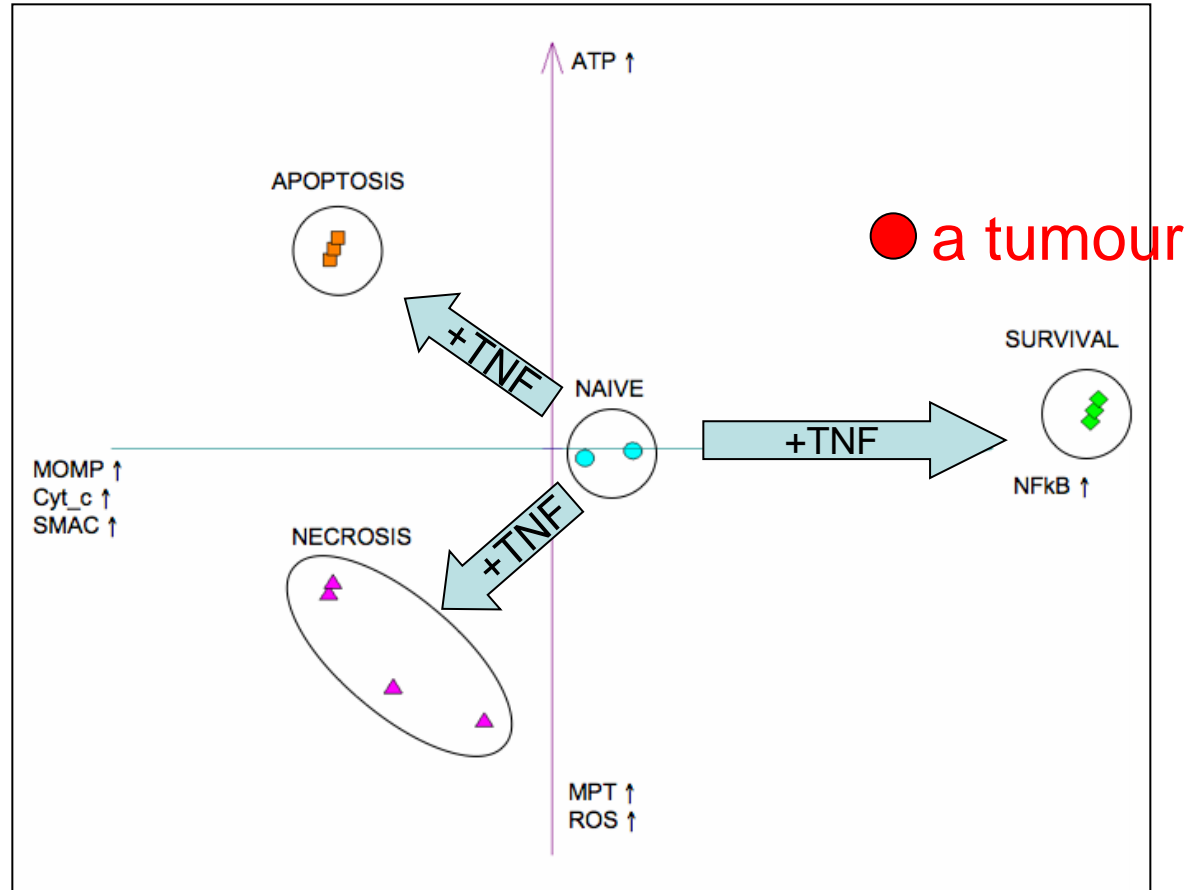
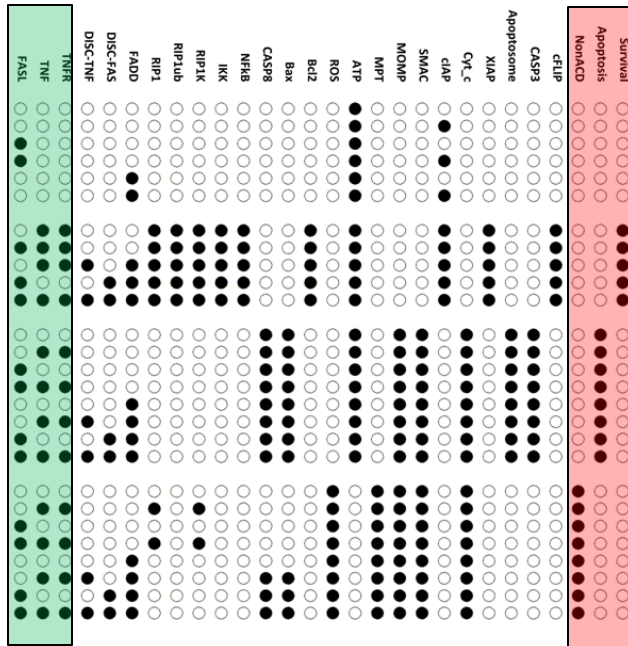
## Influence graph



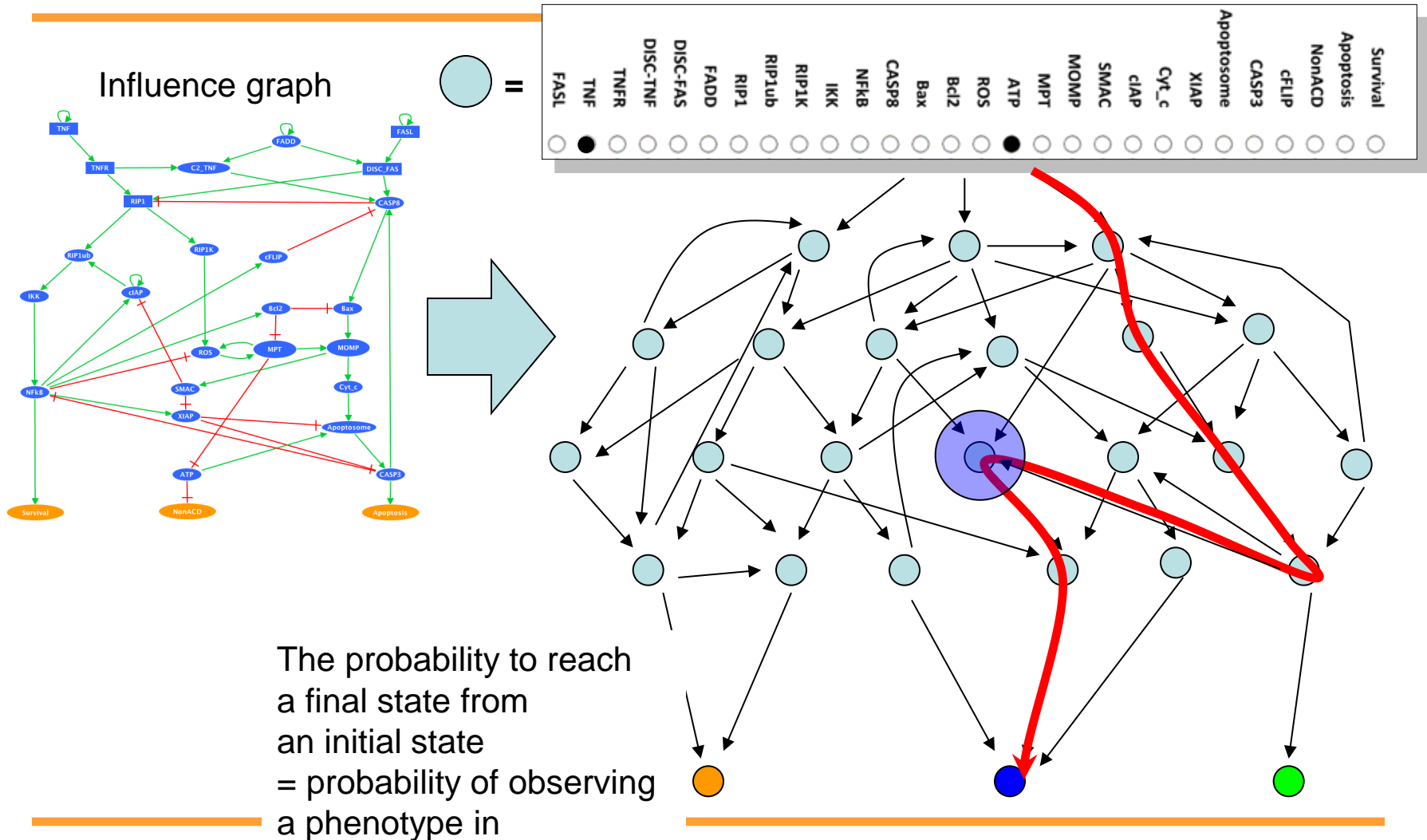
DISC\_TNF' = TNFR and FADD  
 RIP1' = (TNFR or DISC\_FAS) and (NOT CASP8)  
 CASP8' = (DISC\_TNF or DISC\_FAS or CASP3) and (NOT cFlip)  
 RIPub' = RIP1 and cIAP  
 cIAP' = (NFkB or cIAP) and (NOT SMAC)  
 BAX' = CASP8 and (NOT BCL2)  
 ROS' = (RIP1K or MPT) and (NOT NFkB)  
 MPT' = ROS and (NOT BCL2)  
 MOMP' = BAX or MPT  
 NFkB' = IKK and (NOT CASP3)  
 XIAP' = NFkB and (NOT SMAC)  
 Apoptosome' = CYT\_C and ATP and (NOT XIAP)  
 CASP3' = Apoptosome and (NOT XIAP)

TNF' = TNF  
 FADD' = FADD  
 FAS' = FAS  
 TNFR' = TNF  
 RIP1K' = RIP1  
 cFlip' = NFkB  
 IKK' = RIPub  
 BCL2' = NFkB  
 SMAC' = MOMP  
 CYT\_C' = MOMP  
 DISC\_FAS' = FAS and FADD  
 ATP' = NOT MPT

# Structure of attractors: distribution of logical stable states



## Asynchronous state transition graph



# Predicting phenotype probabilities

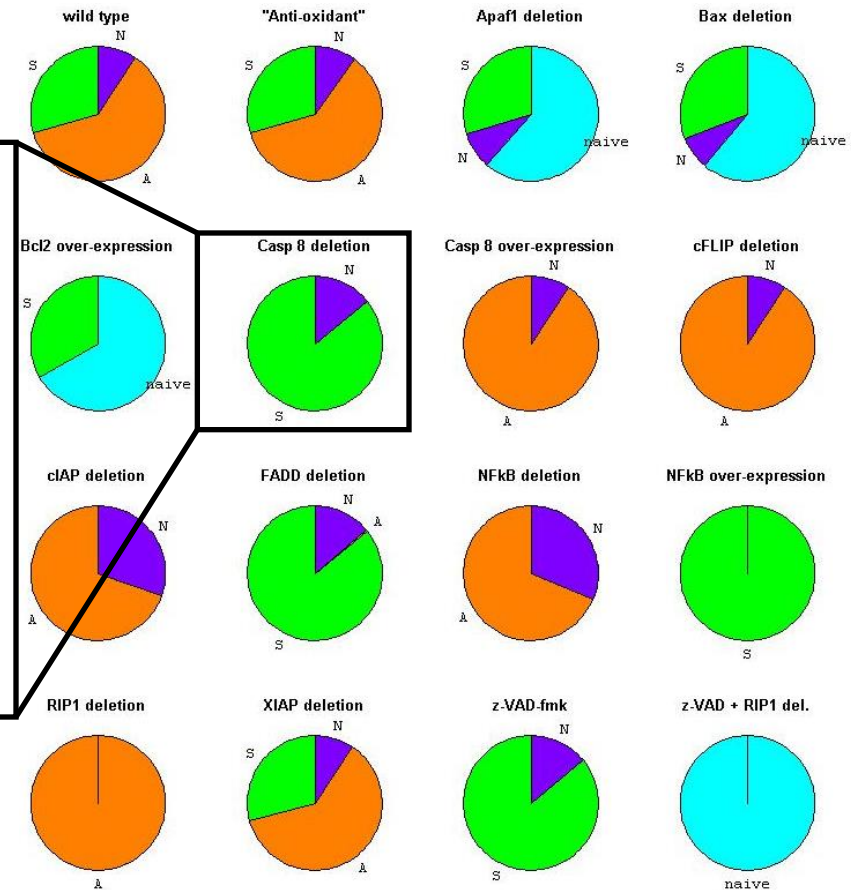
TNF=1

## Example : Caspase 8 deletion

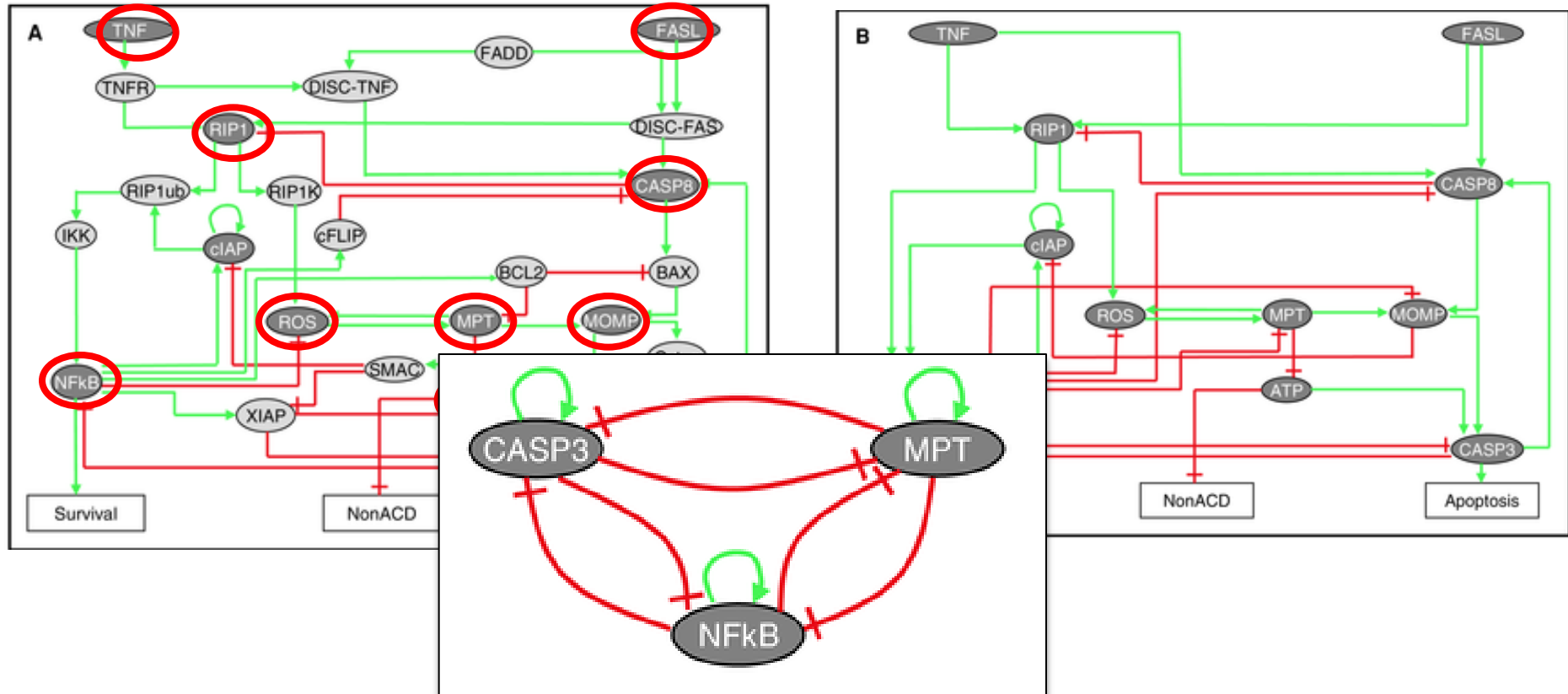
≈ 85% survival (NFκB)  
≈ 15% necrosis  
No apoptosis

### Qualitatively consistent with the literature

“TNF-induced apoptosis is blocked though not necrosis”  
[Kawahara, Ohsawa *et al.*, *J Cell Biol* 1998]  
(Jurkat cells, C8-/-)



# Model reduction of logical models



Naldi A, Remy E, Thieffry D, Chaouiya C. A reduction of logical regulatory graphs preserving essential dynamical properties. *Computational Methods in Systems Biology, Lect. Notes in Bioinformatics* 5688:266-80, 2009.



# “Ligand dosage” experiments

What if the signal was removed...

at which point would the cell commit to one or the other phenotype?

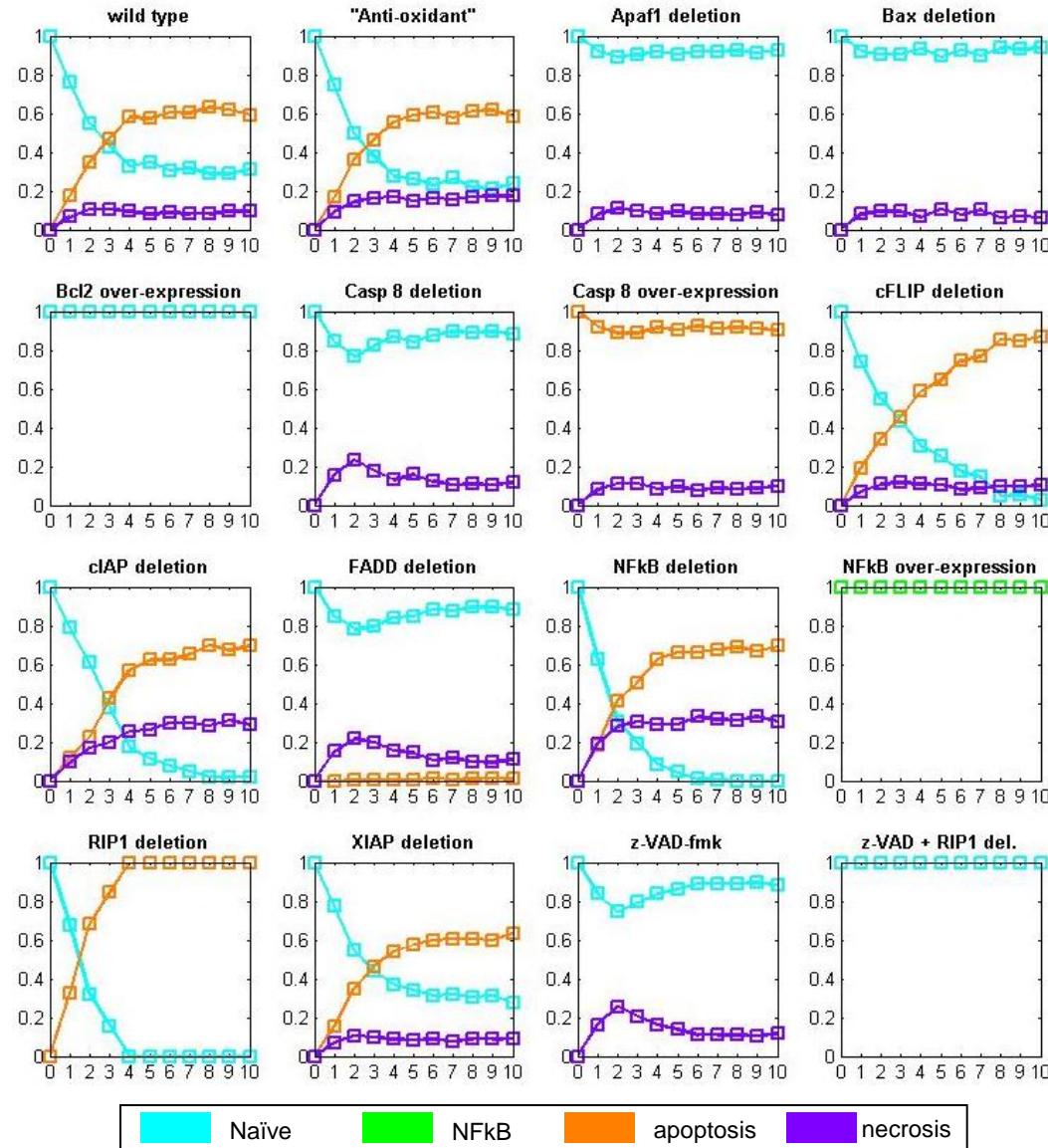
Introduction of “pulse” of TNF instead of constant induction

$t$ : integer

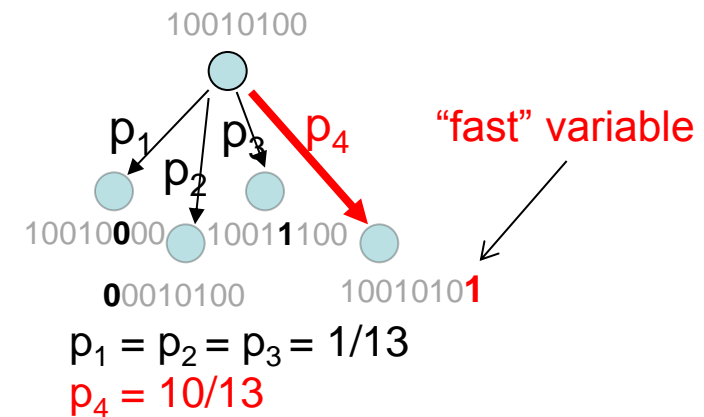
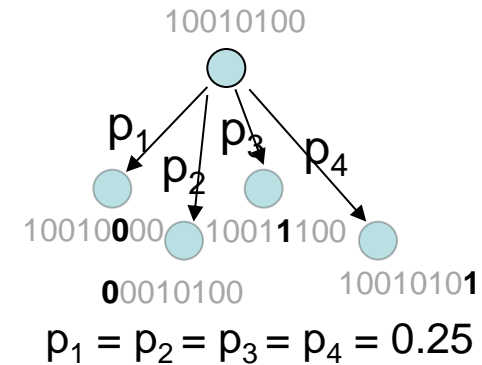
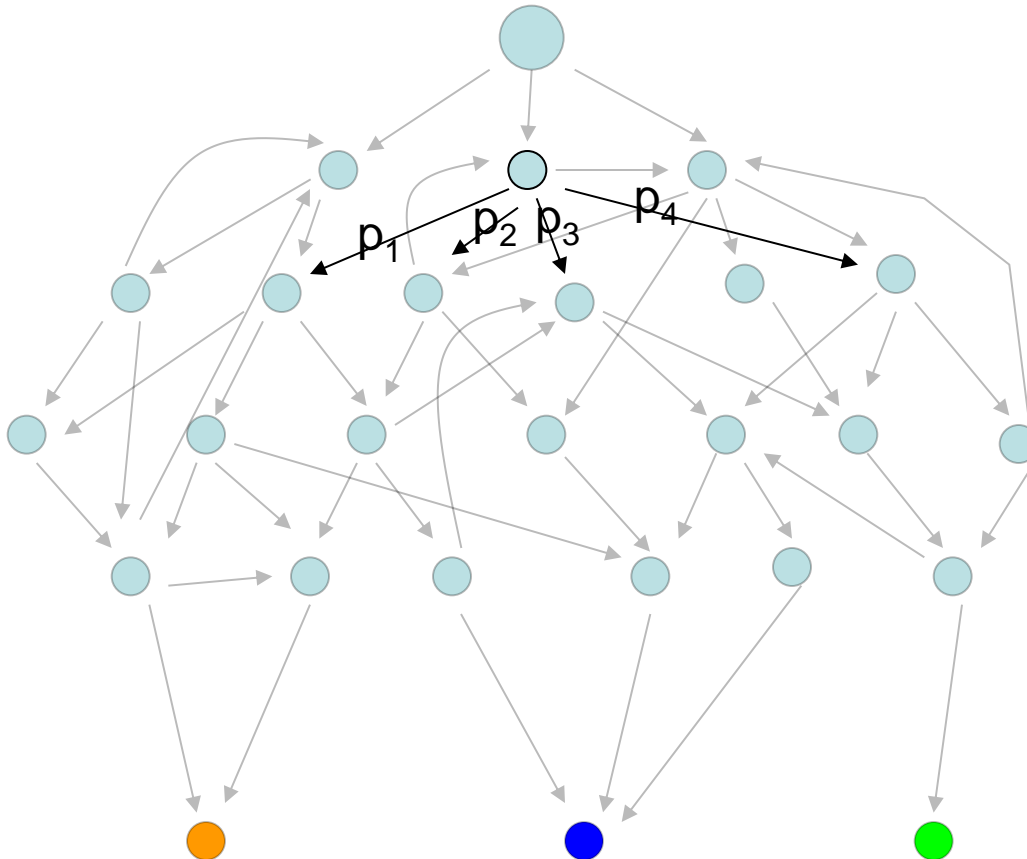
During  $t$  steps, the system evolves with  $TNF=1$

At step  $t+1$ , TNF is switched to 0 (until the end)

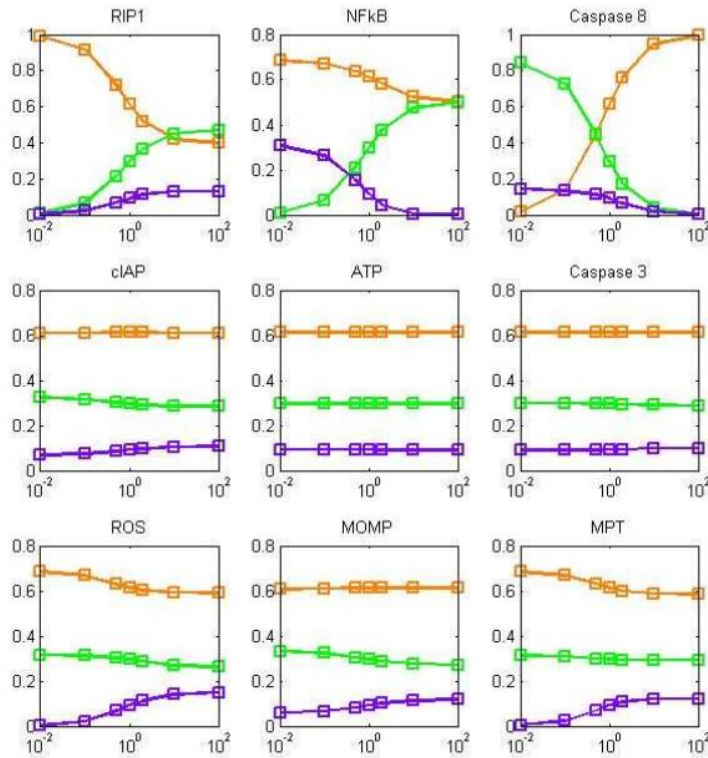
(x-axis  $\rightarrow$  duration of TNF “pulse”)



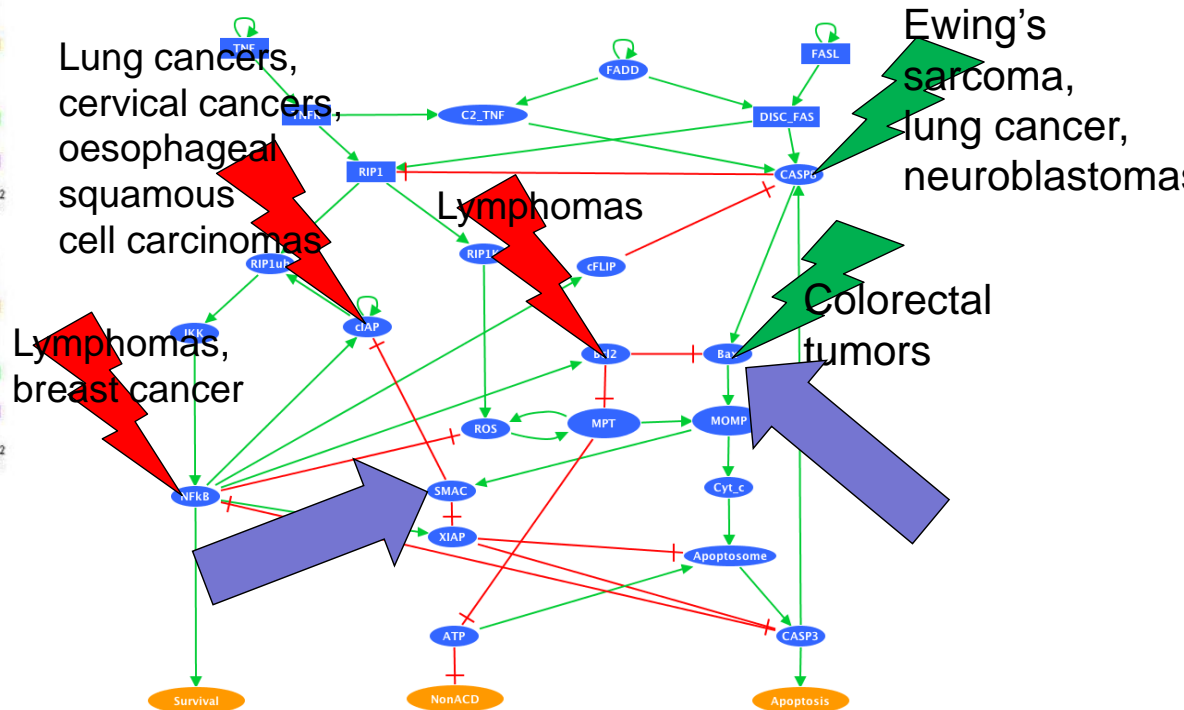
# Sensitivity analysis: testing “rates” of variables



# “Sensitivity” analysis: testing the effect of “rates” of variables



apoptosis  
survival  
necrosis



Zinovyev A, Fourquet S, Tournier L, Calzone L, Barillot E. Cell death and life in cancer: mathematical modeling of cell fate decisions. *Adv Exp Med Biol.* 2012;736:261-74.

# MaBOSS: introducing rates and continuous time

```

node TNFR
{
  logic = TNF;
  rate_up = @logic ? 1.0 : 0.0;
  rate_down = @logic ? 0.0 : 1.0;
}

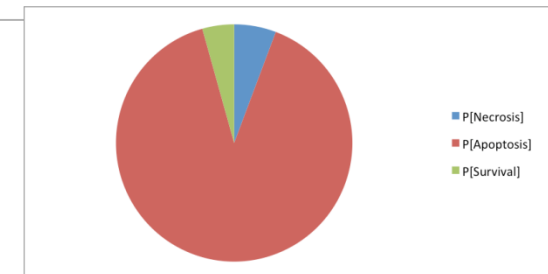
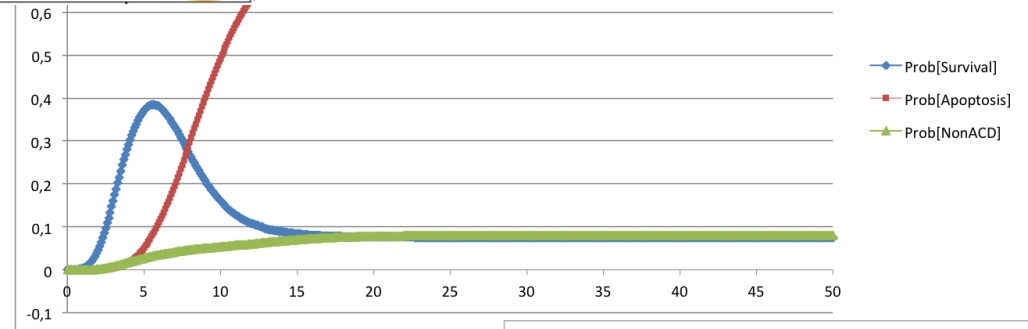
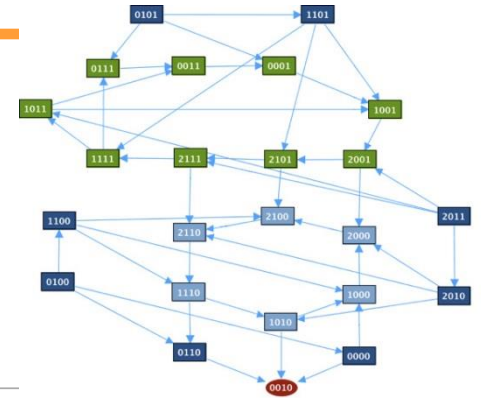
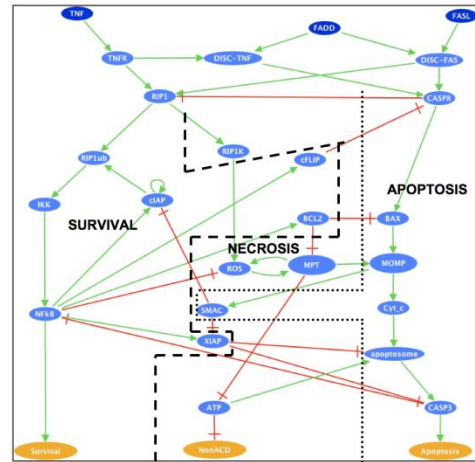
node DISC_TNF
{
  logic = FADD & TNFR;
  rate_up = @logic ? 1.0 : 0.0;
  rate_down = @logic ? 0.0 : 1.0;
}

node DISC_FAS
{
  logic = FASL & FADD;
  rate_up = @logic ? 1.0 : 0.0;
  rate_down = @logic ? 0.0 : 1.0;
}

node FADD
{
  rate_up = 0.0;
  rate_down = 0.0 + 1000*$FADD_del;
}

node RIP1
{
  logic = (DISC_FAS | TNFR) & (!CASP8);
  rate_up = (@logic & (!RIP1_del)) ? 1.0 : 0.0;
  rate_down = (@logic ? 0.0 : 1.0) + 1000*$RIP1_del;
}

node RIP1ub
{
  logic = cIAP & RIP1;
  rate_up = @logic ? 1.0 : 0.0;
  rate_down = @logic ? 0.0 : 1.0;
}
    
```



# Acknowledgements

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INSERM U900 Computational Systems Biology

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**Inna Kuperstein**

**David Cohen**

**Stuart Pook**



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**Boris Zhivotovsky**, Carolinska Institutet

**Gautier Stoll**, Institut Gustav Roussy

**Eric Viara**, Sysra



**Agilent Technologies**