

Mathematical modeling with CellDesigner

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6th Aug. 2017



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Overview

- Introduction of CellDesigner
- What kind of model you can build
- How to build a model with CellDesigner
 - From scratch
 - Import a model, kinetic law and parameters from existing databases

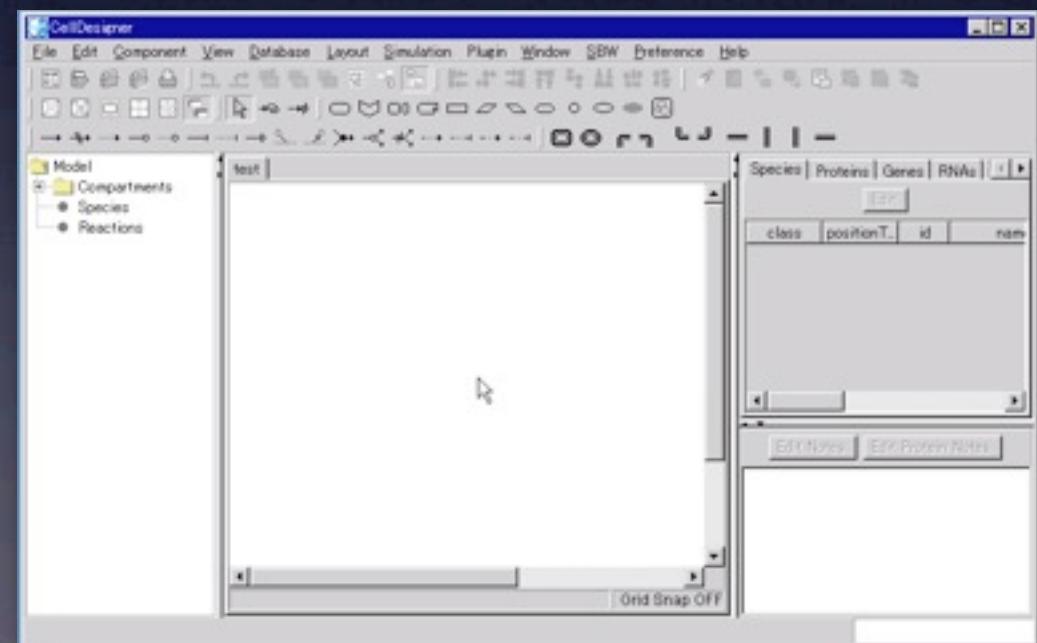
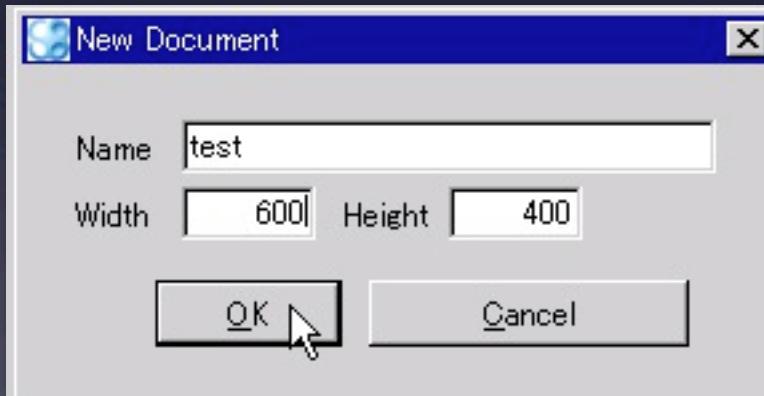
Installation



Create model

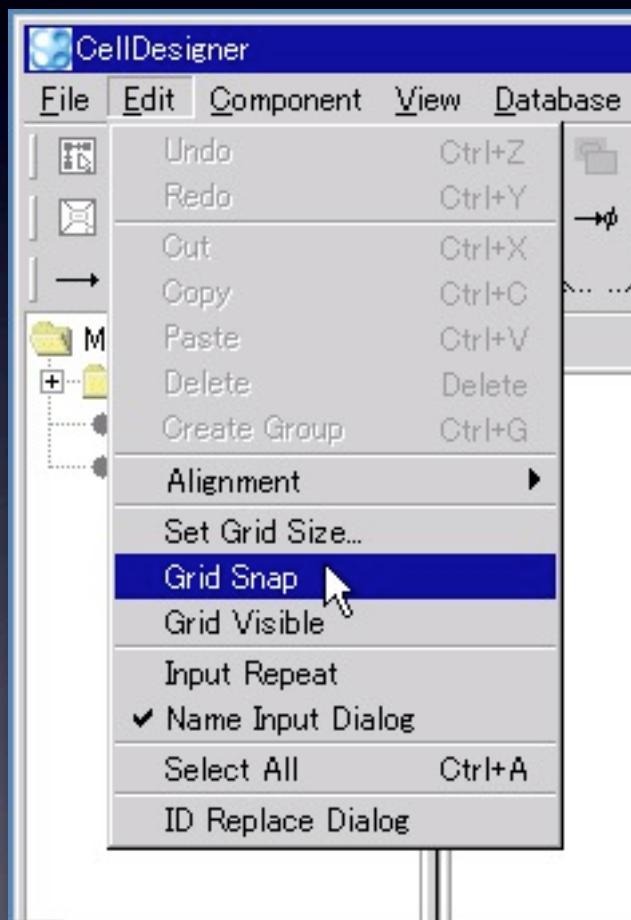
- Create new model:

- [File] → [New] → input title → [OK]



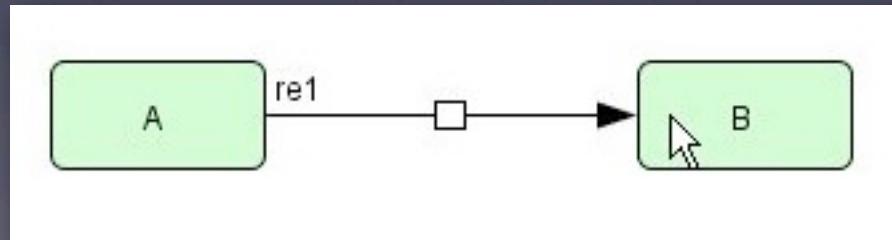
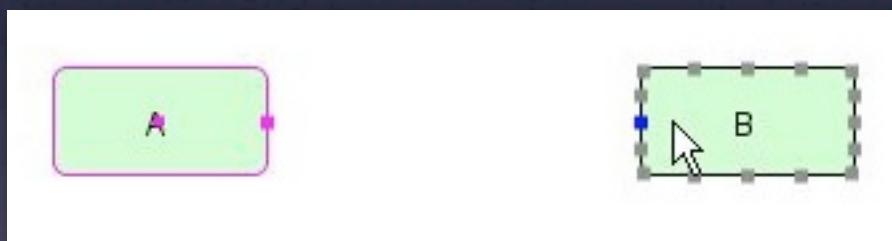
Tips

- Enable [Grid Snap] will help you draw your model much easier



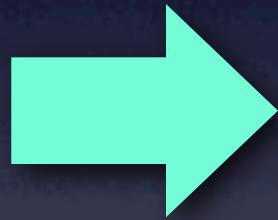
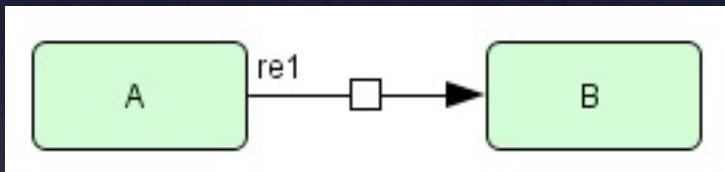
Create Reaction

- Create Protein “A” and “B”
- Draw “State transition” → arrow from “A” to “B”



Simulation (ex1)

- Create following biochemical reaction
- Click [Simulation] → [ControlPanel]
and call SBML ODE Solver

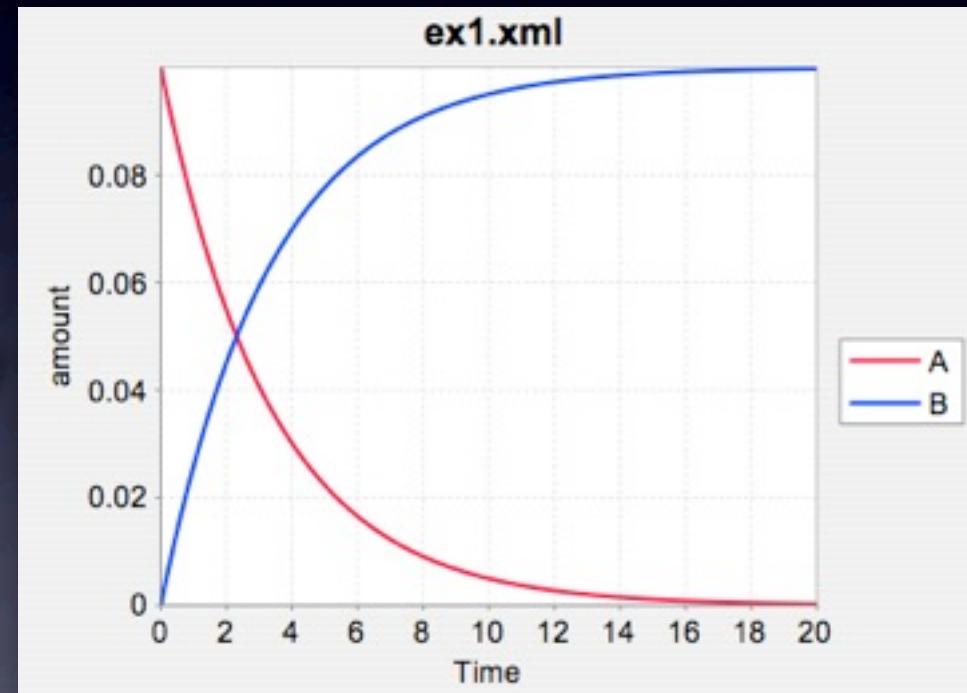


$$v = k[A]$$

$$k = 0.3$$

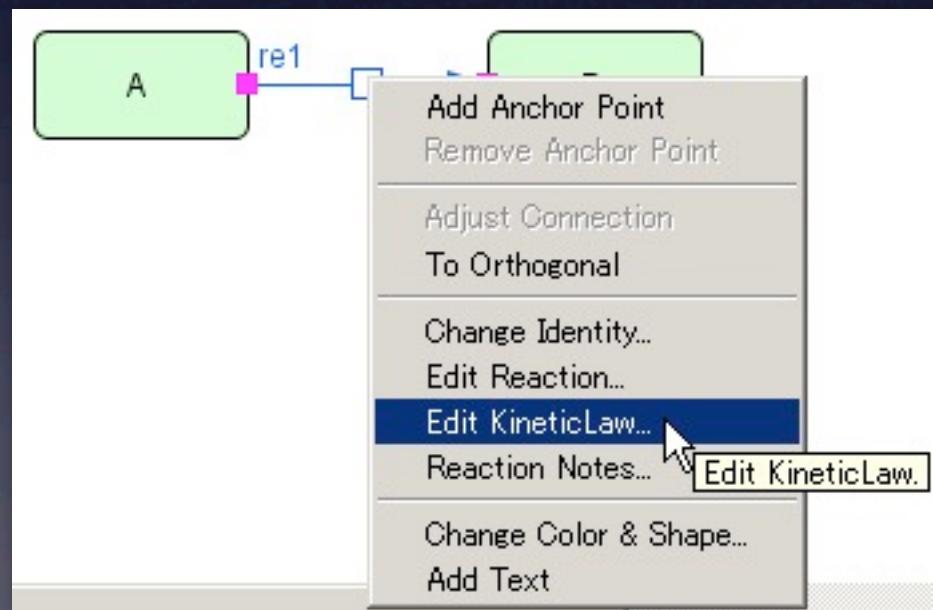
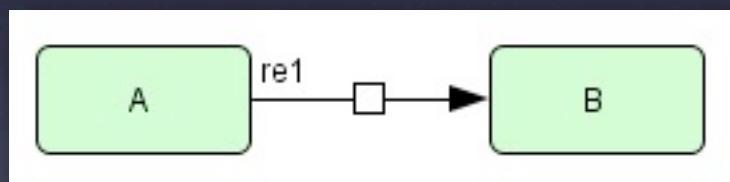
$$A = 0.1$$

$$B = 0$$



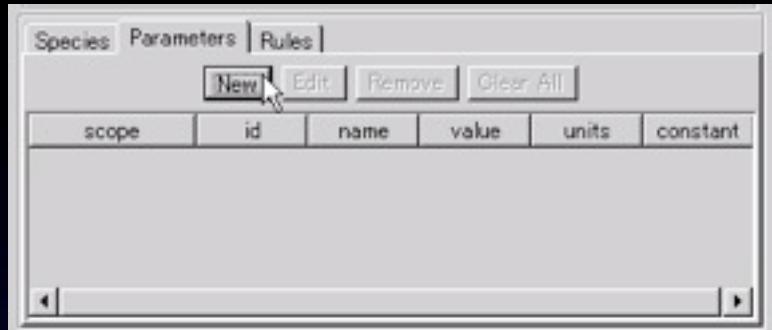
Simulation (ex1)

- Right click on the reaction and select [Edit KineticLaw...]



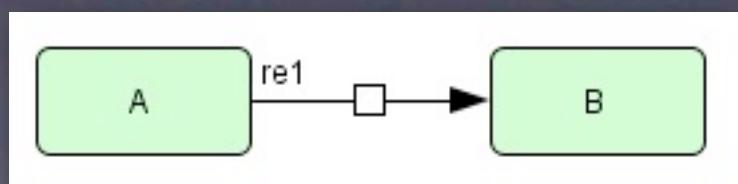
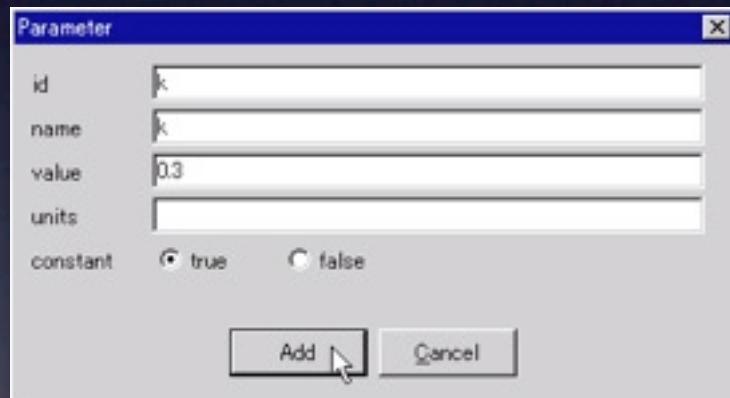
Simulation (ex1)

- Click [New] button on [Parameters] tab



- Input values as follows:

- id: k
- name: k
- value: 0.3



$$v = k[A]$$

$$k = 0.3$$

$$A = 0.1$$

$$B = 0$$

Simulation (ex1)

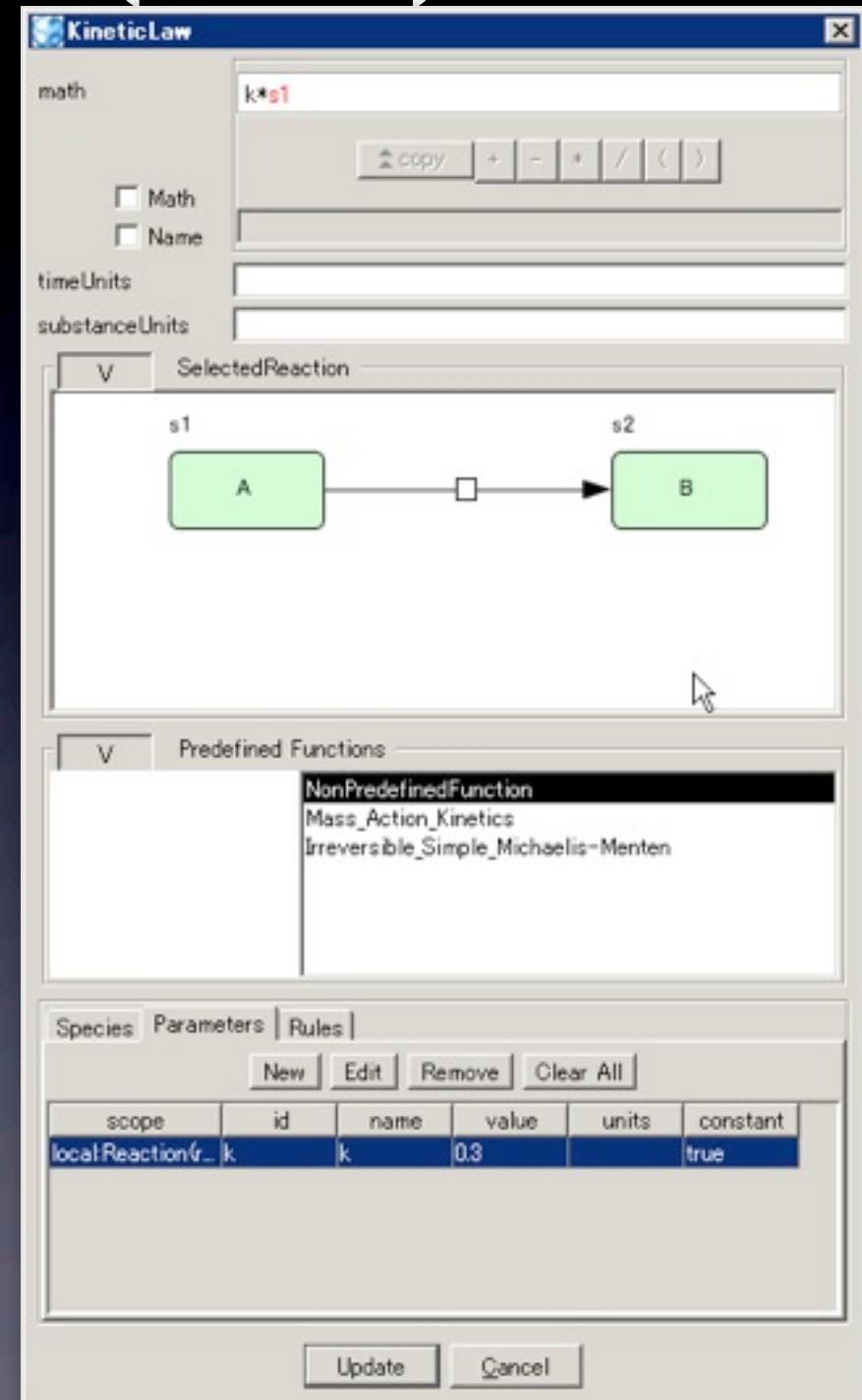
- Click top most text field
- Type $k * (k \text{ times})$
- Select Protein “A”
- Click [Name] checkbox
($k^*s1 \rightarrow k^*A$)

$$v = k[A]$$

$$k = 0.3$$

$$A = 0.1$$

$$B = 0$$



KineticLaw

math
 Math
 Name

+ - * / ()

V SelectedReaction

```

graph LR
    A[A] --> B[B]
    subgraph "SelectedReaction"
        A
        B
    end
    
```

V Predefined Functions

- NonPredefinedFunction
- Mass_Action_Kinetics
- Irreversible_Simple_Michaelis-Menten

Species Parameters Rules

New Edit Remove Clear All

scope	id	name	value	units	constant
local:Reaction	k	k	0.3	substan...	true

Update Cancel

Species

Edit Export

class	id	name
ROTEIN	s1	A
ROTEIN	s2	B

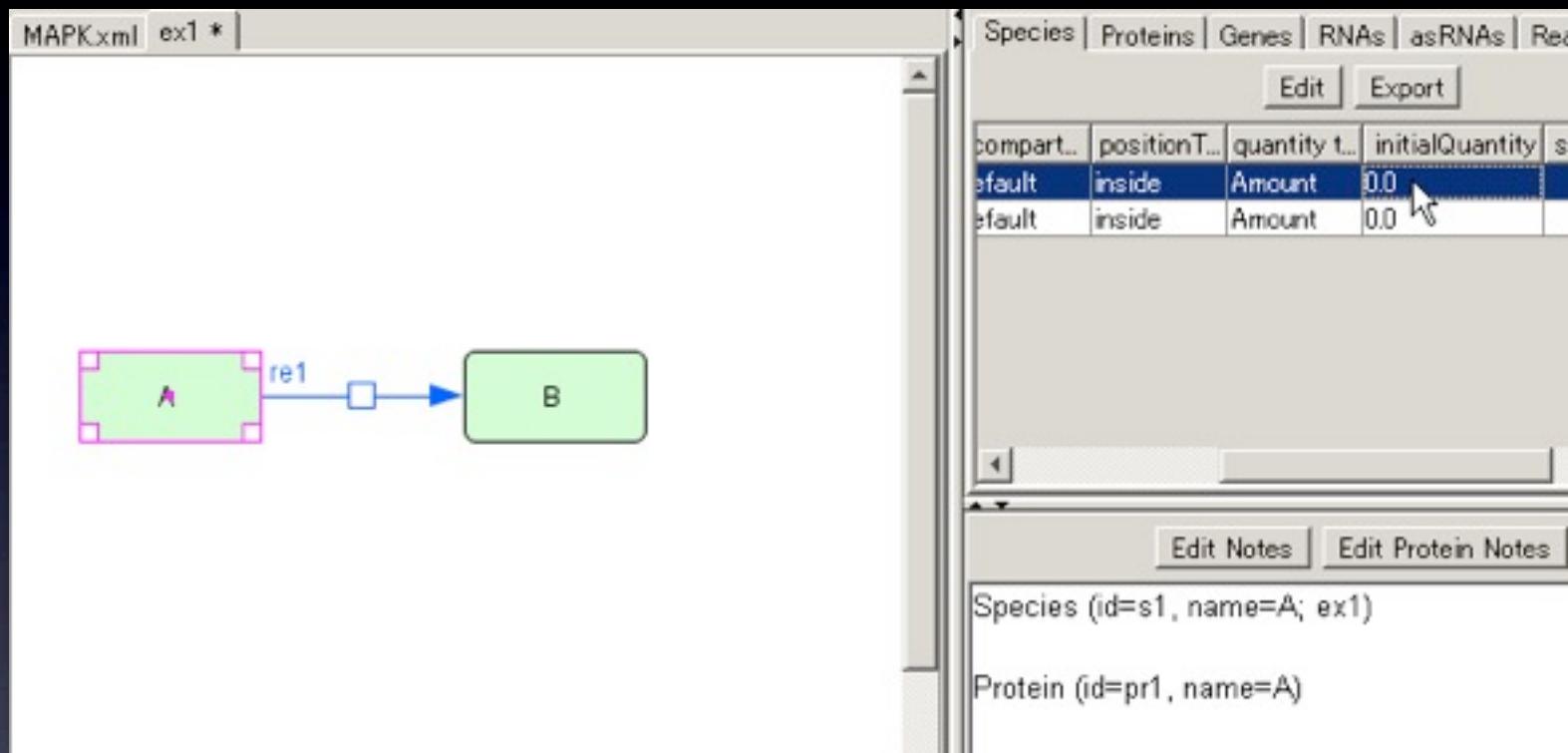
NOTE MIRIAM

Edit Notes

reaction (id=re1, name=; est)

Simulation (ex1)

- Double click [initialQuantity] column for Protein “A”



- Set value as 0.1

$$v = k[A]$$

$$k = 0.3$$

$$A = 0.1$$

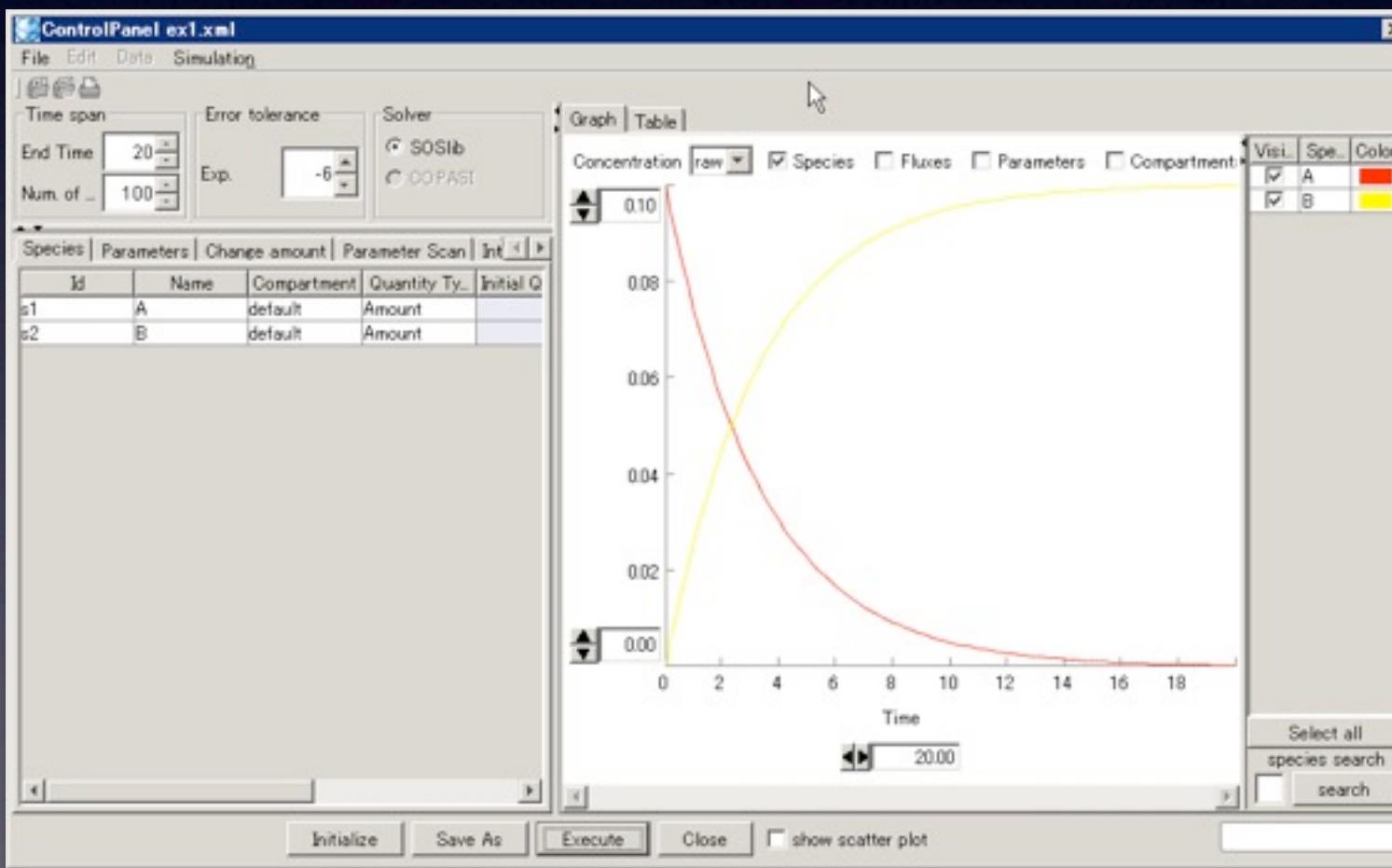
$$B = 0$$

Simulation (ex1)

- Click [Simulation] → [ControlPanel]
- Set [End Time] to 20
- Click [Execute] button

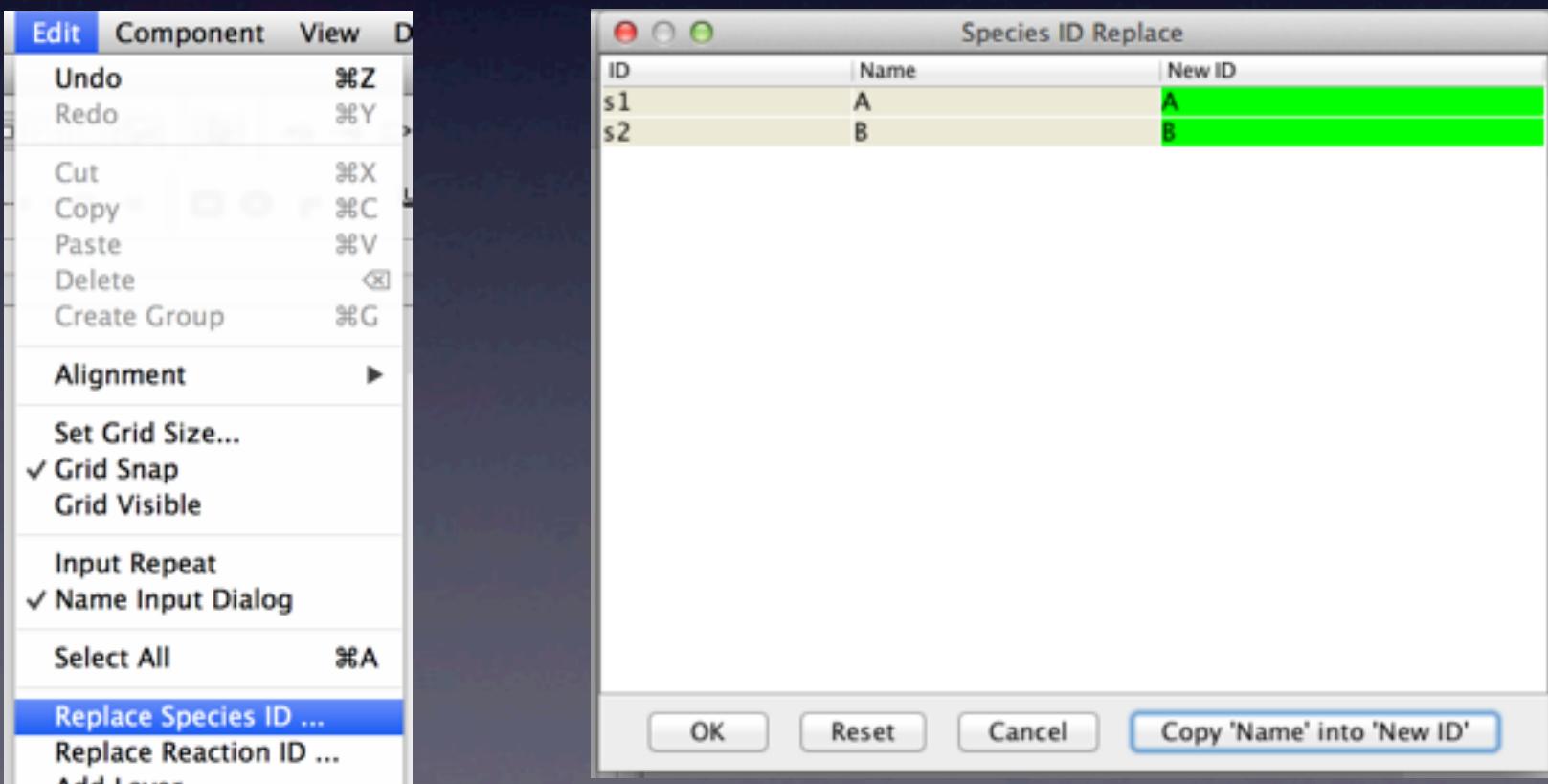
Time span

End Time	20
Num. of Points	100

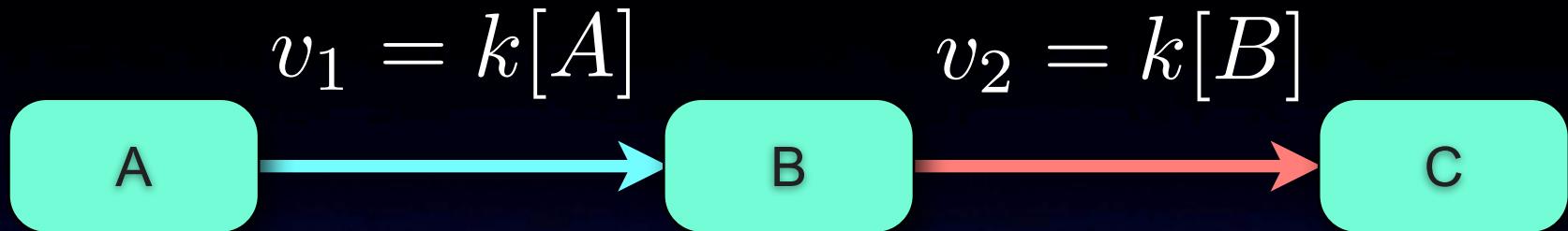


Rename Species ID (s1 → A)

- Click [Edit] → [Replace Species ID]
- Click [Copy ‘Name’ into ...] button
- Use “Species Name” in KineticLaw Editor



Network → Equation



$$\frac{d[A]}{dt} = -k[A]$$

$$\frac{d[B]}{dt} = k[A] - k[B]$$

$$\frac{d[C]}{dt} = k[B]$$

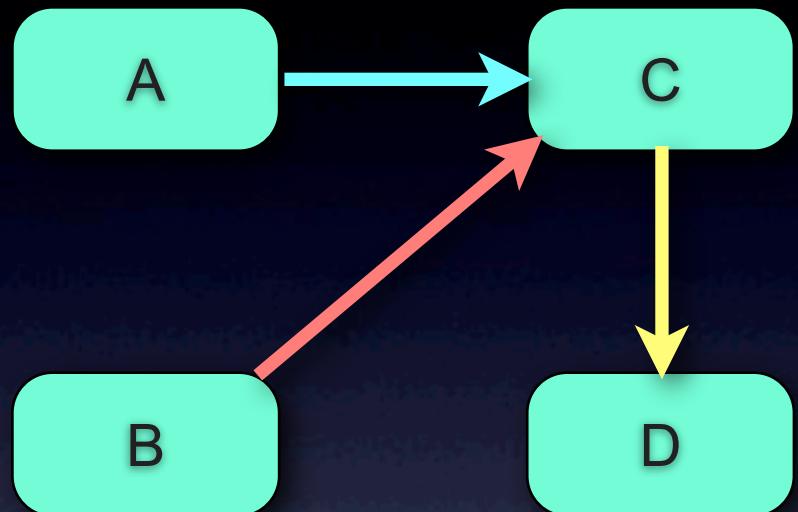
Equation → Network

$$\frac{dA}{dt} = -k_1 A$$

$$\frac{dB}{dt} = -k_2 B$$

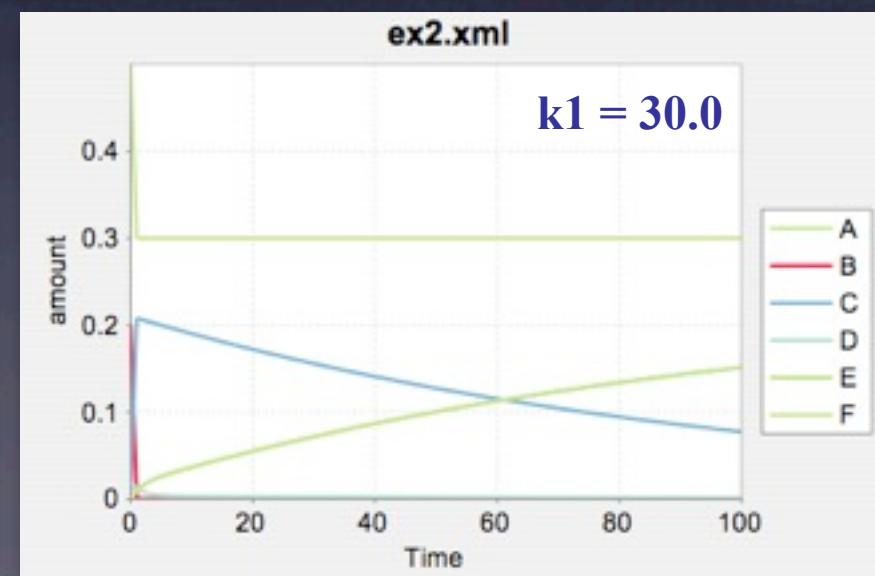
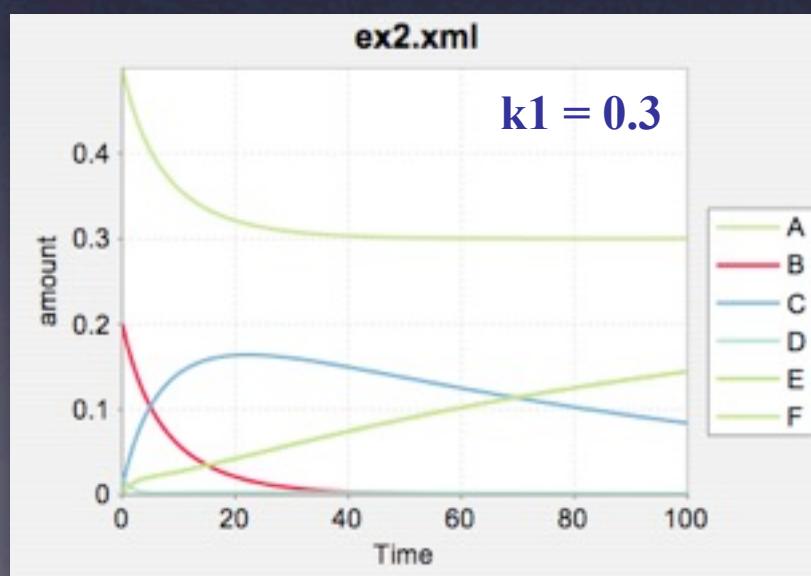
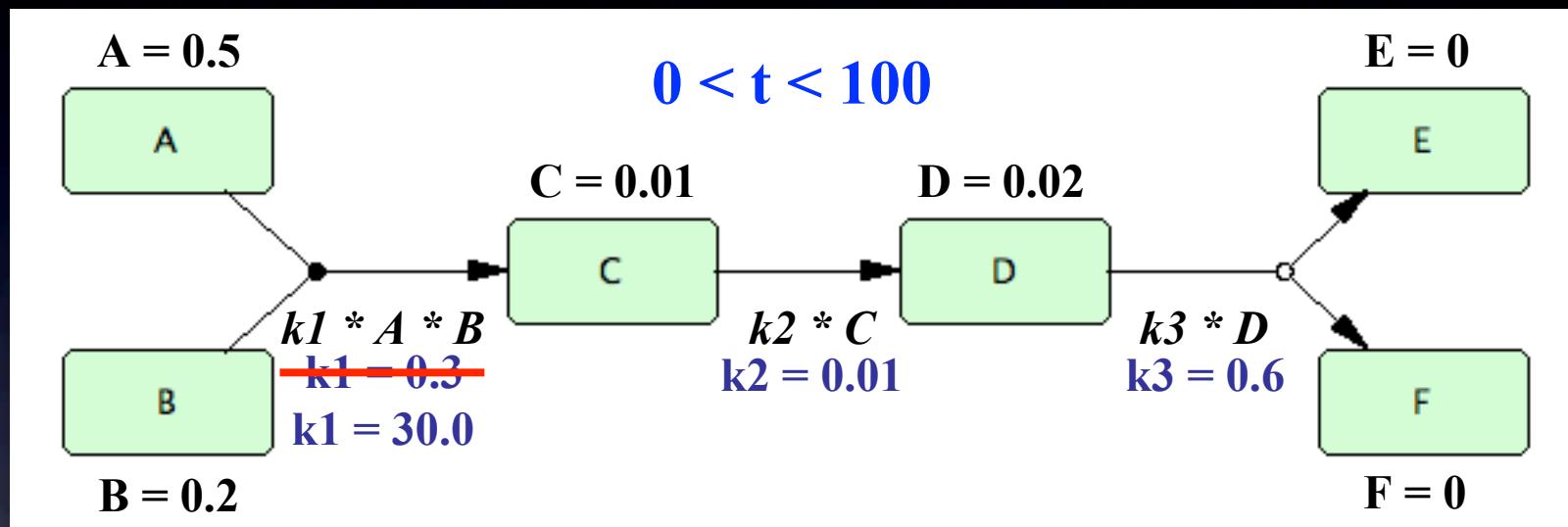
$$\frac{dC}{dt} = k_1 A + k_2 B - k_3 C$$

$$\frac{dD}{dt} = k_3 C$$



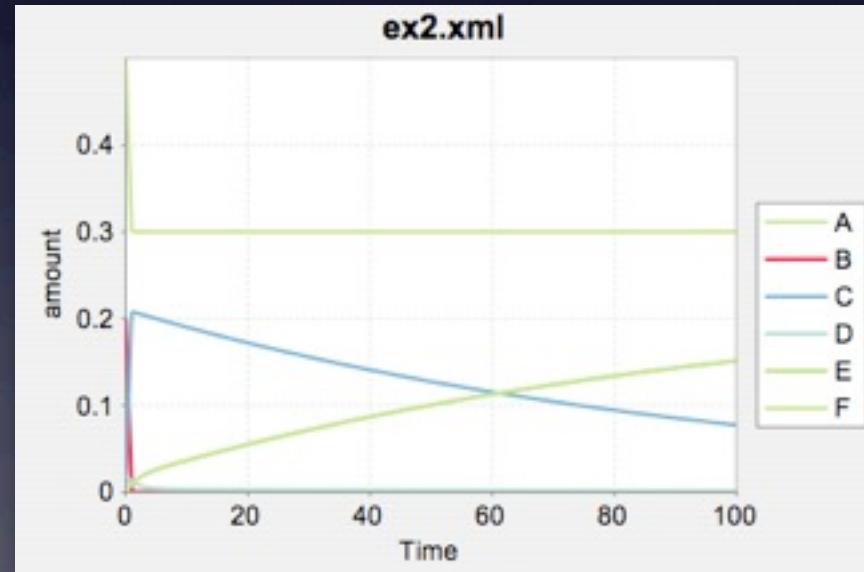
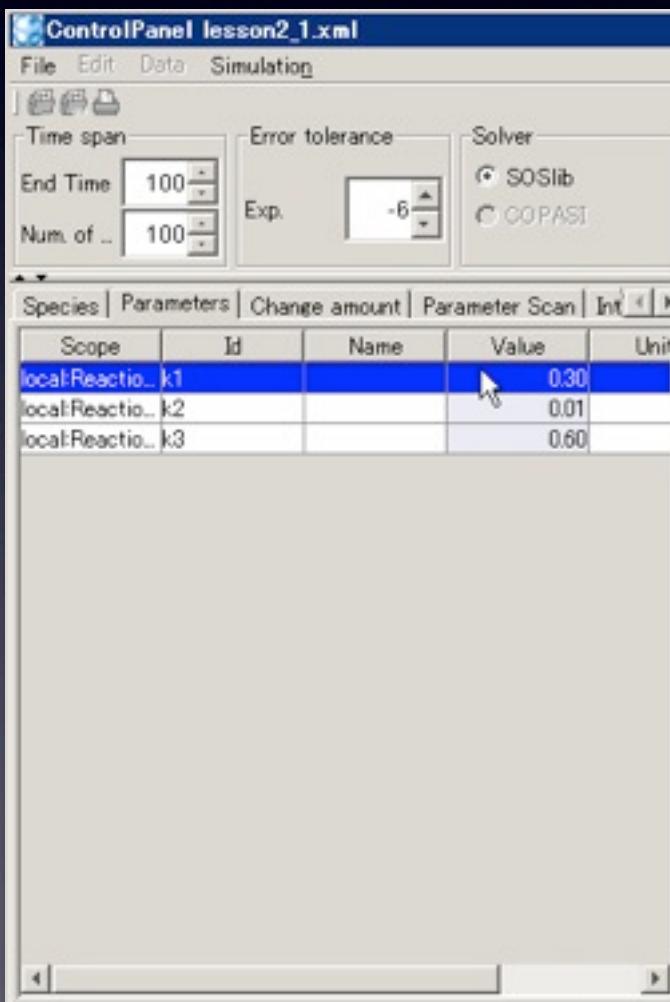
Simulation (ex2)

Change parameter k1 to 30.0



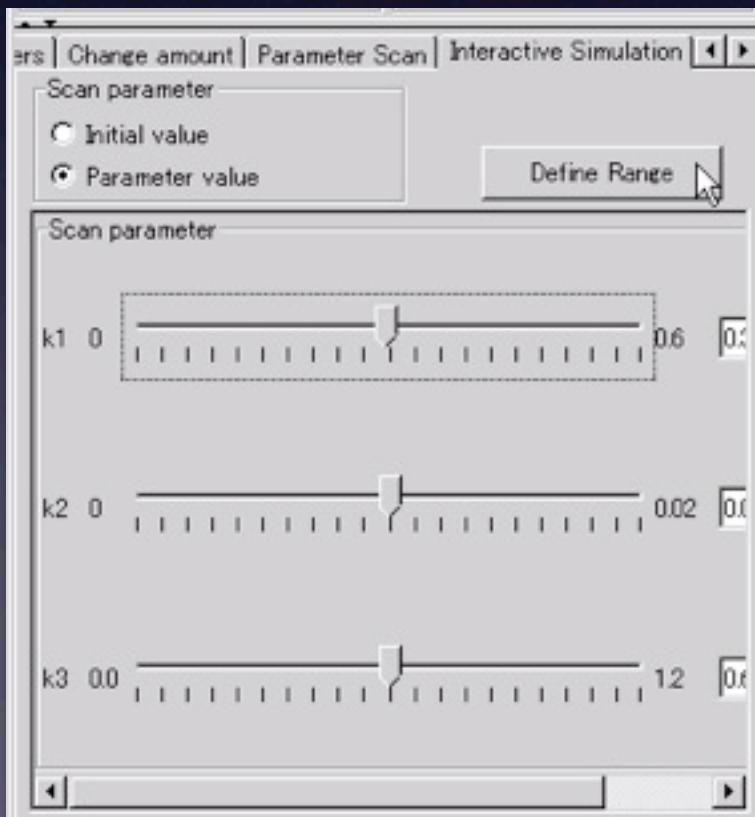
Simulation (ex2)

- Click [Parameters] tab
- Double click [Value] column for k_1
- Change parameter k_1 to 30.0



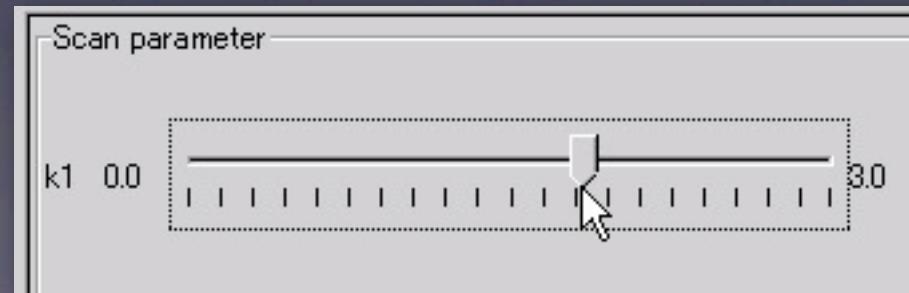
Simulation (ex2)

- Click [Interactive Simulation] tab
- Click [Parameter value] radio button
- Click [Define Range] button
- Click [Max] column for k1 and set value as 3.0



Define Slider Range				
Id	Min	Max	Current	
k1	0.0	3.00	0.30	
k2	0.0	0.02	0.01	
k3	0.0	1.20	0.60	

Drag sliderbar for k1



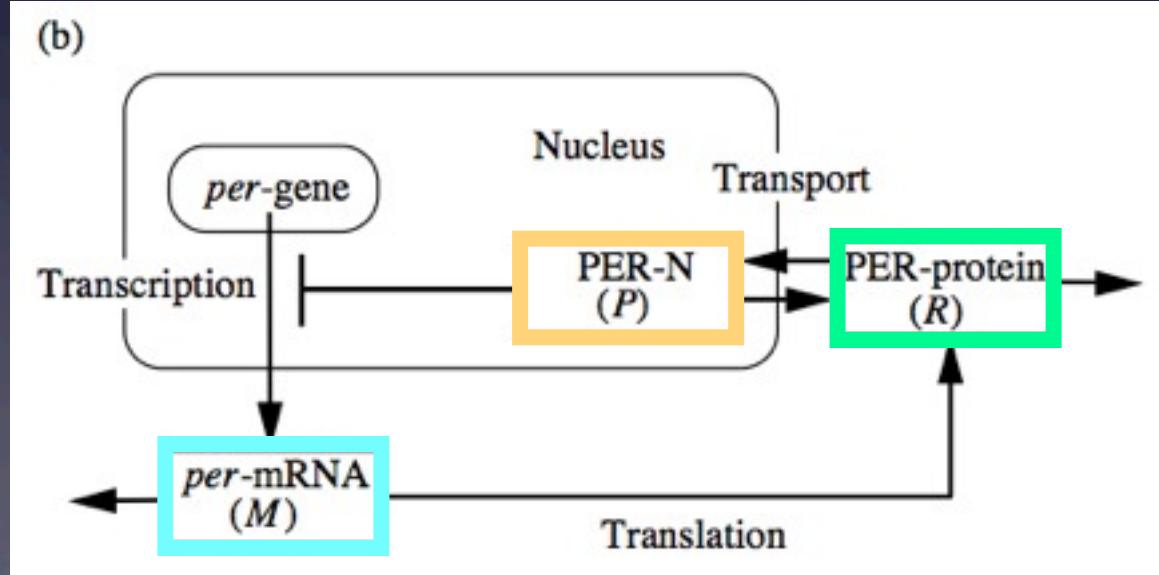
Circadian clock model

- Protein (P) **inhibits** transcription of mRNA (M)
- M is translated to Protein (R)
- P / R will be transported to cytosol / nucleus

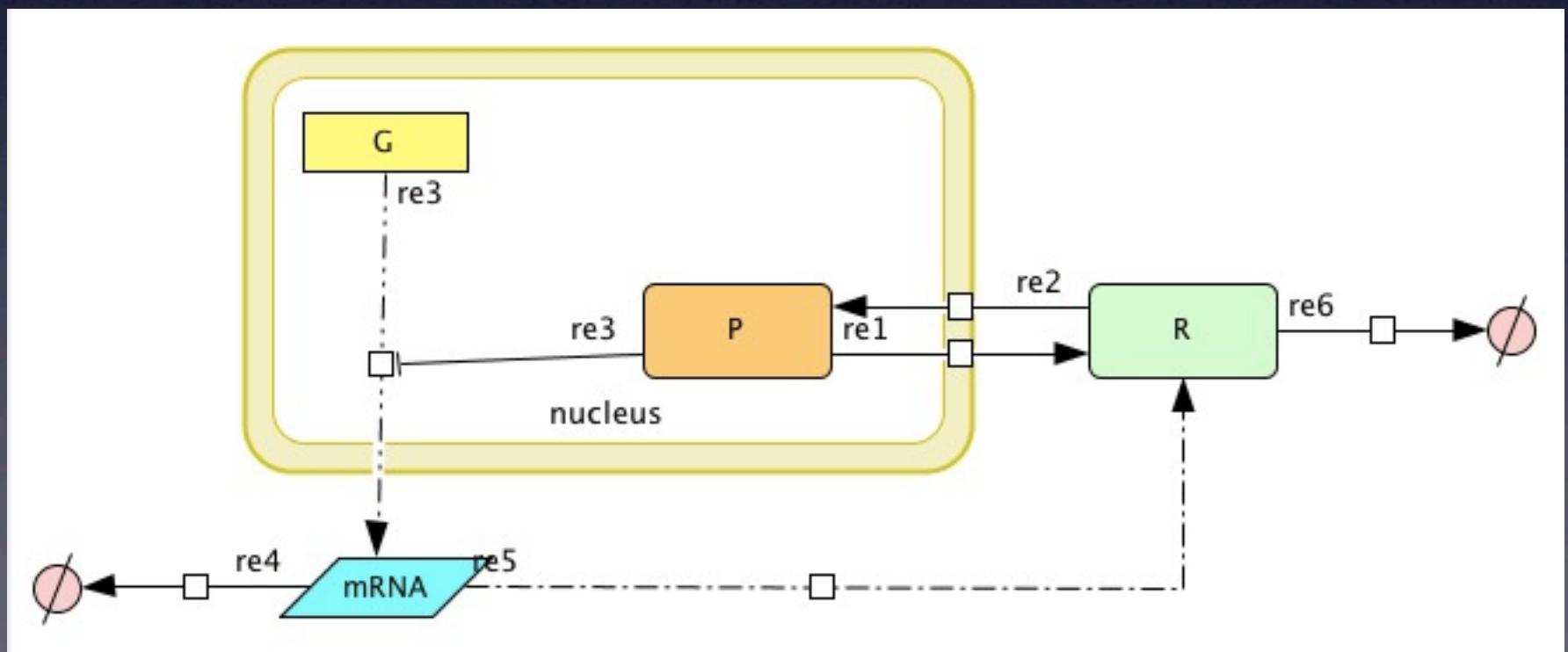
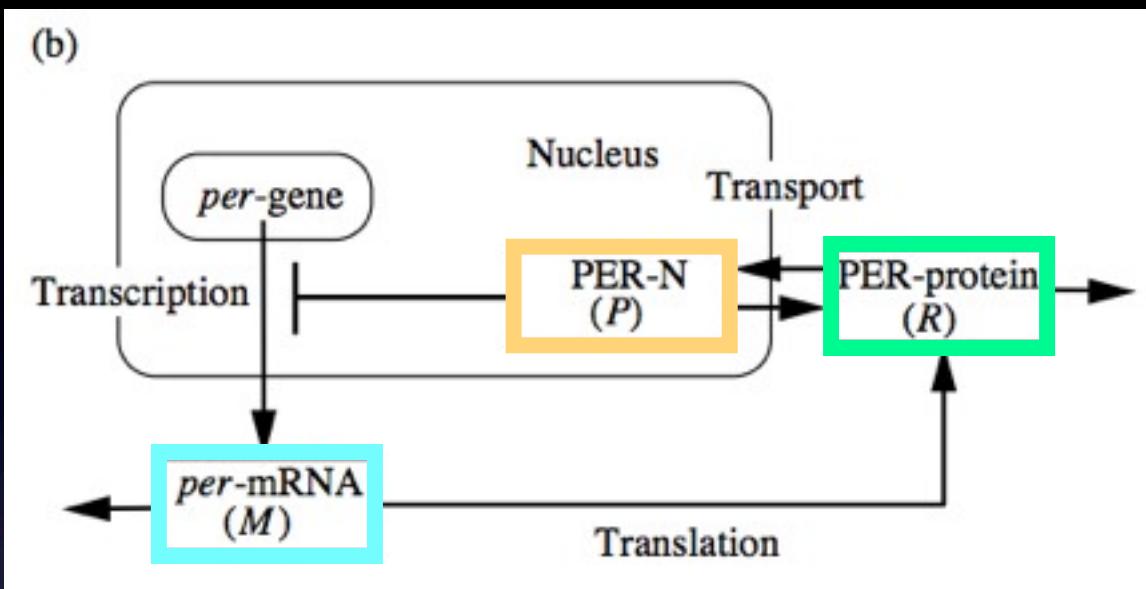
$$\frac{dM}{dt} = \frac{1}{1 + (P/h)^n} - aM - sM$$

$$\frac{dR}{dt} = sM - (d + u)R + vP$$

$$\frac{dP}{dt} = uR - vP$$



Circadian clock model



Circadian clock model

$$\frac{dM}{dt} = \frac{1}{1 + (P/h)^n} - aM - sM$$

$$a = s = d = v = 1.0$$

$$\frac{dR}{dt} = sM - (d + u)R + vP$$

$$u = 0.1$$

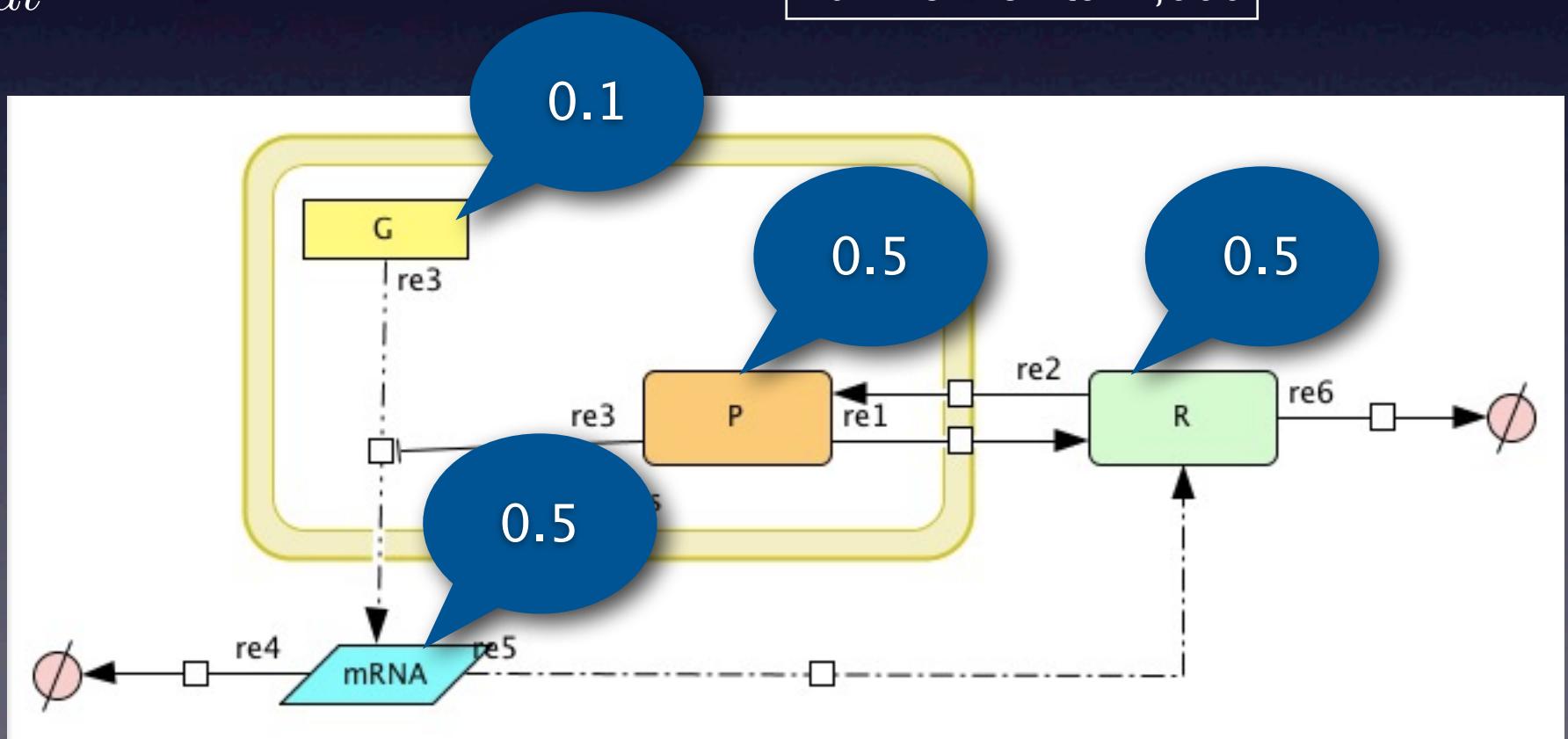
$$\frac{dP}{dt} = uR - vP$$

$$h = 0.01$$

$$x^n = \text{pow}(x, n)$$

$$n = 40$$

End Time: 50
Num. of Points: 1,000



Circadian clock model

$$\frac{dM}{dt} = \frac{1}{1 + (P/h)^n} - aM - sM$$

$$a = s = d = v = 1.0$$

$$\frac{dR}{dt} = sM - (d + u)R + vP$$

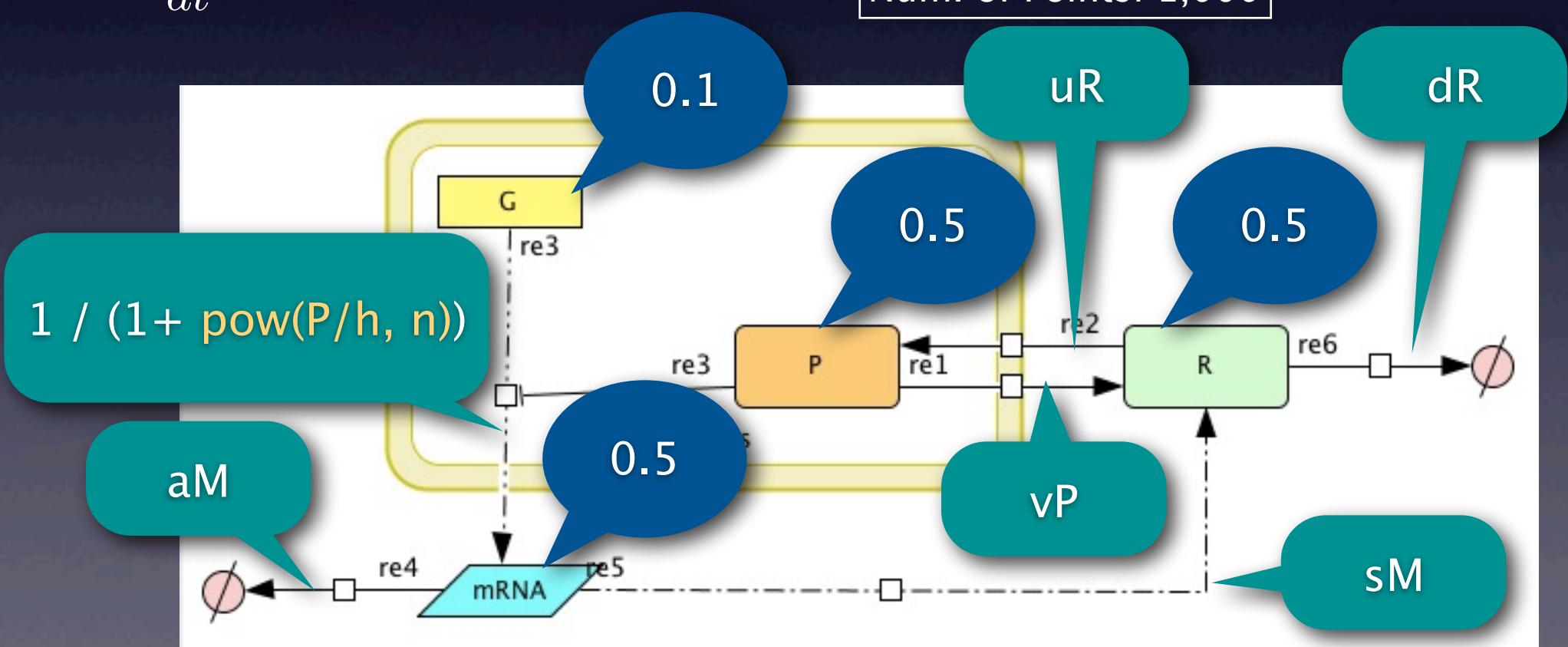
$$u = 0.1$$

$$\frac{dP}{dt} = uR - vP$$

$$h = 0.01$$

$$n = 40$$

End Time: 50
Num. of Points: 1,000



Boundary condition

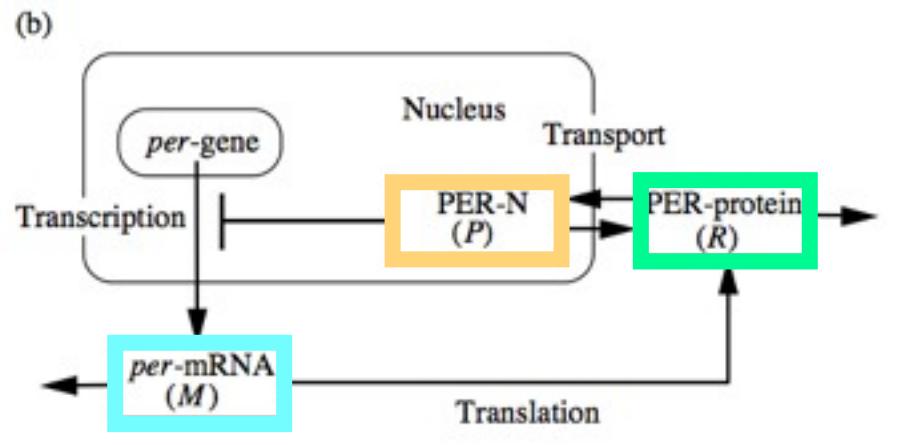


name	speciesType	compart...	positio...	included	quantit...	initialQuantity	sub...	hasO...	b.c.
G		c1	inside		Amount	0.1		true	true
mRNA		default	inside		Amount	0.5		true	false
P		c1	inside		Amount	0.5		true	false
R		default	inside		Amount	0.5		true	false
waste		default	inside		Amount	0.0		true	true
waste2		default	inside		Amount	0.0		true	true

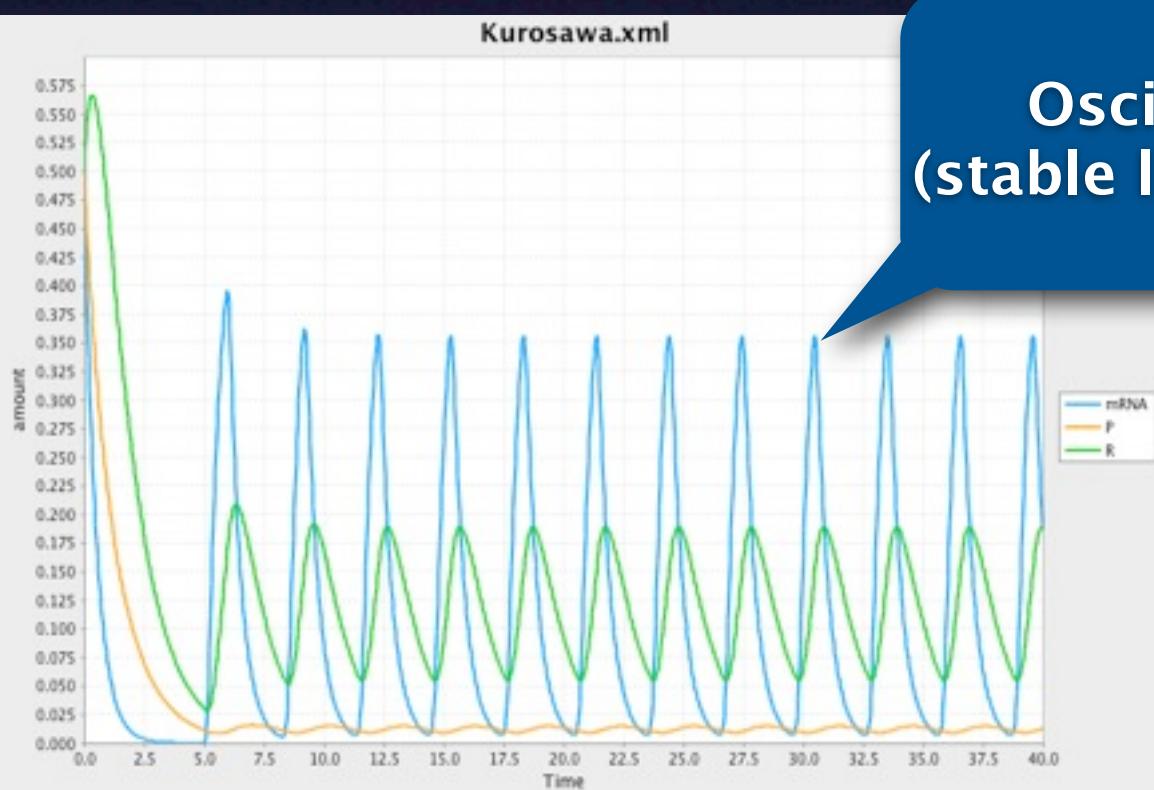
Species

id	s1
name	G
speciesType	<input type="button" value="▼"/>
compartment	c1
initial...	<input checked="" type="radio"/> Amount <input type="radio"/> Concentration 0.1
substanceUnits	<input type="button" value="▼"/>
hasOnlySubstanceUnits	<input checked="" type="radio"/> true <input type="radio"/> false
boundaryCondition	<input checked="" type="radio"/> true <input type="radio"/> false
constant	<input type="radio"/> true <input checked="" type="radio"/> false
	<input type="button" value="Update"/> <input type="button" value="Cancel"/>

Qualitative change by ‘n’



$$\frac{dM}{dt} = \frac{1}{1 + (P/h)^n} - aM - sM$$
$$\frac{dR}{dt} = sM - (d + u)R + vP$$
$$\frac{dP}{dt} = uR - vP$$



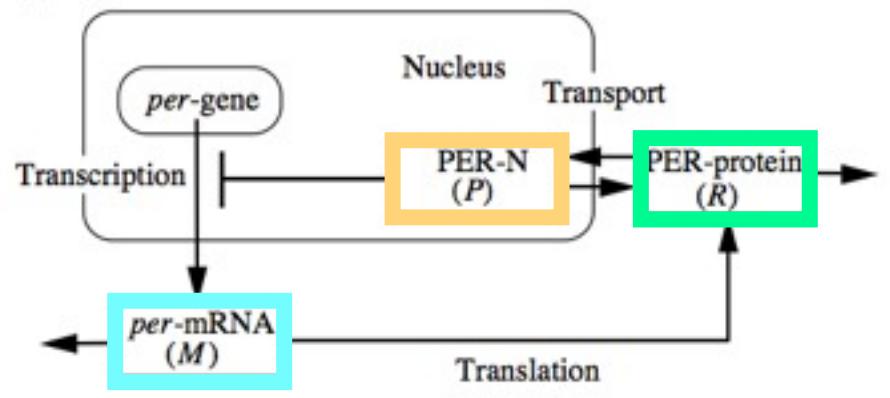
Oscillation
(stable limit cycle)

$n = 40$

End Time: 50
Num. of Points: 1,000

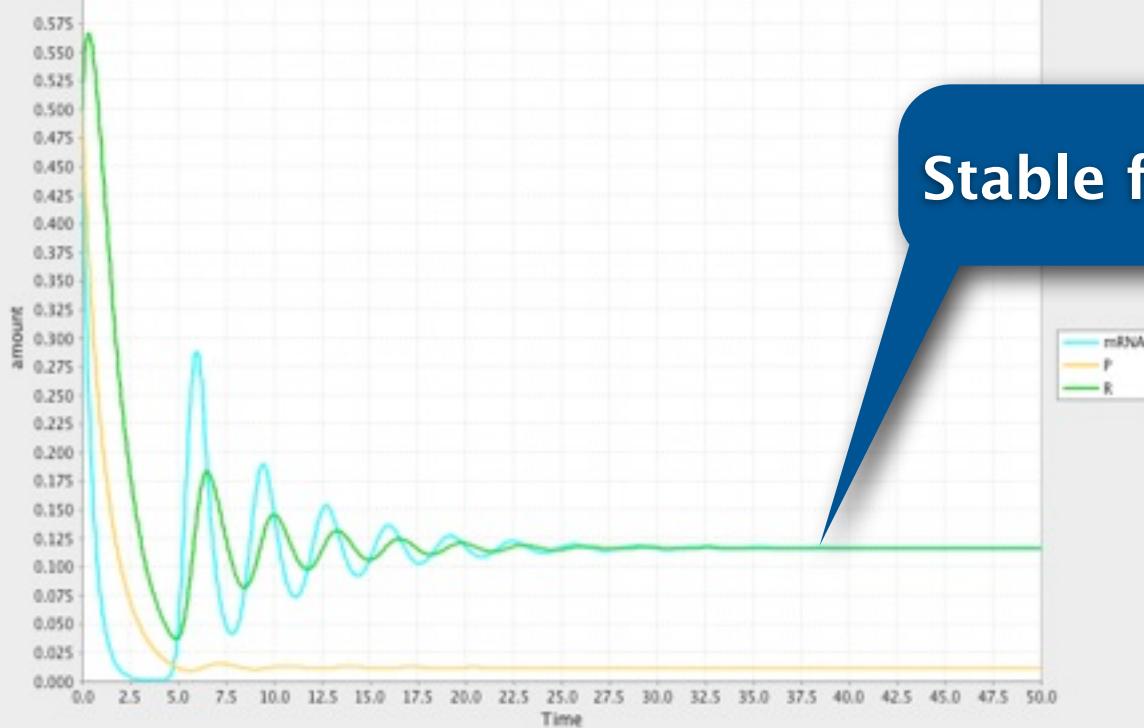
Qualitative change by ‘n’

(b)



$$\frac{dM}{dt} = \frac{1}{1 + (P/h)^n} - aM - sM$$
$$\frac{dR}{dt} = sM - (d + u)R + vP$$
$$\frac{dP}{dt} = uR - vP$$

Kurosawa.xml

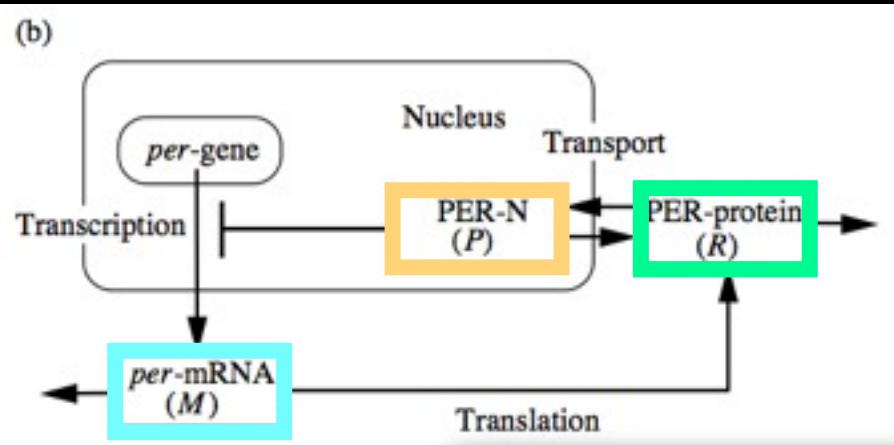


Stable fixed point

$n = 8$

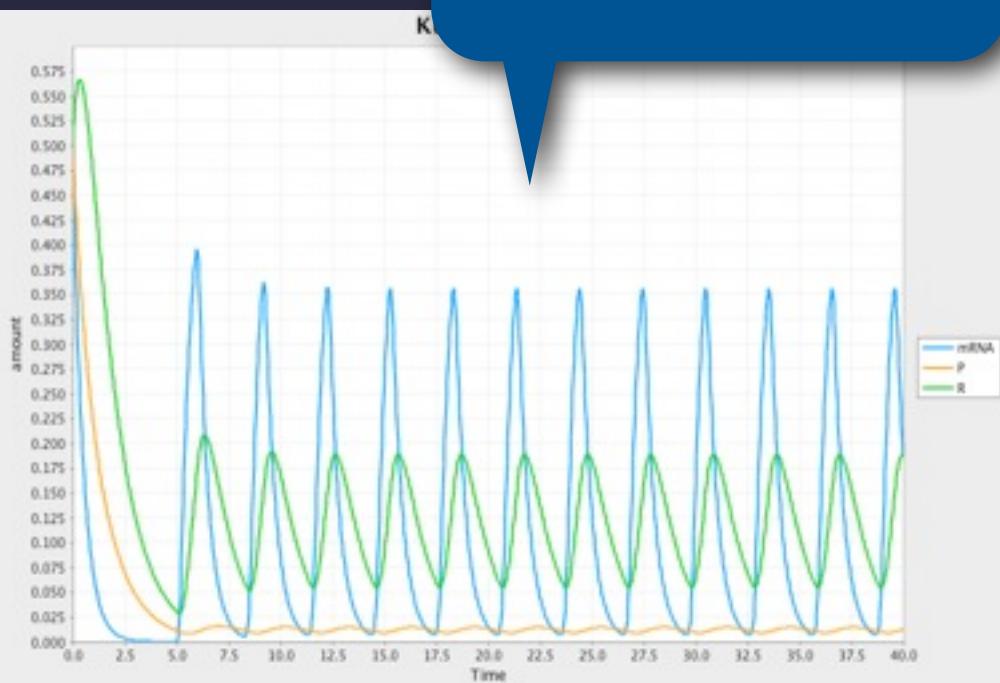
End Time: 50
Num. of Points: 1,000

Why we simulate a model?



$n = 40$

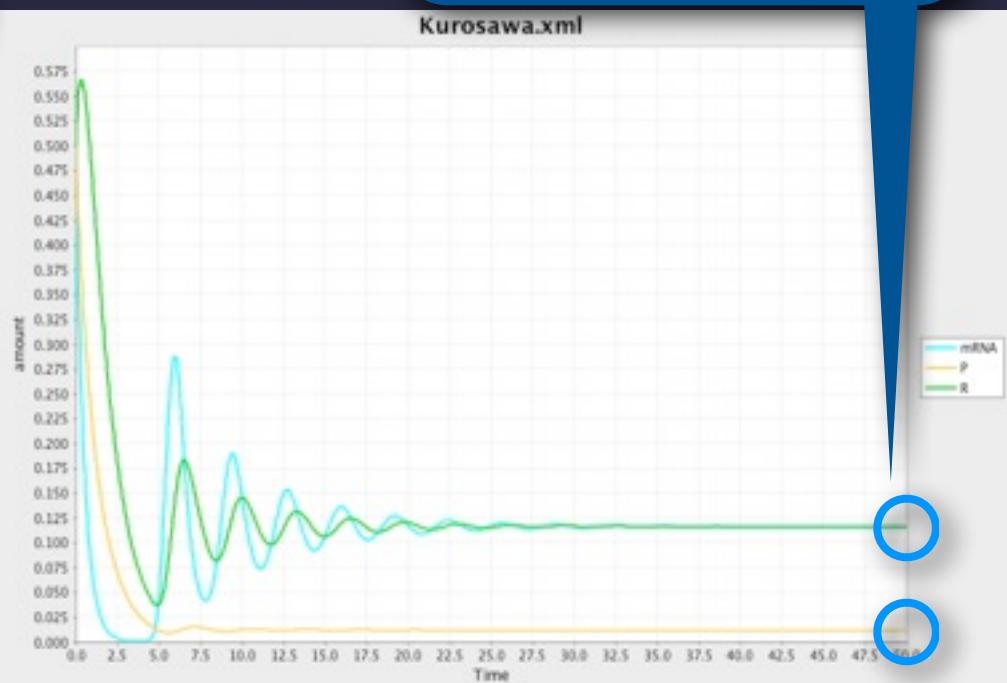
Oscillation
(stable limit cycle)



Mathematical model and
Quantitative evaluation
(Simulation) will reach a
Qualitative conclusion

$n = 8$

Stable fixed point



Database connection



Import model from BioModels.net

Screenshot of the BioModels Database interface:

The page title is "BioModels Database". The URL in the address bar is "http://www.ebi.ac.uk/compneur-srv/biomodels-main/publ-models.do". The page header includes the EMBL-EBI logo, a search bar, and links for "All Databases", "Enter Text Here", "Go", "Reset", "Advanced Search", "Give us Feedback", "Site Index", and "Help".

The left sidebar contains the following links:

- BioModels
- Curated Models
- Non-curated Models
- Search
- Simulate in JWS
- Submit Your Model
- Sign-in
- News
- Model of the Month
- Meetings
- Support
- Contact

The right sidebar features logos for SION-MODELS.NET, Computational Neurobiology, SBML, DOQCS, SBW, SBI, and JWS online.

The main content area is titled "Browse - Curated models". It contains the following text:

The following fields are used to describe a model:

- BioModels ID → A unique string of characters associated with the model, which will never be re-used even if the model is deleted from the BioModels Database.
- Name → The name of the model, as written in the model itself by its creator(s).
- Publication ID → The unique identifier of the reference publication describing the model, specified either as a PubMed identifier (linked to the EBI Medline database), or as a DOI (linked to the original publication through a DOI resolver), or as an URL. Being all published, all models must have one publication identifier, and the same identifier can be shared amongst several models if they have been described in the same publication.
- Last Modified → The date when the model was last modified.

To view a model, simply click on the correspondant BioModels ID provided within the leftmost column of the row corresponding to the model.

Navigation links: Next ↗ | Show All

BioModels ID	Name	Publication ID	Last Modified
BIOMD0000000001	Edelstein1996_EPSP_AChEvent	8863160	2007-09-23T23:24:19
BIOMD0000000002	Edelstein1996_EPSP_AChSpecies	8863160	2007-01-04T23:01:47
BIOMD0000000003	Goldbeter1991_MinMitOscil	1833774	2007-04-30T21:35:17
BIOMD0000000004	Goldbeter1991_MinMitOscil_Explain	1833774	2007-05-14T23:01:13
BIOMD0000000005	Tyson1991_CellCycle_6var	1831270	2007-05-15T18:24:25
BIOMD0000000006	Tyson1991_CellCycle_2var	1831270	2007-05-15T18:26:15
BIOMD0000000007	Novak1997_CellCycle	9256150	2007-05-15T18:32:05
BIOMD0000000008	Gardner1998_CellCycle_Goldbeter	9826676	2007-01-06T10:37:30
BIOMD0000000009	Huang1996_MAPK_ultrasens	8816754	2006-12-29T00:54:48
BIOMD0000000010	Kholodenko2000_MAPK_feedback	10712587	2007-01-10T10:35:07

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Database connection



Import model from BioModels.net

CellDesigner

File Edit Component View Database Layout Simulation Plugin Window

Import model from BioModels.net...

Connect to SGD
Connect to DBGET
Connect to iHOP
Connect to Genome Network
Connect to PubMed
Connect to Entrez Gene

Model
● Compartments
● Species
● Reactions

BioModels.net

ID	Name
BIO MD000000000001	Edelstein1996_EPSP_AChEvent
BIO MD000000000002	Edelstein1996_EPSP_AChSpecies
BIO MD000000000003	Goldbeter1991_MinMitOscil
BIO MD000000000004	Goldbeter1991_MinMitOscil_ExplInact
BIO MD000000000005	Tyson1991_CellCycle_6var
BIO MD000000000006	Tyson1991_CellCycle_2var
BIO MD000000000007	Novak1997_CellCycle
BIO MD000000000008	Gardner1998_CellCycle_Goldbeter
09	Huang1996_MAPK_ultrasens
10	Kholodenko2000_MAPK_feedback
11	Levchenko2000_MAPK_noScaffold
12	Elowitz2000_Repressilator
13	Poolman2004_CalvinCycle
14	Levchenko2000_MAPK_Scaffold
15	Curto1998_purineMetabol
16	Goldbeter1995_CircClock
17	Hoefnagel2002_PyruvateBranches
18	Morrison1989_FolateCycle
19	hodgkin-huxley squid-axon 1952
20	Leloup1999_CircClock
21	Ueda2001_CircClock
22	Rohwer2001_Sucrose
23	Scheper1999_CircClock
24	Smolen2002_CircClock
25	Markevich2004_MAPK_orderedElementary
26	

Description Reference Import Cancel

```
graph TD; subgraph Pathway [MAPK Signaling Pathway]; J1[MAPKKK-P] --> J2[MAPKK-P]; J2 --> J3[MAPKK-P]; J3 --> J4[MAPKK-PP]; J4 --> J5[MAPK-P]; J5 --> J6[MAPK-P]; J6 --> J7[MAPK-PP]; J7 --> J8[MAPK-PP]; J8 --> J9[MAPK-P]; J9 --> J10[MAPKKK-P]; J10 --> J1; end; J1 --> J2; J2 --> J3; J3 --> J4; J4 --> J5; J5 --> J6; J6 --> J7; J7 --> J8; J8 --> J9; J9 --> J10; J10 --> J1; J2 -.-> J5; J3 -.-> J4; J4 -.-> J7; J7 -.-> J9; J9 -.-> J10;
```

SABIO-RK

- Web-accessible database
- <http://sabio.h-its.org/>
- Contains information about biochemical reactions, related kinetic equations and parameters

The screenshot shows the SABIO-RK web interface. At the top, there's a navigation bar with links for CONTACT, HELP, and IMPRINT. Below it is a search bar and a "Search Results" section. The search results table has columns for Reactions, Select Reaction(s) (with a "(De)Select All" button), Kinetic Data for this reaction (with a "Check to View" link), Enzyme ECIF, and Kinetic data for enzymes (with a "Check to View" link). There are two rows of data in the table.

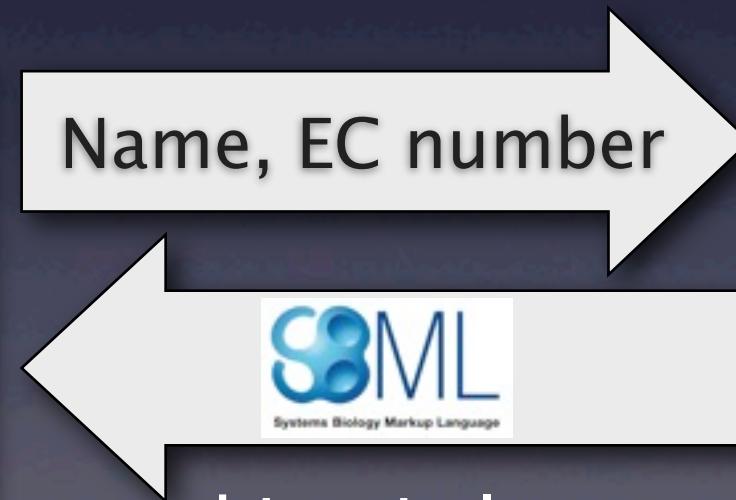
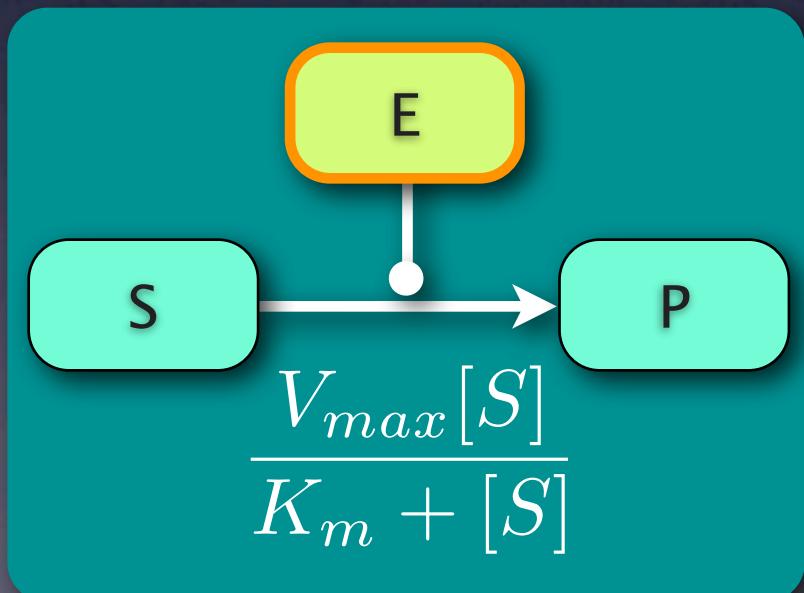
Reactions	Select Reaction(s) (De)Select All	Kinetic Data for this reaction Check to View	Enzyme ECIF	Kinetic data for enzymes Check to View
D-Glucose + ATP <=> D-Glucose-6-phosphate EAL1	<input type="checkbox"/>	View	2.7.3.1 2.7.3.2	View
ATP + Glucose-1<> ADP + Glucose-6 GNGT65	<input type="checkbox"/>	View	2.7.3.1 2.7.3.2	View



CellDesigner ↔ SABIO RK

- Users can import additional information to each object (reaction) on-the-fly
- SBML (Systems Biology Markup Language) is used to exchange information

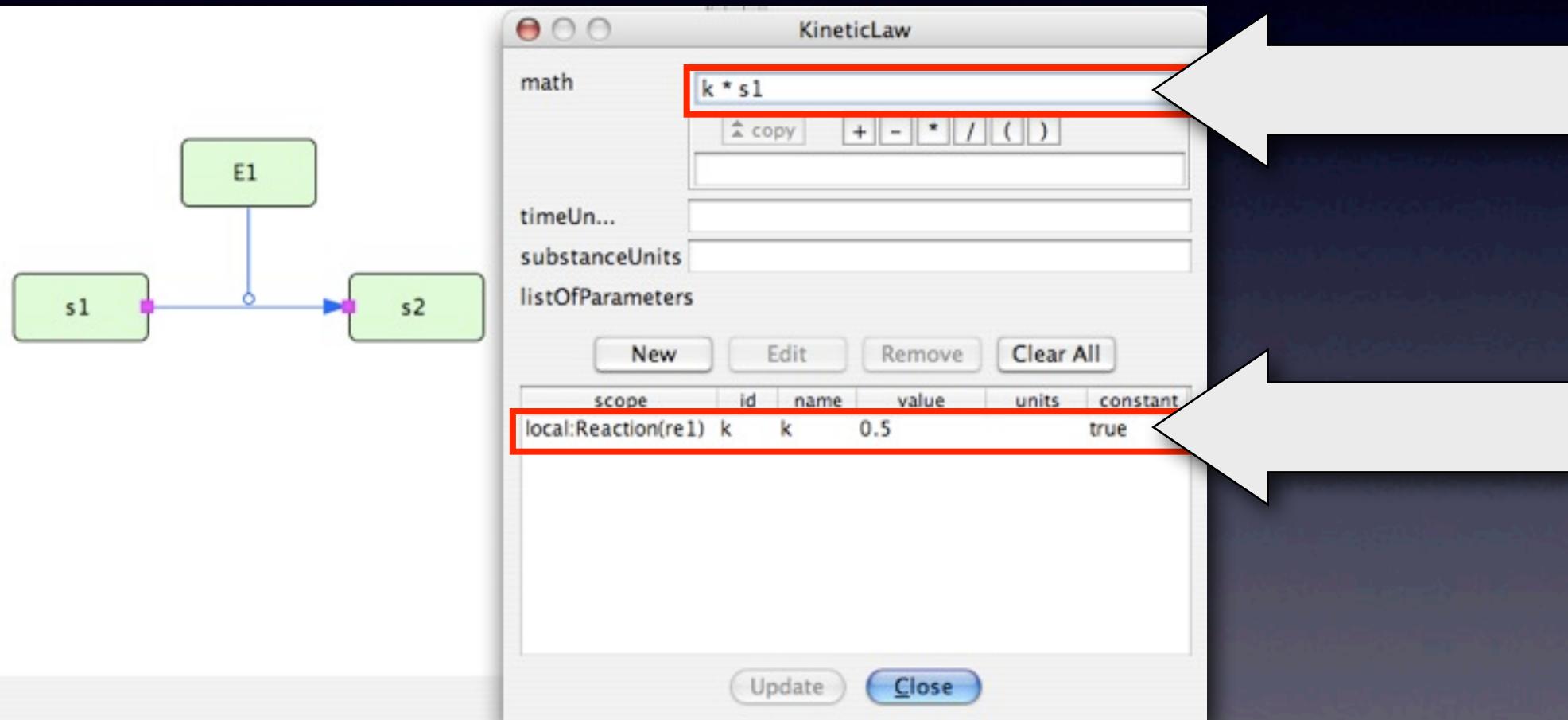
CellDesigner



kinetic law, parameters,
function / unit definitions

Integration

- Import kinetic law, parameters to the model from SABIO-RK



Annotating a model

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Keio University, Japan
6th Aug. 2017



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IGF signaling pathway

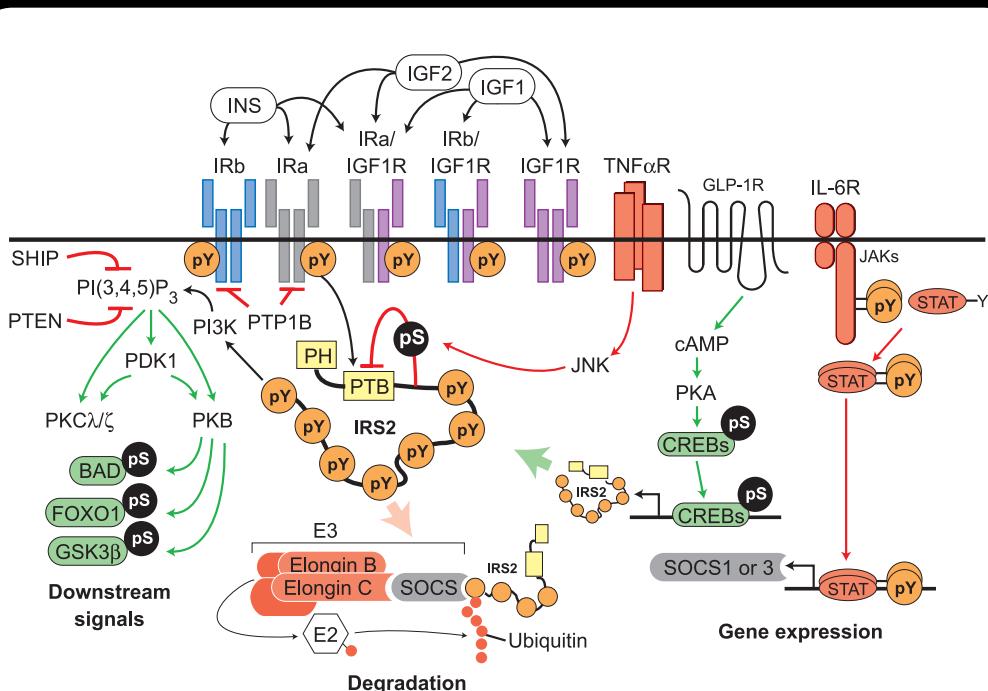
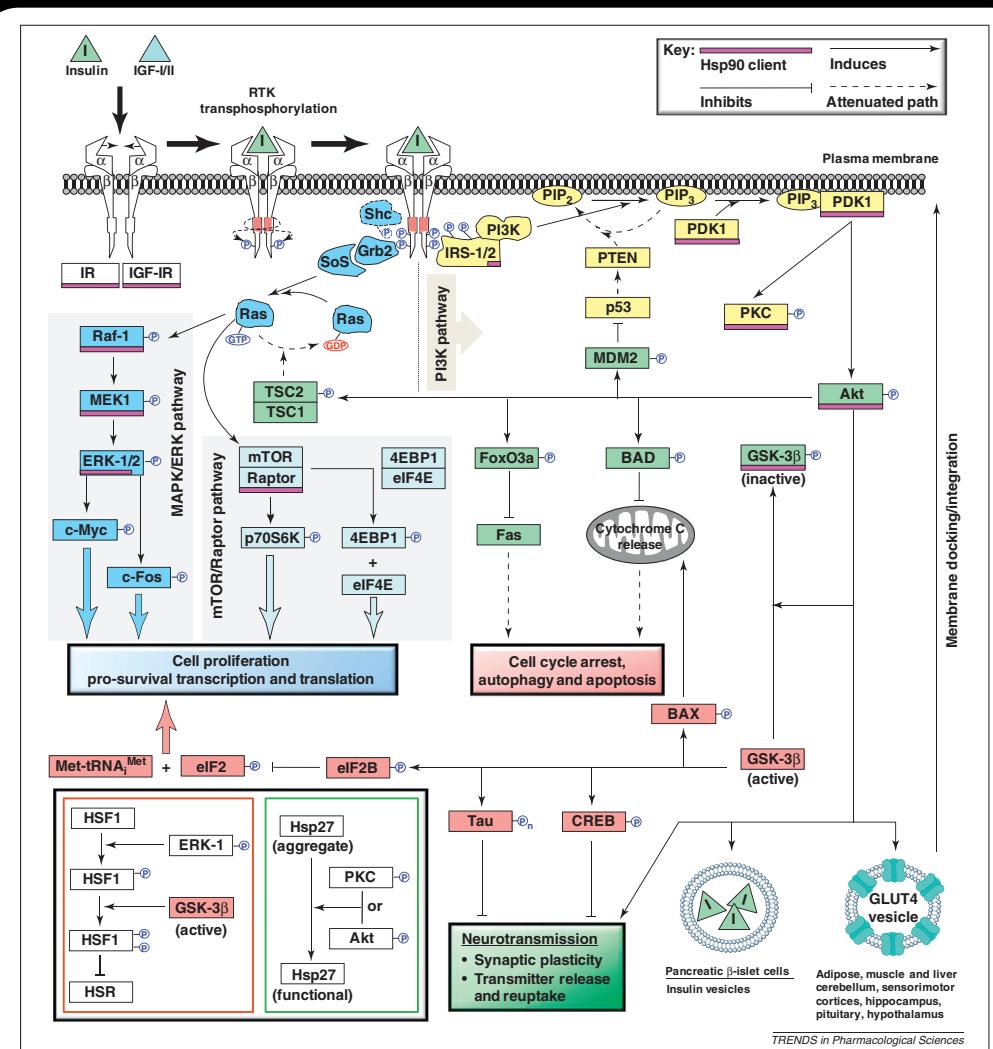


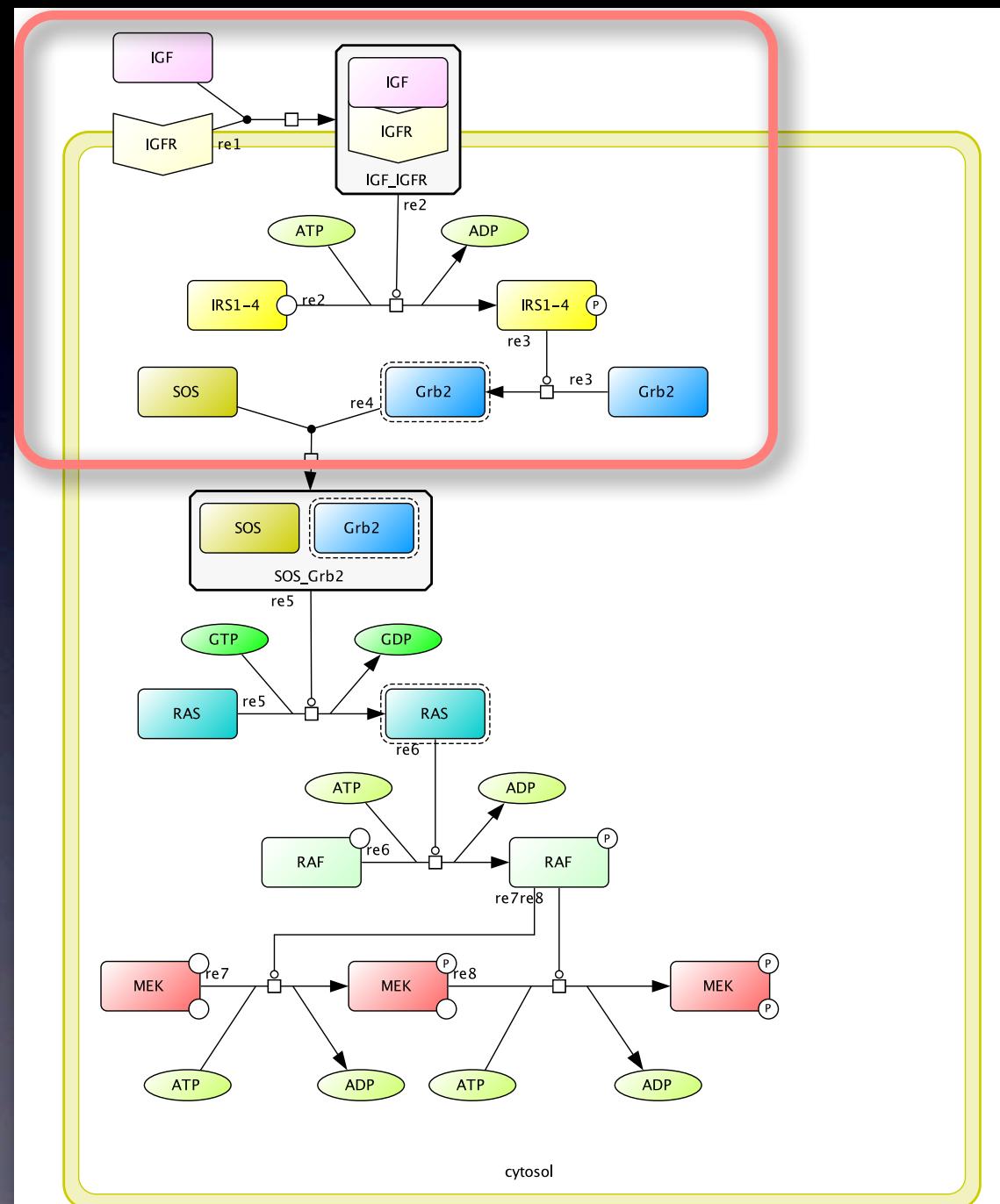
Fig. 1. Regulation of insulin and IGF signaling. Insulin and IGF1 receptors form hybrids that modulate the selectivity and affinity for insulin and insulin-like growth factors (IGF1 and IGF2). Insulin or IGF binding stimulates tyrosine autophosphorylation in the receptor β subunits, which activates the kinase and recruits cellular substrates—IRS1 and IRS2—for tyrosine phosphorylation. Recruitment is regulated by serine phosphorylation of the IRS proteins, which inhibits the interaction between its PTB domain and the phosphorylated receptor. Proinflammatory cytokines increase the synthesis of SOCS1 or SOCS3, which promote ubiquitination and degradation of IRS1 and IRS2. Production of cAMP enhances expression of IRS2 through the activity of phosphorylated CREB. Tyrosine phosphorylation of IRS1 or IRS2 recruits and activates various SH2 domain-containing proteins, including the PI 3-kinase, which activates the PKB cascade. Abbreviations: pY, phosphotyrosine; pS, phosphoserine; PKC λ/ζ , protein kinase C λ or ζ ; E2, ubiquitin conjugating enzymes; TNF α R, tumor necrosis factor- α receptor; GLP-1R, glucagon-like peptide-1 receptor; IL6R, interleukin-6 receptor; for other abbreviations, see the text.



Insulin Signaling in Health and Disease
Science 302 (5651), 2003, 1710.

Heat shock response and insulin-associated neurodegeneration
Trends in Pharmacological Sciences, 33(3), 2012, 129–137

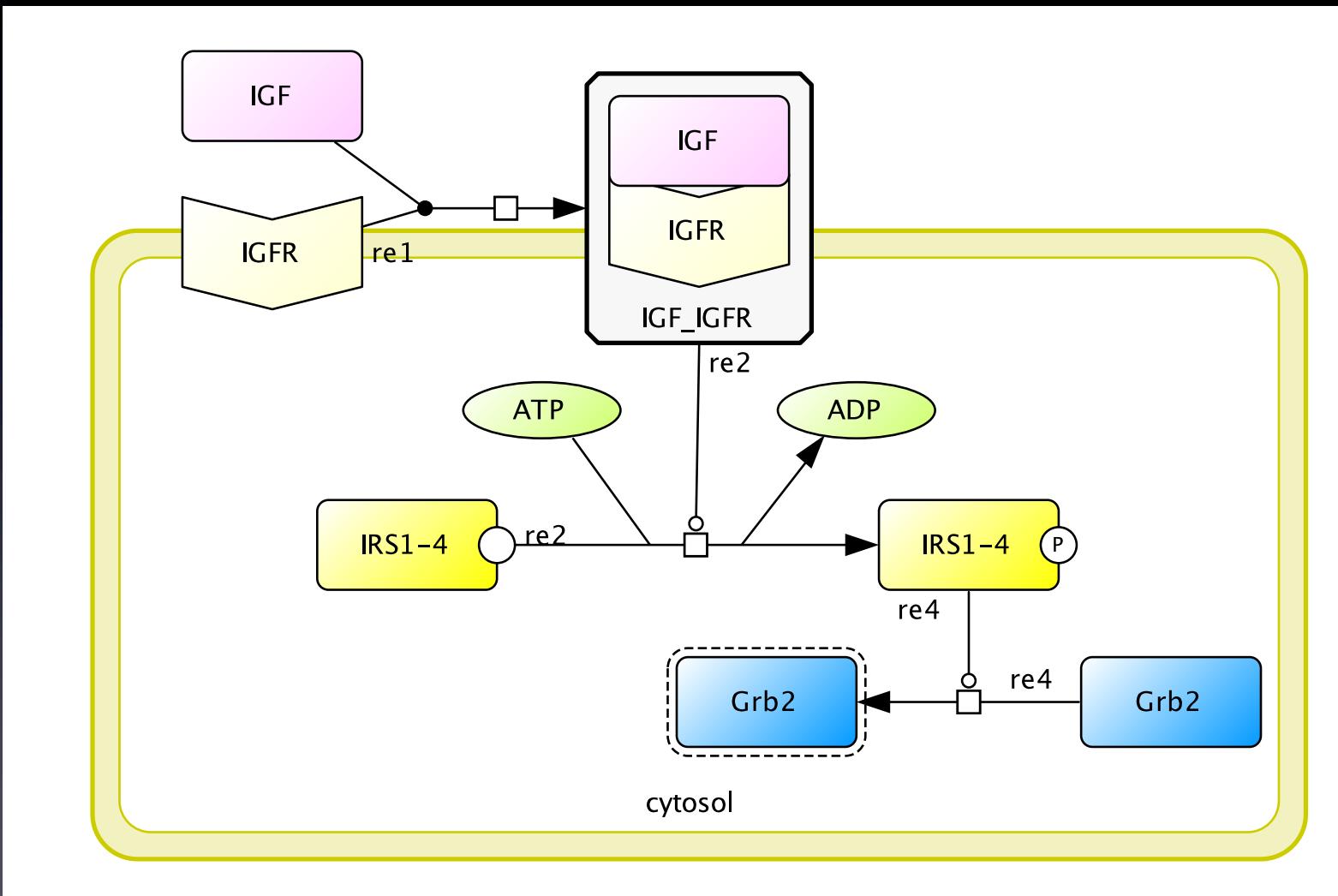
IGF signaling pathway



<http://www.sbgn.org/>
Documents/PD_L1_Examples

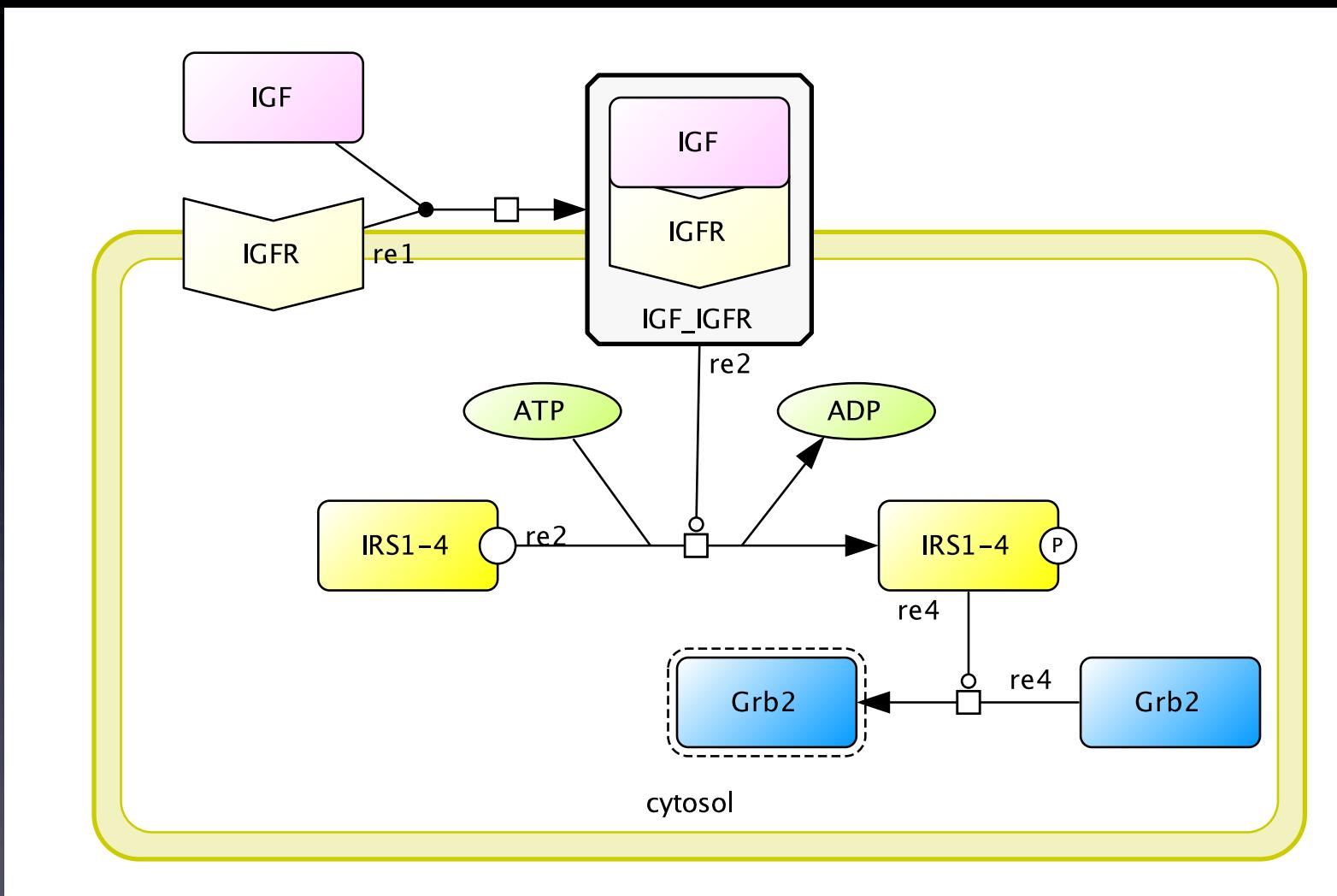
Exercise

- Create a following model on CellDesigner



Exercise

- Search Database from CellDesigner

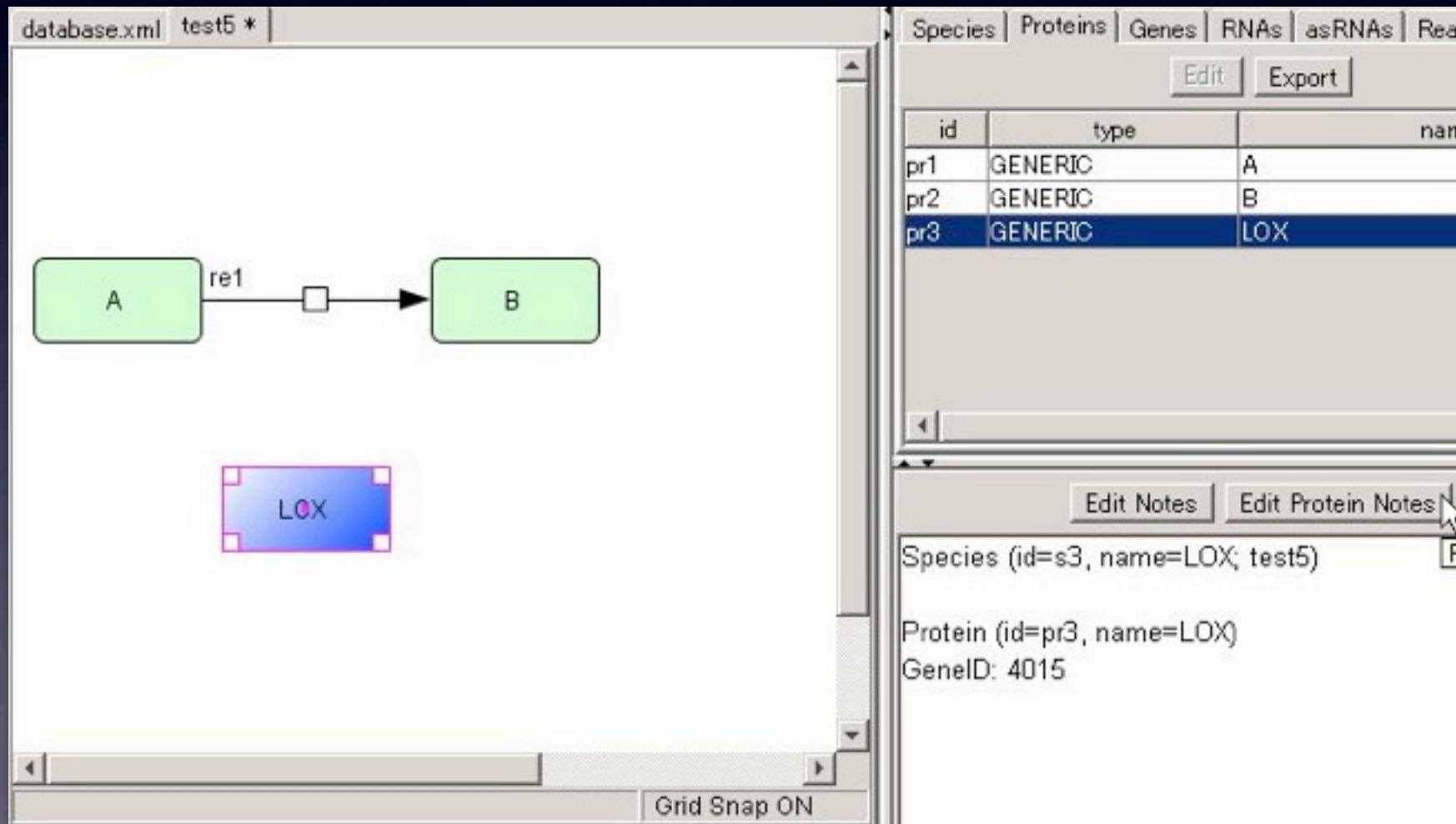


Database Connection



Search Database by Notes:

- PubMed: PMID: 123456
- Entrez Gene: GenelD: 4015

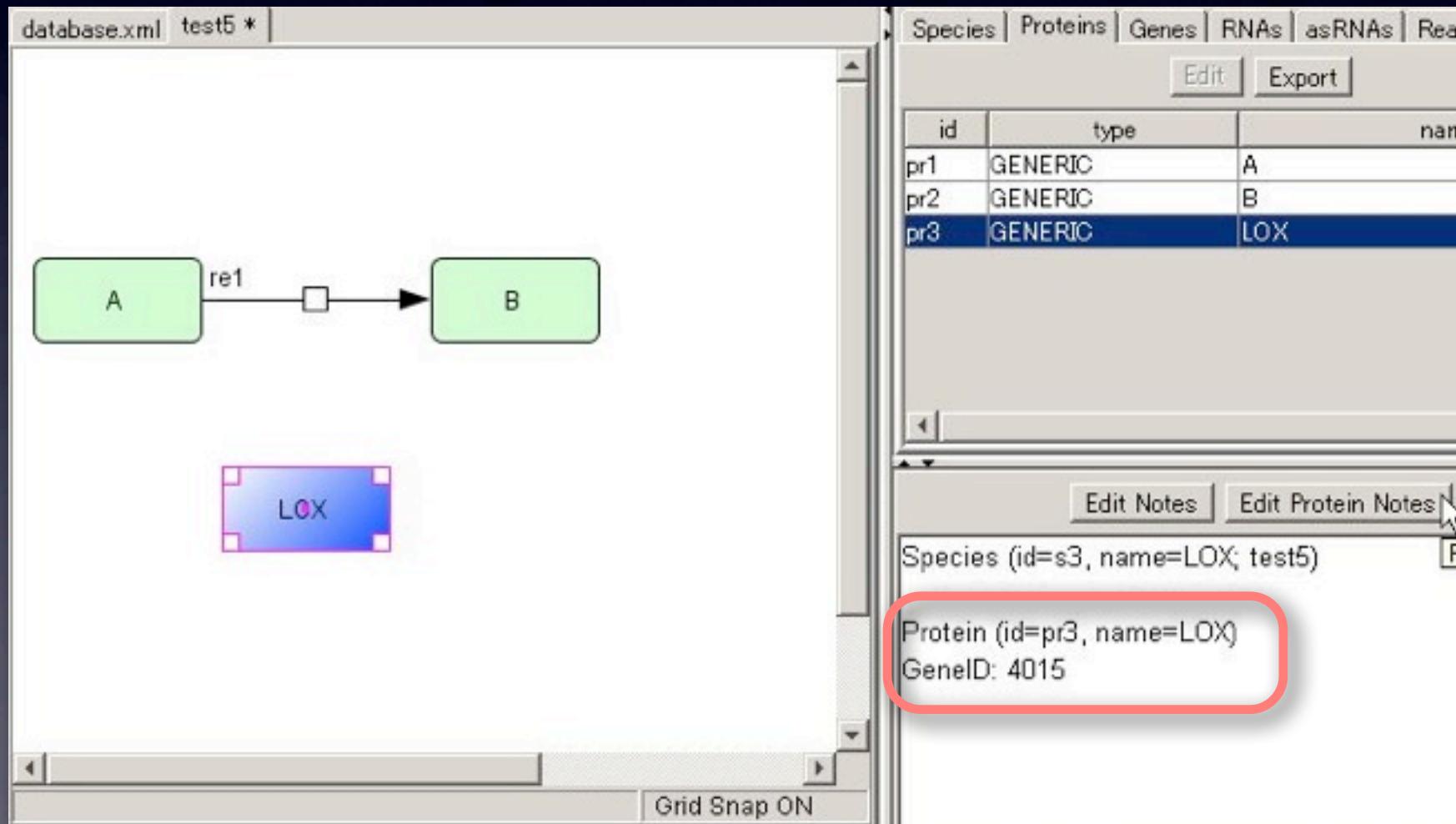


Database Connection



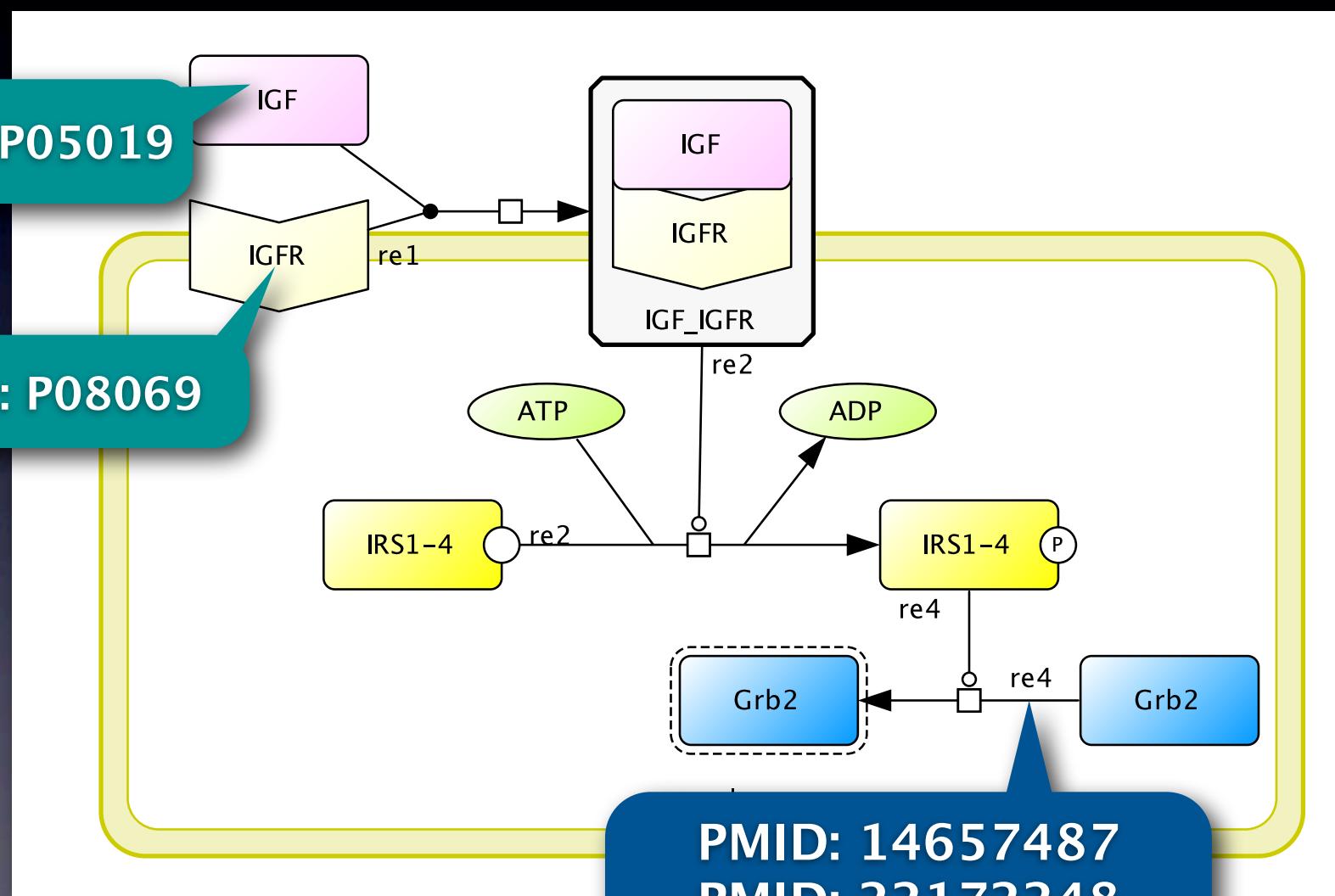
Search Database by Notes:

- PubMed: PMID: 123456
- Entrez Gene: GenID: 4015



Exercise

- Add UniProt ID for Proteins, PubMed ID for reactions and call “Connect to UniProt”



MIRIAM annotation

_computational
BIOLOGY

PERSPECTIVE

Minimum information requested in the annotation of biochemical models (MIRIAM)

Nicolas Le Novère^{1,15}, Andrew Finney^{2,15}, Michael Hucka³, Upinder S Bhalla⁴, Fabien Campagne⁵, Julio Collado-Vides⁶, Edmund J Crampin⁷, Matt Halstead⁷, Edda Klipp⁸, Pedro Mendes⁹, Poul Nielsen⁷, Herbert Sauro¹⁰, Bruce Shapiro¹¹, Jacky L Snoep¹², Hugh D Spence¹³ & Barry L Wanner¹⁴

Most of the published quantitative models in biology are lost for the community because they are either not made available or they are insufficiently characterized to allow them to be reused. The lack of a standard description, lack of stringent reviewing and authors' carelessness are the main causes for incomplete model descriptions. With today's increased interest in detailed biochemical models, it is necessary to define a minimum quality standard for the encoding of those models. We propose a set of rules for curating quantitative models of biological systems. These rules define procedures for encoding and annotating models represented in machine-readable form. We believe their application will enable users to (i) have confidence that curated models are an accurate reflection of their reference descriptions, (ii) search collections of curated models with precision, (iii) quickly identify the biological phenomena that a given curated model or model constituent represents and (iv) facilitate model reuse and composition into large subcellular models.

Box 1 Glossary

Some terms are used in a very specific way throughout the article. We provide here a precise definition of each one.

Quantitative biochemical model. A formal model of a biological system, based on the mathematical description of its molecular and cellular components, and the interactions between those components.

Encoded model. A mathematical model written in a formal machine-readable language, such that it can be systematically parsed and employed by simulation and analysis software without further human translation.

MIRIAM-compliant model. A model that passes all the tests and fulfills all the conditions listed in MIRIAM.

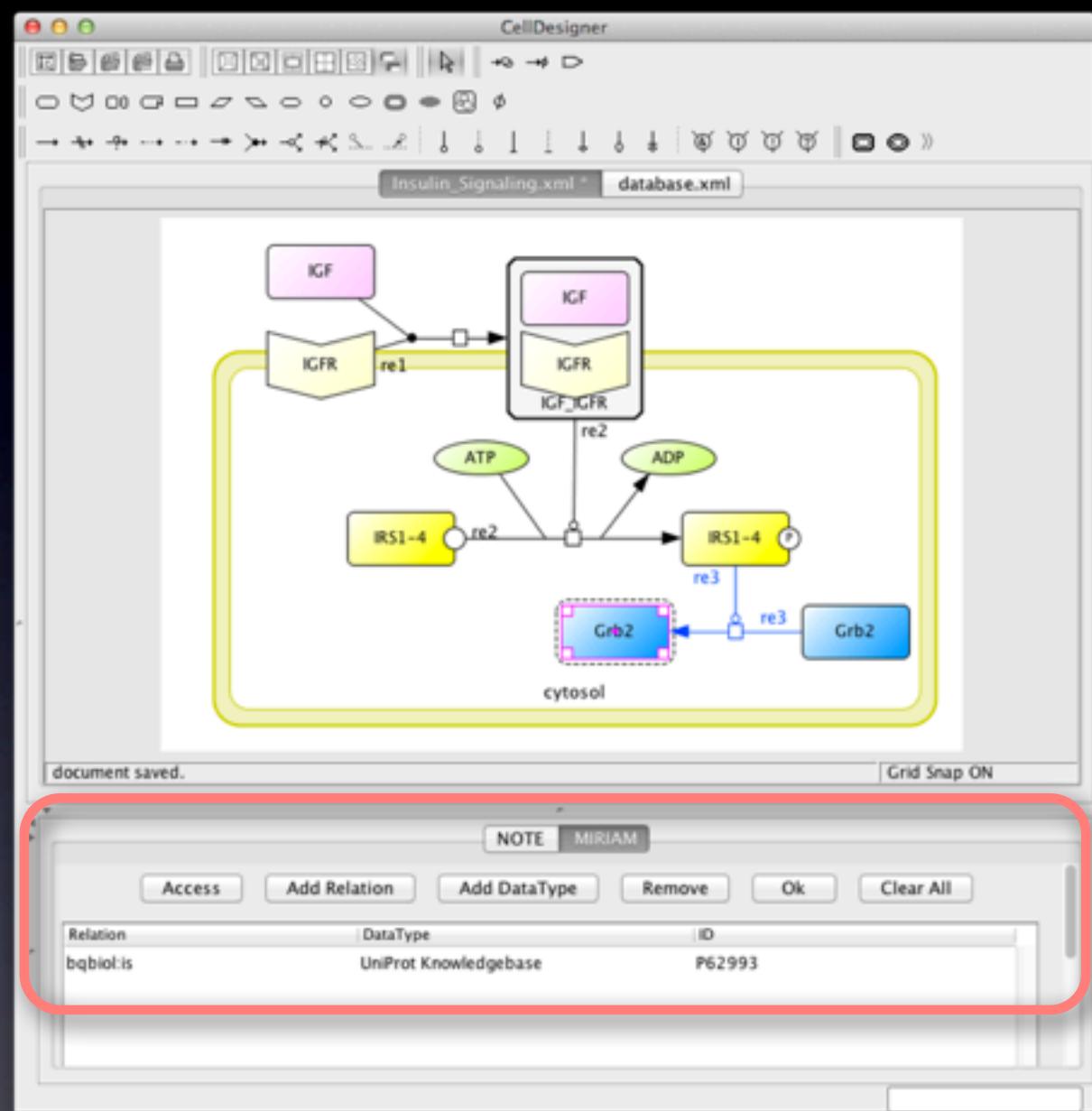
Reference description. A unique document that describes, or references the description of the model, the structure of the model, the numerical values necessary to instantiate a simulation from the model, or to perform a mathematical analysis of the model, and the results one expects from such a simulation or analysis.

Curation process. The process by which the compliance of an encoded model with MIRIAM is achieved and/or verified. The curation process may encompass some or all of the following tasks: encoding of the model, verification of the reference correspondence and annotation of the model.

Reference correspondence. The fact that the structure of a model and the results of a simulation or an analysis match the information present in the reference description.

Published online 6 December 2005; doi:10.1038/nbt1156

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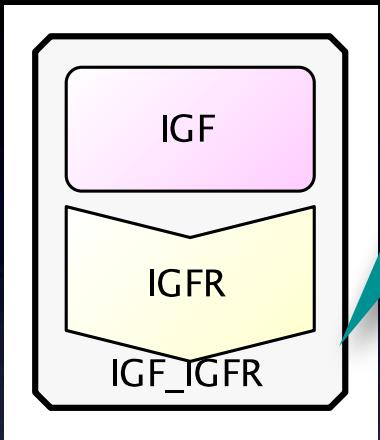
Minimum information requested in the annotation of biochemical models (MIRIAM)

Nature Biotechnology 23, 1509 – 1515 (2005)

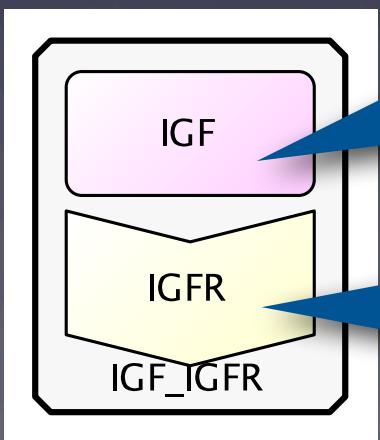
MIRIAM annotation

• <http://www.ebi.ac.uk/miriam/main/qualifiers/>

- is
- hasPart
- isPartOf
- hasVersion (isoform)
- isVersionOf (superclass, parent)



IGF_IGFR hasPart: IGF
IGF_IGFR hasPart: IGFR

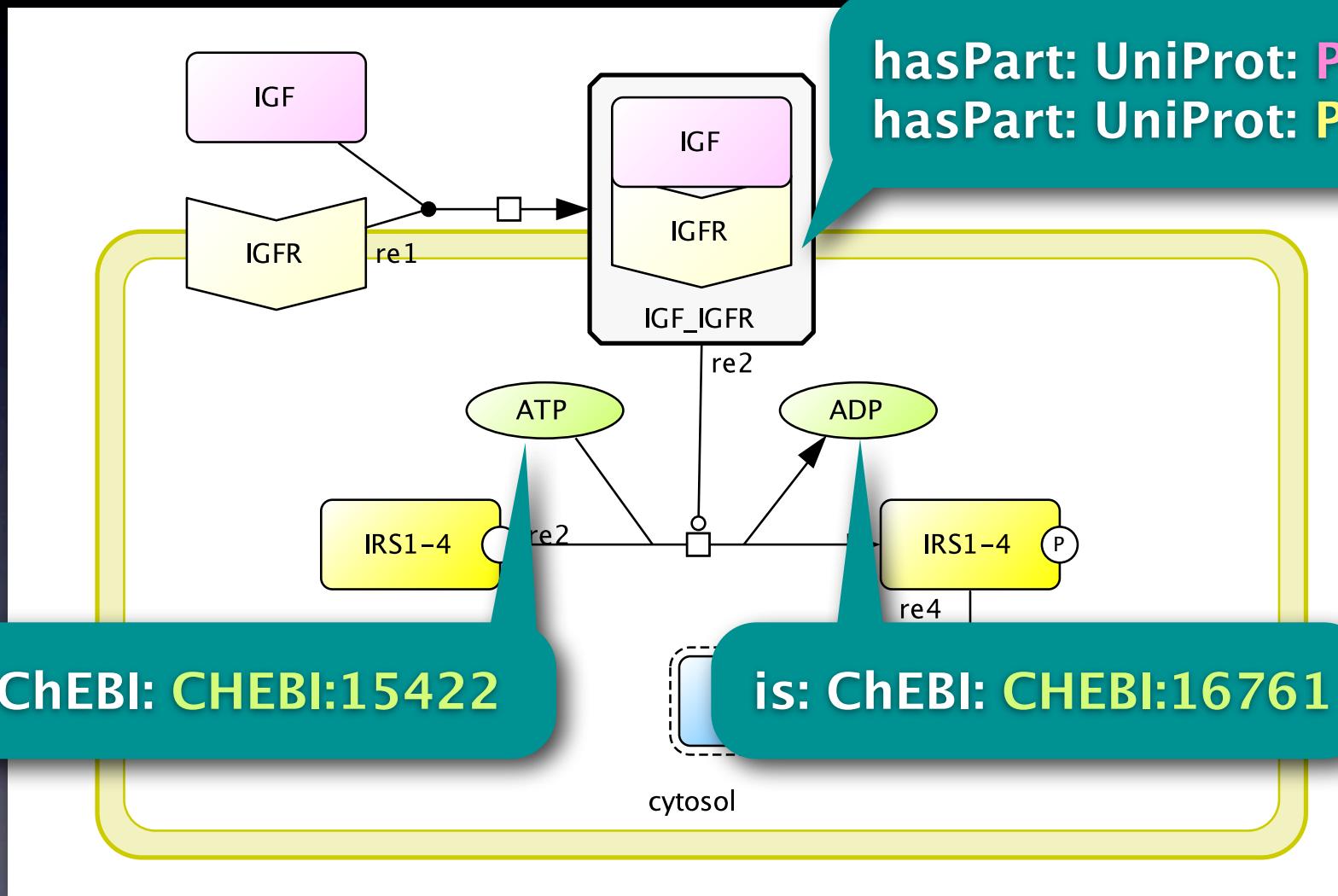


IGF isPartOf: IGF_IGFR

IGFR isPartOf: IGF_IGFR

Exercise

- Add MIRIAM annotation



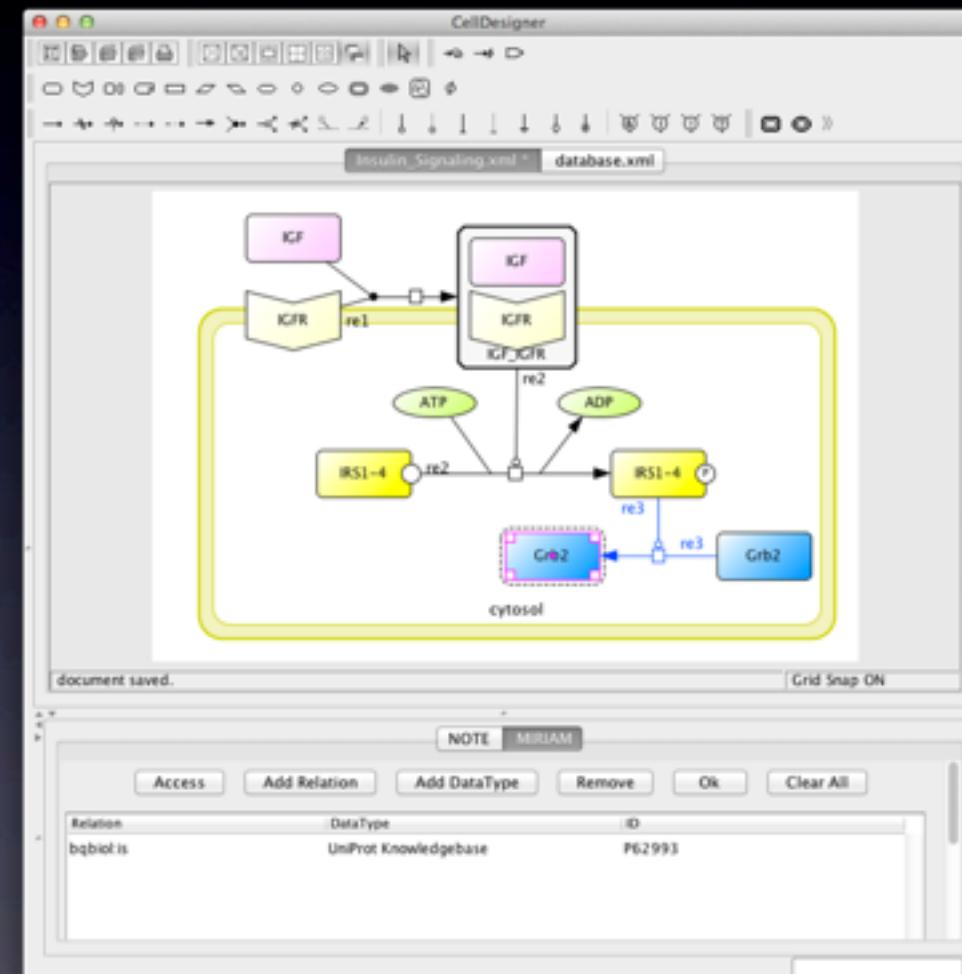
Notes or MIRIAM?

CellDesigner Notes

- Easy to add (text)

MIRIAM

- Tool neutral (SBML)
- Precise annotation



Summary

- Introduction of CellDesigner
- What kind of model you can build
 - Mathematical model
 - Pathway map
- How to build a model with CellDesigner
 - From scratch
 - Import a model from BioModels.net
 - Import kinetic law and parameters from SABIO-RK