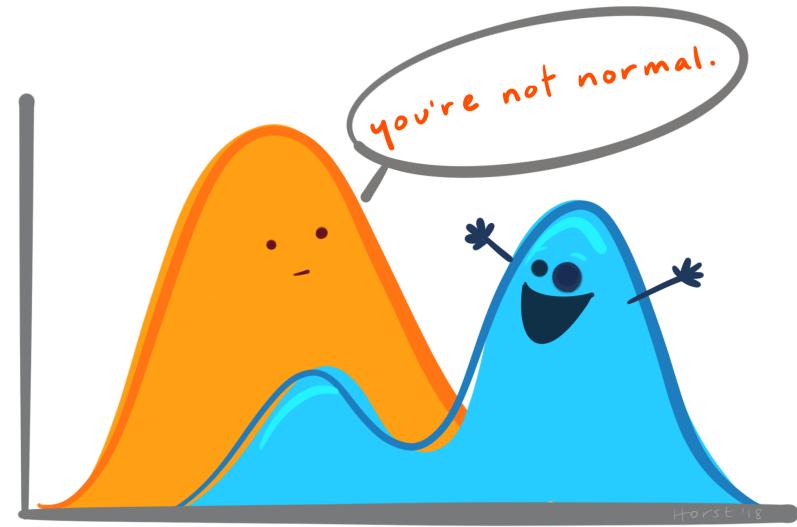


Visualizing biological data

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Did you get the dataset
“gene_expr_pasilla_results.csv” and the
“VizBiological.Rmd” file?



ggmarginal

```
# library(ggExtra)

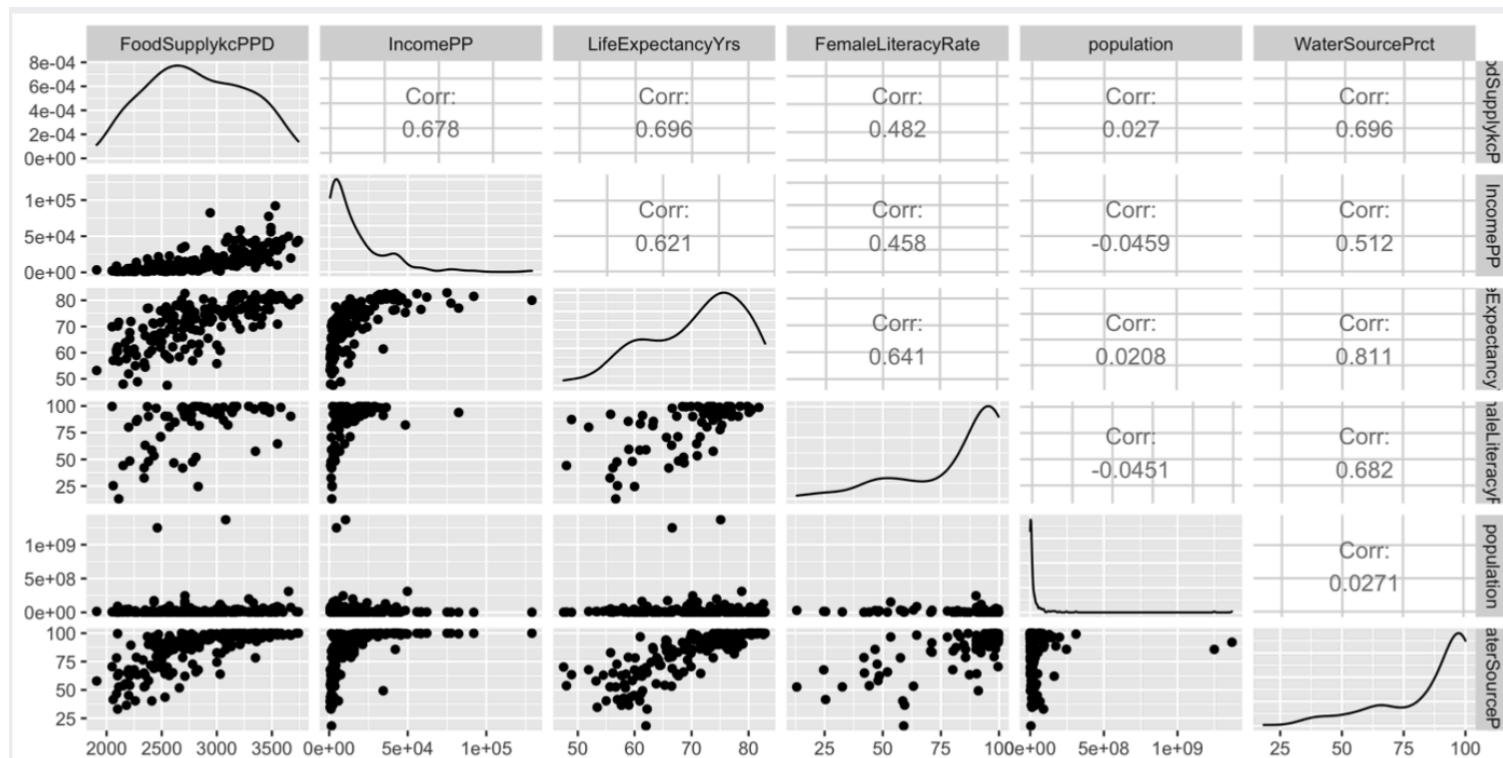
p <- ggplot(data = gapminder2011,
             aes(x = FoodSupplykcPPD,
                  y = LifeExpectancyYrs,
                  color = four_regions)
            ) +
  geom_point(alpha = .4) +
  scale_color_discrete(
    name = "Regions",
    labels = c("Africa", "Americas",
              "Asia", "Europe")
  ) +
  theme(legend.position="bottom") +
  labs(
    x = "Daily Food Supply PP (kc)",
    y = "Life Expectancy (years)",
    title = "Scatterplot"
  )
```

```
ggMarginal(p,
            type = "density",
            margins = "both",
            groupColour = TRUE,
            groupFill = TRUE
          )
```



GGally::ggpairs()

```
# library(GGally)
gapminder2011 %>%
  select(FoodSupplykcPPD:WaterSourcePrct) %>% # specifying which columns to use
  ggpairs()
```



<https://ggobi.github.io/ggally/index.html>

ggtree ()

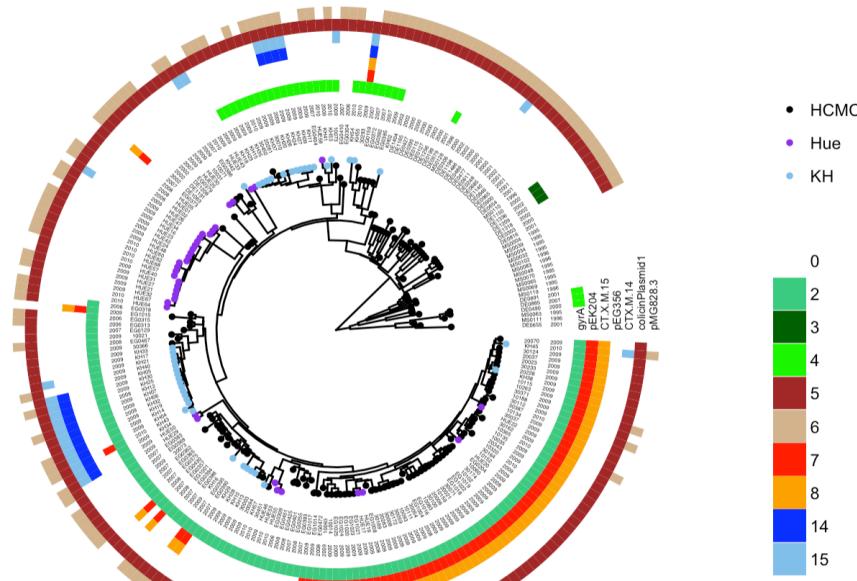


Figure 4.17: **Example of annotating a tree with diverse associated data.** Circle symbols are colored by strain sampling location. Taxa names and sampling years are aligned to the tips. Curated gene information were visualized as a heatmap (colored boxes on the outer circles).

<https://guangchuangyu.github.io/ggtree-book/chapter-ggtree.html>

Genomic data with ggbio

```
library("ggbio")
data("hg19IdeogramCyto", package = "biovizBase")
plotIdeogram(hg19IdeogramCyto, subchr = "chr1")
```



```
library("GenomicRanges")
data("darned_hg19_subset500", package = "biovizBase")
autoplot(darned_hg19_subset500, layout = "karyogram",
         aes(color = exReg, fill = exReg))
```

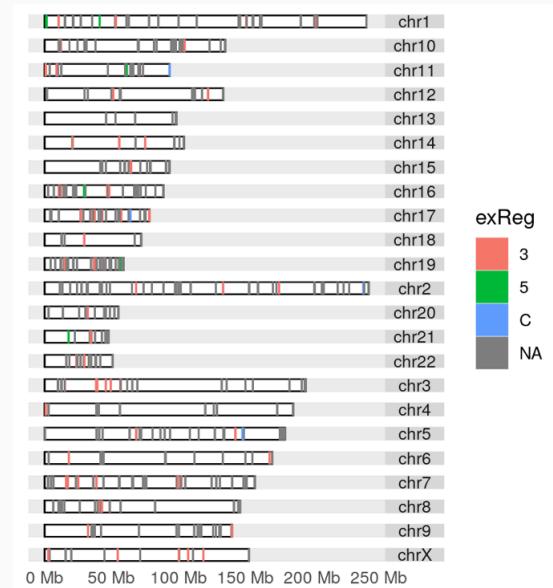


Figure 3.46: Karyogram with RNA editing sites.
exReg indicates whether a site is in the coding
region (C), 3'- or 5'-UTR.

<http://127.0.0.1:12631/library/ggbio/doc/ggbio.pdf>

<https://web.stanford.edu/class/bios221/book/Chap-Graphics.html>

Genomic data with ggbio

```
data("ideoCyto", package = "biovizBase")
dn = darned_hg19_subset500
seqlengths(dn) = seqlengths(ideoCyto$hg19)[names(seqlengths(dn))]
dn = keepSeqlevels(dn, paste0("chr", c(1:22, "X")))
autoplot(dn, layout = "karyogram", aes(color = exReg, fill = exReg))
```

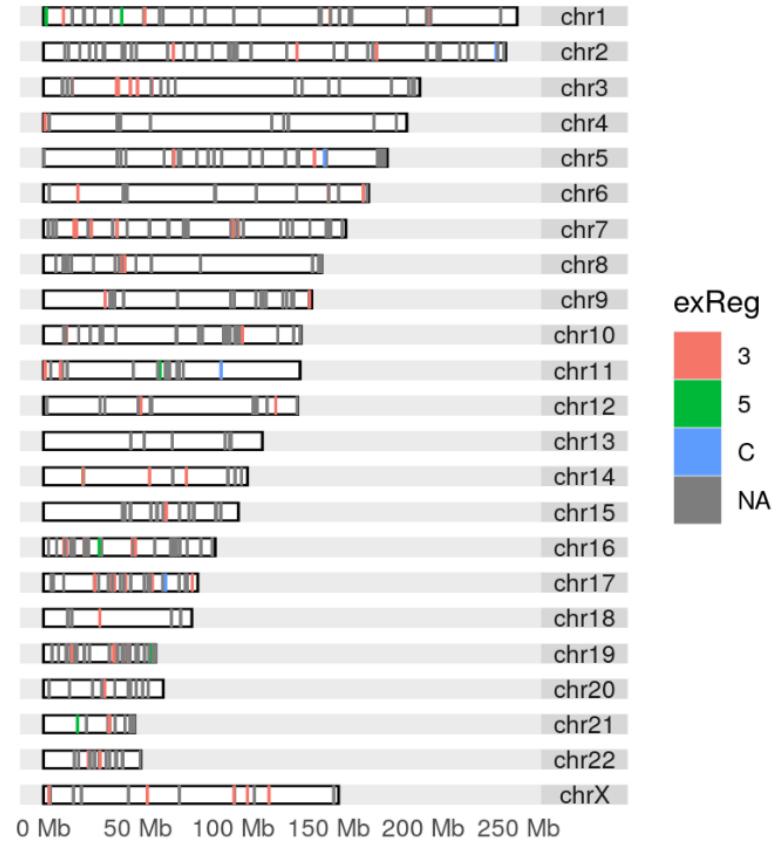


Figure 3.47: Improved version of Figure 3.46.

<http://127.0.0.1:12631/library/ggbio/doc/ggbio.pdf>

<https://web.stanford.edu/class/bios221/book/Chap-Graphics.html>

Gene expression

```
pasilla_data <-  
read_csv("../data/gene_expr_pasilla_results.csv")
```

Gene expression

Pasilla data

```
glimpse(pasilla_data)
```

Rows: 8,377

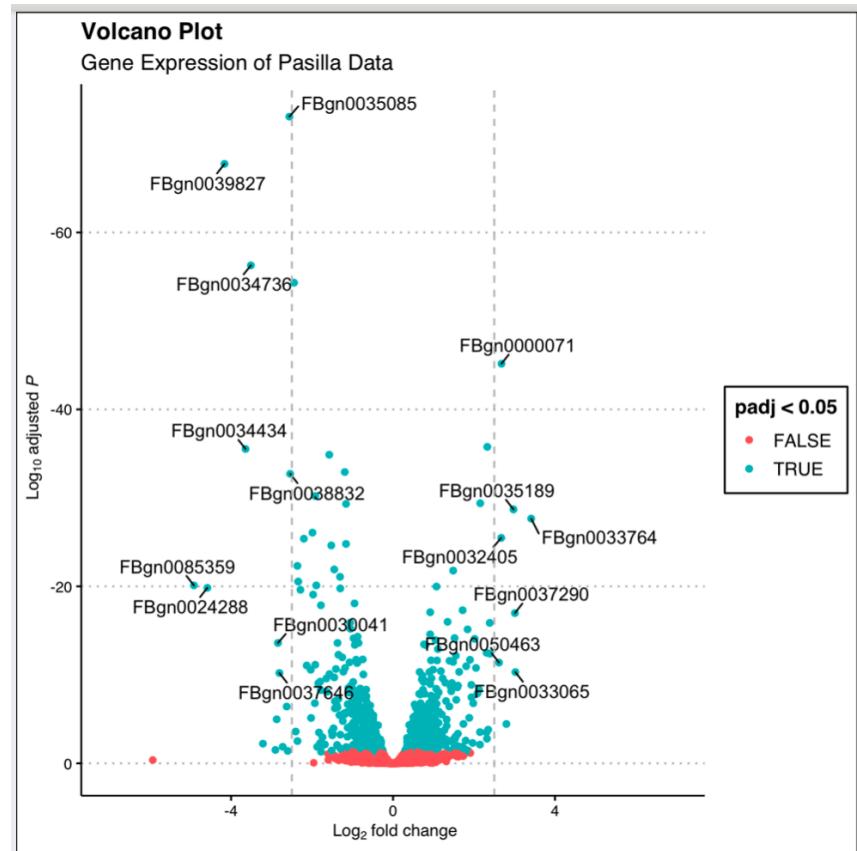
Columns: 15

```
$ gene          <chr> "FBgn0000008", "FBgn0000017", "FBgn00000...
$ baseMean      <dbl> 95.144292, 4352.553569, 418.610484, 6.40...
$ fc            <dbl> 1.0015792, 0.8467929, 0.9300151, 1.15736...
$ log2FoldChange <dbl> 0.002276441, -0.239918944, -0.104673912, ...
$ lfcSE         <dbl> 0.2237287, 0.1263369, 0.1484891, 0.68958...
$ stat          <dbl> 0.01017501, -1.89904084, -0.70492676, 0....
$ pvalue        <dbl> 9.918817e-01, 5.755911e-02, 4.808558e-01...
$ padj          <dbl> 9.972108e-01, 2.880017e-01, 8.268337e-01...
$ treated1      <dbl> 7.607917, 11.938311, 9.143372, 6.479135, ...
$ treated2      <dbl> 7.834912, 12.024557, 9.011505, 6.577240, ...
$ treated3      <dbl> 7.595052, 12.013565, 8.944883, 6.475226, ...
$ untreated1    <dbl> 7.567298, 12.045721, 9.315269, 6.565256, ...
$ untreated2    <dbl> 7.642174, 12.284647, 9.098290, 6.479802, ...
$ untreated3    <dbl> 7.844603, 12.455939, 8.966546, 6.422196, ...
$ untreated4    <dbl> 7.669147, 12.077404, 9.066286, 6.395509, ...
```

```

ggplot(data = pasilla_data,
       aes(x = log2FoldChange,
           y = log10(padj))) +
  geom_point() +
  scale_y_reverse() +
  aes(color = padj < 0.05) +
  ggrepel::geom_text_repel(
    data = pasilla_data_top,
    aes(label = gene), color = "black",
    box.padding = 0.5,
    min.segment.length = 0) +
  xlim(c(-7, 7)) +
  geom_vline(xintercept = c(-2.5, 2.5),
             lty = "dashed", color="grey") +
  ggthemes::theme_clean() +
  labs(
    x = bquote(~Log[2]~ "fold change"),
    y = bquote(~Log[10]~adjusted~italic(P)),
    title = "Volcano Plot",
    subtitle="Gene Expression of Pasilla Data"
)

```



Heatmap with pheatmap::pheatmap()

```
# select expression data
pasilla_heat <- pasilla_data %>%
  select(treated1:untreated4)
# subtract off gene-specific means
pasilla_heat <- pasilla_heat - rowMeans(pasilla_heat)
# calculate standard deviation of each centered gene
sd_gene <- apply(pasilla_heat, 1, sd)
# select top 500 most variable
pasilla_heat <-
  pasilla_heat[order(sd_gene, decreasing = TRUE)[1:500],]

# create annotation data
pasilla_col <- data.frame(
  trt = factor(c(rep("trt", 3), rep("untrt", 4))),
  id = 1:7,
  row.names=colnames(pasilla_heat))
```

```
head(pasilla_heat, n = 3)
```

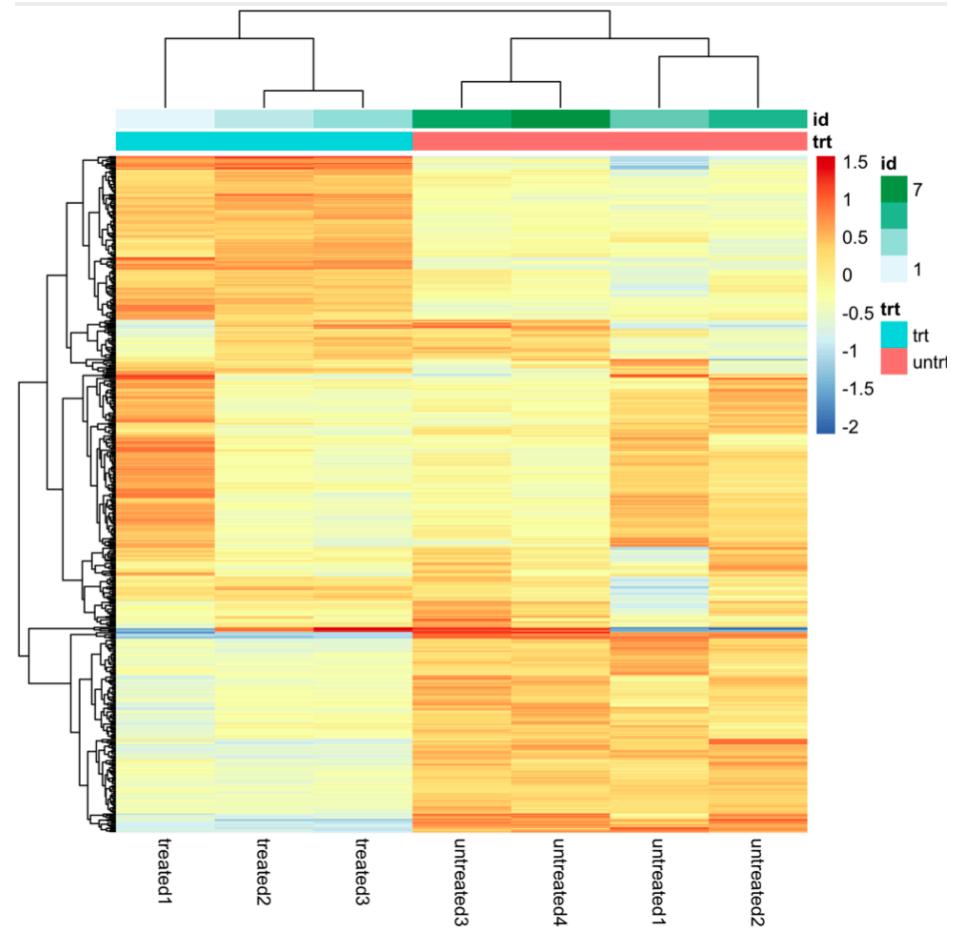
	treated1	treated2	treated3	unt
2390	-1.5997691	0.8713581	1.568570	-1
521	-1.3218267	0.9954861	1.278523	-1
7886	-0.5901012	0.8225366	1.339219	-1
	untreated3	untreated4		
2390	1.338488	1.4253512		
521	1.040472	0.9541077		
7886	1.155933	0.7369965		

```
pasilla_col
```

	trt	id
treated1	trt	1
treated2	trt	2
treated3	trt	3
untreated1	untrt	4

Heatmap with pheatmap::pheatmap()

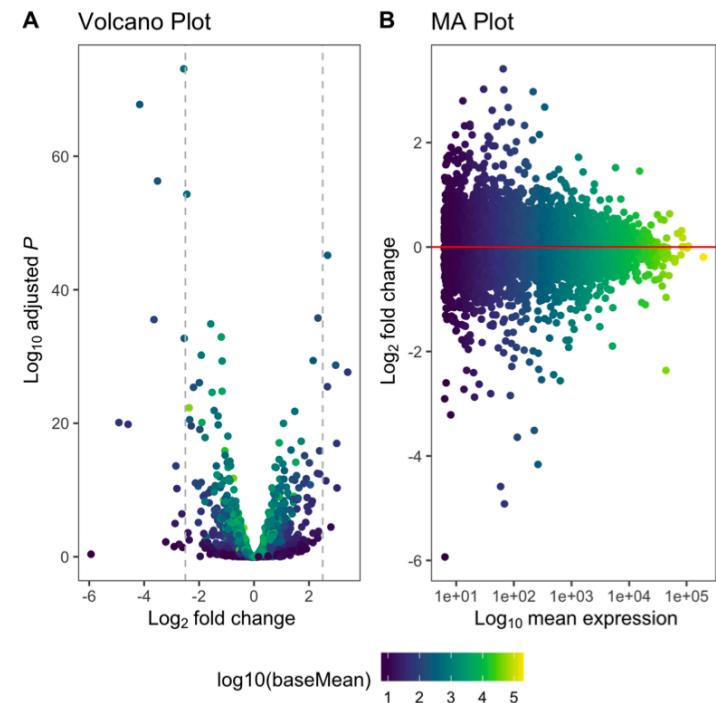
```
pheatmap::pheatmap(  
  mat = pasilla_heat,  
  show_rownames = FALSE,  
  annotation_col = pasilla_col  
)
```



Side by side plot with ggpubr

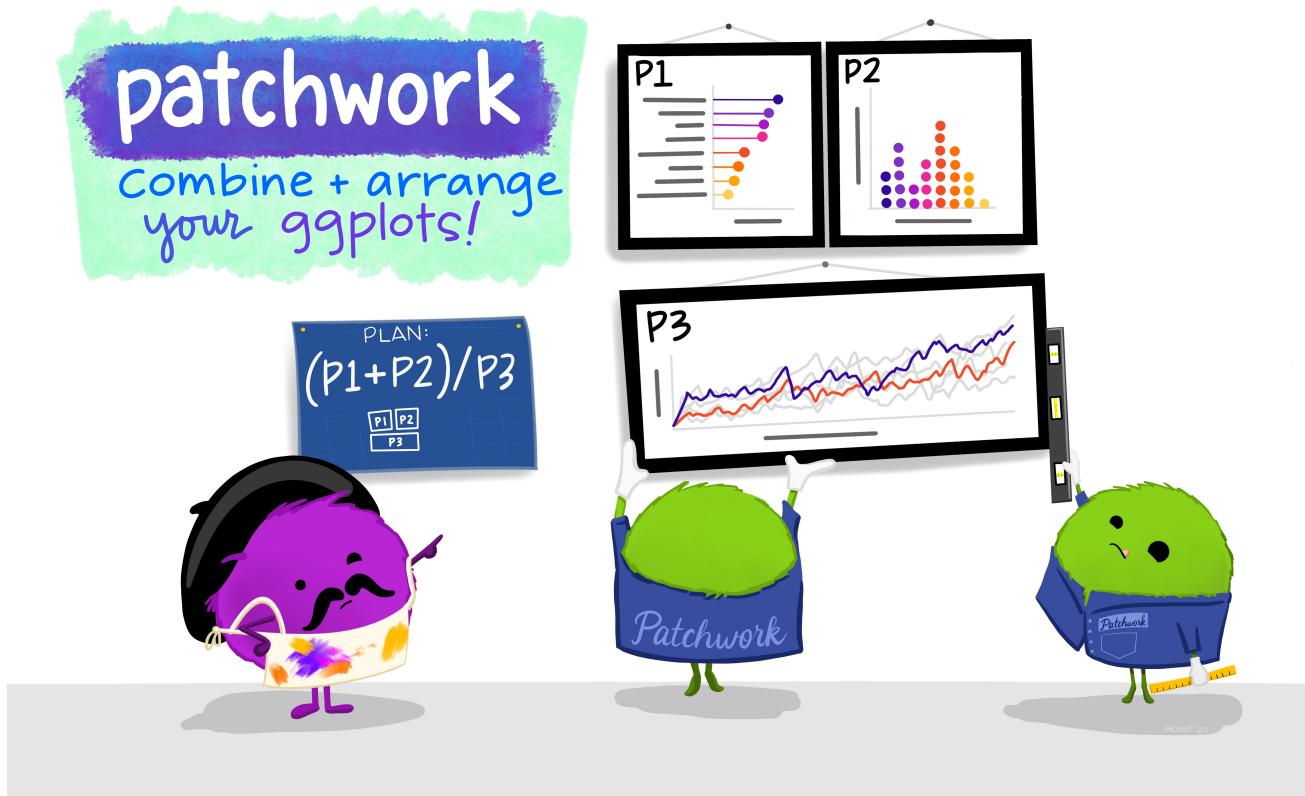
```
p1 <- ggplot(data = pasilla_data,
               aes(x = log2FoldChange,
                   y = -log10(padj),
                   color = log10(baseMean))) +
  geom_point() +
  geom_vline(xintercept = c(-2.5, 2.5),
             lty = 2, color="grey") +
  theme_few() + scale_color_viridis_c() +
  labs(x = bquote(~Log[2]~ "fold change"),
       y = bquote(~Log[10]~adjusted~italic(P)),
       title = "Volcano Plot")
p2 <- ggplot(data = pasilla_data,
               aes(x = baseMean,
                   y = log2FoldChange,
                   color = log10(baseMean))) +
  geom_point() +
  scale_x_log10() +
  geom_hline(yintercept = 0, color = "red") +
  theme_few() + scale_color_viridis_c() +
  labs(y = bquote(~Log[2]~ "fold change"),
       x = bquote(~Log[10]~ "mean expression"),
       title = "MA Plot")
```

```
ggpubr::ggarrange(p1, p2, labels = "AUTO",
                   common.legend = TRUE, legend = "bottom")
```



Other options:

cowplot (<https://wilkelab.org/cowplot/articles/index.html>) and
patchwork (<https://github.com/thomasp85/patchwork>)



Resources

ggbio: visualization toolkits for genomic data

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April 27, 2020

<http://127.0.0.1:12631/library/ggbio/doc/ggbio.pdf>

Modern Statistics for Modern Biology

Susan Holmes, Wolfgang Huber

[3 High Quality Graphics in R](https://web.stanford.edu/class/bios221/book/Chap-Graphics.html)

<https://web.stanford.edu/class/bios221/book/Chap-Graphics.html>

Inspiration and slides for this talk
Thanks!

Open content & slides:

- Jessica Minnier · Meike Niederhausen. bit.ly/berd_ggplot



Artwork by Allison Horst