

# AS-Quant User Manual

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## 1. About

AS-Quant is a computational tool used to detect alternative splicing (AS) events of two biological conditions- specifically, two groups of samples- from RNA-seq data. It can categorize five major types of AS in a comparative and comprehensive manner. AS-Quant also includes a visualization tool which generates plots for both the AS events and the annotation of the whole gene.

## 2. Download

AS-Quant tool can be downloaded directly from [Github](#). Users need to have Python installed on their machine. It can work on both Windows and Linux platforms.

## 3. Required tools

- a. [Python](#) (version 3.0 or higher)
- b. [Samtools 0.1.8\\*](#) [This specific version, provided with the code]

### Required python packages

- [matplotlib](#)  
[`$sudo apt-get install python3-matplotlib` **or** `$pip install matplotlib`]
- [scipy](#)  
[`$sudo apt-get install python3-scipy` **or** `$pip install scipy`]
- [pandas](#)  
[`$sudo apt-get install python3-pandas` **or** `$pip install pandas`]

## 4. Run AS-Quant

AS-Quant is designed to work on both human (hg19/hg38) and mouse (mm10) alternative splicing events. The supplementary data (the five types of alternative splicing exons' dataset and the reformatted annotation) is provided in the project directory in Github.

Users have to run the following two Python files in order to run AS-Quant:

- a. **as\_quant.py**: the main function which the user needs to run.
- b. **make\_plots.py**: generates figures for visual representation of data.

### 4. 1. Run as\_quant.py

Command: `$ python3 as_quant.py [options]`

AS-Quant supports the analysis of multiple samples or replicates (in .bam format) in each group/condition. For eliminating the false positives and ignoring the noises of the samples, **it is recommended to provide input data with at least 50M reads on each sample.**

Users can also select the underlying method [-method option] to determine the significance of each alternative splicing event. Available methods are: chi-squared test (chisquare) and Wilcoxon rank-sum test (ranksum). If there is more than one sample or replicate in each group, Wilcoxon rank-sum test is an additional option to determine the significance of the events.

### List of available options

(\* refers to a mandatory field)

|         |   |
|---------|---|
| -i*     | Input directories. The two directories for the two groups of samples in consecutive order.  |
| -s*     | Species name: hg38 or hg19 for human, mm10 for mouse  |
| -o      | Output directory  |
| -novel  | If the user wants to detect both novel and annotated alternative splicing events. Default is annotated only.                            |
| -method | Statistical method to determine the significance of the potential alternative splicing events: chisquare/ranksum. Default is chisquare. |

as-quant.py will generate several intermediary files in the output directory. After computing the significance of the association between the two conditions, the final results will be written in the spreadsheet named **asquant\_group1\_vs\_group2.xlsx**, with five separate sheets for five different splicing types. The following image shows some of the generated fields in **asquant\_group1\_vs\_group2.xlsx** for event type 'SE':

| chrom | geneName  | splicedExonStart | splicedExonEnd | P_value  | ratioDifference | absoluteRatioDifference | chromRegionLLong                 | eventType |
|-------|-----------|------------------|----------------|----------|-----------------|-------------------------|----------------------------------|-----------|
| chr1  | FN1       | 71603659         | 71603929       | 1.76E-74 | 0.21349749      | 0.21349749              | chr1:FN1:71603659-71603929       | SE        |
| chr11 | SRSF2     | 116851647        | 116851751      | 6.62E-23 | -0.167381288    | 0.167381288             | chr11:SRSF2:116851647-116851751  | SE        |
| chr1  | FN1       | 71613762         | 71614035       | 1.01E-19 | 0.092372387     | 0.092372387             | chr1:FN1:71613762-71614035       | SE        |
| chr11 | RTN4      | 29706409         | 29708770       | 1.32E-17 | 0.181502776     | 0.181502776             | chr11:RTN4:29706409-29708770     | SE        |
| chr6  | HNRNPA2B1 | 51461919         | 51462055       | 2.67E-14 | -0.103489035    | 0.103489035             | chr6:HNRNPA2B1:51461919-51462055 | SE        |
| chr15 | CBX5      | 103215021        | 103215121      | 1.41E-13 | -0.213653927    | 0.213653927             | chr15:CBX5:103215021-103215121   | SE        |
| chr16 | APP       | 84971407         | 84971461       | 3.5E-12  | 0.143653298     | 0.143653298             | chr16:APP:84971407-84971461      | SE        |
| chr6  | PHB2      | 124716424        | 124716430      | 1.58E-11 | -0.141349525    | 0.141349525             | chr6:PHB2:124716424-124716430    | SE        |
| chr7  | SNRNP70   | 45381824         | 45383184       | 1.97E-10 | -0.089632714    | 0.089632714             | chr7:SNRNP70:45381824-45383184   | SE        |
| chr15 | SERHL     | 83102624         | 83102644       | 2.31E-09 | -0.24611139     | 0.24611139              | chr15:SERHL:83102624-83102644    | SE        |
| chr6  | LUC7L2    | 38568846         | 38568917       | 4.48E-07 | -0.29741836     | 0.29741836              | chr6:LUC7L2:38568846-38568917    | SE        |
| chr9  | YIPF2     | 21592527         | 21592588       | 7.82E-07 | -0.248413685    | 0.248413685             | chr9:YIPF2:21592527-21592588     | SE        |
| chr5  | VPS29     | 122356793        | 122356805      | 7.87E-07 | -0.074800107    | 0.074800107             | chr5:VPS29:122356793-122356805   | SE        |
| chr2  | NSFL1C    | 151502454        | 151502460      | 1.53E-06 | -0.169609706    | 0.169609706             | chr2:NSFL1C:151502454-151502460  | SE        |
| chr3  | UBE2D3    | 135456600        | 135456644      | 1.97E-06 | -0.078878856    | 0.078878856             | chr3:UBE2D3:135456600-135456644  | SE        |
| chr17 | SRSF3     | 29039453         | 29039909       | 3.05E-06 | -0.140916809    | 0.140916809             | chr17:SRSF3:29039453-29039909    | SE        |
| chr2  | RBM39     | 156177632        | 156177906      | 1.29E-05 | -0.144285895    | 0.144285895             | chr2:RBM39:156177632-156177906   | SE        |
| chr6  | EP58      | 137591417        | 137591492      | 1.32E-05 | -0.188849143    | 0.188849143             | chr6:EP58:137591417-137591492    | SE        |
| chr12 | MTA1      | 113120210        | 113120261      | 1.48E-05 | -0.167204912    | 0.167204912             | chr12:MTA1:113120210-113120261   | SE        |
| chr5  | SETD8     | 124445968        | 124446084      | 1.52E-05 | -0.165478951    | 0.165478951             | chr5:SETD8:124445968-124446084   | SE        |

The column 'P\_value' defines the significance of the detected events. The results are sorted based on *P*-values. The splicing events with '**absoluteRatioDifference**' $\geq 1$  and **P\_value**  $< 0.05$  are considered to be significant between two conditions. The genes which have the average read coverage  $< 10$  on all the samples, are removed from the analysis. In the 'Ratio difference' column, a large positive ratio difference indicates a potential splicing event in group 2 (condition 2), whereas a negative ratio difference with a large absolute value indicates a potential splicing event in group 1 (condition 1).

## 4.2. Running AS-Quant with provided sample data

We provided sample data 'sample\_input\_mouse' in our GitHub repository to test AS-Quant, where group1 and group2 are two directories containing the input bam files.

Command: **\$ python3 as\_quant.py -s mm10 -i /<dir>/sample\_input\_mouse/group1 /<dir>/sample\_input\_mouse/group2 -o sample\_output**

## 4.3. Run make\_plots.py

AS-Quant provides a visualization tool, makeplots.py, which generates plots for both the AS events and the annotation of the whole gene. To run the visualization tool, users need to enter the following command:

**\$ python3 make\_plots.py -s species -o outputdirectory -i input1 input2**

Example: **\$ python3 make\_plots.py -s mm10 -i sample\_input\_mouse/group1 sample\_input\_mouse/group2 -o annotation\_plot**

Next, make\_plots.py will ask the users to enter the region of interest for which they want to generate the annotation plot. The format is specific:

**Chom:GeneName:RegionStart-RegionEnd**

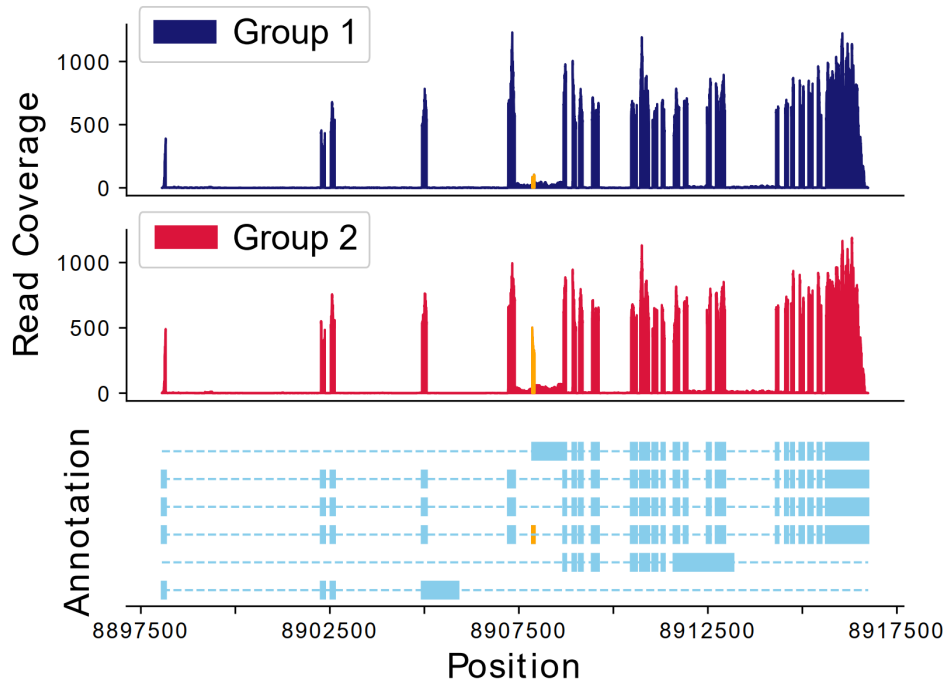
**make\_plots.py parameter descriptions**

|               |                                 |
|---------------|---------------------------------|
| Chrom :       | Name of the chromosome          |
| GeneName :    | Name of the gene                |
| RegionStart : | Starting position of the region |
| Region End :  | End position of the region      |

Example: **chr1:Tceb1:16641724-16643478**

The users can generate multiple plots, one by one, by providing one input region at a time. The code exits when the user puts the text 'Exit' and presses Enter.

Make\_plots.py will generate the read coverage plot for the given gene along with the whole annotation plot with all exons' information of that gene. The figure below shows an example of the read coverage plot generated by AS-Quant.



The first two subplots of the figure represent the read coverage of the two biological conditions. The bottom subplot shows the gene annotation and the exon information of that gene.