

Exercise 3 - Basic Models

WLS, GLM and Logistic

O'Malley

2025-01-19

Table of Contents

Submission	2
Setup	2
1. Heteroskedasticity Testing	3
2. Weighted Least Squares (WLS) Model	5
3. Logistic Regression	9
4. Decision Trees	11

```
knitr::opts_chunk$set(echo=T, warning=F, message=F)
library(lmtest)

## Loading required package: zoo

##
## Attaching package: 'zoo'

## The following objects are masked from 'package:base':
##
##   as.Date, as.Date.numeric

library(tidyverse)

## — Attaching core tidyverse packages ————— tidyverse
2.0.0 —
## ✓ dplyr      1.1.4      ✓ readr      2.1.5
## ✓ forcats   1.0.0      ✓ stringr   1.5.1
## ✓ ggplot2    3.5.1      ✓ tibble    3.2.1
## ✓ lubridate  1.9.3      ✓ tidyr     1.3.1
## ✓ purrr      1.0.2

## — Conflicts —————
tidyverse_conflicts() —
## ✗ dplyr::filter() masks stats::filter()
## ✗ dplyr::lag()     masks stats::lag()
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all
conflicts to become errors
```

```
library(tree)
```

```
## Warning: package 'tree' was built under R version 4.3.3
```

Submission

Download the **Ex3_BasicModels_YourLastName.Rmd** R Markdown file and save it with your own last name. Complete all your work in that template file, **Knit** the corresponding Word or PDF file. Your knitted document **must display your R commands**. Submit your knitted homework document. No need to submit the .Rmd file, just your knitted file.

Also, please prepare your R Markdown file with a **professional appearance**, as you would for top management or an important client.

Please, write all your interpretation narratives outside of the R code chunks, with the appropriate formatting and businesslike appearance. I write all my comments inside of the R code chunk to suppress their display until I print the solution, but you should not do this. I will read your submission as a report to a client or senior management. Anything unacceptable to that audience is unacceptable to me.

Setup

This analysis will be done with the **Hitters{ISLR}** baseball player dataset, using AtBat, Hits, Walks, PutOuts, Assists and HmRun as predictors and player **Salary** as the outcome variable. Let's start with an OLS model and we will then test for heteroskedasticity.

```
# Done for you
```

```
library(ISLR) # Contains the Hitters dataset
```

```
# Enter the commands below in the R Console window, but NOT in the R Markdown file. Inspect the data and the description of each predictor, to familiarize yourself with the data
```

```
# ?Salaries  
# View(Salaries)
```

```
# This dataset has several records with omitted data, Let's remove them  
Hitters=na.omit(Hitters)
```

```
# Fit an OLS model to start with  
fit.ols <- lm(Salary ~ AtBat+Hits+Walks+PutOuts+Assists+HmRun, data=Hitters)  
summary(fit.ols)
```

```
##  
## Call:  
## lm(formula = Salary ~ AtBat + Hits + Walks + PutOuts + Assists +  
##      HmRun, data = Hitters)  
##
```

```
## Residuals:
##      Min       1Q   Median       3Q      Max
## -920.3 -215.7  -47.7   175.4  2007.9
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept) 124.48415    72.75876   1.711 0.088308 .
## AtBat       -2.43104     0.66358  -3.664 0.000302 ***
## Hits        8.98051     1.97223   4.553 8.17e-06 ***
## Walks       6.34231     1.41170   4.493 1.07e-05 ***
## PutOuts     0.25462     0.08960   2.842 0.004847 **
## Assists     0.06698     0.19649   0.341 0.733485
## HmRun       7.02439     3.61990   1.940 0.053418 .
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 378.8 on 256 degrees of freedom
## Multiple R-squared:  0.311, Adjusted R-squared:  0.2949
## F-statistic: 19.26 on 6 and 256 DF,  p-value: < 2.2e-16
```

As the output shows, there are 4 significant predictors: AtBat, Hits, Walks and PutOuts, and 2 non-significant predictors: Assists and HmRun.

1. Heteroskedasticity Testing

1.1 Conduct a **Breusch-Pagan** test for Heteroskedasticity for the **fit.ols** model above.

```
bptest(fit.ols) # Breusch-Pagan test

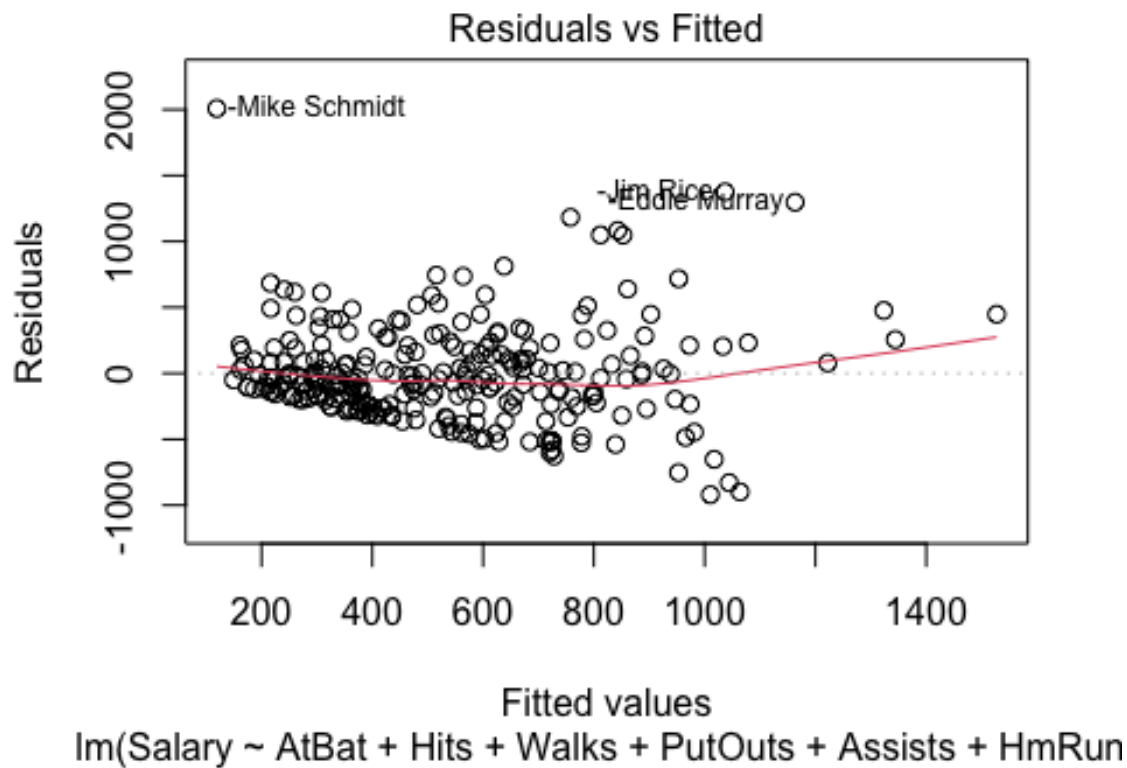
##
## studentized Breusch-Pagan test
##
## data: fit.ols
## BP = 15.456, df = 6, p-value = 0.01699
```

Analysis

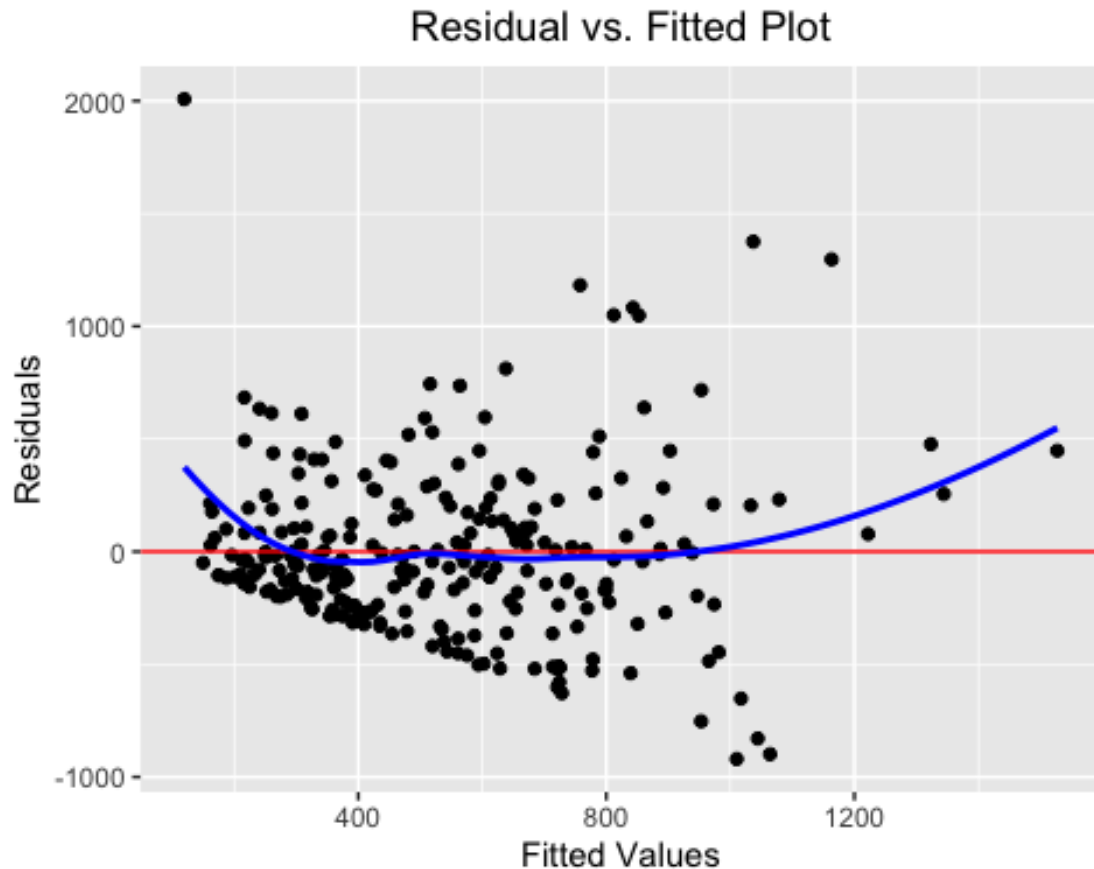
After conducting the Breusch-Pagan test, we can conclude that there is heteroscedasticity in our model. Since $p(0.01699) < 0.05$, we reject the null hypothesis that homoscedasticity exists in our model.

1.2 Display the first residual plot for **fit.ols** by using `which=1`.

```
plot(fit.ols, which = 1)
```



```
residuals_df <- data.frame(fitted = fitted(fit.ols), residuals =
resid(fit.ols)) # create data frame
ggplot(residuals_df, aes(fitted, residuals)) +
  geom_point() +
  geom_hline(yintercept = 0, color = "red") +
  geom_smooth(method = "loess", color = "blue", se = FALSE) +
  labs(x = "Fitted Values", y = "Residuals", title = "Residual vs. Fitted
Plot") +
  theme(plot.title = element_text(hjust = 0.5))
```



1.3 Is there a problem with Heteroskedasticity? Why or why not? In your answer, please refer to **both**, the BP test and the residual plot.

Analysis

Yes, there appears to be heteroscedasticity in the model. The Breusch-Pagan test causes us to reject the H_0 that homoscedasticity exists in the model because $p(0.01699) < 0.05$ and the residual plot shows a cone-patterned dispersion of residuals, which supports the existence of heteroscedasticity in the model.

2. Weighted Least Squares (WLS) Model

2.1 Set up the parameters of the WLS model. Use the `abs()` and `residuals()` functions compute the absolute value of the residuals from the OLS model **fit.ols** and store the results in a vector object named **abs.res**. Then use the `fitted()` function to extract the fitted (i.e., predicted) values from **fit.ols** and store the results in a vector object named **fitted.ols**. The run an `lm()` model using the predicted values in **fitted.ols** as a predictor of the absolute value of the residuals in **abs.res**.

Technical tip: Because you are using one data vector to predict another data vector, you don't need the `data=` parameter.

As a sanity check, display the first 10 rows of the fitted() values of **lm.abs.res**

```
abs.res <- abs(residuals(fit.ols)) # extract absolute residuals
fitted.ols <- fitted(fit.ols) # extract fitted values

lm.abs.res <- lm(abs.res ~ fitted.ols) # second OLS Model

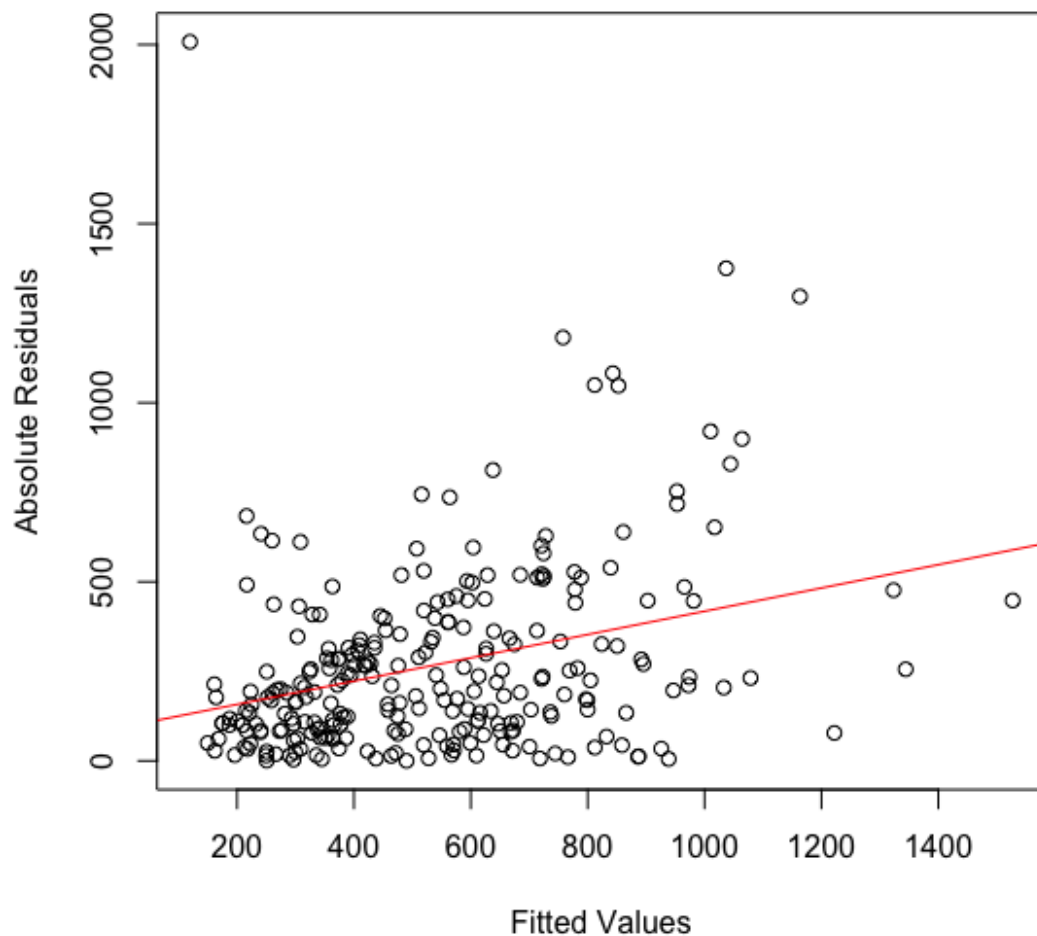
fitted(lm.abs.res)[1:10] # sanity check
```

##	-Alan Ashby	-Alvin Davis	-Andre Dawson	-Andres Galarraga
##	270.2217	406.4748	291.4550	285.5487
##	-Alfredo Griffin	-Al Newman	-Argenis Salazar	-Andres Thomas
##	270.8002	149.4738	141.2410	162.5797
##	-Andre Thornton	-Alan Trammell		
##	257.5593	342.6095		

2.2 To visualize the lm.abs.res regression line, plot the **fitted.ols** vector against the **abs.res** vector. Then draw a red line using the abline() function for the **lm.abs.res** regression object.

```
plot(fitted.ols, abs.res, xlab = "Fitted Values", ylab = "Absolute
Residuals", main = "Fitted vs. Absolute Residuals Plot")
abline(lm.abs.res, col = "red")
```

Fitted vs. Absolute Residuals Plot



2.3 Specify and Run the WLS Model. First, a vector named **wts** equal to the inverse squared predicted values of **lm.abs.res** (use `wts <- 1/fitted(lm.abs.res)^2`).

Then fit the WLS regression model using the same predictors you used in `ols.fit`, but using **wts** as the weights. Name this regression object **wls.fit**. Display the summary results.

While we are at it, also fit a similar weighted GLM model (**WGLM**), by using the `glm()` function and storing the results in an object named **fit.wglm**. Then display the `summary()` results for the WGLM.

```
wts <- 1/fitted(lm.abs.res)^2 # calculate weights

wls.fit <- lm(Salary ~ AtBat+Hits+Walks+PutOuts+Assists+HmRun, data=Hitters,
weights = wts) # WLS Model
summary(wls.fit) # summary statistics
```

```
##
## Call:
## lm(formula = Salary ~ AtBat + Hits + Walks + PutOuts + Assists +
##     HmRun, data = Hitters, weights = wts)
##
## Weighted Residuals:
##      Min       1Q   Median       3Q      Max
## -2.0512 -1.0607 -0.2793  0.6520 13.6904
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept) 259.2218    61.2878   4.230 3.26e-05 ***
## AtBat       -2.6758     0.6914  -3.870 0.000138 ***
## Hits         8.4446     2.2715   3.718 0.000247 ***
## Walks        4.4277     1.5889   2.787 0.005723 **
## PutOuts      0.2953     0.1157   2.553 0.011257 *
## Assists      0.4160     0.2022   2.057 0.040679 *
## HmRun       10.4194     4.0230   2.590 0.010150 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 1.542 on 256 degrees of freedom
## Multiple R-squared:  0.1747, Adjusted R-squared:  0.1554
## F-statistic: 9.034 on 6 and 256 DF,  p-value: 5.81e-09

fit.wglm <- glm(Salary ~ AtBat+Hits+Walks+PutOuts+Assists+HmRun,
data=Hitters, weights = wts) # WGLM Model
summary(fit.wglm) # summary statistics

##
## Call:
## glm(formula = Salary ~ AtBat + Hits + Walks + PutOuts + Assists +
##     HmRun, data = Hitters, weights = wts)
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept) 259.2218    61.2878   4.230 3.26e-05 ***
## AtBat       -2.6758     0.6914  -3.870 0.000138 ***
## Hits         8.4446     2.2715   3.718 0.000247 ***
## Walks        4.4277     1.5889   2.787 0.005723 **
## PutOuts      0.2953     0.1157   2.553 0.011257 *
## Assists      0.4160     0.2022   2.057 0.040679 *
## HmRun       10.4194     4.0230   2.590 0.010150 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for gaussian family taken to be 2.378078)
##
##      Null deviance: 737.69  on 262  degrees of freedom
## Residual deviance: 608.79  on 256  degrees of freedom
```



```
## AIC: 3897.9
##
## Number of Fisher Scoring iterations: 2
```

2.3 Observe the similarities and differences between the OLS, WLS and WGLM model and provide a brief commentary of your observations.

Analysis

The r-squared value has decreased from the original OLS model to the WLS, although the WLS model now accounts for heteroscedasticity. Two statistically insignificant variables - Assists and HmRun - have now become significant to both the WLS and WGLM models. Finally, in the WGLM model, Residual Deviance < Null Deviance, which means the model is more accurate than random guessing - a good sign for the WGLM model.

3. Logistic Regression

3.1 Download the **myopia.csv** file to your working directory. Then read it using `read.table()` with the parameters `header=T`, `row.names=1`, `sep=","`. Store the dataset in an object named **myopia**.

Dataset documentation at: <https://rdrr.io/cran/aplore3/man/myopia.html> Please note that **myopic** is coded as 1 (Yes), 0 (No) (not 1 and 2)

For sanity check, list the first 10 rows and 8 columns of this dataset.

```
read.table("myopia.csv", header = TRUE, row.names = 1, sep = ",") -> myopia #
assign table to vector
```

```
myopia[1:10, 1:8] # sanity check
```

```
##      study.year myopic age female spheq    al    acd    lt
## 1         1992      1   6      1 -0.052 21.89 3.690 3.498
## 2         1995      0   6      1  0.608 22.38 3.702 3.392
## 3         1991      0   6      1  1.179 22.49 3.462 3.514
## 4         1990      1   6      1  0.525 22.20 3.862 3.612
## 5         1995      0   5      0  0.697 23.29 3.676 3.454
## 6         1995      0   6      0  1.744 22.14 3.224 3.556
## 7         1993      0   6      1  0.683 22.33 3.186 3.654
## 8         1991      0   6      1  1.272 22.39 3.732 3.584
## 9         1991      0   7      0  1.396 22.62 3.464 3.408
## 10        1991      0   6      1  0.972 22.74 3.504 3.696
```

3.2 Fit a logistic model to predict whether a kid is **myopic**, using `age + female + sports.hrs + read.hrs + mommy + dadmy` as predictors. **Tip:** use `family="binomial"(link="logit")`. Store the results in an object named **myopia.logit**. Display the `summary()` results. Then display the `summary()` results.

```

myopia.logit <- glm(myopic ~ age + female + sports.hrs + read.hrs + mommy +
dadmy, data = myopia, family = binomial(link = "logit")) # Logreg model
summary(myopia.logit) # summary statistics

##
## Call:
## glm(formula = myopic ~ age + female + sports.hrs + read.hrs +
##      mommy + dadmy, family = binomial(link = "logit"), data = myopia)
##
## Coefficients:
##              Estimate Std. Error z value Pr(>|z|)
## (Intercept) -8.83835     2.21222  -3.995 6.46e-05 ***
## age          -0.03073     0.29219  -0.105   0.916
## female       -0.15787     0.46980  -0.336   0.737
## sports.hrs   -0.13993     0.03507  -3.990 6.60e-05 ***
## read.hrs      0.79920     0.09929   8.049 8.35e-16 ***
## mommy         2.93733     0.54288   5.411 6.28e-08 ***
## dadmy         2.77087     0.54069   5.125 2.98e-07 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for binomial family taken to be 1)
##
##      Null deviance: 480.08  on 617  degrees of freedom
## Residual deviance: 131.00  on 611  degrees of freedom
## AIC: 145
##
## Number of Fisher Scoring iterations: 8

```

3.3 For interpretation purposes, display the log-odds alongside the odds. Use the `coef()` function to extract the log-odds coefficients from **myopia.logit** and save them in a vector object named **log.odds**. Then use the `exp()` function to convert the log-odds into odds and store the results in a vector object named **odds**. Then enter the options(`scipen=4`) command to minimize the use of scientific notation. Finally, list the log-odds and odds side by side. To do this, use the `cbind()` function to bind the two vectors into one table and name the columns “Log-Odds” and “Odds” respectively. Embed the `cbind()` function inside the `print()` function with the parameter `digits=2` to get a more compact display.

```

log.odds <- coef(myopia.logit) # Logodds coefficients
odds <- exp(log.odds) # calculate odds
print(cbind("Log-Odds" = log.odds, "Odds" = odds), digits = 2)

##              Log-Odds      Odds
## (Intercept)  -8.838 1.5e-04
## age          -0.031 9.7e-01
## female       -0.158 8.5e-01
## sports.hrs   -0.140 8.7e-01
## read.hrs      0.799 2.2e+00
## mommy         2.937 1.9e+01
## dadmy         2.771 1.6e+01

```

3.4 Provide a brief interpretation of both, the log-odds and odds effects of read.hrs and mommy. Please refer to the respective variable measurement units in your discussion.

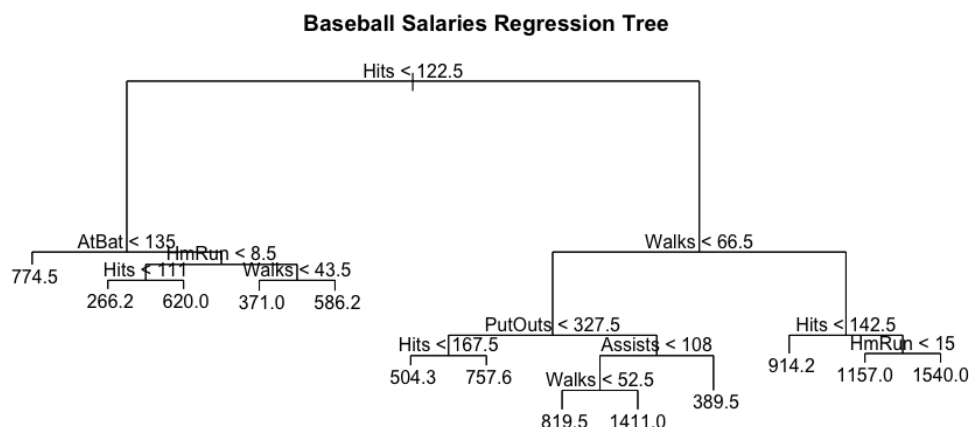
Analysis

On average, holding everything else constant, there is a positive relationship between read.hrs and myopia (Log-Odds = 0.799) and a one unit increase in read.hrs multiplies the odds of myopia by 2.22 (increase of 122.2%). On average, holding everything else constant, there is a stronger positive relationship between mommy and myopia (Log-Odds = 2.937) and since mommy is a binary variable, if the value is 1, then the odds of myopia are multiplied by 18.86 (increase of 1787.5%).

4. Decision Trees

4.1 Regression Tree. Load the **{tree}** library. Then fit the regression model **ols.fit** above, but this time as a **regression tree** using the **tree()** function and save the results in an object named **fit.tree.salary**. Then plot the tree using the **plot()** and **text()** functions (use the **pretty=0** parameter). Also use the **title()** function to title you tree diagram “**Baseball Salaries Regression Tree**”.

```
tree(fit.ols) -> fit.tree.salary # regression tree model
# plot creation
plot(fit.tree.salary)
text(fit.tree.salary, pretty = 0)
title("Baseball Salaries Regression Tree")
```



4.2 Classification Tree. Fit the Logistic model **myopia.logit**, but this time as a **classification tree** using the **tree()** function and save the results in an object named **fit.tree.myopia**. Then plot the tree using the **plot()** and **text()** functions (use the **pretty=0** parameter). Also use the **title()** function to title you tree diagram “**Myopia Classification Tree**”.

```
fit.tree.myopia <- tree(myopia.logit) # classification tree model

# plot creation
plot(fit.tree.myopia)
text(fit.tree.myopia, pretty = 0)
title("Myopia Classification Tree")
```

