

Introduction of a Biologically Plausible Color Descriptor to a Neurodynamical Model of the Primary Visual Cortex

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Abstract

We present a computational model of color description & processing in the primary visual cortex (V1), inspired by current neurobiological understanding. This understanding posits single and double opponent neurons as fundamental to low level color processing. We offer a novel representation of color by defining these cells' responses and the connections between them, within the framework of a dynamical model of neuronal activity. Our model reproduces perceptual experiences in a number of cases, offering credence to said biological theories.

Index Terms

primary visual cortex, striate cortex, V1, receptive field, single-opponent, double-opponent, color assimilation, color induction

I. INTRODUCTION

COLOR induction and contrast are two related, opposing, perceptual phenomena. The former is a change in perceived color "toward" a nearby color, while the latter is a change of one color "away" from the nearby color. Neurophysiological research suggests that these phenomena may arise as early in primate vision as the primary visual cortex (V1). It is proposed that the boundaries between two colored regions drive these effects. Specifically, research in the field describes neurons which fire selectively to boundaries between certain colors, so called double-opponent cells, and identifies them as being critically related to the color perceived.

Within, we propose a computational model designed around the current understanding of this biology. We present two implementations, one more biologically accurate, and another more computationally elegant. We explore the behavior of these models with respect to what they can teach us about the assumed biological theories, as well as their application to the field of computer vision.

II. STATE OF THE ART

A. Neurobiology

//TODO This understanding treats color and shape as intrinsically connected and, as a consequence, predicts perceptual phenomena such as color induction to arise very early in visual processing.

Historically, it was widely believed that color and shape are two distinct aspects of visual perception. Truly, this line of thought is intuitive: one can perceive the color of a flat surface which occupies our full field of vision, despite its lack of 'shape', likewise we can see the shape of achromatic objects, as in black and white film. This theory of perception innervated neurophysiological understanding and was supported by neurobiological findings, such as that the lateral geniculate nucleus (LGN), the pathway which carries information from the retina to the primary visual cortex (V1), consists of three entirely distinct layers; two (the parvocellular and koniocellular) carrying only color information, and one (the magnocellular) dealing purely in achromatic contrast (edge) information. Based on early anatomical observations, it was proposed that these three LGN pathways for color and contrast are then processed into two separate streams in V1, one for color and the other for form. This separate handling of color and form confirmed for researchers that, as suspected, these two perceptual concepts are processed separately in the brain.

Research in the past decade or so has seen a shift from this thinking, however. Psychophysical observations, such as those in Figure 1, influenced researchers to consider that color and form are more intrinsically related than previously thought. In these examples, we see that the perceived color of the inner square is highly dependent on the background and, most importantly, how the inner square *contacts* the background. The color perceived is not just determined by the physical properties of the surface, but also by the context in which the surface is viewed. Furthermore, we can observe that this contextual influence comes largely from the boundary edges of the surface: in example (c), the dulling color assimilation effect witnessed in (b) is almost

entirely negated by simply adding a thin border. That is, by removing contact between the inner square and the background, the effect of the context is significantly reduced. In fact, "the color appearance of a region may be more dependent on color contrast at the boundary of the region than it is on the spectral reflectance of the region's interior." [5, p.572]



Fig. 1. Psychophysical example emphasizing the effect of edge contrast in color perception. In all cases, the center square is the same physical color. On (a) the green background, however, it appears much brighter than it does on (b) the red background. (c) Adding a thin black border negates the assimilation effect, increasing the perceived brightness of the center square. (d) Blurring the background, on the other hand, seems to enhance assimilation, lowering the perceived brightness at the center.

With these psychophysical observations in mind, much research has been devoted to elucidating the exact neurobiological mechanisms behind the perception of color. The current view holds that while the LGN does indeed carry color and contrast information through distinct pathways to the striate cortex (V1), once there, color and form become deeply intertwined as they are processed further. To explain the simultaneous processing of color and form in V1, the literature proposes two broad classifications of neurons: single-opponent cells, & double-opponent cells (Johnson et al. Color and Orientation in V1).

Opponency, in neurobiology, refers to antagonistic inputs of information: one source of input excites the neuron, while another inhibits it. If the two sources provide equal input, for example, they would cancel out and the neuron would not fire. Furthermore, removal of stimuli from the antagonist input also excites the neuron. **//TODO Discuss how this is ON/OFF.**

//TODO Include examples of ON/OFF center surround

With respect to cells in the early visual system, we are here specifically referring to chromatic and spatial opponency, as will be detailed below. Briefly, single-opponent cells respond best to large areas of color, while double-opponent cells respond only to the boundaries between such areas.

Single-Opponent Neurons

Single-opponent neurons are built using the classical center/surround receptive fields. The ON receptive field exciting the cell when presented with a particular color in the center, the OFF receptive field inhibiting the cell when another color is present in the surround, or exciting it when that color is *removed* from the surround. This other color is called its opponent, and thus colors come in *opponent pairs*. In the human visual system, we recognize three axes of color opponency: red vs. green, blue vs. yellow, and light vs. dark. To construct such concepts of color, we contrast activity of our three types of cone cells, each having its own specific spectral response function. By contrasting relative activity with neighboring cones, the retina can already begin to construct signal resembling psychological concepts of color. **//TODO What a mess.. see how others explain this.**

//TODO add image of simple behavior: gradient response to isoluminant chromatic boundary

//TODO add image of simple behavior: no response to intensity changes

//TODO add image of ON/OFF differences: R-G cell responds to fullscreen green light removed (not red removed)

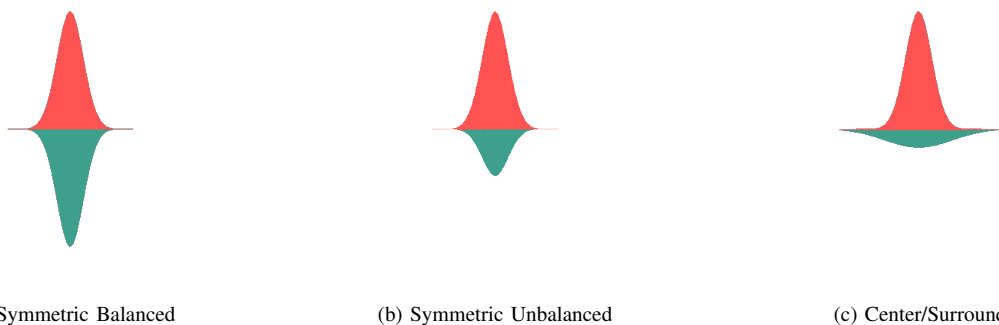


Fig. 2. Visualization of a variety of single-opponent (SO) receptive fields.

Achromatic Single-Opponent Neurons: Sometimes called non-opponent cells, they have no chromatic nor spatial opponency. Instead, they amalgamate all color input in a balanced manner so as to only respond to changes in luminosity. Technically, we can consider non-opponent cells to be a special case of single-opponent cells.

//TODO are NO cells the same as SO cells? What about achromatic double-opponent cells?

//TODO add image depicting example receptive field(s)

//TODO add image of simple behavior: gradient response to ((mono)chromatic) intensity changes

//TODO add image of simple behavior: no response to isoluminant chromatic boundaries

Double-Opponent Neurons

Point: Double-opponent neurons are edge detectors.

Story: Double-opponent neurons are a topic of confusion in the field of neurobiology. All researchers on the topic seem to agree that the input to the neuron is that of two single-opponent neurons itself. In this sense, the term "double-opponent" can be thought of as indicating that the dimensionality of color opponency has been doubled. However, another camp of researchers take the definition a step further and suggest that the two SO inputs are spatially offset. By this definition, the term "double-opponent" is thought to indicate that the cell is sensitive to opponency in two *different* dimensions, color and space.

The distinction is non-trivial as the response patterns differ, and thus the interpretation of their role in vision differs. Truly, both types of cells may, and likely do [//TODO Cite studies which find these cells], exist in the primate visual system. However, for the purpose of our work we only consider the latter definition; double-opponent cells' receptive fields are both chromatically and spatially antagonistic.

//TODO present the differences between the two DO configurations

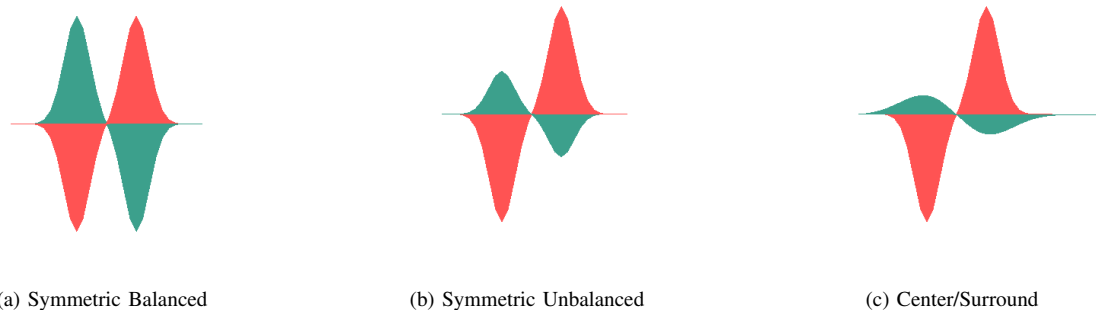


Fig. 3. Visualization of a variety of double-opponent (DO) receptive fields.

Orientation Selectivity: Research shows that cells identified as double-opponent are orientation selective. That is, they respond most strongly when stimulated by a border of particular orientation, less so with variation from that preferred orientation, and weekly, if at all, to borders orthogonal to the preferred orientation. This is intuitive given the organization of the receptive field as previously defined. By arranging the components of the receptive field to be spatially antagonistic, there will be one orientation of border which best separates the two components, and another orientation, precisely perpendicular (//TODO correct term?), which does not separate the two components at all. This can be easily visualized, as in Figure ??.

//TODO add image depicting vertical vs. diagonal vs. horizontal receptive fields & ideal vs. unideal stimuli

//TODO add image of simple behavior: peak activity AT sharp edge

//TODO PARAGRAPH ON SINGLE OPPONENT ORIENTATION SELECTIVITY (possible but not considered here)

Spatial Frequency Selectivity: Another important feature of double-opponent cells is that they are inherently sensitive to borders of specific size, or scale. Technically, we refer to such preference of 'scale' as a cell's spatial frequency selectivity. Each double-opponent neuron will be selective to a particular spatial frequency and excited less so by stimuli of spatial frequency deviating from this preference. Again, this is quite intuitively a product of the design of the neuron's receptive field and easily visualized, as in Figure ??.

//TODO CONTRAST WITH SINGLE-OPPONENT'S LACK OF SPATIAL FREQ. SELECTIVITY (show Shapley's graphs) [erm.. don't we have spatial frequency for SO??]

Achromatic Double-Opponent Neurons: As we saw with single-opponent cells, while we frequently discuss opponency as being a thing of 'color', one of the axes of the opponent process theory is 'lightness' vs 'darkness'. This is easily achieved by balancing cone inputs into the neuron's antagonistic receptive fields. Such a cell is often referred to as an *achromatic double-opponent* cell. It behaves just as the previously described double-opponent cells, with specific orientation and spatial frequency selectivity, but responds to luminosity borders rather than chromatic borders. For more details on achromatic receptive fields, please refer back to the section on *Achromatic Single-Opponent Neurons*.

//TODO add image of Shapley response curves for SO, & DO

Hypercolumns

Thus far we have defined two broad classes of neurons in the primate visual system: single-opponent and double-opponent. Each class has a number of parameters which dictate what stimuli the cell should respond to: opponent color, orientation selectivity, and spatial frequency. A single cell can only be sensitive to a specific combination of settings for each parameter. However, for each parameter it is important to have neurons sensitive to the full range of parameters. Thus, it is necessary to have a collection of neurons which together cover the complete combinatorial set of settings for all parameters.

Furthermore, as may be obvious, the receptive field of a neuron in V1 is directly **connected/related/tied** (???) to a set of neurons in the retina. That is, neurons in V1 are sensitive only to activity at a particular physical location on the retina; V1 is *retinotopic*, or retinally mapped.

The implication is that for each retinal position, in V1 it is necessary to have a large number of both single-opponent and double-opponent cells, selective to the full gamut of parameters possible for each type of cell. Biologically, these collections of cells are organized in *hypercolumns*, a physical grouping of cells in which neuronal connections 'up' and 'down' within the column are much more dense than extensions to cells in other columns. Within a single hypercolumn, all cells are sensitive to a specific retinal location, but together express sensitivity to the full range of parameters.

//TODO mention relative abundance of SO, & DO

CO Blobs

//TODO PARAGRAPH ON WHERE DO CELLS ARE

//TODO PARAGRAPH ON WHERE SO CELLS ARE

//TODO PARAGRAPH ON PROPOSED CONNECTIONS BETWEEN DO & SO

Neurobiology Summary

//TODO Summarize the key points from all THAT.

To recapitulate: single-opponent neurons fire best to full field stimulation while double-opponent neurons fire only at the boundaries between particular colors. Both cell types can be found in color preferring and achromatic flavors.

B. Computational Modeling:

The model described in this research is an extension of that presented by Penacchio *et al.* [?], itself based on work by Z. Li [2], [3]. Before detailing our implementation, it is important to review prior art, these works and others. As our goal is to model the behavior of color preferring and achromatic single-opponent and double-opponent neurons, we will also review other computational endeavors tackling the issue of color and/or form from a biologically inspired perspective.

Li's Neurodynamical Model for Segmentation (1999)

In Li's original work, a neurodynamical model was presented which focused on the global region segmentation using only local interactions between neurons. In the interest of simplicity, Li's implementation dealt only with the nature of these interactions, ignoring where the stimulus, or information, might be coming from. Conceptually, Li defined neurons by the physical position in the image and the 'feature' to which they are sensitive. She then defined the connections between these neurons such that those physically close to each other, and sensitive to similar features, interacted most strongly. Stimulation of one neuron, then, positively stimulated (excited) similar neurons nearby, and negatively stimulated (inhibited) dissimilar neurons nearby. Such excitation and inhibition cascades through the network naturally, producing a large scale response pattern defined by local stimuli.

In the model presented, Li used oriented bars as features, though expressed that any logical feature could be reasonably considered in its place. This choice was biologically inspired by neurons sensitive to specifically oriented bars, the aforementioned double-opponent cells. When considering such features, inter-neuronal connections can be logically deduced: two neurons positively interact most when both 1) are sensitive to similarly oriented bars and 2) are co-located along that same orientation. Two neurons negatively interact most when either of these two conditions is not met.

//TODO include graphic or oriented bars which positively & negatively interact

By defining the neuronal connectivity in this manner, patterns of neurons sensitive to co-located and co-aligned bars positively interact with each other to enhance their collective response to the stimuli. Similarly, isolated stimuli negatively interact with their neighbors and are silenced. From these local interactions, global features are enhanced if they satisfy the neuronal connectivity rules, and noise is suppressed. Li showed that this method can be used to enhance contours and identify boundaries between regions for which normal segmentation methods struggle.

Penacchio, Otazu, & Dempere-Marco's Neurodynamical Model for Brightness Induction (2013)

Li's work laid the foundation for Penacchio *et al.* who extended the model to a usable framework which:

- 1) Uses real black & white images /movies as input.
- 2) Utilizes discrete wavelet transforms to extract edges (more on this later).
- 3) Added multi scale support.
- 4) Summarizes the results into an output 'perceptual image'.

Their research was focused on observing brightness induction (BI) arising from such a neurodynamical model.

- No color, just black & white *edges*
- Generalized to real images (edges vs lines)
- Added scales
- Dynamical processing
- **Avoid detail, save that for *Method..?***
- Extension of Z. Li's edge detection work
- Uses DWT to extract oriented edges in grayscale
 - ..in our context, it's essentially a luminance sensitive double-opponent cell.

Itti, Koch, & Niebur's Model for Saliency (1999)

- Opponent color transformations
- No double-opponent cells
- Center & surround using scales
- Has scales, but collapses them into one (right?)
- *No dynamical processing*

Zhang, Barhomi, & Serre's Biologically Inspired Color Descriptor (2013)

- Single & double-opponent color using weights
- Has center/surround (Gabor filters for DO, gaussians for SO?)
- No scales
- *No dynamical processing*

Spitzer & Barkan's Model of Color Induction (2005)

- Single & double-opponent color transformations using receptive fields
- Center & surround receptive fields
- *No dynamical processing*

Comparison of Models

These models, though approaching different problems in different ways, can be compared to each other in terms of our current work. In Table II-B we lay out the features which are implemented in our Proposed Model, and we contrast and compare the implementation of features in prior art. Here it is easy to see that the goal of our work is to combine the dynamical processing of Li and Penacchio with descriptions of color at scale and orientation inspired by the works of Itti, Zhang, and Spitzer.

//TODO describe what exactly qualifies for 'colors', 'scales', and 'orientations' in this general comparison

To summarize, the purpose of this project is to **feed opponent color information into a neurodynamical model** sensitive to edges & surfaces in a biologically inspired manner.

//TODO We need to properly summarize the State of the Art before proceeding.

- What does the biology predict?
- What have computational models done?
- Where do computational models fall short? (A: no dynamical color model)
- Thus, where does our work fit into the field?

III. METHOD

We present a computational model designed to be representative of the aforementioned biology. The implementation of this model can be conceived of as two distinct parts:

- *Description of Visual Information*: transformation of image(s) into a biologically meaningful representation.
- *Neurodynamical Processing*: iterative processing of the dynamic interactions between the neurons modeled.

TABLE I
COMPARISON OF SOME OF THE RELEVANT MODELS. EACH HAS VERY DIFFERENT GOALS, AND THUS TAKES A VERY DIFFERENT APPROACH.

	Proposed Model	Li [3]	Penacchio [4]	Itti [1]	Zhang [7]	Spitzer [6]
Dynamical	Y	Y	Y	N	N	N
Colors	Y	N	N	Y	Y	Y
Scales	Y	N	Y	Y	N	N
Orientations	Y	Y	Y	Y	N	N
SO	Y	N	N	Y	Y	Y
DO	Y	N	Y (achromatic)	Y	Y	Y
SO RF	Gaussian / DWT	N/A	N/A	Gaussian Pyramid	None	Gaussian
DO RF	Gabor-like / DWT	N/A	DWT	Gaussian Pyramid	None	Gabor
Goal	Color Induction	Saliency	Brightness Induction	Saliency	Color Descriptor	Color Induction

In the interest of clarity, the categories are defined below.

- Dynamical: Does the work attempt to model neurodynamical processes?
- Colors: Does the work consider chromatic differences as features?
- Scales: Does the work separate information at different scales?
- Orientations: If incorporating edge/boundary features, does the work distinguish between orientations?
- SO: Does the work attempt to model single-opponent cell behavior, directly or indirectly?
- DO: Does the work attempt to model double-opponent cell behavior, directly or indirectly?
- SO RF: If modeling single-opponent cells, how are their receptive fields defined?
- DO RF: If modeling double-opponent cells, how are their receptive fields defined?
- Goal: Generally speaking, what was the purpose of the modeling effort?

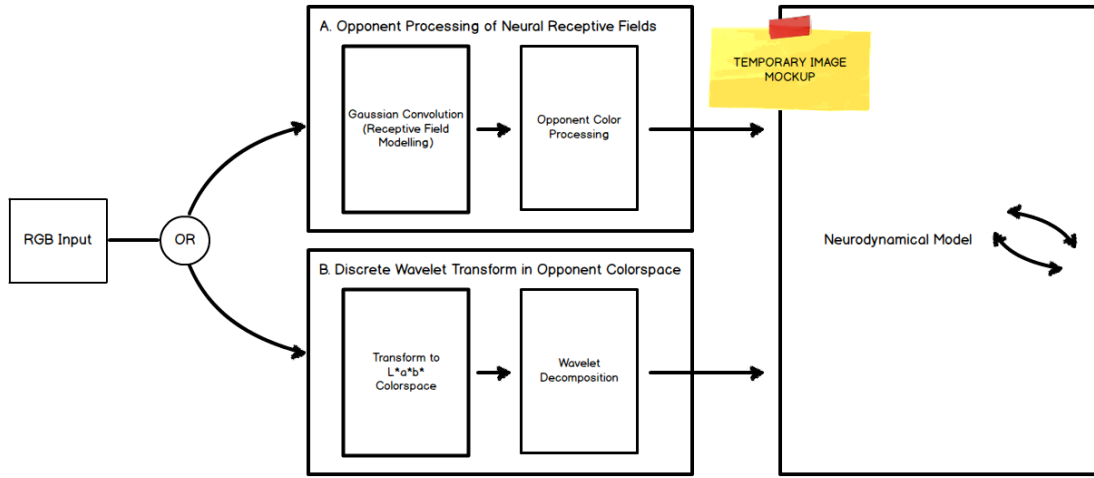


Fig. 4. High level schematic of model components. The point of emphasis here is that neurodynamical model is agnostic to how the data is processed upfront, as long as the format is correct. The two methods for data preparation (read: color description) are independent of each other and analyzed separately.

Description of Visual Information

The opponent-process theory defines three axes visual information, obtained by processing of cone activity from the retina. These axes are Red-Green (R-G), Blue-Yellow (B-Y), and Light-Dark (L-D). Neurobiological research corroborates such processing of color information (*//TODO cite this*). *//TODO MORE ON OPPONENT-PROCESS THEORY*

//TODO PARAGRAPH ON SINGLE AND DOUBLE OPPONENT CELLS

In accordance with this understanding of biology, the image transformation process should 1) separate opponent color components from each other, and 2) separate surface information from boundary information. We describe two different approaches to this end. The first approach, herein referred to in proximity as the "*Opponent Processing of Neural Receptive Fields*", represents an effort to explicitly transform raw visual data in strict accordance with neurobiology. Essentially attempting to mimic the information processing pathways of the retina, lateral geniculate nucleus (LGN), and striate cortex (V1). The second approach, termed with equal verbosity, is the "*Discrete Wavelet Transform in Opponent Colorspace*". Here we stray from the details of the biological pathways in an attempt to achieve the same end, through more computationally efficient means.

In both, the input is a normal RGB image, and the output to the neurodynamical model is a 5 dimensional matrix containing 6 opponent color channels (R, G, B, Y, L, & D), each deconvoluted into its surface and oriented edge components, and 1 or more scales. In this work we generalize oriented edges into 3 orientations: horizontal, diagonal, & vertical. Before processing by the neurodynamical model, this data is normalized so as to circumvent differences between the data output by these two methods.

A. Opponent Processing of Neural Receptive Fields

Neural Receptive Fields: //TODO REFACTOR INTO PARAGRAPH ON RECEPTIVE FIELDS

Opponent Processing: //TODO REFACTOR INTO PARAGRAPH ON OPPONENT PROCESSING

Processing starts by convolving each of the L, M, and S cone spectral response functions with two different gaussian filters. These convolutions simulate integration of information in a neuron's receptive field. One convolution is used to build the excitatory component of the receptive field (referred to as the center), the other convolution, the inhibitory (referred to as the surround).

Single-opponent cells' receptive fields are the classical center-surround configuration: symmetric, centered, circular gaussians. Typically, the center is smaller and weighed significantly more heavily than the surround, as in Figure ?? (c). Double-opponent receptive fields are markedly more complex: asymmetrical, off-center, elongated gaussian filters. Similarly, these filters can be adjusted to different sizes, shapes, and weights. For our tests we worked with symmetric balanced receptive fields, as in Figure ?? (a).

In our implementation, for convenience, we utilize the R, G, and B channels of the raw image as an approximation of L, M, and S cone activity, respectively. It should be noted that a preprocessing step could be performed to better match RGB to LMS so as to be more biologically consistent. The impact is likely negligible.

To construct opponent color channels, we contrast center (c) and surround (s) convolutions of the L, M, and S channels into three pairs: two color [6] and one achromatic.

$$R(c, s, \sigma) = \max(0, L(c, \sigma) - M(s, \sigma)) \quad (1)$$

$$G(c, s, \sigma) = \max(0, M(c, \sigma) - L(s, \sigma)) \quad (2)$$

$$B(c, s, \sigma) = \max(0, S(c, \sigma) - \frac{L(s, \sigma) + M(s, \sigma)}{2}) \quad (3)$$

$$Y(c, s, \sigma) = \max(0, \frac{L(c, \sigma) + M(c, \sigma)}{2} - S(s, \sigma)) \quad (4)$$

We also introduce two new opponent channels, lightness (L) and darkness (D):

$$L(c, s, \sigma) = \max(0, \frac{L(c, \sigma) + M(c, \sigma) + S(c, \sigma)}{3} - 0.5) \quad (5)$$

$$D(c, s, \sigma) = |\min(0, \frac{L(c, \sigma) + M(c, \sigma) + S(c, \sigma)}{3} - 0.5)| \quad (6)$$

//TODO consider $\frac{L(c)+M(c)+S(c)}{3} - \frac{L(s)+M(s)+S(s)}{3}$

//TODO consider $(L(c) + M(c)) - (L(s) + M(s))$ (Rolf & Deco)

It is important to note that the L & D opponent channels do not incorporate surround effects as the color opponent channels do. More work can be done in this regard.

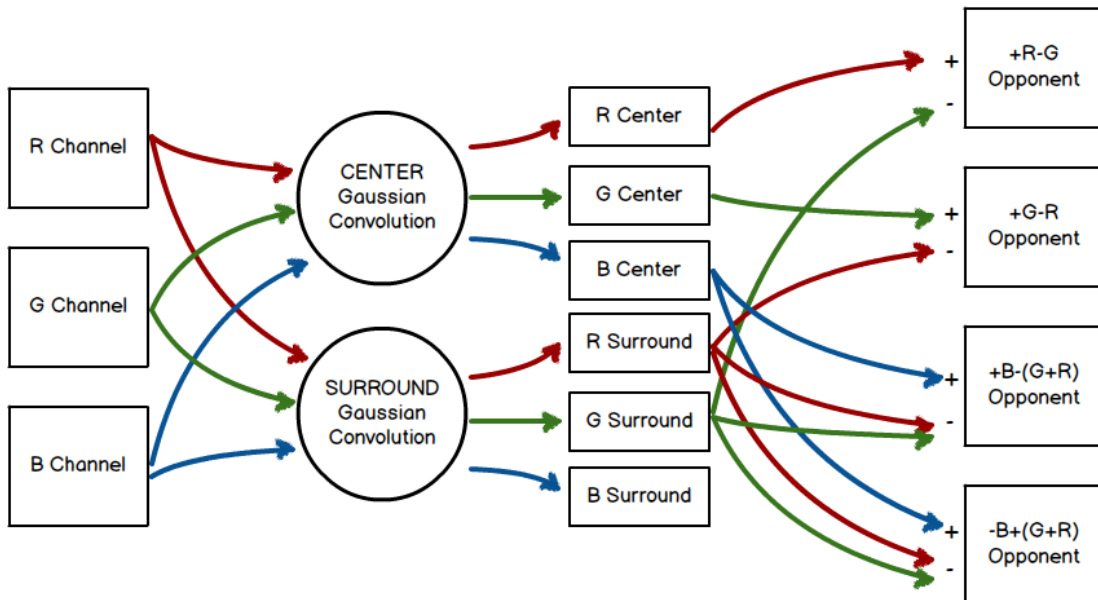


Fig. 5. Depiction of opponent processing of center/surround receptive fields. The RGB channels are convolved with gaussians and then combined to obtain opponent color signal.

//TODO ADD L & D to Figure 5

//TODO PARAGRAPH ON DOUBLE-OPPONENT ORIENTATIONS

//TODO PARAGRAPH ON SCALE

1) Considerations:

- CON: Relatively slow
 - Could be improved with Gabor instead of gaussian for DO cells.
 - Could be improved by reusing SO info -> more biological
 - It's just an upfront cost, the neurodynamical processing is the most expensive.
- PRO: more receptive field control (explicit RF definitions)
- CON: requires tweaking of receptive fields
- PRO: more true to biology (combination of signal rather than numeric transformation of colorspace)
- CON: requires decision on meaningful RGB combination (should be elucidated from biology)
 - Pre-transformation to LMS might be valuable/meaningful.

//TODO show original & decomposed image

Summary

This first of two approaches to color representation aims to be true to the biology at all cost. We explicitly model the receptive fields of both single-opponent and double-opponent neurons. These receptive fields are used to transduce retinal activity of L, M, and S cones into the cell activity expected in V1.

B. Discrete Wavelet Transform in Opponent Colorspace

Previous work by Penacchio *et al.* [?] utilized a discrete wavelet transform (DWT) to decompose a greyscale image into its oriented edge components, at scales. In the context of our current research, this could be thought of as representing achromatic double-opponent cells; the response is greatest at luminosity boundaries, and nonexistent on surfaces or at chromatic changes. In this work we extend their approach to the opponent colorspace and examine its applicability as a replacement of the previously detailed *Opponent Processing of Neural Receptive Fields*.

Opponent Colorspace: This transformation begins, not with the convolution step, but instead with a colorspace transformation. We convert the image from RGB to CIE $L^*a^*b^*$. This colorspace separates luminance from its two opponent color channels. The two color channels are 0 centered, with both positive and negative values possible. The luminance channel, L^* , scales from 0 to 100, 0 being total darkness, 100 being pure brightness. We subtract 50 from this channel to obtain a 0 centered range of luminance. We consider positive values to define 'lightness' and negative values 'darkness'. From the opponent color channels, we take positive values in the a^* channel as denoting 'redness' and negative values 'greenness', while positive values in the b^* channel indicate 'blueness' and negative values 'yellowness'.

Discrete Wavelet Transform: Similar to the aforementioned use of gaussian filters, the application of the DWT can be interpreted as representative of the processing in a cell's receptive field. That is, the activity of a particular cell is defined not just by its own activity, but also of the activity of the cells around it. The key difference here being that the DWT is applied in the opponent colorspace, while in our previous approach the convolutions are applied to the raw L, M, and S input, before deriving opponent colors.

As all color channels are 0 centered, we can easily split them into independent opponent components:

$$Li_{do} = \max(0, DWT(L^* - 50)) \quad (7)$$

$$Li_{so} = \max(0, DWT(L^* - 50)_{residual}) \quad (8)$$

$$Da_{do} = |\min(0, DWT(L^* - 50))| \quad (9)$$

$$Da_{so} = |\min(0, DWT(L^* - 50)_{residual})| \quad (10)$$

$$R_{do} = \max(0, DWT(a^*)) \quad (11)$$

$$R_{so} = \max(0, DWT(a^*)_{residual}) \quad (12)$$

$$G_{do} = |\min(0, DWT(a^*))| \quad (13)$$

$$G_{so} = |\min(0, DWT(a^*)_{residual})| \quad (14)$$

$$B_{do} = \max(0, DWT(b^*)) \quad (15)$$

$$B_{so} = \max(0, DWT(b^*)_{residual}) \quad (16)$$

$$Y_{do} = |\min(0, DWT(b^*))| \quad (17)$$

$$Y_{so} = |\min(0, DWT(b^*)_{residual})| \quad (18)$$

//TODO FIGURE: show diagram of oriented DWT filter (use TikZ)

//TODO FIGURE: show original & decomposed image
 //TODO PARAGRAPH ON INTERPRETATION OF COMPONENTS (DO & SO)
 //TODO PARAGRAPH ON WORKFLOW: $RGB \rightarrow L^*a^*b^* (-50) \rightarrow DWT \rightarrow ON/OFF$
 //TODO PARAGRAPH ON PROS/CONS

Neurodynamical Processing

Stage I of the proposed method focuses on the description of color in a biologically meaningful form, of which we've described two distinct approaches. Stage II is to further process this data in an iterative computational model of neurodynamical processes, as defined in Equations 19 and 20. The input stimulus, from Stage I, at time τ , $I_{i\gamma\sigma\theta}^\tau$ drives the model and mainly determines its response. The normalization of the stimulus at each time step is extended from that proposed by Li [3] and Penacchio [4]. We normalize within each color channel (γ), scale (σ), and orientation (θ). At the first time step, $I_{i\gamma\sigma\theta}^1$ is used to bootstrap the model and set as the initial excitation response, $x_{i\gamma\sigma\theta}$.

$x_{i\gamma\sigma\theta}$ can be viewed as a model of retinotopic excitation hypercolumns in V1, i defining the retinotopic location of the hypercolumn. Similarly, $y_{i\gamma\sigma\theta}$ may be interpreted as its retinotopic inhibitory counterpart. $\dot{x}_{i\gamma\sigma\theta}$ and $\dot{y}_{i\gamma\sigma\theta}$, then, are the change in excitatory and inhibitory membrane potentials, respectively, and follow the equations:

$$\dot{x}_{i\gamma\sigma\theta} = -\alpha_x x_{i\gamma\sigma\theta} \quad (19)$$

$$\begin{aligned} & -g_y(y_{i\gamma\sigma\theta}) \\ & - \sum_{\gamma' \neq \text{opp}(\gamma), \Delta_\sigma, \Delta_\theta \neq 0} \Psi(\gamma', \Delta_\sigma, \Delta_\theta) g_y(y_{i\gamma\sigma + \Delta_\sigma + \Delta_\theta}) + J_0 g_x(x_{i\gamma\sigma\theta}) \\ & + \sum_{j \neq i, \gamma' \neq \text{opp}(\gamma), \sigma' \neq \sigma, \theta' \neq \theta} J_{[i\gamma\sigma\theta, j\gamma'\sigma'\theta']} g_x(x_{j\gamma'\sigma'\theta'}) \\ & + I_{i\gamma\sigma\theta}^\tau \\ & + I_0 \end{aligned}$$

$$\begin{aligned} \dot{y}_{i\gamma\sigma\theta} = & -\alpha_y y_{i\gamma\sigma\theta} \quad (20) \\ & + g_x(x_{i\gamma\sigma\theta}) \\ & + \sum_{j \neq i, \gamma' \neq \text{opp}(\gamma), \sigma' \neq \sigma, \theta' \neq \theta} W_{[i\gamma\sigma\theta, j\gamma'\sigma'\theta']} g_x(x_{j\gamma'\sigma'\theta'}) \\ & + I_c \end{aligned}$$

where $\alpha_x x_{i\gamma\sigma\theta}$ and $\alpha_y y_{i\gamma\sigma\theta}$ model the decay to the resting potential, $g_x(x)$ and $g_y(y)$ are sigmoid-like functions modeling cells' firing rates in response to membrane potentials x and y , respectively, $\Psi(\gamma', \Delta_\sigma, \Delta_\theta) \leq 1$ is the spread of inhibition within a hypercolumn, $J_0 g_x(x_{i\gamma\sigma\theta})$ is self-excitation, and I_c is background noise [3]. Any cell can interact with another by exciting it or inhibiting it, via monosynaptic excitation through excitatory-excitatory horizontal connections, or disinaptic inhibition through excitatory-inhibitory connections, respectively [4]. Such interactions are modeled between cells within and across hypercolumns; color, scale, and orientation, as well as between cells within and across cell classes; double-opponent and single-opponent. These cellular interactions are defined by $J_{[i\gamma\sigma\theta, j\gamma'\sigma'\theta']}$ (excitatory) and $W_{[i\gamma\sigma\theta, j\gamma'\sigma'\theta']}$ (inhibitory).

Orientation (θ): The excitatory and inhibitory interactions between orientation specific double-opponent cells are exactly as those defined by Penacchio [4] and Li [3]. Single-opponent cells, introduced in our work, are non-directional; they have no preferred orientation θ . $\Delta\theta$ is thus not computable and so this class of cells do not *inhibit* the activity of other cells. Single-opponent cells do not contribute to W . In order to explore 'fill in' effects, however, we support parameterization of *excitatory* weights to and from single-opponent and double-opponent cells. This weight functions as $\Delta\theta$ and is equal for all preferred orientations of the double-opponent cells. These interactions are modeled by a circular-symmetric gaussian in J .

Scale (σ): As in Penacchio [4], we decompose the input signal into signal at different scales. This reflect single-opponent and double-opponent cells' spatial frequency selectivity derived from their receptive fields. While this confers a level of scale invariance to the model, the primate visual cortex is not completely scale invariant and nor should our model be. Both image transformation methods presented in Stage I inherently represent activity at lower spatial frequencies with higher response.

Color (γ): We introduce a novel concept to the neurodynamical model framework developed by Li [3] and Penacchio [4]: color. Unfortunately, the structural organization of color preferring cells and the connections between them is not well understood and leading theories are still quite controversial [?]. We herein define a set of *logical* rules of interaction opponent color cells (including achromatic cells).

If a cell is stimulated, it excites nearby cells of the same orientation and scale preference, in any color channel other than its opponent. Conversely, a stimulated cell will inhibit nearby cells of the same orientation and scale preference, only in its opponent color channel. For example, stimulation of a DO cell preferring 'yellow' vertical edges will excite a DO cell preferring 'red' vertical edges at the same location. It will equally excite 'green', 'light', and 'dark' cells preferring the same orientation in the same location. It will not, however, excite cells of its opponent color 'blue'. In fact, it will inhibit activity in 'blue' cells preferring this same orientation in the same location.

We propose that these rules of interaction are logical as only opponent colors are mutually exclusive. While a bluish yellow line is not permissible in the opponent colorspace, a bluish green, bluish red, bright blue, or dark blue line is entirely plausible. Each of the possible combinations are equally probable and so the excitatory-excitatory connections between cells of these color preferences are equally weighed. These excitatory and inhibitory rules are encoded in J and W , respectively.

Activity Normalization (I_0): The general and local normalization of activities [?] has been extended from its used by Li [3] and Penacchio [4]. We normalize excitatory membrane potentials within double-opponent cells of similar location (i), color (γ), scale (σ), and orientation (θ) preferences. Likewise we normalize single-opponent cells of similar location (i), color (γ), scale (σ). We found it important to normalize the activity of these two classes of neurons independently as the information encoded by each is radically different: localized edge information by double-opponent cells, and extensive surface information by single-opponent cells.

IV. EXPERIMENTS

Analyze Data Transformation

- Opponent Receptive Fields
 - does SO behave as expected
 - does DO behave as expected
 - does Itti space behave as expected
- DWT
 - does SO behave as expected (as Itti?)
 - does DO behave as expected
 - ($L*a*b*$ is trusted)
- What are the differences between these outputs?
 - Show side by side LDRGBY/LAB+- convolutions
- What are general shortcomings?
 - OFF receptive fields function in time: removing green excites red. Our RFs don't reflect this.

Analyze Neurodynamical Model

- We can trust the concept because of Otazu 2013
- We need to analyze how color works
 - Connections between color channels
 - * What are our assumptions? (& why?)
 - * How does it modify the results over no connections?
 - * How does it modify the results over full connections (including between opponents)?
 - Connections between SO and DO

V. RESULTS

TODO

VI. CONCLUSIONS

TODO

APPENDIX A APPENDIX TITLE

TODO

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NOTES

State of the Art - Biology - Notes:

- 1) What is color?
 - Subjective
 - Correlates to reflectance patterns
- 2) Historical view → separation of color & shape
 - Parallel/modular/segregated processing [5]
 - Intuitive
 - Black & white movies work fine (Shapley 2011)
 - Full field color can be seen fine
 - LGN research suggested parvocellular & koniocellular has color, magnocellular has contrast (edges)
 - Similarly, V5 was 'motion'
- 3) Current view → integration of color & shape
 - All information is processed as one information stream (too strong??)
 - Color opponency
 - Discuss LMS & opponent color theory
 - Retinal receptive fields & horizontal cells
 - LGN information reflects opponent colors (no spatial opponency)
 - SINGLE OPPONENT CELLS RESPOND BEST TO FULL FIELD COLOR
 - Spatial opponency
 - LGN information upgraded to include spatial opponency
 - Double-opponent cells: color & spatially opponent
 - Spatial frequency sensitivity
 - Orientation sensitivity
 - Shapley shows most V1 cells are double-opponent
 - DOUBLE OPPONENT CELLS RESPOND BEST TO COLOR BOUNDARIES
 - DO & SO roles
 - If there are SO cells in V1, they aren't just a stepping stone, but encode valuable information. Thus, they likely work in concert with DO cells (more numerous (Shapley))
 - DO cells detect edges → saliency? (Z. Li)
 - Interactions (hypercolumns, CO blobs, etc.)
 - Not well understood =(
 - Retinotopic
 - Hypercolumns (Z. Li?)
 - What does Shapley think of CO blobs (youtube Q & A)?

*Method - DWT & $L^*a^*b^*$ - Notes:*

- Process:
 - Convert image from RGB to $L^*a^*b^*$
 - Subtract 50 from L^* to center it on 0 (a^* and b^* are already zero centered)
 - Apply DWT at each scale
- 1) the wavelet signal at each scale is the DO response in that channel
- 2) the wavelet residual at each scale is the SO response in the channel
 - To recover R, G, B, Y, L, & D we take positive and negative values of the R-G, B-Y, & L-D channels.
- This implementation comes with obvious deviations from the biology:
 - 1) By transforming RGB to $L^*a^*b^*$ at the pixel level, we lose receptive field integration
 - 2) The brain doesn't translate to opponent colors and then find edges
 - this can be formalized as the difference between
 - a) the addition of convolutions
 - b) the convolution of additions
- What are the advantages?
 - 1) Computationally efficient & relatively fast
 - 2) ...?

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