# Ascorbate (Vitamin C), its Antioxidant Chemistry

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## **Ascorbic Acid Structure**

# AscH<sub>2</sub> is a Di-acid

HO OH OH OH OH OH OH OH OH OH AscH2 AscH2 AscH2 OH OH OH AscA AscA AscA Asc
$$^{OH}$$

At pH 7.4, 99.95% of vitamin C will be present as AscH<sup>-</sup>; 0.05% as AscH<sub>2</sub> and 0.004% as Asc<sup>2-</sup>. Thus, the antioxidant chemistry of vitamin C is the chemistry of AscH<sup>-</sup>.

# Forms of Ascorbate

From Bors W, Buettner GR. (1997) The vitamin C radical and its reactions in *Vitamin C in Health and Disease*, ed. by L. Packer and J. Fuchs, Marcel Dekker, Inc., New York, Chapter 4, pp75-94.

#### Kinetics of AscH<sup>-</sup> Reactions

Radical	$k_{obs}/M^{-1}s^{-1}$ (p	oH 7.4)
HO●	1.1 x 10 <sup>10</sup>	
RO <sup>●</sup> (tert-butyl alkoxyl radical)	1.6 x 10 <sup>9</sup>	
ROO <sup>●</sup> (alkyl peroxyl radical, e.g. CH <sub>3</sub> OO <sup>●</sup> )	1-2 x 10 <sup>6</sup>	These rate constants are
CI <sub>3</sub> COO•	1.8 x 10 <sup>8</sup>	for the reaction:
GS® (glutathiyl radical)	6 x 10 <sup>8</sup> (5.6)	A 11 D. A . D.11
UH <sup>•-</sup> (Urate radical)	1 x 10 <sup>6</sup>	$AscH^{\scriptscriptstyle{-}} + R^{\scriptscriptstyle{\bullet}} \to Asc^{\scriptscriptstyle{\bullet}} + RH$
TO <sup>•</sup> (Tocopheroxyl radical)	2 x 10 <sup>5 b</sup>	AscH- reacts rapidly with
Asc <sup>•-</sup> (dismutation)	2 x 10 <sup>5</sup>	these and similar oxidants making it an outstanding
CPZ <sup>•+</sup> (Clorpromazine radical action)	1.4 x 10 <sup>9</sup> (5.9)	
Fe(III)EDTA/ Fe(II)EDTA	≈10 <sup>2</sup>	donor antioxidant.
O <sub>2</sub> •-/HO <sub>2</sub> •	2.7 x 10 <sup>5</sup>	
Fe(III)Desferal®/Fe(II)Desferal®	Very slow	

 $<sup>^{\</sup>rm a}$  Estimated  $k_{\rm obs}$  for TO+ when in a biological membrane.

Adapted from: Buettner GR, Jurkiewicz BA. (1996) Catalytic metals, ascorbate, and free radicals: combinations to avoid. *Rad Research* **145:**532-541.

<sup>&</sup>lt;sup>b</sup> k is pH dependent, thus this is kobs at pH 7.4.

#### AscH is a Donor Antioxidant

AscH<sup>-</sup> donates a hydrogen atom (H• or H+ + e<sup>-</sup>) to an oxidizing radical to produce the resonance-stabilized tricarbonyl ascorbate free radical. AscH• has a pK<sub>a</sub> of -0.86; thus, it is not protonated in biology and will be present as Asc•-.

#### Dismutation of Ascorbate Radical

$$k_{\text{obs}}$$
 (7.4) = 1.4 x 10<sup>5</sup> M<sup>-1</sup> s<sup>-1</sup>

This rate constant increases by a factor of »10 when phosphate is present.\*

This dismutation reaction is the principal route to the elimination of the Asc•- in vitro. However, in vivo it is thought that reducing enzymes are involved in the removal of this radical, resulting in the recycling of ascorbate.\*\*

<sup>\*</sup>Reviewed in: Bors W, Buettner GR. (1997) The vitamin C radical and its reactions in *Vitamin C in Health and Disease*, ed. by L. Packer and J. Fuchs, Marcel Dekker, Inc., New York, Chapter 4, pp75-94.

<sup>\*\*</sup>Hossain MA, Asada K. (1985) Monodehydroascorbate reductase from cucumber is a flavin adenine dinucleotide enzyme. *J Biol Chem.* **260**:12920-12926.

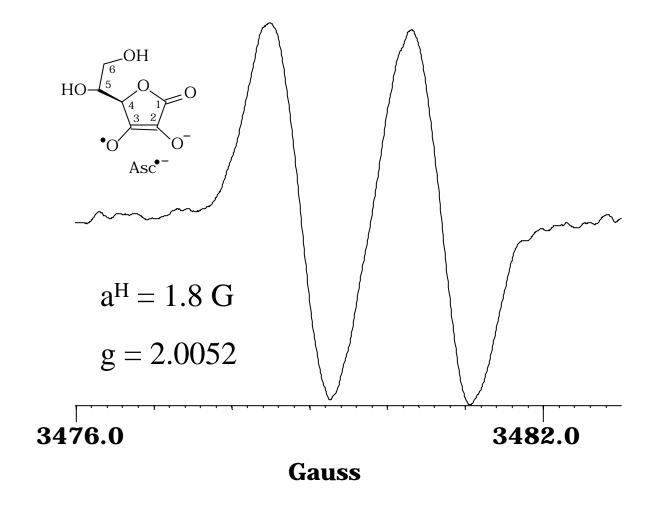
# The Dismutation of Ascorbate Radical is an Equilibrium Reaction

$$K = \frac{\left[ DHA \right] \left[ AscH_2 \right]_{total}}{\left[ Asc^{\bullet} \right]^2 \left[ H^+ \right] \left[ 1 + H^+ / K_{AscH_2} \right]} = 5 \times 10^{14} M^2$$

Because of this equilibrium reaction, Asc•- can be formed if both, DHA and AscH- are present. Note that the equilibrium constant, K, for this dismutation reaction is pH-dependent. Thus, the higher the pH the more Asc•- would be formed; however, at higher pH DHA is more unstable. ( $K_{AscH2}$  is the first ionization constant of ascorbic acid.)

Reviewed in: Bors W, Buettner GR. (1997) The vitamin C radical and its reactions in *Vitamin C in Health and Disease*, ed. by L. Packer and J. Fuchs, Marcel Dekker, Inc., New York, Chapter 4, pp75-94.

## EPR Detection of Asc



The ascorbate radical is usually observed as a simple doublet species by EPR.

The intensity of the EPR spectrum of Asc•- can be used as an indicator of oxidative stress in vitro and in vivo.

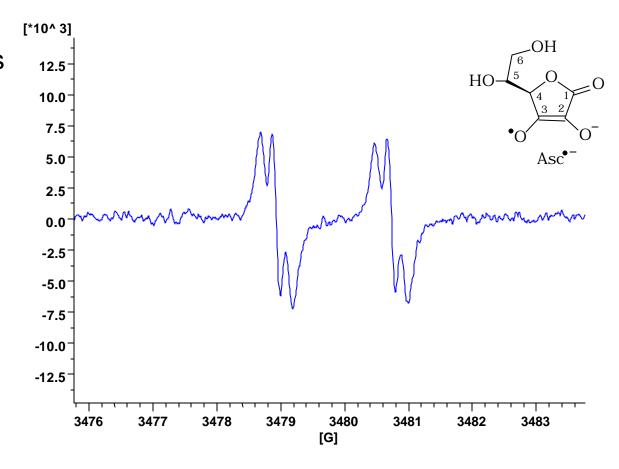
# **Higher Resolution EPR**

With appropriate instrument settings a more detailed spectrum can be observed by EPR.

$$a^{H_{\Delta}}(1) = 1.76 G$$

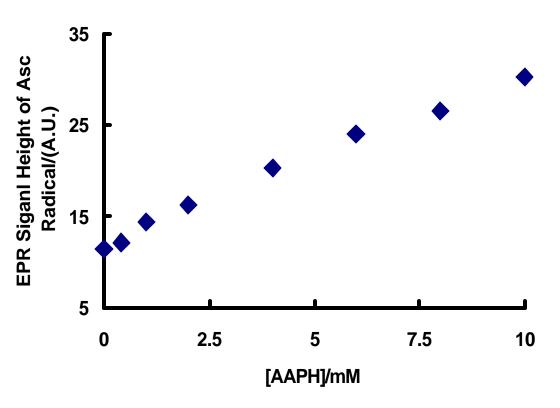
$$a_{5}^{H}(1) = 0.07 G$$

$$a_{6}^{H}(2) = 0.19 G$$



#### Asc<sup>--</sup>, Real Time Marker of Oxidative Stress

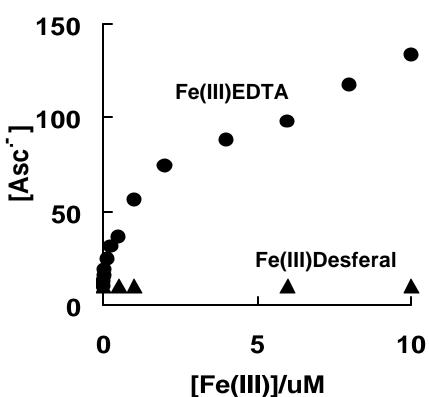
#### **Ascorbate Radical in Plasma**



[Asc•-]<sub>ss</sub> is proportional to the rate of ascorbate oxidation.

[Asc•-]<sub>ss</sub> in plasma is directly proportional to oxidative flux: EPR signal height of Asc•- (arbitrary units) versus AAPH concentration. The solutions contained 58 µM ascorbate in plasma and various amounts of the free radical-generator AAPH. From: Buettner GR, Jurkiewicz BA. (1993) The ascorbate free radical as a marker of oxidative stress: An EPR study. Free Radic Biol Med 14: 49-55.

# Asc<sup>--</sup>, as an indicator for adventitious transition metals



The oxidation of ascorbate is very slow in the absence of catalytic metals. This plot shows [Asc\*-]<sub>ss</sub> with varying [Fe(III)] in two different chelating environments. Because the iron in EDTA is freely accessible to reductants, Fe(III)EDTA is an excellent catalyst for AscH oxidation. In contrast Fe(III) in Desferal is not accessible to reductants and thus Fe(III)Desferal does not catalyze AscH oxidation.

EPR as well as UV-Vis spectroscopy ( $\epsilon_{265}$  = 14,500 M<sup>-1</sup> cm<sup>-1</sup> for AscH<sup>-</sup>) have been used to determine the metal content of buffer solutions.

Buettner GR. (1988) In the absence of catalytic metals, ascorbate does not autoxidize at pH 7: Ascorbate as a test for catalytic metals. *J Biochem Biophys Meth* **16:** 20-40.

Buettner GR. (1990) Ascorbate oxidation: UV absorbance of ascorbate and ESR spectroscopy of the ascorbyl radical as assays for iron. Free Rad Res Comm 10: 5-9\

# Thermodynamics of Ascorbate

The unpaired electron of Asc $^{\bullet-}$  resides in the  $\pi$ -system that includes the tri-carbonyl moiety of ascorbate. This results in a weakly oxidizing and weakly reducing radical. Due to its  $\pi$ -character Asc $^{\bullet-}$  does not react with oxygen to form dangerously oxidizing peroxyl radicals. Thermodynamically, it is relatively unreactive with a one-electron reduction potential of only +282 mV. It is considered to be a terminal, small-molecule antioxidant.

Buettner GR, Jurkiewicz BA. (1993) The ascorbate free radical as a marker of oxidative stress: An EPR study. Free Radic Biol Med 14: 49-55.

Buettner GR. (1993) The pecking order of free radicals and antioxidants: Lipid peroxidation, α-tocopherol, and ascorbate. Arch Biochem Biophy. **300:**535-543.

# The Pecking Order

Note that the donor antioxidants are found in the middle of the "pecking order".

Buettner GR. (1993) The pecking order of free radicals and antioxidants: Lipid peroxidation, α-tocopherol, and ascorbate. *Arch Biochem Biophys.* **300**:535-543.

Redox Couple	E°'/mV
(one-electron reductions)	
$HO^{\bullet}$ , $H^{\dagger}/H_2O$	+ 2310
RO <sup>•</sup> , H <sup>+</sup> /ROH (aliphatic alkoxyl radical)	+ 1600
ROO <sup>•</sup> , H <sup>+</sup> /ROOH (alkyl peroxyl radical)	+ 1000
GS*/GS (glutathione)	+ 920
PUFA <sup>•</sup> , H <sup>+</sup> /PUFA-H ( <i>bis</i> -allylic-H)	+ 600
TO, H <sup>+</sup> /TOH (tocopherol)	+ 480
$H_2O_2$ , $H^+/H_2O$ , $HO^{\bullet}$	+ 320
Asc <sup></sup> , H <sup>+</sup> /AscH <sup></sup> (Ascorbate)	+ 282
$CoQ^{\bullet -}$ , $2H^{+}/CoQH_{2}$	+ 200
Fe(III) EDTA/Fe(II) EDTA	+ 120
CoQ/CoQ*	<del>-</del> 36
$O_2/O_2^{\bullet-}$	<b>-</b> 160
Paraquat/Paraquat •	<b>-</b> 448
Fe(III)DFO/Fe(II)DFO	<b>-</b> 450
RSSR/RSSR*- (GSH)	<del>-</del> 1500
$H_2O/e^-$ aq	<del>-</del> 2870

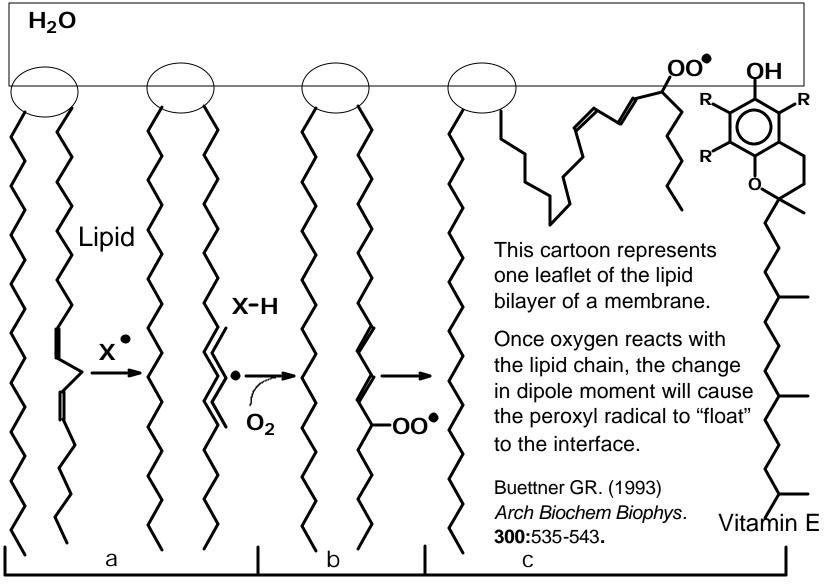
## C and E as Co-antioxidants

As seen in the thermodynamic pecking order above, the tocopherol radical, TO<sup>•</sup>, is more oxidizing then Asc•. It is thought that ascorbate contributes to the recycling of TO back to TOH.

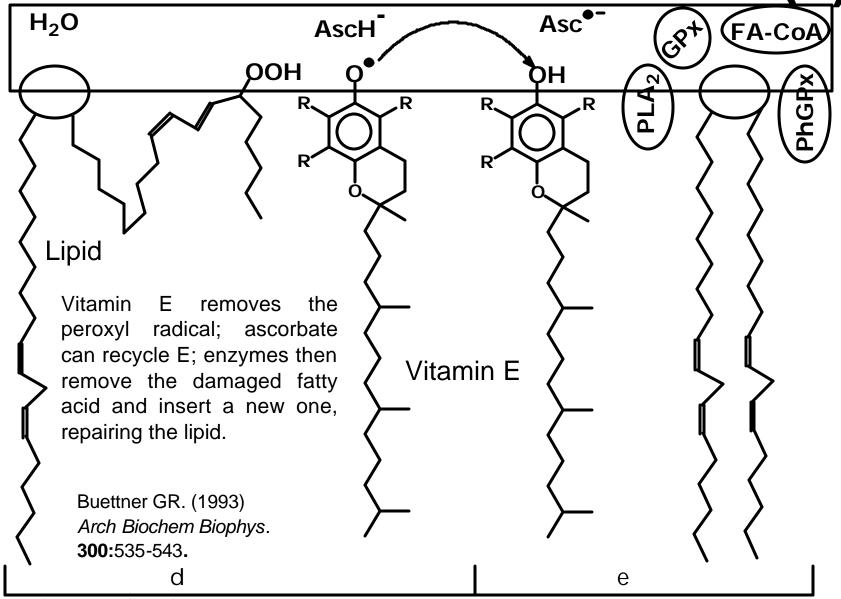
#### TO + AscH ® TOH + Asc · ·

This mechanism is clearly important in protecting LDL from unwanted oxidations, because LDL lacks enzymes that could recycle TO. But its importance in cells and tissues is still being debated.

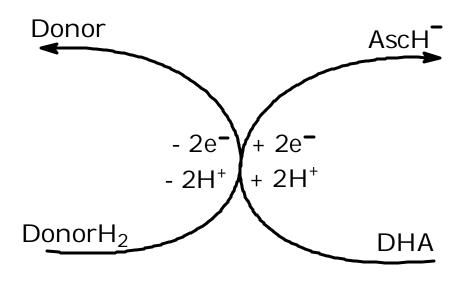
# C and E as Co-Antioxidants (1)



C and E as Co-Antioxidants (2)



## Recycling of Ascorbate



The recycling of ascorbate appears to be an enzymedependent process. The two electrons required can come from GSH.

May JM, Qu Z. Li X, (2001) Requirement for GSH in **recycling** of ascorbic acid in endothelial cells. Biochemical Pharmacology. 62(7):873-81.

Vethanayagam JG, Green EH, Rose RC, Bode AM. (1999) Glutathione-dependent ascorbate recycling activity of rat serum albumin. Free Radical Biology & Medicine. 26:1591-8.

Mendiratta S, Qu ZC, May JM. (1998) Enzyme-dependent ascorbate recycling in human erythrocytes: role of thioredoxin reductase. Free Radical Biology & Medicine. 25:221-8.

## **Ascorbate, Summary**

Ascorbate is a versatile, water soluble, donor, antioxidant.

Thermodynamically, it can be considered to be the terminal small molecule antioxidant.

$$OH$$
 $OOO$ 
 $OOO$