

The background features a complex, light blue illustration of the CRISPR-Cas system. It includes a central DNA double helix, various Cas proteins (RuvC, REC2, PI, REC1, sgRNA), and a target DNA sequence. The entire scene is set against a grid pattern.

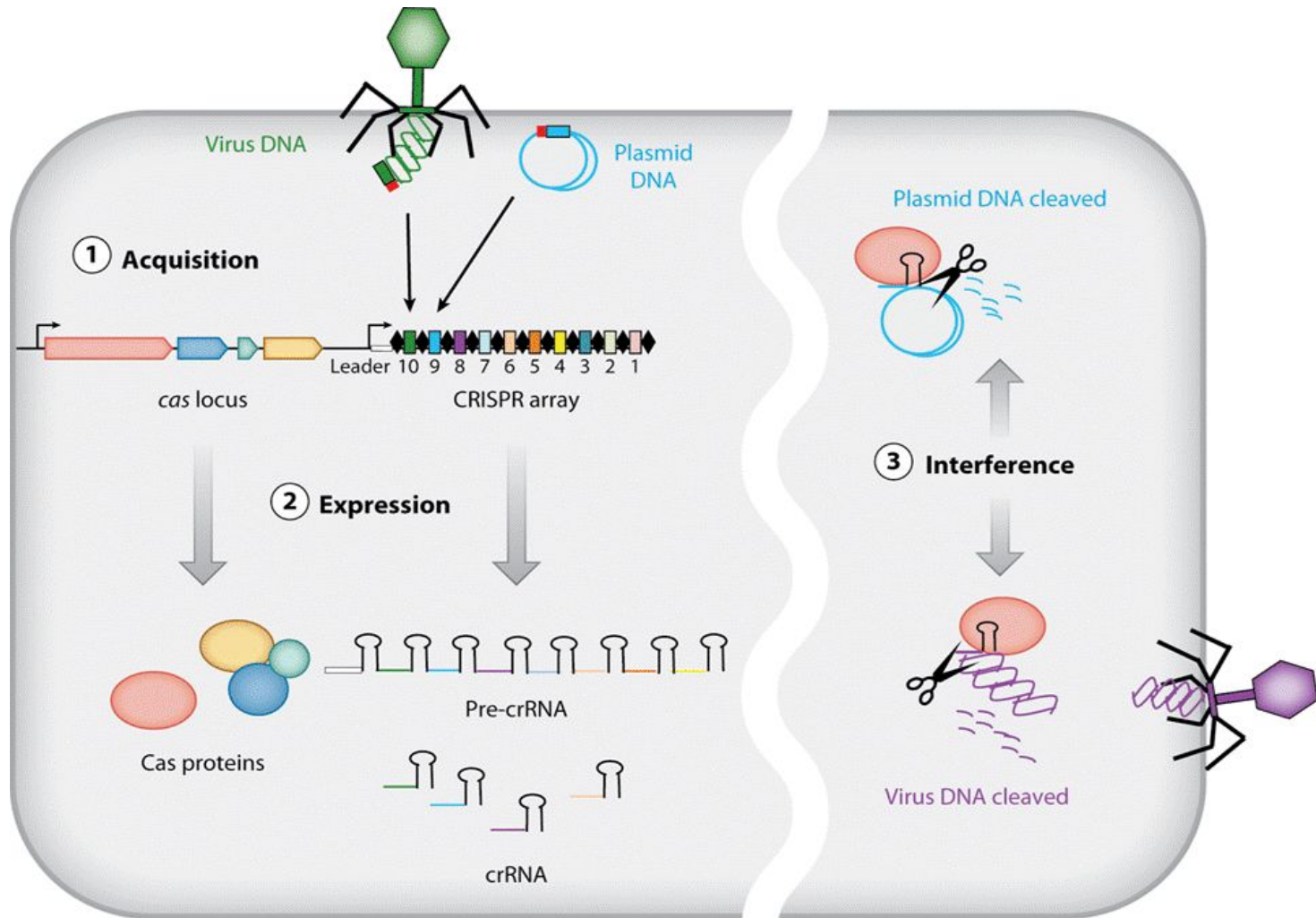
Identification of Conserved Regions in CRISPR Protein Family

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Introduction

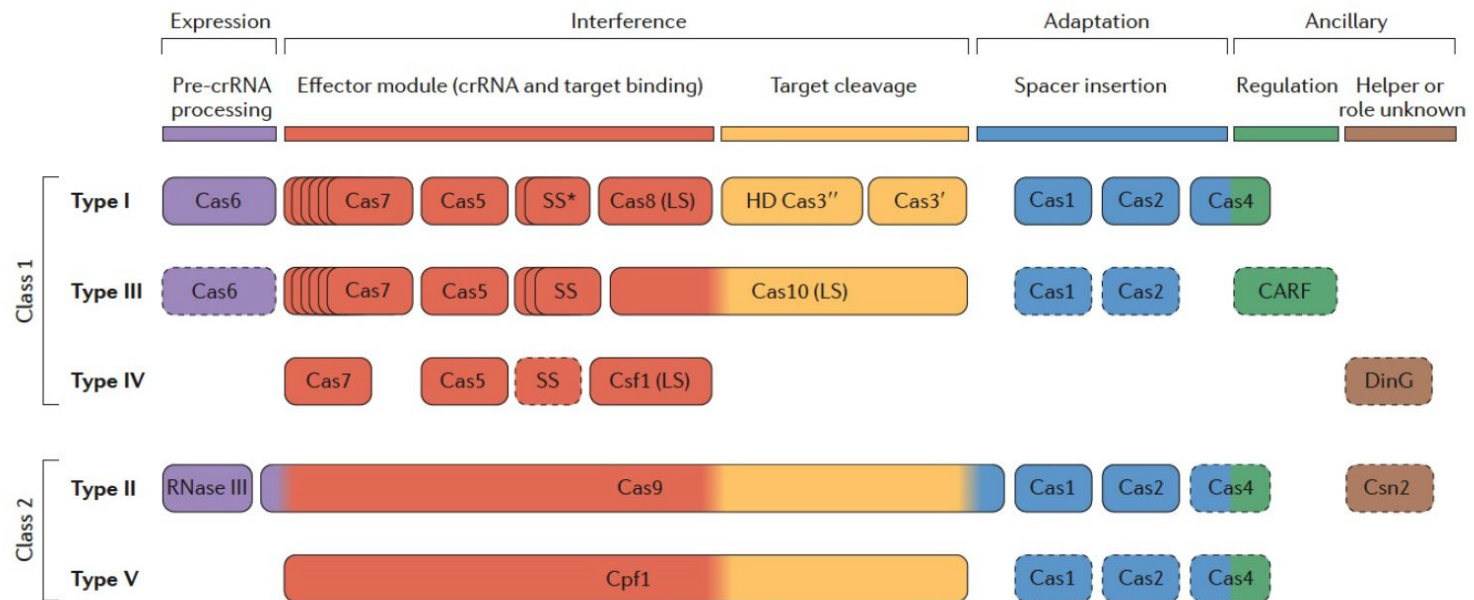
- CRISPR/Cas is virtually used by all kinds of bioengineering applications that benefit from sequence-level recognition
 - Genome Editing
 - In vitro (for transfection and genome editing)
 - In vivo (for studies and therapeutics)
 - Localization studies
 - Much more
- All of above is based on a single *Cas* protein, out of many possible candidates : *Cas9*
 - Initially chosen for its relative simplicity (single protein required for application)
- Much of current publication is focused on applications of *Cas9*, rather than the CRISPR/Cas system itself
 - Size of *Cas9*, as well as its PAM can be limiting for applications
- We attempt 3 different modeling approaches to identify conserved regions and motifs in different CRISPR/Cas protein, to broaden potential *Cas* proteins that may be used for future applications (and cuz, science)

CRISPR/Cas9



Goal of Project

- To identify conserved regions shared by Cas9 and other Cas proteins



Three Methods for Motif Identification

- Pairwise Sequence Alignment
- Gibbs Sampling
- Profile HMM

Data (Input)

- *Cas1* through *Cas10*
- Various species for each
- NCBI
- Amino Acid sequences used
 - Much more useful for identifying conserved regions in distantly related, divergent sequences

Method - Sequence Alignment

- Overview : compare 2 sequences, and identify which regions they are the most similar (and likely to have common ancestors)
- Alignment Methods
 - Semi-Global Alignment (Needleman-Wunsch)
 - Maps entire query sequence to reference sequence, with assumption that query sequence is shorter than the reference (begins later, ends earlier)
 - Local Alignment (Smith-Waterman)
 - Identifies only the highest similar regions between the query and reference sequence
- Parameters
 - BLAST default parameters (BLOSUM62, end gap, affine gap, and gap extension penalty)
- Considers biophysical properties of amino acids into account
 - Discrimination between different types of substitution, such as I→L vs I→W

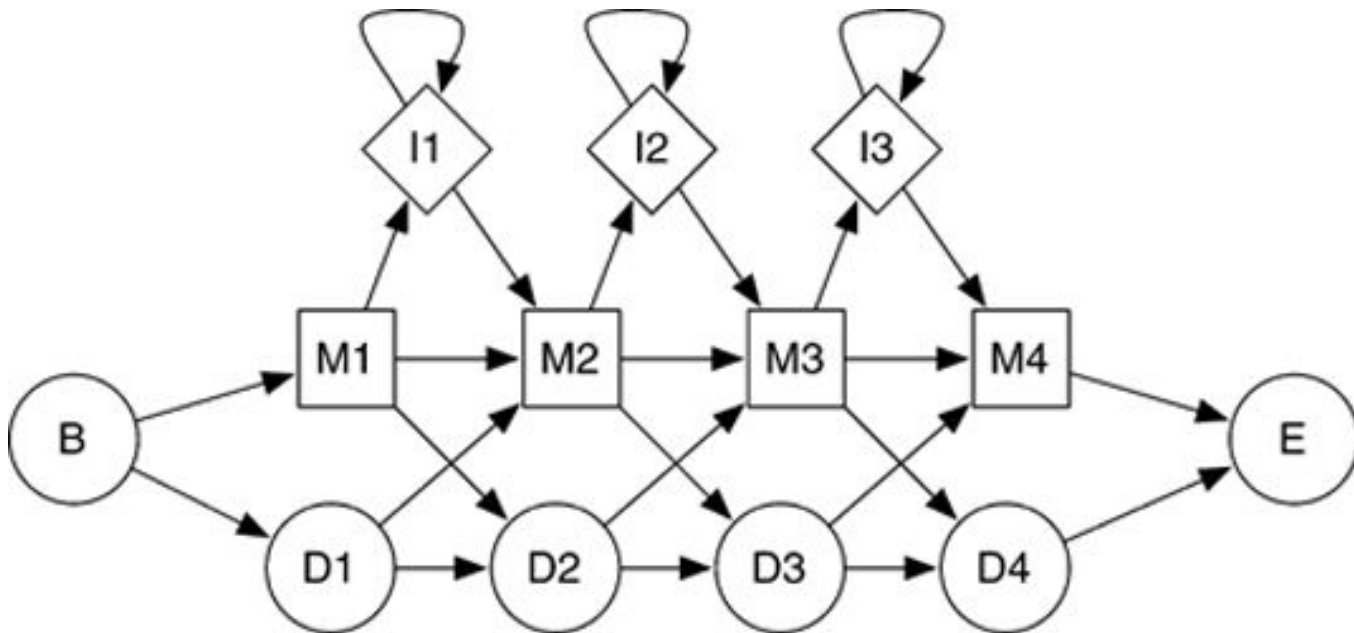
Method - Gibbs Sampling



- Method overview
 - Step 1: find motif in a sequence collection
 - Step 2: Recognize the motif in another collection
 - If the recognition succeeds, it helps to locate the conserved regions between the two sequence collections

Method - HMM

- profile-HMM
- Built for each marked domain in Cas5/9



source:<http://hmmer.org/>

Results

Results - Sequence Alignment

Individual *Cas* sequences mapped to *Cas9*

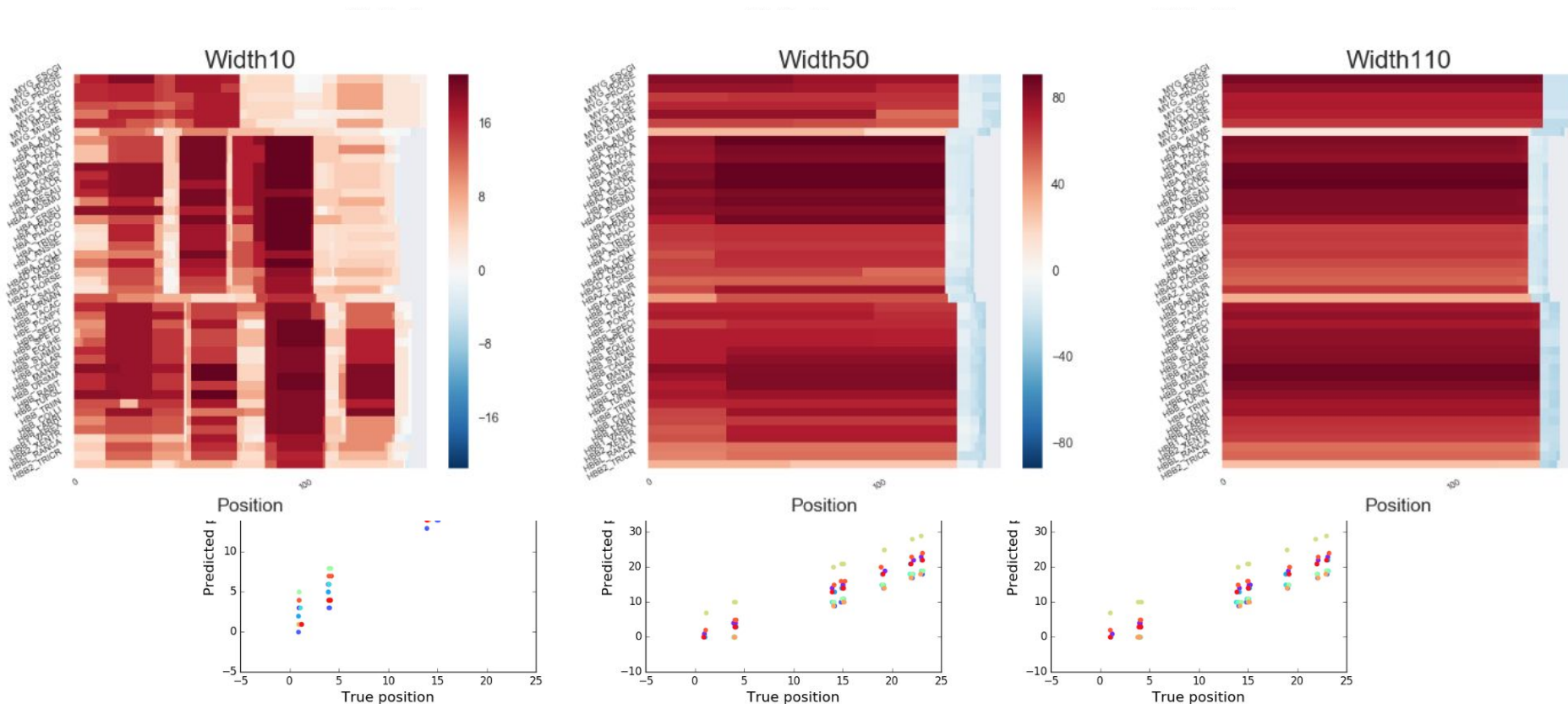
Darker color means higher sequence similarity (higher avg score per position)

Purple : local alignment, Green = semi-global alignment



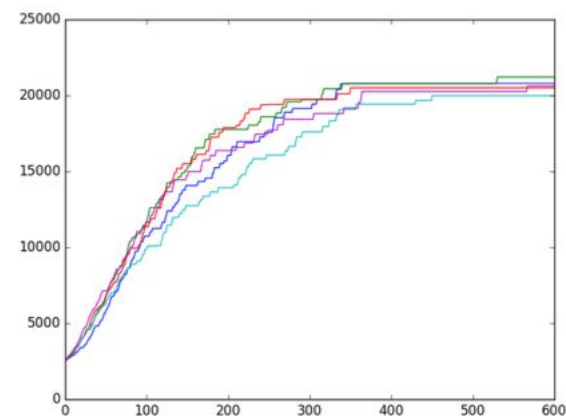
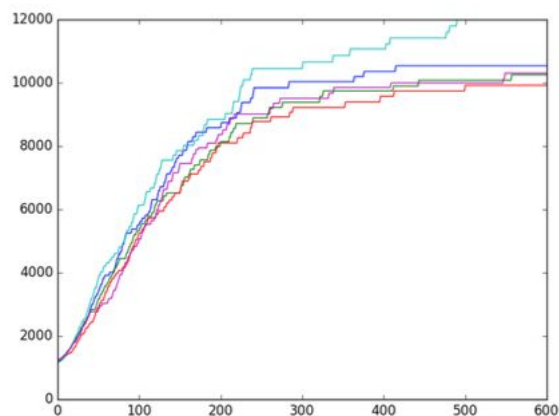
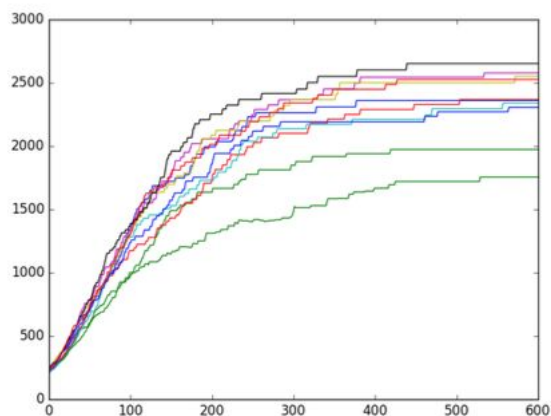
Results - Gibbs Sampling

- Simulated data & globin data (positive control)
 - Robust to motif width
 - Capable to recover the position of the motif



Results - Gibbs Sampling

- Cas 5 vs. Cas 9 - training & recognition

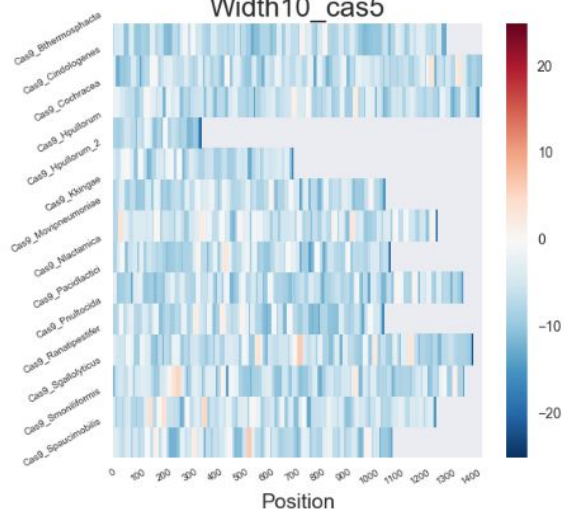


Width = 10

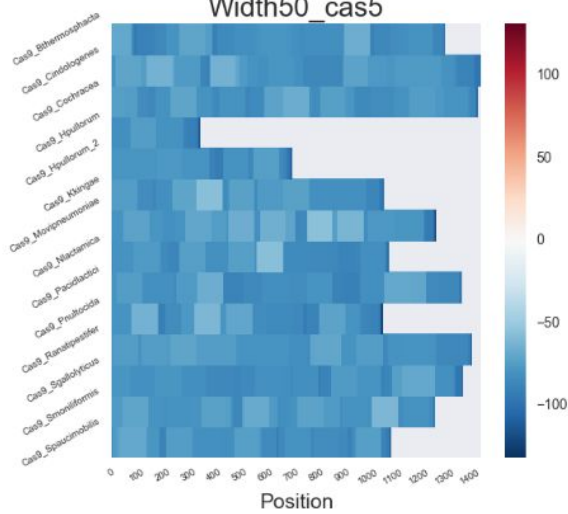
Width = 50

Width = 110

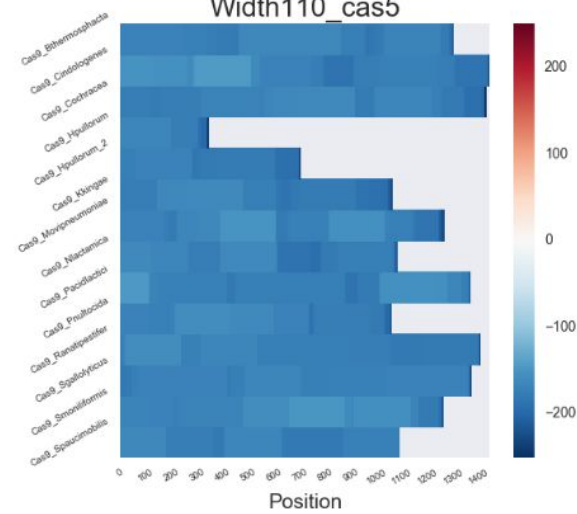
Width10_cas5



Width50_cas5



Width110_cas5



Results - HMM

Search for domain IPR003615 (from *Cas9*) in all collected *Cas* family protein

- Can find similar domains but only in other *Cas9*

```
# hmmsearch :: search profile(s) against a sequence database
# HMMER 3.1b2 (February 2015); http://hmmer.org/
# Copyright (C) 2015 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
# - - - - -
# query HMM file:                IPR003615.hmm
# target sequence database:      ../../../../genes/complete_amino_acids.fa
# - - - - -
```

```
Query:      IPR003615  [M=50]
Scores for complete sequences (score includes all domains):
--- full sequence ---   --- best 1 domain ---   -#dom-
E-value  score  bias    E-value  score  bias    exp  N  Sequence                                Description
-----  -
7.4e-31   97.3   4.4      2e-30   95.9   4.4      1.8  1  Cas9_Sgallolyticus.fasta_1
1.1e-25   80.7   3.5      2.7e-25  79.5   3.5      1.7  1  Cas9_Pacidlactici.fasta_1
2e-23     73.5   1.0      5.7e-23  72.1   1.0      1.9  1  Cas9_Bthermosphacta.fasta_1
5.7e-19   59.3   0.1      1.8e-18  57.6   0.1      2.0  1  Cas9_Cindologenes.fasta_1
8.9e-16   49.0   0.0      2.2e-15  47.8   0.0      1.7  1  Cas9_Pnultocida.fasta_1
```

Overall Results

- Sequence alignment performs well on evolutionarily close sequence, but performs poorly on highly divergent or short sequences
- Gibbs sampling approach is not capable to find conserved region between *Cas5/7* and *Cas9*, even if it is sensitive enough in positive control (simulated & globin)
- Profile HMM can find conserved domains in the same protein as the source in different species but cannot find similar domain in different proteins in the *Cas* family

The fact that this protein family is highly divergent through a long evolutionary timeframe (prokaryote), with limited data affected performance of each model

Conclusion

- More data is needed
- Input sequences represent billions of years in evolutionary distance (compared to Globin protein family of eukaryote)
- Some success with relatively closely related sequences, but divergent sequences showed limited success in identification of conserved regions
- CRISPR/Cas is far too divergent for a single method, especially if verified on eukaryotic evolutionary relationship, to work with direct application
- Future directions
 - Reduce size of alphabet
 - Add structural information into motif model
 - Obtain and test with more data
 - Develop models specifically designed for distantly related/divergent protein families

Questions