Summarization and Bivariate Analysis Week 4

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0.1 Recap

Last week: Data Management and Manipulation

Today: Basic Descriptive Analysis

0.2 Understanding Your Data

- You must understand your data before you can perform an analysis
- Often, the data arrangement determines the analysis plan
- Poor preparation leads to wasted time

0.3 Simple Data Analysis



0.4 Packages

- gmodels
- stats
- gtsummary
- e1071
- ggplot2
- car

0.5 Special Function for NA

To identify missing values, R typically does not interpret variable == NA properly. In order to identify NA values, the function is is.na(variable.

To identify *present* values (i.e. non-NA values), you have to use the *NOT* operator !. !is.na(variable)

0.6 MIMIC Data Usage

```
df.dxwide <- df.dx %>%
  pivot_wider(
    id_cols = c(subject_id, stay_id),
    names_from = seq_num,
    values_from = c(icd_code, icd_version, icd_title)
)

df.ed <- df.ed %>% mutate(hlos = outtime-intime)

df.ed <- df.ed %>% mutate(across(c("gender", "race"), as.factor))

df.triage <- df.triage %>% mutate(fever = if_else(temperature >= 100.4, 1, 0))
```

0.7 Categorical Data

- Usually represented as frequencies and percentages
- Chi-square tests used to evaluate two categorical variables
- Fisher's exact tests for small sample sizes
- Any number of categories can be evaluated

0.8 Basic Code

0.8.1 Frequencies Crosstab and Chi-square Test

```
library(stats)
library(gmodels)
crosstab1 <- table(df$var1, df$var2, useNA = "ifany")
CrossTable(crosstab1)</pre>
```

```
chisq.test(df$var1, df$var2, correct=FALSE)
```

- var1 and var2 should be categorical variables. var1 will be the *rows* and var2 will be the *columns* of the table
- useNA = "ifany" will add missing values to the frequency table if they exist. Other options include "no" and "always"
- correct = FALSE applies a continuity correction for use when a categorical variable was derived from a continuous variable

0.9 Example w/ MIMIC Data

```
# Dichotomizing Race to White/Non-White
df.ed <- df.ed %>%
  mutate(race_white = case_when(
    race == "WHITE" ~ 1,
    race == "WHITE - BRAZILIAN" ~ 1,
    race == "WHITE - OTHER EUROPEAN" ~ 1,
    .default = 0
))
library(stats)
library(gmodels)
table(df.ed$race_white, df.ed$gender, useNA = "ifany")
```

```
F M
0 64 16
1 58 84
```

```
chisq.test(df.ed$race_white, df.ed$gender, correct=FALSE)
```

```
Pearson's Chi-squared test

data: df.ed$race_white and df.ed$gender

X-squared = 31.692, df = 1, p-value = 1.807e-08
```

Note: A warning will be given when cell sizes are small but the analysis will proceed nonetheless.

0.10 CrossTable Command from library(gmodels)

```
library(gmodels)
CrossTable(x, y, ...)
```

x refers to your row object

y refers to your column object

By default, CrossTable will only output frequencies and percentages.

To perform a statistical analysis, you must specify:

- chisq = TRUE for standard chi-square test
- fisher = TRUE for Fisher's exact test for low cell sizes
- mcnemar = TRUE for paired data

0.11 CrossTable Example

```
library(gmodels)
CrossTable(df.ed$race_white, df.ed$gender, chisq = TRUE)
```

Total Observations in Table: 222

I	df.ed\$gende	r	
<pre>df.ed\$race_white </pre>	F	M	Row Total
0	64 l	16	80
I	9.131	11.140	l I
I	0.800	0.200	0.360
I	0.525	0.160	l I
I	0.288	0.072	l l
1	58	84	142
I	5.144	6.276	l l
I	0.408	0.592	0.640
I	0.475	0.840	l l
I	0.261	0.378	l l
Column Total	122	100	222
I	0.550	0.450	l l

Statistics for All Table Factors

0.12 Bivariate Data Analysis With Continuous Variables

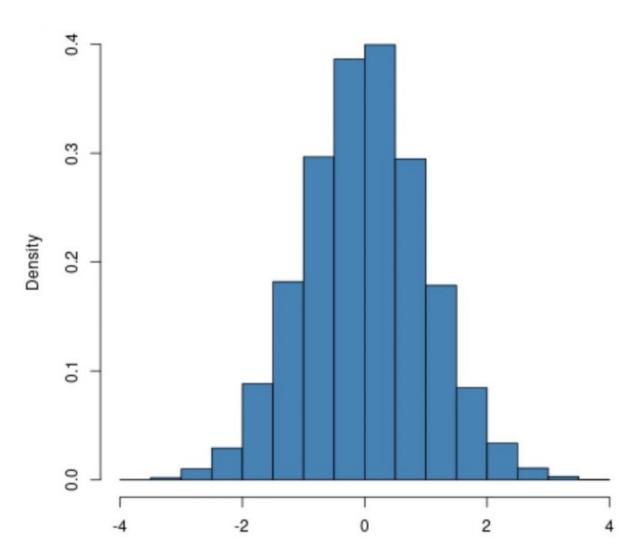
$Dependent\ Variable = Continuous$

- Independent Variable = Continuous
- Pearson correlation

- Spearman correlation
- Independent Variable = Categorical
- t-test
- ANOVA
- Mann-Whitney U (Rank Sum)
- Kruskal-Wallis Test

0.13 Behold, the Distribution!

Normal Distribution



Distributional assumptions of normality must be evaluated!

1 Basic Analysis

1.1 Single Variable Univariate Statistics

Using tidyverse and the summarise() command, we can output summary statistics of continuous variables

```
df.ed %>%
    summarise(
    obs = n(),
    min = min(hlos),
    max = max(hlos),
    mean = mean(hlos),
    sd = sd(hlos),
    median = median(hlos),
    p25 = quantile(hlos, probs = 0.25),
    p75 = quantile(hlos, probs = 0.75)
)
```

```
obs min max mean sd median p25 1 222 4 mins 4460 mins 485.7803 mins 509.7573 350.5 mins 253.25 mins p75 1 526.0875 mins
```

1.2 Tidyverse Scoped Verbs

Tidyverse can incorporate "scoped verbs" to operate on multiple variables in one block of code. These include across(), pick(), and a modified summarise_all() to apply a function across a selection/all variables in a data frame.

```
df.ed %>%
  select(hlos) %>%
  summarise_all(list(
    n = length,
    min = min,
    max = max,
    mean = mean,
    sd = sd,
    median = median,
    p25 = ~quantile(., probs = 0.25),
    p75 = ~quantile(., probs = 0.75)
    ))
```

```
n min max mean sd median p25
1 222 4 mins 4460 mins 485.7803 mins 509.7573 350.5 mins 253.25 mins
p75
1 526.0875 mins
```

Note that the quantile call is a function that can be used for any value, so it requires a ~ operator to tag it as a function. This comes from the purrr package within tidyverse that contains additional programming shorthand tools.

1.3 Many univariates at once

```
df.ed %>%
  select(hlos, intime, outtime) %>%
   summarise_all(list(
    n = length,
    min = min,
   max = max,
   mean = mean,
   sd = sd,
   median = median,
   p25 = ~quantile(., probs = 0.25),
   p75 = ~quantile(., probs = 0.75)
   ))
```

```
hlos_n intime_n outtime_n hlos_min
                                              intime_min
                                                                  outtime_min
    222
              222
                        222
                              4 mins 2112-09-17 18:46:00 2112-09-17 19:50:00
  hlos_max
                                                         hlos_mean
                     intime_max
                                        outtime_max
1 4460 mins 2201-10-30 10:48:00 2201-10-30 12:25:00 485.7803 mins
          intime_mean
                             outtime_mean hlos_sd intime_sd outtime_sd
1 2157-09-10 17:55:54 2157-09-11 02:01:41 509.7573 700146546 700150031
 hlos_median
                    intime_median
                                       outtime_median
                                                          hlos_p25
1 350.5 mins 2150-03-08 17:20:00 2150-03-08 23:59:54 253.25 mins
           intime_p25
                              outtime_p25
                                               hlos_p75
                                                                  intime_p75
1 2142-05-15 16:41:45 2142-05-15 23:50:30 526.0875 mins 2177-11-24 14:14:00
          outtime_p75
1 2177-11-24 19:51:10
```

Note: summarise and summarise_all will not work if missing values are present. A straight tidyverse solution requires advanced programming.

Characteristic	$ m N=222^{\it 1}$
temperature	98.10 (97.60, 98.50)
Unknown	26
heartrate	90 (77, 104)
Unknown	24
resprate	18 (16, 18)
Unknown	23
o2sat	98 (97, 100)
Unknown	24
sbp	136 (116, 155)
Unknown	$23^{'}$
dbp	72 (62, 83)
Unknown	23

 $[\]overline{^{1}\text{Median (Q1, Q3)}}$

1.4 gtsummary for quick statistics

```
library(gtsummary)

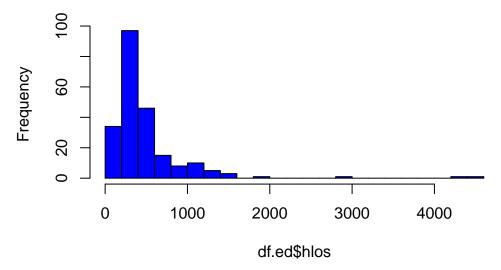
df.triage %>%
   select(temperature:dbp) %>%
   tbl_summary()
```

2 Checking Normality Visually

2.1 via Histograms

```
df.ed$hlos <- as.numeric(df.ed$hlos)
hist(df.ed$hlos, col = "blue", breaks = 30)</pre>
```

Histogram of df.ed\$hlos



Widely used method

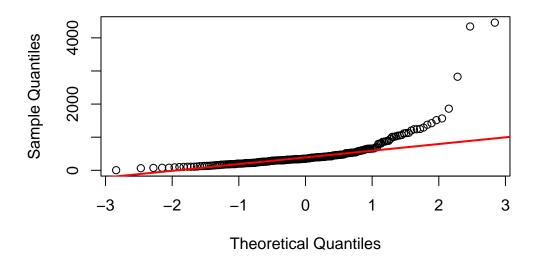
If the histogram is bell-shaped and symmetrical, it suggests normality

Dependent on the number of bins (breaks) and can be subjective

2.2 via Q-Q plots

```
qqnorm(df.ed$hlos, pch = 1, main = "Quantile-Quantile Plot of HLOS")
qqline(df.ed$hlos, col = "red", lwd = 2)
```

Quantile-Quantile Plot of HLOS



Points should fall close to a straight diagonal line

Deviations from the line suggest departures from normality

- Points curving upwards: skewed to the right (positive skew)
- Points curving downwards: skewed to the left (negative skew)
- Points spread out more than the line: flatter than normal distribution
- Points bunched together closer than the line: heavier tails than normal distribution

3 Checking Normality Numerically

3.1 Shapiro-Wilk

```
library(stats)

swt <- shapiro.test(df.ed$hlos)
print(swt)</pre>
```

Shapiro-Wilk normality test

```
data: df.ed$hlos
W = 0.56083, p-value < 2.2e-16
```

Good for smaller samples

A p-value greater than 0.05 indicates that the data does not deviate significantly from the normal distribution

3.2 Kolmogorov-Smirnov

```
library(stats)
kst <- ks.test(df.ed$hlos, "pnorm")</pre>
```

Warning in ks.test.default(df.ed\$hlos, "pnorm"): ties should not be present for the one-sample Kolmogorov-Smirnov test

```
print(kst)
```

Asymptotic one-sample Kolmogorov-Smirnov test

data: df.ed\$hlos
D = 0.99997, p-value < 2.2e-16
alternative hypothesis: two-sided</pre>

Good for larger samples

A p-value greater than 0.05 indicates that the data does not deviate significantly from the normal distribution

3.3 Skewness & Kurtosis

```
library(e1071)

skewness <- skewness(df.ed$hlos)
kurtosis <- kurtosis(df.ed$hlos)

print(c("Skewness:", skewness, "Kurtosis:", kurtosis))</pre>
```

[1] "Skewness:" "4.81774459323537" "Kurtosis:" "31.1513455581506"

Kind of arbitrary, but these assess the asymmetry and tail heaviness of a distribution Skewness:

- Absolute values close to 0 suggest symmetry
- Positive values indicate a right-skewed distribution
- Negative values indicate a left-skewed distribution

Kurtosis:

- A value of 3 indicates a normal distribution
- Values greater than 3 indicate heavier tails (leptokurtic)
- Values less than 3 indicate lighter tails (platykurtic)

4 Bivariate Analyses

4.1 Two continuous variables (1 dependent, 1 independent)

4.2 Scatter Plots

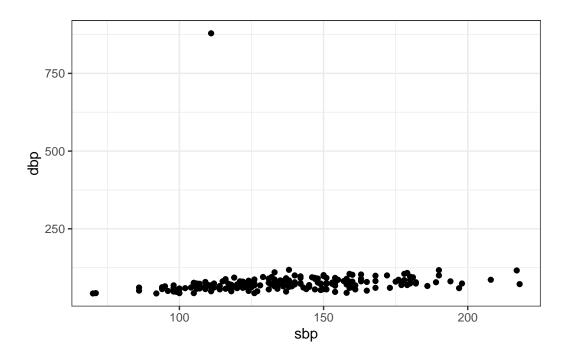
Two-variable scatterplots can be generated with ggplot2 within the tidyverse

This is a highly customizable package that will be discussed in more detail later.

```
library(ggplot2)

# Simple Plots
ggplot(df.triage, aes(sbp, dbp)) +
    geom_point() +
    theme_bw()
```

Warning: Removed 23 rows containing missing values or values outside the scale range (`geom_point()`).

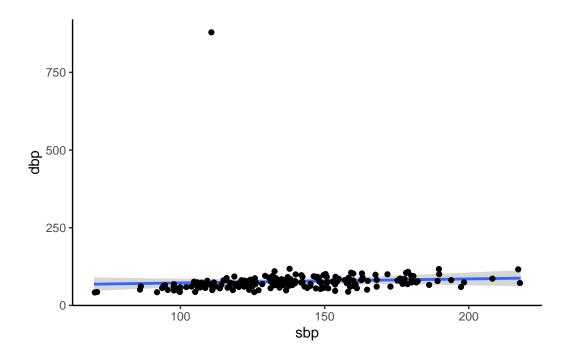


```
# With Regression Lines
ggplot(df.triage, aes(sbp, dbp)) +
  geom_smooth(method = "lm") +
  geom_jitter() +
  theme_classic()
```

`geom_smooth()` using formula = 'y ~ x'

Warning: Removed 23 rows containing non-finite outside the scale range (`stat_smooth()`).

Removed 23 rows containing missing values or values outside the scale range (`geom_point()`).



4.3 Pearson Correlation

```
# Calculate correlations
cor.test(df.triage$temperature, df.triage$sbp, method = "pearson")
```

Pearson's product-moment correlation

```
cor.test(~ temperature + sbp, data = df.triage, method = "pearson")
```

Pearson's product-moment correlation

the parametric method

The correlation_coefficient value will be between -1 and 1. - 1: Perfect positive correlation (variables increase/decrease together). - 0: No linear correlation (no relationship between variables). - -1: Perfect negative correlation (variables increase/decrease in opposite directions).

Correlation strength: - 0.0 to 0.2: Very weak or negligible - 0.2 to 0.4: Weak - 0.4 to 0.6: Moderate - 0.6 to 0.8: Strong - 0.8 to 1.0: Very strong

The p-value indicates the probability of observing the correlation by chance. Smaller p-values (usually < 0.05) suggest a statistically significant correlation.

4.4 Spearman Correlation

```
cor.test(~ temperature + sbp, data = df.triage, method = "spearman")
```

Warning in cor.test.default(x = mf[[1L]], y = mf[[2L]], ...): Cannot compute exact p-value with ties

Spearman's rank correlation rho

The non-parametric method

It is suitable for measuring associations between variables measured on an ordinal scale (e.g., rankings, grades, levels).

It reflects the strength and direction of monotonic relationships, even if they are not perfectly linear.

Rho values range from -1 (perfect negative correlation) to 1 (perfect positive correlation), with 0 indicating no correlation.

the p-value indicates the statistical significance of the correlation

4.5 Two variables (1 continuous dependent, 1 categorical independent)

4.6 Summary Statistics

gtsummary will provide the easiest and fastest method for quick bivariate statistics

```
library(gtsummary)

df <- left_join(df.ed, df.triage, by = "stay_id")

df %>%
    select(race_white, temperature:dbp) %>%
    tbl_summary(by = "race_white")
```

Characteristic	$0 \text{ N} = 80^{1}$	$1 \text{ N} = 142^{1}$
temperature	98.00 (97.40, 98.20)	98.20 (97.70, 98.70)
Unknown	13	13
heartrate	84 (72, 96)	96 (80, 106)
Unknown	11	13
resprate	18 (16, 18)	18 (16, 20)
Unknown	11	12
o2sat	99 (97, 100)	98 (96, 99)
Unknown	11	13
sbp	142 (121, 163)	133 (116, 151)
Unknown	11	12
dbp	74 (61, 82)	72(62, 83)
Unknown	11	12

 $[\]overline{^{1}\text{Median}}$ (Q1, Q3)

4.7 One-sample T-Test

```
oneSamp.res <- t.test(df$age, mu = 75)
oneSamp.res</pre>
```

4.8 Two Samples

Equality of Variances - Levene's Test

```
library(car)
lev.res1 <- leveneTest(age ~ inptDeath, data = df)
lev.res1
lev.res2 <- leveneTest(age ~ gender, data = df)
lev.res2</pre>
```

4.9 T-test for two groups

```
age.ttest.res <- t.test(age ~ inptDeath, data = df)
age.ttest.res
gend.ttest.res <- t.test(age ~ gender, data = df)
gend.ttest.res</pre>
```

4.10 Two-samples T-test

Requiring two separate dataframes with the same variables.

4.10.1 Unpaired Equal Variance

```
# subset to get our "two samples"
df.white <- subset(df, race_white == 1)
df.nonwhite <- subset(df, race_white == 0)

# Welch's T-Test, equal variance assumed
twoSampEq.res <- t.test(df.white$hlos, df.nonwhite$hlos, paired = FALSE)
twoSampEq.res</pre>
```

```
Welch Two Sample t-test

data: df.white$hlos and df.nonwhite$hlos

t = -1.0625, df = 131.26, p-value = 0.29

alternative hypothesis: true difference in means is not equal to 0

95 percent confidence interval:
   -233.30843   70.25974

sample estimates:
mean of x mean of y

456.4021   537.9265
```

4.10.2 Unpaired Unequal Variance

```
# Welch's T-Test, unequal variance
twoSampUnEq.res <- t.test(df.white$hlos, df.nonwhite$hlos, paired = FALSE, var.equal = FALSE
twoSampUnEq.res</pre>
```

Welch Two Sample t-test

```
data: df.white$hlos and df.nonwhite$hlos
t = -1.0625, df = 131.26, p-value = 0.29
alternative hypothesis: true difference in means is not equal to 0
95 percent confidence interval:
   -233.30843    70.25974
sample estimates:
mean of x mean of y
   456.4021    537.9265
```

4.11 Paired t-test

Requiring one data frame with two columns to be compared.

```
# Paired t-test
paired.res <- t.test(df$baselineMeasure, df$followupMeasure, paired = TRUE)
paired.res</pre>
```

5 Analysis of Continuous Data w/ Polychotomous Categorical

5.1 ANOVA

5.2 Equality of Variance Test

```
# Bartlett's Test
age.bartl.res <- bartlett.test(age ~ admission_type, data = df)
age.bartl.res</pre>
```

age is set as the "dependent variable" and is continuous admission_type is the "independent variable" and is categorical w/ more than 2 categories.

5.3 One-way ANOVA

```
one.way <- aov(hlos ~ admission_type, data = df)
summary(one.way)
head(one.way)</pre>
```

5.4 Two-way ANOVA [Multivariate!]

```
two.way <- aov(hlos ~ admission_type + race, data = df)
summary(two.way)</pre>
```

5.5 Test of Medians

```
# Mann-Whitney U / Wilcoxon Rank Sum Test
    # two groups
wilcox <- wilcox.test(hlos ~ race, data = df)
wilcox
# Kruskal-Wallis Test
    # more than two groups
krusk <- kruskal.test(hlos ~ admission_type, data = df)
krusk</pre>
```