



CHAPTER 12

Infiltrations of PRGF to Treat Ligament and Tendon Injuries in the Hip and Pelvis

AUTHORS

Kraeutler MJ.¹, Garabekyan T.², Mei-Dan O.¹

¹ University of Colorado School of Medicine, Department of Orthopedics, Aurora CO 80045

² Southern California Hip Institute, North Hollywood CA 91602

SUMMARY

In recent years, platelet-rich plasma (PRP) has gained popularity within the orthopaedic community as a treatment modality to enhance tissue healing. This chapter aims to concisely present the current indications for PRP injections in the treatment of hip and pelvic pathologies and to describe some novel applications for PRP which have not yet been reported in the literature. With regard to hip and pelvic pathologies, PRP injections are used most commonly as a non-operative intervention, and have been described in the literature to treat osteoarthritis of the hip joint as well as tendinopathy of the hamstrings, adductor longus, and gluteus medius. In contrast, most of the surgical applications of PRP for the hip are

novel, with few reported studies in the literature. Because of the increasing awareness of PRP's beneficial effects on musculoskeletal healing and thus the growing number of indications for its use, this review also describes some novel applications for PRP, including osteitis pubis, post-microfracture of the hip, tears of the rectus femoris, and avulsion of the sartorius muscle.

1. INTRODUCTION

Platelet-rich plasma (PRP) has gained popularity within the last decade among the orthopaedic community as a treatment modality to enhance tissue healing. The term platelet-rich plasma may be applied to any fraction of autologous blood that contains a higher concentration of platelets than baseline¹. Thus, this term is non-specific and factors such as the concentration of platelets and leukocytes as well as centrifugation methods have differed between studies. DeLong et al² developed the PAW classification system to aid in comparing different protocols of PRP preparation. This classification system is based on the absolute number of platelets (P), the method of platelet activation (A), and the presence/absence of white cells (W).

Recently, PRP has been utilized for numerous musculoskeletal indications such as rotator cuff repair^{3,4}, patellar tendinopathy⁵, knee osteoarthritis⁶, lateral epicondylitis⁷, osteochondral lesions of the talus^{8,9}, and many other orthopaedic conditions. PRP induces musculoskeletal healing through a number of effects. As a treatment modality for tendon healing, PRP enhances the mobilization of circulation-derived cells¹⁰. This may include inflammatory cells that secrete cytokines and growth factors as well as fibroblast-like cells that synthesize matrix. Compared to serum, PRP has been shown to significantly increase the deposition of a collagen-rich extracellular matrix¹¹, with higher collagen I content compared to placebo¹². Interestingly, PRP-treated tendon tears have actually been shown to contain fewer blood vessels compared to placebo¹², possibly indicating a more physiological healing process. Once PRP enhances the early phase of regeneration, mechanical stimulation is required to promote organized collagen synthesis and remodeling during new tendon development¹³.

With muscle strains or contusions, the hematoma that originates contains approximately 94% red blood cells, 4% platelets, and < 1% leukocytes¹⁴. Compared to whole blood, PRP contains higher concentrations of certain growth factors, in par-

ticular platelet-derived growth factor (PDGF) and transforming growth factor- β (TGF- β)¹⁵. Thus, PRP is theorized to replace the hematoma with a high concentration of platelets and growth factors to promote healing. Furthermore, PRP has been shown to promote angiogenesis through activation of PRP-releasate (PRP-r)¹⁶. In comparison to tendon and muscle healing, little is known on the mechanisms of PRP in promoting healing of articular cartilage, though this likely involves multiple biological processes including apoptosis, extracellular matrix synthesis, angiogenesis, and inflammation¹⁷. Because of the increasing awareness of PRP's beneficial effects on musculoskeletal healing and thus the growing number of indications for its use, we present a chapter of the current indications for platelet-rich plasma injections to augment the conservative and surgical treatment of hip and pelvic ligament and tendon pathologies and describe some novel applications for PRP¹⁸. Although several studies have described the use of PRP for some of these pathologies, other indications for PRP discussed in this chapter have not been published previously. As such, these indications are not yet evidence-based.

2. DECISION-MAKING

After performing an appropriate patient history and physical examination, advanced imaging is typically obtained to better characterize the suspected pathology. With muscle or tendon tears, magnetic resonance imaging (MRI) or ultrasound (US) should be used to determine the exact location and extent of the injury. Depending on the pathology, PRP may be used as a conservative treatment measure or as adjunctive treatment during surgery. When used as a conservative treatment option, PRP may be applicable for tendinopathic changes or partial tendon tears in which the tendon ends are not retracted¹.

The cost of platelet-rich plasma treatment is certainly a factor in the decision-making process. In-

insurance companies still do not recognize PRP as standard of care, and thus PRP must be paid by patients out of pocket. It has been estimated recently that the cost of PRP is \$500 to \$1,500 per application¹⁹. It is important to have an open discussion regarding the cost of PRP injections, given that it may be prohibitively expensive for some patients.

3. PREPARATION AND APPLICATION TECHNIQUES

A sample of whole blood is collected in a sodium citrate tube in order to delay clotting of the blood sample. Once the whole blood sample is collected, centrifugation allows separation of the sample into its component cells and serum. Either one or two centrifugation steps may be used depending on the final product desired. A "soft" spin separates the whole blood sample into three layers: an upper layer consisting mainly of plasma and platelets, a very thin middle layer known as the "buffy coat" that is highly concentrated in WBCs, and a bottom layer consisting mainly of red blood cells (RBCs)²⁰. A second, "hard" spin may be used to further concentrate the platelets^{20,21}. Following centrifugation, approximately 10% of the initial whole blood volume remains as PRP concentrate¹. The platelets in the PRP concentrate are activated with calcium chloride and/or autologous or bovine thrombin. These additions are used to initiate the clotting cascade, the release of growth factors from the platelets, and the formation of a fibrin scaffold²¹. Autologous thrombin has been shown to have a lower clot strength compared to bovine thrombin or calcium chloride, with bovine thrombin having the quickest clot initiation time²². In an equine model, calcium chloride activation of PRP has been shown to result in greater release of platelet-derived growth factor compared to autologous or bovine thrombin²³. Calcium chloride also provides the advantage of not using bovine or other non-autologous materials.

Therapeutic doses of PRP require 2.5-5 times the baseline concentration of platelets^{24,25}, though higher concentrations than this have an inhibitory effect on healing²⁶. The white blood cell concentration may also be controlled, with leukocyte-rich PRP (L-PRP) and leukocyte-poor PRP (P-PRP) both being used in the literature. For production of L-PRP, the entire layer of the buffy coat and few RBCs are transferred to an empty sterile tube, while the upper layer and only the superficial buffy coat are transferred for production of P-PRP²⁰. Plasma rich in growth factors (PRGF) is a term used to describe a leukocyte-poor PRP which is separated manually (direct visualization using a fine pipette) from the lower fraction of the plasma containing the highest concentration of platelets and growth factors, avoiding the thin WBC layer. PRGF techniques have been shown to produce lower concentrations of growth factors compared to standard PRP kits by 3-4 fold²⁷ which, according to many studies, may serve as the optimal ratio for tissue healing.

When a tendon or muscle is injured, healing proceeds through three processes: inflammation/degeneration, regeneration, and fibrosis¹. Although L-PRP has been shown to contain the highest levels of growth factors and cytokines²⁸, it induces catabolic effects and a significantly greater acute inflammatory response and thus may actually prolong the healing process²⁸⁻³¹. Thus, the inclusion of white cells defeats the purpose of PRP. On the other hand, P-PRP induces mainly anabolic changes, and while this is generally a beneficial outcome, it could also result in scar tissue formation due to these anabolic effects^{28,31}. Still, no randomized or prospective clinical studies have been performed to compare outcomes between leukocyte-rich versus leukocyte-poor PRP.

4. NON-OPERATIVE APPLICATIONS

Platelet-rich plasma injections are used most commonly as an adjunct to conservative treatment. In cases of chronic tendinopathy or osteoarthritis, PRP is typically indicated when first-line treatment (physical therapy, rest) fails. For professional athletes, in-season PRP may be used to reduce pain and improve function as an interim solution until the offseason when the athlete can undergo surgical intervention. This is particularly true for hip labral tears and chronic tendinopathies.

As detailed below, PRP injections have been used to treat tendinopathy of the hamstrings, adductor longus, and gluteus medius. These injections are typically performed under US guidance in the clinic. The number of injections used differs by study, though most reports describe the use of a single PRP injection for a variety of hip pathologies. Prior to injection, patients should fast for a minimum of three hours and should limit water intake to 8 ounces. In addition, patients should avoid the use of non-steroidal anti-inflammatory drugs (NSAIDs) for at least two days prior to and five days following injection, as these medications have been shown to impair platelet function³².

Hamstring tendinopathy

Chronic tendinopathy or partial tears of the proximal hamstring tendons are common injuries among athletes. These injuries often occur while running, particularly when accelerating. Following severe hamstring injuries, many high level athletes may struggle with sitting on a bike or returning to running. Corticosteroid injections should be avoided as they may result in further tendon weakening and progression to high-grade tearing. PRP injections can be used to facilitate healing when there is partial thickness involvement or tendinosis without retraction. Complete tears, especially if chronic and retracted, are best treated with surgical repair.

A Hamid et al³³ conducted a randomized controlled trial to compare PRP therapy plus a rehabilitation program versus rehabilitation alone in patients with acute hamstring injuries (table 1). Patients in the PRP group were given a single intra-lesional injection of PRP, without addition of an activating agent, under US guidance at an average of 4.6 days following injury. Time to return to play was significantly lower in the PRP group (mean 26.7 days) versus the control group (mean 42.5 days). In addition, the PRP group had significantly lower pain severity scores at all time points up to 7 weeks following intervention.

STUDY	LEVEL OF EVIDENCE	PATHOLOGY	CONTROL GROUP(S)	OUTCOMES MEASURED
A Hamid et al, 2014 ³³	II	Acute hamstring injuries	Rehabilitation without PRP	Return to play time, pain severity score, pain interference score
Hamilton et al, 2015 ³⁴	I	Hamstring injuries	Platelet-poor plasma, no injection	Return to play time, reinjury rate
Fader et al, 2015 ³⁵	IV	Chronic proximal hamstring tendinopathy	N/A	VAS pain
Wetzel et al, 2013 ³⁶	III	Proximal hamstring injuries	NSAIDs, physical therapy	VAS, NPRS
Davenport et al, 2015 ³⁷	I	Proximal hamstring tendinopathy	Autologous whole blood	MHHS, Hip Outcome Score-ADL, IHOT-33
Dallaudière et al, 2014 ³⁹	IV	Upper and lower limb tendinopathy	N/A	WOMAC, ultrasound lesion size
Mautner et al, 2013 ⁴²	IV	Chronic tendinopathy	N/A	VAS, assessment of functional pain, overall satisfaction

TABLE 1. CLINICAL STUDIES ON PRP TREATMENT FOR HIP AND PELVIC PATHOLOGIES.

VAS = visual analog scale, NSAIDs = non-steroidal anti-inflammatory drugs, NPRS = Nirschl Phase Rating Scale, MHHS = Modified Harris Hip Score, ADL = activities of daily living, IHOT-33 = International Hip Outcome Tool 33, WOMAC = Western Ontario and McMaster Universities Osteoarthritis Index, OA = osteoarthritis, HA = hyaluronic acid, HHS = Harris Hip Score, AVN = avascular necrosis.

In another randomized controlled trial, Hamilton et al³⁴ compared PRP, platelet-poor plasma (PPP), and no injection in professional athletes with MRI-positive hamstring injuries. All patients underwent a standardized rehabilitation program. Time to return to sport was found to be significantly lower in the PRP group (mean 21 days) compared with the PPP group (mean 27 days). No significant difference in re-injury rate was noted between the three groups at 2 months or 6 months following intervention.

Fader et al³⁵ performed a retrospective case series of 18 patients with chronic proximal hamstring tendinopathy. Each patient received a single PRP injection by US guidance. Patients had chronic hamstring pain symptoms for an average of 32.6 months prior to their injection and all patients had attempted other non-surgical treatments such as cortisone injections and physical therapy prior to injection. Based on a visual analog scale (VAS) for pain, patients had an average improvement in pain of 63% at 6 months following PRP injection.

In another study, Wetzel et al³⁶ performed a retrospective cohort study comparing twelve cases of proximal hamstring injuries treated by a single PRP injection and five patients treated with traditional conservative treatment (TCT) consisting of NSAIDs and physical therapy. Patients in the PRP and TCT groups presented at an average of 9.6 and 7.8 months after injury, respectively. At an average follow-up of 4.5 months, the PRP group demonstrated significantly improved VAS ($p < 0.01$) and Nirschl Phase Rating Scale (NPRS) scores ($p < 0.01$) compared to pre-treatment. At an average follow-up of 2 months, the TCT group did not show the same degree of improvement in VAS ($p = 0.06$) or NPRS scores ($p = 0.06$). However, due to the small sample sizes and differences in follow-up times, it is difficult to discern these outcome differences.

Davenport et al³⁷ conducted a double-blind, randomized controlled trial comparing a single injection of PRP versus autologous whole blood (WB) for the treatment of proximal hamstring tendinopathy. At follow-up times of 2, 6, and 12 weeks and 6 months, no significant differences were ob-

served between groups with regard to the Modified Harris Hip Score, Hip Outcome Scores for activities of daily living (ADL) and sport-specific function, and International Hip Outcome Tool 33 (IHOT-33). However, compared to baseline, the PRP group demonstrated significant improvements in ADL and IHOT-33 scores, whereas the WB group showed significantly decreased pain with 15-minute sitting at 6 months.

Although conflicting results exist, the majority of published studies have reported successful outcomes of PRP injections for hamstring injuries. The authors use PRP injections for acute hamstring injuries including, in rare cases, complete tears with a small amount of retraction in patients with low activity levels or when patients opt out of surgery.

Adductor tendinopathy/athletic pubalgia

Adductor tendinopathy typically presents with groin pain and is often seen in soccer players due to the frequency of running and cutting movements involved in this sport³⁸. Athletic pubalgia is a more general term involving groin pain, often in athletes, with adductor tendinopathy being a frequent concomitant pathology in these patients. Good outcomes have been shown following adductor tenotomy with and without hernioplasty³⁸, though PRP provides a non-operative treatment option for this pathology.

Adductor longus tendinopathy is a common indication for PRP treatment. Dallaudière et al³⁹ performed a retrospective case series of 408 consecutive patients treated by a single ultrasound-guided PRP injection for tendinopathy of upper (medial and lateral epicondylar tendons, i.e. golfer's and tennis elbow, respectively) and lower (patellar, Achilles, hamstring, adductor longus, and peroneal tendons) limbs. Patients with hamstring and adductor longus tendinopathy demonstrated significantly improved Western Ontario and McMaster Universities Osteoarthritis Index (WOMAC) scores at 6 weeks and a mean 20.2 months following injection ($p < 0.001$). Ultrasound was also used to assess lesion size at baseline and 6 weeks post-

injection, with hamstring and adductor longus tendon lesion size decreasing from an average of 21.2 mm to 2.6 mm during this time ($p < 0.001$).

Interestingly, there is a high incidence of athletic pubalgia symptoms in patients with symptomatic femoroacetabular impingement (FAI)⁴⁰. Ham-moud et al⁴⁰ showed that 39% of professional athletes presenting with concomitant AP and FAI experienced symptom resolution with surgical treatment of FAI alone. Larson et al⁴¹ demonstrated a return to unrestricted activity in 89% of patients with surgical management of both athletic pubalgia and intra-articular hip pathologies, compared to only 25% in patients undergoing surgical treatment of athletic pubalgia alone.

The authors inject the origin of the adductor longus (AL) tendon for chronic tendinopathy or acute tears (fig. 1). We first exclude the pubic symphysis as the source of the pain (using a lidocaine test) due to its anatomical proximity to the AL. It is also important to conduct full range of motion (ROM) evaluation of the hip joint to exclude concomitant FAI. FAI results in reduced ROM which in turn places increased stress on the AL origin and may result in chronic microtears and tendinopathy. In these cases, PRGF may not be a beneficial long-term solution as the offending mechanism is still present.

When electing to perform PRGF injections for adductor tendinopathy, it is important to have the patient shave the groin area a few days prior to injection for ease of US guidance. These injections can be very painful, though due to the superficial location of the pathology, lidocaine cannot be used. It is recommended that nitric oxide be used as an inhaled anesthetic if possible.

Gluteus medius tendinopathy

The literature is currently lacking in reported outcomes of PRP injection for gluteus medius tendinopathy. However, in a multicenter, retrospective review of 180 patients with chronic tendinopathy⁴², 16 patients underwent US-guided PRP injections for gluteus medius tendinopathy with 13 patients demonstrating moderate improvement to complete resolution of symptoms at an average follow-up of 15 months post-injection. However, the PRP injection methodology of this study was non-uniform in that 60% of all patients received one injection, 30% received two, and 10% received three or more injections.

The authors use a series of three injections of PRGF for chronic indications⁸ such as gluteus medius or minimus tendinopathy (fig. 2). It is important to



FIG. 1
Ultrasound-guided PRP injection of the adductor longus near its origin on the pubic body.



FIG. 2
Ultrasound-guided PRP injection of the gluteus medius near its insertion on the greater trochanter.

conduct a comprehensive physical examination to determine that the reported pain is not referred from the lower back or hip joint. In some cases, a lidocaine test can help to confirm or exclude the gluteus as the primary pathology. It is instrumental to review a high resolution MRI of the region and to determine pre-injection if gluteal bursitis is causing the patient's pain or if tendinopathic changes/tears are present. It is also important to note which tendons are involved, as the minimus tendon is deeper and thus requires proper US guidance during injection. From our experience, the majority of gluteal tendon tears are in the medial/deeper side of the tendon.

As mentioned with hamstring tendinopathy, many practitioners would use steroid injections for greater trochanteric bursitis. However, this is usually a misdiagnosis or a secondary pathology as gluteal tendon tears are often the primary pain source. Although steroid injections can improve pain for a few months, they can also result in atrophy and weakening of the gluteal tendons.

Sartorius avulsion

The sartorius originates on the anterior superior iliac spine (ASIS) apophysis, which begins to ossify between the ages of 13-15 years and fuses with the ilium between the ages of 21-25 years. An ASIS apophyseal avulsion injury occurs most commonly during running with the hip in extension and knee in flexion, or during a kicking motion⁴³. Pain and a tearing sensation are the most common symptoms.

Recently, a 16 year old male presented to our clinic with anterolateral left hip pain following a lacrosse injury in which the patient made a cut while sprinting and felt and heard a pop around his hip. MRI showed a proximal sartorius avulsion with a few millimeters of distraction and significant soft tissue and bone edema in the surrounding area (fig. 3). The patient underwent US-guided PRGF injection (fig. 4) and reported complete resolution of symptoms within a week. He returned to full activity with our clearance at five weeks post-injection. A similar technique can be used for other apophyseal avulsion injuries such as that of the hamstrings or rectus femoris.

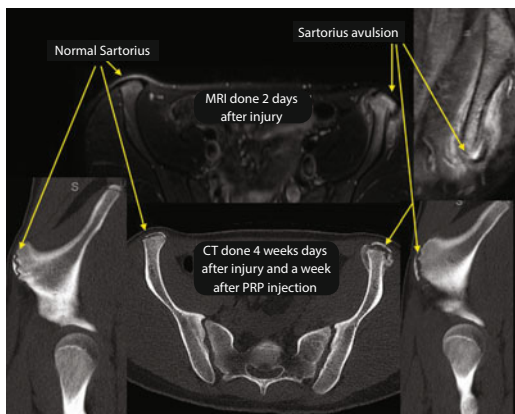


FIG. 3 MRI demonstrating avulsion of the sartorius muscle from its origin on the anterior superior iliac spine (ASIS, top images). CT images comparing a normal sartorius origin on the ASIS (bottom left) and a sartorius avulsion (bottom right).



FIG. 4 Ultrasound-guided PRP injection of Hamstrings near its origin on the anterior superior iliac spine.

5. SURGICAL APPLICATIONS

As described above, when used in a non-operative setting, platelet-rich plasma is applied as a liquid injectable. However, in an intraoperative setting, PRP may be injected as a liquid or gel⁴⁴, or delivered as a fibrin matrix (platelet-rich fibrin matrix, PRFM)⁴⁵. Most of the surgical applications of PRP for hip and pelvic pathology are novel, with few studies currently in the literature.

Adjuvant therapy for tendon repair

Platelet-rich plasma has been shown to improve healing in patients with acute ruptured Achilles tendons^{12,46}. Sánchez et al⁴⁶ showed an earlier recovery of range of motion, return to gentle running, and return to training activities in 12 athletes who underwent open suture repair of a complete Achilles tendon rupture with PRP versus standard open suture repair. Alsousou¹² compared biopsy samples of acute ruptured Achilles tendons treated with PRP versus a control group receiving no treatment. The PRP group demonstrated significantly higher collagen I content and a significantly lower modified Bonar score, which indicates improved early tendon healing.

The authors use PRGF injections in patients undergoing surgical repair of the hamstring tendons when the tendon tissue is found to be of low quality and enhancement of the surgical repair is required. Another common indication for PRGF within our practice is in professional athletes for whom there is a need for an expedited recovery and return to play. This is most commonly indicated for the hamstrings, rectus femoris, or a sartorius avulsion.

6. CONCLUSIONS

This chapter describes some of the established as well as novel applications of platelet-rich plasma or plasma rich in growth factors (PRGF) for the treatment of hip and pelvic pathologies. Although the outcomes of many of these applications have not been described in the literature, particularly in high-level studies, from our experience we have found that symptomatic and functional outcomes are successful in the majority of patients. As indications for PRP continue to expand, it will become increasingly important for future studies to state specific methodologies used in the preparation of PRP in order to recognize ideal preparation techniques and the ideal number of PRP injections for each pathology. Leukocyte-poor PRP has the advantages of a reduced inflammatory response and mainly anabolic changes compared to leukocyte-rich PRP, though further, high quality studies are necessary to determine outcome differences between these two PRP preparations.

1. Mei-Dan O, Carmont MR. Novel applications of platelet-rich plasma technology in musculoskeletal medicine and surgery. *Operative Techniques in Orthopaedics*. 2012;22:56-63.
2. DeLong JM, Russell RP, Mazzocca AD. Platelet-rich plasma: the PAW classification system. *Arthroscopy : the journal of arthroscopic & related surgery : official publication of the Arthroscopy Association of North America and the International Arthroscopy Association*. 2012;28:998-1009.
3. Castricini R, Longo UG, De Benedetto M, et al. Platelet-rich plasma augmentation for arthroscopic rotator cuff repair: a randomized controlled trial. *The American journal of sports medicine*. 2011;39:258-65.
4. Mei-Dan O, Carmont MR. Autologous blood products in rotator cuff repair. *Medicine and sport science*. 2012;57:65-75.
5. Liddle AD, Rodriguez-Merchan EC. Platelet-Rich Plasma in the Treatment of Patellar Tendinopathy: A Systematic Review. *The American journal of sports medicine*. 2015;43:2583-90.
6. Meheux CJ, McCulloch PC, Lintner DM, Varner KE, Harris JD. Efficacy of Intra-articular Platelet-Rich Plasma Injections in Knee Osteoarthritis: A Systematic Review. *Arthroscopy : the journal of arthroscopic & related surgery : official publication of the Arthroscopy Association of North America and the International Arthroscopy Association*. 2016;32:495-505.
7. Mishra AK, Skrepnik NV, Edwards SG, et al. Efficacy of platelet-rich plasma for chronic tennis elbow: a double-blind, prospective, multicenter, randomized controlled trial of 230 patients. *The American journal of sports medicine*. 2014;42:463-71.
8. Mei-Dan O, Carmont MR, Laver L, Mann G, Maffulli N, Nyska M. Platelet-rich plasma or hyaluronate in the management of osteochondral lesions of the talus. *The American journal of sports medicine*. 2012;40:534-41.
9. Kraeutler MJ, Chahla J, Dean CS, et al. Current Concepts Review Update. *Foot & ankle international*. 2017;38:331-42.
10. Kajikawa Y, Morihara T, Sakamoto H, et al. Platelet-rich plasma enhances the initial mobilization of circulation-derived cells for tendon healing. *Journal of cellular physiology*. 2008;215:837-45.
11. Visser LC, Arnoczky SP, Caballero O, Kern A, Ratcliffe A, Gardner KL. Growth factor-rich plasma increases tendon cell proliferation and matrix synthesis on a synthetic scaffold: an in vitro study. *Tissue engineering. Part A*. 2010;16:1021-9.
12. Alsousou J, Thompson M, Harrison P, Willett K, Franklin S. Effect of platelet-rich plasma on healing tissues in acute ruptured Achilles tendon: a human immunohistochemistry study. *Lancet*. 2015;385 Suppl 1:S19.
13. Virchenko O, Aspenberg P. How can one platelet injection after tendon injury lead to a stronger tendon after 4 weeks? Interplay between early regeneration and mechanical stimulation. *Acta orthopaedica*. 2006;77:806-12.
14. Mei-Dan O, Lippi G, Sanchez M, Andia I, Maffulli N. Autologous platelet-rich plasma: a revolution in soft tissue sports injury management? *The Physician and sportsmedicine*. 2010;38:127-35.
15. Lee JW, Kwon OH, Kim TK, et al. Platelet-rich plasma: quantitative assessment of growth factor levels and comparative analysis of activated and inactivated groups. *Archives of plastic surgery*. 2013;40:530-5.
16. Kakudo N, Morimoto N, Kushida S, Ogawa T, Kusumoto K. Platelet-rich plasma releasate promotes

- angiogenesis in vitro and in vivo. *Medical molecular morphology*. 2014;47:83-9.
17. Dallari D, Stagni C, Rani N, et al. Ultrasound-Guided Injection of Platelet-Rich Plasma and Hyaluronic Acid, Separately and in Combination, for Hip Osteoarthritis: A Randomized Controlled Study. *The American journal of sports medicine*. 2016;44:664-71.
 18. Kraeutler MJ, Garabekyan T, Mei-Dan O. The use of platelet-rich plasma to augment conservative and surgical treatment of hip and pelvic disorders. *Muscles, ligaments and tendons journal*. 2016;6:410-19.
 19. Samuelson EM, Odum SM, Fleischli JE. The Cost-Effectiveness of Using Platelet-Rich Plasma During Rotator Cuff Repair: A Markov Model Analysis. *Arthroscopy : the journal of arthroscopic & related surgery : official publication of the Arthroscopy Association of North America and the International Arthroscopy Association*. 2016;32:1237-44.
 20. Dhurat R, Sukesh M. Principles and Methods of Preparation of Platelet-Rich Plasma: A Review and Author's Perspective. *Journal of cutaneous and aesthetic surgery*. 2014;7:189-97.
 21. Arnoczky SP, Sheibani-Rad S. The basic science of platelet-rich plasma (PRP): what clinicians need to know. *Sports medicine and arthroscopy review*. 2013;21:180-5.
 22. Ghassab S, Dulin J, Bertone AL. Thromboelastographic Clot Characteristics of Autologous Equine Blood Products After Activation by Autologous Thrombin, Bovine Thrombin, or Calcium Chloride. *Veterinary surgery : VS*. 2015;44:970-5.
 23. Textor JA, Tablin F. Activation of equine platelet-rich plasma: comparison of methods and characterization of equine autologous thrombin. *Veterinary surgery : VS*. 2012;41:784-94.
 24. Graziani F, Ivanovski S, Cei S, Ducci F, Tonetti M, Gabriele M. The in vitro effect of different PRP concentrations on osteoblasts and fibroblasts. *Clinical oral implants research*. 2006;17:212-9.
 25. Weibrich G, Hansen T, Kleis W, Buch R, Hitzler WE. Effect of platelet concentration in platelet-rich plasma on peri-implant bone regeneration. *Bone*. 2004;34:665-71.
 26. Anitua E, Sanchez M, Nurden AT, Nurden P, Orive G, Andia I. New insights into and novel applications for platelet-rich fibrin therapies. *Trends in biotechnology*. 2006;24:227-34.
 27. Weibrich G, Kleis WK, Hitzler WE, Hafner G. Comparison of the platelet concentrate collection system with the plasma-rich-in-growth-factors kit to produce platelet-rich plasma: a technical report. *The International journal of oral & maxillofacial implants*. 2005;20:118-23.
 28. Cavallo C, Filardo G, Mariani E, et al. Comparison of platelet-rich plasma formulations for cartilage healing: an in vitro study. *The Journal of bone and joint surgery. American volume*. 2014;96:423-9.
 29. Dragoo JL, Braun HJ, Durham JL, et al. Comparison of the acute inflammatory response of two commercial platelet-rich plasma systems in healthy rabbit tendons. *The American journal of sports medicine*. 2012;40:1274-81.
 30. McCarrel TM, Minas T, Fortier LA. Optimization of leukocyte concentration in platelet-rich plasma for the treatment of tendinopathy. *The Journal of bone and joint surgery. American volume*. 2012;94:e143(1-8).
 31. Zhou Y, Zhang J, Wu H, Hogan MV, Wang JH. The differential effects of leukocyte-containing and pure platelet-rich plasma (PRP) on tendon stem/progenitor cells - implications of PRP application for the clinical treatment of tendon injuries. *Stem cell research & therapy*. 2015;6:173.
 32. Schippinger G, Pruller F, Divjak M, et al. Autologous Platelet-Rich Plasma Preparations: Influence of Non-steroidal Anti-inflammatory Drugs on Platelet Function. *Orthop J Sports Med*. 2015;3:2325967115588896.
 33. A Hamid MS, Mohamed Ali MR, Yusof A, George J, Lee LP. Platelet-rich plasma injections for the treatment of hamstring injuries: a randomized controlled trial. *The American journal of sports medicine*. 2014;42:2410-8.

34. Hamilton B, Tol JL, Almusa E, et al. Platelet-rich plasma does not enhance return to play in hamstring injuries: a randomised controlled trial. *British journal of sports medicine*. 2015;49:943-50.
35. Fader RR, Mitchell JJ, Traub S, et al. Platelet-rich plasma treatment improves outcomes for chronic proximal hamstring injuries in an athletic population. *Muscles, ligaments and tendons journal*. 2014;4:461-6.
36. Wetzel RJ, Patel RM, Terry MA. Platelet-rich plasma as an effective treatment for proximal hamstring injuries. *Orthopedics*. 2013;36:e64-70.
37. Davenport KL, Campos JS, Nguyen J, Saboeiro G, Adler RS, Moley PJ. Ultrasound-Guided Intratendinous Injections With Platelet-Rich Plasma or Autologous Whole Blood for Treatment of Proximal Hamstring Tendinopathy: A Double-Blind Randomized Controlled Trial. *Journal of ultrasound in medicine : official journal of the American Institute of Ultrasound in Medicine*. 2015;34:1455-63.
38. Mei-Dan O, Lopez V, Carmont MR, et al. Adductor tenotomy as a treatment for groin pain in professional soccer players. *Orthopedics*. 2013;36:e1189-97.
39. Dallaudiere B, Pesquer L, Meyer P, et al. Intratendinous injection of platelet-rich plasma under US guidance to treat tendinopathy: a long-term pilot study. *Journal of vascular and interventional radiology : JVIR*. 2014;25:717-23.
40. Hammoud S, Bedi A, Magennis E, Meyers WC, Kelly BT. High incidence of athletic pubalgia symptoms in professional athletes with symptomatic femoroacetabular impingement. *Arthroscopy : the journal of arthroscopic & related surgery : official publication of the Arthroscopy Association of North America and the International Arthroscopy Association*. 2012;28:1388-95.
41. Larson CM, Pierce BR, Giveans MR. Treatment of athletes with symptomatic intra-articular hip pathology and athletic pubalgia/sports hernia: a case series. *Arthroscopy : the journal of arthroscopic & related surgery : official publication of the Arthroscopy Association of North America and the International Arthroscopy Association*. 2011;27:768-75.
42. Mautner K, Colberg RE, Malanga G, et al. Outcomes after ultrasound-guided platelet-rich plasma injections for chronic tendinopathy: a multicenter, retrospective review. *PM & R : the journal of injury, function, and rehabilitation*. 2013;5:169-75.
43. Boyd KT, Peirce NS, Batt ME. Common hip injuries in sport. *Sports Med*. 1997;24:273-88.
44. Cervellin M, de Girolamo L, Bait C, Denti M, Volpi P. Autologous platelet-rich plasma gel to reduce donor-site morbidity after patellar tendon graft harvesting for anterior cruciate ligament reconstruction: a randomized, controlled clinical study. *Knee surgery, sports traumatology, arthroscopy : official journal of the ESSKA*. 2012;20:114-20.
45. Rodeo SA, Delos D, Williams RJ, Adler RS, Pearle A, Warren RF. The effect of platelet-rich fibrin matrix on rotator cuff tendon healing: a prospective, randomized clinical study. *The American journal of sports medicine*. 2012;40:1234-41.
46. Sanchez M, Anitua E, Azofra J, Andia I, Padilla S, Mujika I. Comparison of surgically repaired Achilles tendon tears using platelet-rich fibrin matrices. *The American journal of sports medicine*. 2007;35:245-51.

