

CHAPTER 8

A New Approach to Treat Joint Injuries: Combination of Intra-Articular and Intraosseous Injections of Platelet Rich Plasma

AUTHORS

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SUMMARY

This chapter deals with the scientific rationale which underpins a new procedure for the treatment of severe knee osteoarthritis, namely, a combination of intra-articular and intraosseous injections of Platelet Rich Plasma. Intraarticular infiltration of platelet rich plasma is a promising treatment for knee osteoarthritis, but it still has some therapeutic limitations in severe osteoarthritis. Intraosseous infiltration delivers platelet rich plasma into the subchondral bone, acting on this tissue and consequently on cartilage-bone communication. Thus, this technique involves a new route of delivering platelet rich plasma that could be applied not only for severe osteoarthritis but also for other joint pathologies in which the

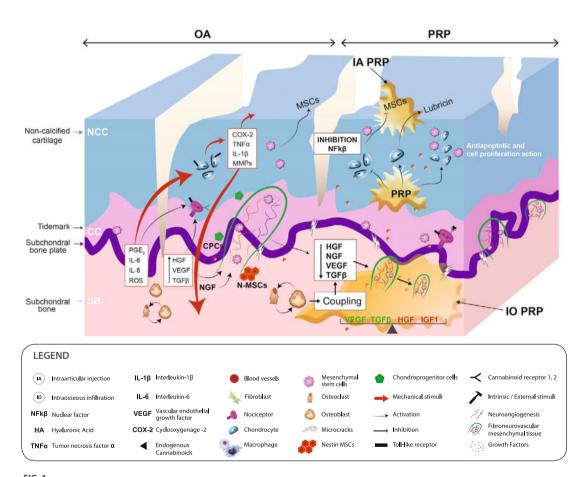
subchondral bone is critical in its etiology such as osteonecrosis, osteochondral lesions, and bone marrow lesions.

This chapter explores some of the recent insights and observations concerning the involvement of subchondral bone in the pathophysiology of osteoarthritis and additionally we will describe a new technique of platelet rich plasma infiltration for the treatment of severe knee osteoarthritis.

1. INTRODUCTION

Subchondral bone has always been present in the equation of the cartilage repair process and OA¹⁻³ but it has suffered neglect for decades as an important player in the etiopathogenesis of OA^{3,4}. There is an increasingly recognized communication between the subchondral bone and articular cartilage based on the changes that the subchondral bone undergoes in patients with severe OA, including microcracks and structural defects, vascularization of channels, nerve growth, and a

progressive replacement of the subchondral marrow with fibroneurovascular mesenchymal tissue (figure 1)⁵⁻⁸. As it is yet to be established precisely which of the joint tissues or structures is the primary driver of knee osteoarthritis (KOA), and therapeutic strategies targeting solely one cell or tissue target may well prove to fail⁹, it is advisable that approaches to severe KOA treatment should be aimed at reaching several joint tissues with the objective of reducing joint inflammation, controlling pain, improving joint functionality, and restoring the homeostasis of joint tissues.



Targeting the osteoarthritic subchondral bone with Intraosseous infiltration of PRP. This schematic drawing illustrates the outside-in (AC-SB) and inside-out (SB-AC) flow of mediators and cells. SB as a point of egress of morphogens and cells, through the channels and vessels breaching the osteochondral junction, partially recruited by the osteoarthritic synovial fluid 3,7. This cartilage cell invasion might be facilitated by the loss of aggrecans, collagen II cleavage, and disruption of water tissue distribution 8 of the articular cartilage as well as by the secretion by MSCs of fibrinolytic enzymes 22. The excessive presence of TGFB1 and VEGF in OA subchondral bone 3,7 could be a driving factor for changes in osteoblast-osteoclast coupling thereby leading to a bone remodelling imbalance 28, 30, 38, NGF expression, and fibroneurovascular growth, changes that additionally might well contribute to overlying cartilage degradation, pain, and an osteoarthritic joint 25, 33, 47, 48, 49. (Reprinted with permission from Sánchez, M. et al.) ¹⁶

In recent years, several clinical trials using intra-articular infiltrations of plasma rich in growth factors have shown promising results¹⁰⁻¹⁴; however, there are still some concerns about whether this form of administration is able to reach the deeper layers of the cartilage and subchondral bone, thereby possibly limiting the growth factors therapeutic potential especially in severe osteoarthritis^{11,15}.

In light of recent studies reporting the importance of subchondral bone in the pathogenesis of osteoarthritis and cartilage-subchondral bone communication^{16,17} we proposed a combination of intra-articular and intraosseous injections to treat severe osteoarthritis¹⁸ In so doing, it is possible to expand the effective range of PRPs by not only acting on the subchondral bone and consequently on its cartilage communications, but also on mesenchymal stem cells, to modulate the affected tissue regeneration^{19,20}.

This chapter is based on two manuscripts published recently^{16,18} an it will explore some of the recent insights and observations concerning the involvement of subchondral bone in the pathophysiology of osteoarthritis.

2. THE ROLE OF SUBCHONDRAL BONE (SB) IN PATHOPHYSIOLOGY AND CLINICAL SYMPTOMS OF OSTEOARTHRITIS

2.1. The subchondral bone-articular cartilage functional unit

Subchondral bone has always been present in the equation of OA pathogenesis, and more than 40 years ago, partially inspired by the 1827 proposal by surgeon Dr. P.P. Physick on the SB as an effective shock absorber, Radin et al^{2,21} suggested a cause-effect connection among mechanical loading, subchondral bone sclerosis, and osteoarthri-

tis. Subchondral bone is the layer of bone which lies immediately below the calcified cartilage (figure 1)22, and consists of two different anatomical entities, one called subchondral or cortical plate which is nonporous and poorly vascularized cortical bone, and the SB which contains bone marrow (fatty) and trabecular bone^{23,24}. Together with the articular cartilage (AC), it forms the osteochondral functional unit, which undergoes mechanical stresses that trigger adaptive cell responses and establish a crosstalk among them to adjust their architecture to ongoing physical and biochemical challenges^{7,25}. In the functionality of the osteochondral unit, articular cartilage provides an elastic, gliding, smooth frictionless surface, while subchondral bone, a very low viscoelastic structure, together with periarticular muscles and ligaments, acts as shock absorber structures, accounting for 30% and 50% of the total absorbing energy and only 1-3% for the AC^{23,26}. Besides the pivotal shock absorbing function, SB is a source of vessels whose perfusion rate enables an important nutritional route for AC but any damage to this microvasculature affects venous bony circulation thereby altering cartilage and chondrocyte function^{5,23,27}.

2.2. SB turnover and structural changes in OA

The osteochondral unit in an OA joint undergoes several structural changes including loss of articular cartilage, development of inflamed synovium, calcified cartilage thickening and tidemark duplication, undermineralization of bone, sclerosis and stiffness of SB, bone marrow lesions (BMLs), cysts, osteophyte, and a localized bone marrow replacement by fibroneurovascular tissue (figure 1)^{5-7,28}.

Despite the high turnover of SB in OA, an uncoupling between bone formation and resorption at the same site leads to an increase in bone volume without a concomitant increase in bone mineralization pattern^{3,28,29}. This SB sclerosis is characterized by an increase of the osteoid volume, and a decrease of calcium bound to collagen fibre, and is associated with a gain of trabecular thickness, loss of trabecular number, and a trabecular network more separated and less interconnected^{29,30}. It has

been suggested that sclerotic subchondral bone, localized at subchondral plate, could decrease the load transfer to the underlying bone tissue leading to osteoporotic-like changes⁵. Moreover, SB can undergo microdamage, such as microcracks and clefts, that modify SB stiffness and reduce the shock-absorbing capacity of SB, thereby making chronic a microdamage context and perpetuating an accelerated bone remodelling, which impairs normal mineralization of bone once it has been deposited, most likely by a modified osteoblastic phenotype^{5,24,31}. Magnetic resonance imaging (MRI) has helped to detect subchondral bone marrow edema-like lesions (BMLs), which have been found to be associated with pain and disease progression in KOA³², and together with bone attrition, are strong indicators of a structural deterioration in knee and hip osteoarthritis^{5,33}.Several studies conducted in human knee and hip OA paralleling MRI bone marrow edema lesion (BMLs) studies with histological analysis of SB retrieved at the time of joint replacement, revealed microfractures and increased bone remodelling, subchondral ingrowth of fibrovascular tissue and increased vascularity, various types of bone marrow fibrosis32,34,35 as well as numeric and topographic alterations in native mesenchymal stem cells (MSCs)33. These observations were confirmed in rodent models of OA7,36.The increased activity of osteoclasts in OA cause channels to extend from SB to AC, passing across the calcified tissues into the noncalcified articular cartilage²⁵. The neurovascular invasion of those new-formed channels is accompanied by a new fibroneurovascular mesenchymal tissue within the channel along with cells such as macrophages, osteoclasts, osteoblasts, and endothelial cells, which interact to stimulate angiogenesis and growth of sympathetic and sensory nerves⁷ and reach the noncalcified cartilage (figure 1), a finding which has been supported by animal models of OA7.

2.3. Cellular interactions and molecular crosstalk in osteochondral unit in OA

There is now good evidence that even in a nondiseased joint, naturally occurring pores and holes enable communication between SB and AC via diffusion of small molecules^{6,37,38}. This communication may be exacerbated by structural changes seen early in the osteochondral unit in OA.The increased osteoclastic activity in the OA subchondral plate 28 may increase the permeability of bonecartilage interface by inducing channel formation in the tidemark, in addition to the existent aberrant fibroneurovascular tissue and vasculature. and mechanical stress-induced microcracks^{7,24,39}. Reinforcing this view, Pan et al.37 have demonstrated the diffusion of small-size molecules between SB and AC by utilizing the FLIP method (Imaging method based on fluorescence loss, which quantifies diffusivity of small molecules) with sodium fluorescein in the distal femur of mice, and this communication is greatly increased in osteoarthritic joints of the mice model⁶. Therefore, the presence of these connections enables an elevated crosstalk among chondrocytes, osteoblasts, osteoclasts and MSCs through biological factors and signalling pathways (figure 1).

Several in vitro and in vivo studies have demonstrated that osteoblasts from sclerotic subchondral bone show an altered phenotype. In an in vitro study, Westacott et al 40 reported that osteoblasts in OA-affected bone exhibited a different phenotype, whose activity can degrade articular cartilage in vitro. Supporting this observation, Hilial et al.⁴¹ reported that osteoblasts from OA subchondral bone have an abnormal metabolism with increased levels of PGE2 and TGF\$ (figure 1). Using a co-culture model of OA subchondral bone osteoblasts with chondrocytes, Sanchez et al reported that osteoblasts induced a catabolic response of chondrocytes including a decrease in aggrecan, type II collagen and SOX-9, and an increase of MMP-3 and MMP-13 among other mediators^{42,43}. Moreover, osteoblasts from scletoric subcondral bone have an elevated TGFB expression²⁹ and under cyclical compression express proangiogenic factors such as VEGF,FGF, and IL-8⁴⁴. Hepatocyte growth factor (HGF) is a pleiotropic morphogen present in articular cartilage but produced by osteoarthritic subchondral bone osteoblats, osteoclasts, and MSCs45-47, with likely implications in both the chondrocyte

anabolic state and the proliferation of an invasive fibroneurovascular tissue in SB5,7,47, the latter when an uncoupling osteoclast-osteoblast activity may lead to an overexpression of HGF (figure 1)45. The excessive presence of TGFB1 and VEGF in OA subchondral bone likely stemmed from a dysregulated osteocyte7,48 could be a driving factor for changes in osteoblast-osteoclast coupling thereby leading to a bone remodelling imbalance^{5,30}, NGF expression⁴⁹, and fibroneurovascular growth changes that additionally might well contribute to overlying cartilage degradation^{30,48}, pain^{7,24,25} and an osteoarthritic joint^{30,48}. In a recent study, Zhen et al. showed that by inhibiting TGF-β signalling in a specific population of MSCs present at the SB (Nestin positive MSCs), the severity of OA was reduced, a change associated with improvement of bone parameters, cartilage structure and joint function without affecting TGFB signalling in AC⁴⁸. Moreover, in a recent study, Campbell et al (2016) reported functional and gene expression perturbations in native MSCs which could lead to further damage escalation³³. These findings are in accordance with previous studies that have shown that the decrease of MSCs in the synovial fluid, in low degree OA, suggests clinical improvement⁵⁰. MSCs from osteoarthritic bone marrow have been reported to be substantially reduced in yield and proliferative activity besides showing a weakened chondrogenic and adipogenic activity and increased osteogenic activity⁵¹. However, in vitro studies indicate that the inclusion of growth factors, as a supplementary culture medium, can be beneficial in reverting their chondrogenic activity⁵².

2.4. Subchondral bone as a tissue target in OA treatment

The realization of the biological and mechanical connection between AC and SB has lead to numerous in vivo animal studies that have shown that targeting SB with some drugs can have protective structural effects on cartilage⁴. Blocking or limiting the bone remodelling with alendronate⁵³, zoledronic acid⁵⁴ or improving the microstructure and quality of subchondral bone in osteoarthritic

and osteoporotic rabbits with parathyroid hormone⁴, may prevent cartilage degradation and OA progression. Moreover, Sagar et al⁵⁵ reported a reduction in pain behaviour after a subcutaneous treatment with osteoprotegerin in a monosodium iodoacetate (MIA) rat model of OA pain, and Pelletier et al⁵⁶ demonstrated that an oral strontium ranelate treatment in an experimental osteoarthritic dog model reduced the progression of structural changes including the subchondral bone. Despite the fact that the translation of these promising observations in preclinical research to human clinical trials has often failed, as indicated by a recent metaanalysis of clinical trial with risedronate in knee osteoarthritis57, recent clinical trials are raising expectations. For instance, using zoledronic in patients with clinical KOA associated with bone marrow lesions (BMLs) assessed by MRI, Laslett et al⁵⁸ reported a beneficial effect on pain and on BML evolution at 6 months. In participants from the osteoarthritis initiative, Laslett et al.59 demonstrated significant pain reduction during the first 3 years of treatment with biphosphonates. Two more clinical trials have shown positive structural effects of strontium ranelate on KOA, one improving the joint space narrowing 60 and the other reducing the loss of cartilage volumes concurrent with the decrease of BMLs at 3 years of follow up⁶¹.

Infiltrations of platelet rich plasma (PRP) into the bone marrow cavity of femur of young and old ovariectomized-SAMP8 age-related osteoporotic female mice have been reported to up-regulate osteogenesis and down-regulate adipogenesis⁶². The increase of fat tissue mass in BM is correlated with decreased bone mineralization in aged SAMPS8 mice^{62,63}, bone demineralization that occurs in osteoarthritic subchondral bone together with cysts²⁴. Moreover, improvement of bone mineral density in PRP-treated osteoporotic mice concurred with both histological sections of the bone samples showing more trabecular bone areas and more intense calcium staining and a suppression of bone resorption process as evidenced by the decrease of RANKL transcript⁶². In a trial on 13 healthy volunteers, Philippart et al⁶⁴ reported fatigue on the first day as the only clinical adverse effect after a self-stimulation of BM of the iliac crest by injected autologous platelet rich plasma⁶⁴. Supporting these findings, Figure 2 shows the histological analysis of cartilage and SB from a patient suffering from severe KOA who underwent intraosseous infiltrations of PRGF. Eight months later, the patient had not improved clinically and underwent a knee replacement. During

the surgery, we took this sample of cartilage and subchondral bone from the femoral condyle in which 5cc of PRGF had been infiltrated intraosseously. Part of the biopsy showed a good gross appearance, with pearly areas similar to the original hyaline cartilage, though histological study revealed a fibrocartilage repair tissue. Another area showed nearly exposed bone.

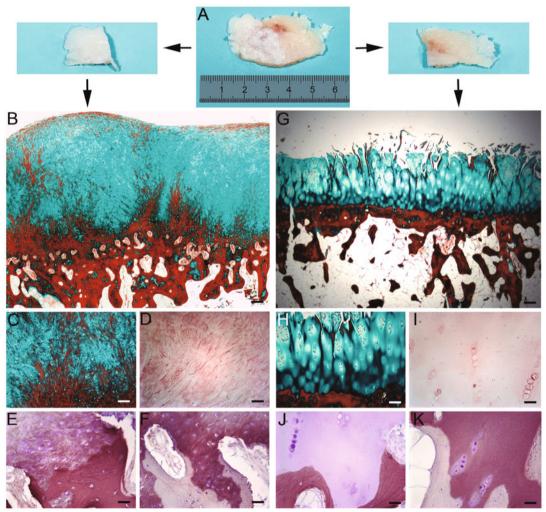


FIG. 2
Fibrocartilage repair tissue after intraosseus PRGF infiltrations in the treatment of human knee osteoarthritis: a histological study. (A) Macroscopic morphology of the sample. The sample was divided into two pieces. The fragment on the left corresponds to fibrocartilage repair tissue (B to F) while the right-hand fragment shows osteoarthritic cartilage (G to K). B and G show panoramic images of the sample (Masson's trichrome staining). In photomicrographs C and H, details of the structure of articular cartilage are observed (Masson's trichrome staining). The presence of elastic fibres is demonstrated by Orcein staining (D and I). These fibres can be seen in D, while they are absent in I. An immunohistochemical study was performed to detect the presence of type I (E and J) and type II (F and K) collagen. In all samples (E to K), both subchondral bone (always positive for type I and negative for type II collagens) and cartilage are observed. In fibrocartilage (E and F) both types of reactivity are observed, while in the degenerated cartilage, only type II collagen positivity is shown (K). Histologically, the pearly area (the left-hand side of the sample) is fibrocartilage repair tissue, while the right-hand side of the sample displays an osteoarthritic area with loss of cartilage surface integrity. (Reprinted with permission from Sánchez. M, et al.) ¹⁶

3. INTRAOSSEOUS INFILTRATIONS OF PLASMA RICH IN GROWTH FACTORS

In light of the aforementioned research and others not mentioned here due to space limitation, and the significant clinical improvement obtained in some but not all patients with KOA treated with intraarticular infiltrations of PRP^{11,13,14,65} our group

arrived at the strategy of combining another drug delivery route, namely, the intraosseous infiltrations combined with intraarticular infiltrations of PRP^{18-20,66,67}. (Figure 3)

The procedure is carried out in the operating room under a 4-5 degree of sedation of the patient. In addition, local anesthesia is conducted into the periosteum of condyle and tibial plateau by injecting 2mL of 2% mepivacaine. Intraosseous infiltrations are performed with a 13G trocar used

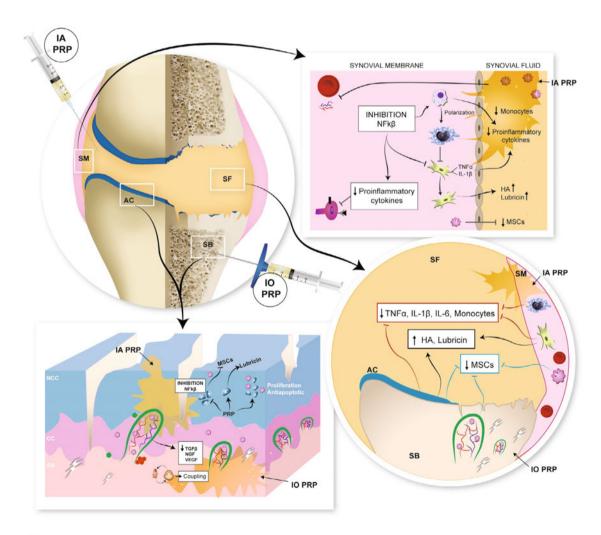
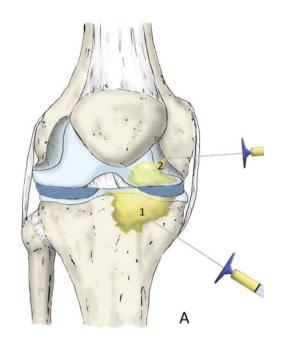


FIG. 3
Depiction of a new strategy to treat severe knee OA by targeting different knee joint structures such as synovial membrane (SM), synovial fluid (SF), articular cartilage (AC), noncalcified cartilage (NCC) and calcified cartilage (CC), and subchondral bone (SB) with intra-articular injections (IA) and intraosseous infiltrations (IO) of platelet rich plasma (PRP). This procedure reduces pain and mesenchymal stem cells (MSC) in SF, besides significantly improving knee joint function of patients with severe OA (Reprinted with permission from Sánchez, M. et al.) 19

for bone biopsy, and the control of trocar placement is facilitated using a fluoroscope (Figure 4, 5, and 6)18. The first treatment includes one PRP intraarticular infiltration and two PRP intraosseous infiltrations (in femoral condyle and tibial plateau). Two more weekly intraarticular infiltrations are performed. The group of Sanchez et al. have found after a 6 month follow-up, a significant pain reduction and decrease of MSC and CFU-F in synovial fluid with no adverse effects^{19,20,66}. We have been performing intraosseous infiltrations of PRGF since 2003 applying them regularly at the condyle and tibial tunnels in the arthroscopic reconstruction of anterior cruciate ligament, and in osteochondral injuries and osteonecrosis of the hip and knee⁶⁸.

4. DISCUSSION AND FUTURE PERSPECTIVES

Intraarticular delivery is an alternative modality to convey PRP in patients with KOA and it has been shown to be safe and efficacious in improving clinical symptoms^{11,13,65}. This route of drug delivery reaches the synovial membrane (SM) and the AC, which is sometimes inefficiently targeted by systemic drug delivery. Intraarticular delivery circumvents systemic toxicity and its side effects, offers an excellent bioavailability, and does not present molecular size limitation, in contrast to the systemically delivered molecules entering the joint via capillaries of the subsynovium^{69,70}. Nevertheless, intraarticular therapy faces other challenges when treating chronic nonsystemic sterile-inflammatory conditions as in the case of KOA. One significant challenge is a short joint dwell time of drugs, since the lymphatic drainage clears proteins in a few hours. This is not the case of PRGF, since it acts as a dynamic liquid scaffold with a fibrin network from where GFs are gradually released into the tissue^{71,72}. Moreover, the increasingly recognized role of SB in the pathophysiology of OA^{3,7,24,28} might make the intraarticular route insufficient to tackle all the joint tissues involved in KOA.



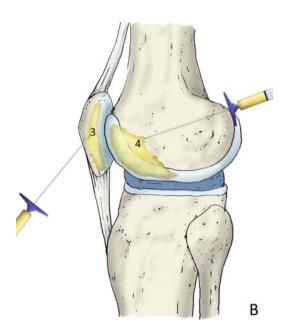


FIG. 4

(A) The platelet-rich plasma (PRP) intraosseous infiltration of a knee with severe femorotibial osteoarthritis is performed into the medial tibial plateau (1) and medial femoral condyle (2). (B) If the patient presents with femoropatellar osteoarthritis, the approach is external and the patella (3) and trochlea (4) are infiltrated. Before these intraosseous injections are performed, conventional knee intraarticular infiltration of PRP is conducted. (Reprinted with permission from Sánchez, M. et al.) 18

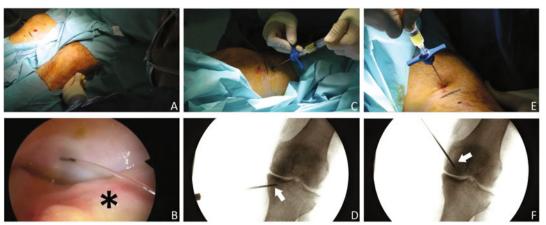


FIG. 5
After the patient is positioned supine on the operating room table, (A) intra-articular infiltration is performed into the joint through the external patellar wing, centred in the central region of the patella in the craniocaudal plane; (B) the infiltration is directed into the midpoint area of the femoropatellar region using an external approach and preventing infiltration into the synovial membrane, (asterisk). (C, D) Intraosseous tibial plateau infiltration is conducted into the medial tibial plateau, just to its middle area. The arrow indicates the trocar. (E, F). With respect to intraosseous femoral condyle infiltration, a trocar (arrow) is applied to the thickness of the medial femoral condyle, as far as the middle area of the medial condyle. (Reprinted with permission from Sánchez, M. et al.) ¹⁸



FIG. 6
(A) Communications between cartilage and subchondral bone are more pronounced in degenerated cartilage. (B) The platelet-rich plasma infiltrated into subchondral bone flows through the degenerated zones, and because of its viscous consistency, (C) it remains in the area, creating a matrix (asterisk). (Reprinted with permission from Sánchez, M. et al.) 18

Intraosseous delivery strategy for local, prolonged, and sustainable release of GFs has been proven to be efficacious in some musculoskeletal pathology, non-union fractures, osteoporosis, and bone fracture healing among them^{73,74}. Over the past 30 years, surgical experience in cartilage defect has revealed that only when the subchondral bone is involved through bone marrow stimulating procedures such as transcortical Pridie drilling and microfractures, is a temporary functional fibrocartilage tissue synthesized, with no serious adverse effect reported⁷⁵. There is good in vitro and in vivo evidence that events in the subchondral bone concur with and have a direct effect on

the overlying articular cartilage^{4,27,29,76}. Moreover, although the titles and much of the text of Liu et al⁶³ and Philippart et al⁶⁴ papers are not focused on osteoarthitis, these studies shed important light on the role that intraosseous infiltrations of PRGF might play in subchondral bone homeostasis by targeting both osteoblast-osteoclast coupling and mesenchymal stem cell responses, as well as in its safety.

The combination of intra-articular and intraosseous injections of PRP is an in situ local biological "joint-centric" approach to treat severe KOA addresses the SM, SF and superficial zone of AC by

intraarticular injections of PRGF, and deep zones of AC and SB through PRGF intraosseous infiltrations (figure 3)¹⁸⁻²⁰. These PRGF infiltrations convey a mimetic biomaterial embedded with a pool of growth factors acting as a smart scaffold77 which might sustain a gradual delivery of growth factors at the dysfunctional and deregulated tissues as a niche therapy. Rebuilding a physiologicalhomeostatic network at knee organ failure tissue level, as is the case of severe knee OA, must be an orderly process, which entails a daunting therapeutic task. Our hypothesis is that the concurrent presence and a balanced ratio between plateletsecreted TGFB-1 and VEGF, and plasma growth factors such as IGF-1 and HGF⁷⁸⁻⁸¹, all conveyed by PRGF intraosseous infiltrations, might reduce or buffer the excess of TGFB in SB and restore HGF activity synthesized by subchondral bone cells. This modulatory effect of PRGF on TGFB-1 signaling pathway might shrink the fibroneurovascular tissue that replaces the bone marrow of OA subchondral bone, an explanation which parallels the antifibrotic mechanism already reported to be exerted by PRP on several cell phenotypes⁷⁹⁻⁸¹. This new reestablished homeostatic balance between TGFB1 and HGF^{39,48} would reduce the synthesis of NGF, VEGF and other inflammatory mediators thereby contributing as well to modulate the aberrant fibroneurovascular tissue and to alleviate pain and hyperalgesia⁸².

In spite of a wealth of preclinical and clinical publications on PRP, many uncertainties remain regarding the ultimate molecular mechanism/s, the variability in its composition mainly due to the presence/absence of leukocytes, the platelet concentration, the donors age, and the manner in which PRPs are applied to the damaged tissues⁸³. Moreover, we need to delve into the particular effect of PRP on MSCs in addition to the systemic effect that this procedure might entail since few human studies have been carried out regarding PRP treatments and systemic effects^{84,85}.

5. CONFLICT OF INTERESTS

The authors declare the following competing financial interest(s): One author is the Scientific Director of and two authors are scientists at BTI Biotechnology Institute, a dental implant company that investigates in the fields of oral implantology and PRGF-Endoret technology.

- Pridie K, Gordon G. A method of resurfacing osteoarthritic knee joints. Paper presented at: Journal of Bone and Joint Surgery-British Volume 1959.
- Radin EL, Rose RM. Role of subchondral bone in the initiation and progression of cartilage damage. Clinical orthopaedics and related research. 1986:34-40.
- Lajeunesse D, Reboul P. Subchondral bone in osteoarthritis: a biologic link with articular cartilage leading to abnormal remodeling. Current opinion in rheumatology. 2003;15:628-33.
- Bellido M, Lugo L, Roman-Blas JA, et al. Improving subchondral bone integrity reduces progression of cartilage damage in experimental osteoarthritis preceded by osteoporosis. Osteoarthritis and cartilage / OARS, Osteoarthritis Research Society. 2011:19:1228-36.
- Lajeunesse D. Subchondral bone involvement in the pathophysiology of osteoarthritis. Understanding Osteoarthritis from Bench to Bedside. 2011:69-83
- Pan J, Wang B, Li W, et al. Elevated cross-talk between subchondral bone and cartilage in osteoarthritic joints. Bone. 2012;51:212-7.
- 7. Suri S, Walsh DA. Osteochondral alterations in osteoarthritis. Bone. 2012;51:204-11.
- Stolz M, Gottardi R, Raiteri R, et al. Early detection of aging cartilage and osteoarthritis in mice and patient samples using atomic force microscopy. Nat Nanotechnol. 2009;4:186-92.
- Martel-Pelletier J, Wildi LM, Pelletier JP. Future therapeutics for osteoarthritis. Bone. 2012;51:297-311.

- 10. Sanchez M, Guadilla J, Fiz N, Andia I. Ultrasound-guided platelet-rich plasma injections for the treatment of osteoarthritis of the hip. Rheumatology (Oxford). 2012;51:144-50.
- 11. Sanchez M, Fiz N, Azofra J, et al. A randomized clinical trial evaluating plasma rich in growth factors (PRGF-Endoret) versus hyaluronic acid in the short-term treatment of symptomatic knee osteoarthritis. Arthroscopy: the journal of arthroscopic & related surgery: official publication of the Arthroscopy Association of North America and the International Arthroscopy Association. 2012;28:1070-8.
- 12. Kon E, Filardo G, Di Martino A, Marcacci M. Platelet-rich plasma (PRP) to treat sports injuries: evidence to support its use. Knee surgery, sports traumatology, arthroscopy: official journal of the ESSKA. 2011;19:516-27.
- 13. Vaquerizo V, Plasencia MA, Arribas I, et al. Comparison of intra-articular injections of plasma rich in growth factors (PRGF-Endoret) versus Durolane hyaluronic acid in the treatment of patients with symptomatic osteoarthritis: a randomized controlled trial. Arthroscopy: the journal of arthroscopic & related surgery: official publication of the Arthroscopy Association of North America and the International Arthroscopy Association. 2013;29:1635-43.
- 14. Wang-Saegusa A, Cugat R, Ares O, Seijas R, Cusco X, Garcia-Balletbo M. Infiltration of plasma rich in growth factors for osteoarthritis of the knee short-term effects on function and quality of life. Archives of orthopaedic and trauma surgery. 2011;131:311-7.
- 15. Khoshbin A, Leroux T, Wasserstein D, et al. The efficacy of platelet-rich plasma in the treatment of symptomatic knee osteoarthritis: a systematic review with quantitative synthesis. Arthroscopy: the journal of arthroscopic & related surgery: official publication of the Arthroscopy Association of North America and the International Arthroscopy Association. 2013;29:2037-48.

- Sanchez M, Anitua E, Delgado D, et al. A new strategy to tackle severe knee osteoarthritis: Combination of intra-articular and intraosseous injections of Platelet Rich Plasma. Expert opinion on biological therapy. 2016:16:627-43.
- 17. Zhen G, Cao X. Targeting TGFbeta signaling in subchondral bone and articular cartilage homeostasis. Trends in pharmacological sciences. 2014;35:227-36.
- 18. Sanchez M, Fiz N, Guadilla J, et al. Intraosseous infiltration of platelet-rich plasma for severe knee osteoarthritis. Arthrosc Tech. 2014;3:e713-7.
- 19. Sánchez M, Delgado D, Sánchez P, et al. Combination of Intra-Articular and Intraosseous Injections of Platelet Rich Plasma for Severe Knee Osteoarthritis: A Pilot Study. BioMed research international. 2016;2016.
- 20. Muinos-Lopez E, Delgado D, Sanchez P, et al. Modulation of Synovial Fluid-Derived Mesenchymal Stem Cells by Intra-Articular and Intraosseous Platelet Rich Plasma Administration. Stem cells international. 2016;2016:1247950.
- 21. Radin EL, Paul IL, Rose RM. Role of mechanical factors in pathogenesis of primary osteoarthritis. Lancet. 1972;1:519-22.
- 22. Neuss S, Schneider RK, Tietze L, Knuchel R, Jahnen-Dechent W. Secretion of fibrinolytic enzymes facilitates human mesenchymal stem cell invasion into fibrin clots. Cells, tissues, organs. 2010;191:36-46.
- 23. Imhof H, Sulzbacher I, Grampp S, Czerny C, Youssefzadeh S, Kainberger F. Subchondral bone and cartilage disease: a rediscovered functional unit. Invest Radiol. 2000;35:581-8.
- 24. Burr DB, Gallant MA. Bone remodelling in osteoarthritis. Nature reviews. Rheumatology. 2012;8:665-73.
- 25. Mapp Pl, Walsh DA. Mechanisms and targets of angiogenesis and nerve growth in osteoarthritis. Nature reviews. Rheumatology. 2012;8:390-8.

- 26. Brandt KD, Radin EL, Dieppe PA, van de Putte L. Yet more evidence that osteoarthritis is not a cartilage disease. Annals of the rheumatic diseases. 2006;65:1261-4.
- Malinin T, Ouellette EA. Articular cartilage nutrition is mediated by subchondral bone: a long-term autograft study in baboons. Osteoarthritis and cartilage / OARS, Osteoarthritis Research Society. 2000;8:483-91.
- 28. Karsdal MA, Bay-Jensen AC, Lories RJ, et al. The coupling of bone and cartilage turnover in osteoarthritis: opportunities for bone antiresorptives and anabolics as potential treatments? Annals of the rheumatic diseases. 2014;73:336-48.
- 29. Henrotin Y, Pesesse L, Sanchez C. Subchondral bone and osteoarthritis: biological and cellular aspects. Osteoporosis international: a journal established as result of cooperation between the European Foundation for Osteoporosis and the National Osteoporosis Foundation of the USA. 2012;23 Suppl 8:S847-51.
- Zhao W, Wang T, Luo Q, et al. Cartilage degeneration and excessive subchondral bone formation in spontaneous osteoarthritis involves altered TGF-β signaling. Journal of Orthopaedic Research. 2015;DOI:10.1002/jor.23079.
- 31. Burr DB, Radin EL. Microfractures and microcracks in subchondral bone: are they relevant to osteoarthrosis? Rheum Dis Clin North Am. 2003;29:675-85.
- 32. Felson DT, Chaisson CE, Hill CL, et al. The association of bone marrow lesions with pain in knee osteoarthritis.

 Annals of internal medicine. 2001;134:541-9.
- 33. Campbell TM, Churchman SM, Gomez A, et al. Mesenchymal Stem Cell Alterations in Bone Marrow Lesions in Patients With Hip Osteoarthritis. Arthritis Rheumatol. 2016;68:1648-59.
- 34. Taljanovic MS, Graham AR, Benjamin JB, et al. Bone marrow edema pattern in advanced hip osteoarthritis: quantitative assessment with magnetic resonance imaging and correlation with clinical examination, radiographic findings, and histopathology. Skeletal Radiol. 2008;37:423-31.

- 35. Hunter DJ, Gerstenfeld L, Bishop G, et al. Bone marrow lesions from osteoarthritis knees are characterized by sclerotic bone that is less well mineralized. Arthritis research & therapy. 2009;11:R11.
- Mapp PI, Avery PS, McWilliams DF, et al. Angiogenesis in two animal models of osteoarthritis.
 Osteoarthritis and cartilage / OARS, Osteoarthritis Research Society. 2008;16:61-9.
- 37. Pan J, Zhou X, Li W, Novotny JE, Doty SB, Wang L. In situ measurement of transport between subchondral bone and articular cartilage. Journal of orthopaedic research: official publication of the Orthopaedic Research Society. 2009;27:1347-52.
- 38. Lyons TJ, McClure SF, Stoddart RW, McClure J. The normal human chondro-osseous junctional region: evidence for contact of uncalcified cartilage with subchondral bone and marrow spaces. BMC Musculoskelet Disord. 2006;7:52.
- 39. Tat SK, Lajeunesse D, Pelletier J-P, Martel-Pelletier J. Targeting subchondral bone for treating osteoarthritis: what is the evidence? Best Practice & Research Clinical Rheumatology. 2010;24:51-70.
- 40. Westacott CI, Webb GR, Warnock MG, Sims JV, Elson CJ. Alteration of cartilage metabolism by cells from osteoarthritic bone. Arthritis and rheumatism. 1997;40:1282-91.
- 41. Hilal G, Martel-Pelletier J, Pelletier JP, Ranger P, Lajeunesse D. Osteoblast-like cells from human subchondral osteoarthritic bone demonstrate an altered phenotype in vitro: possible role in subchondral bone sclerosis. Arthritis and rheumatism. 1998;41:891-9.

- 42. Sanchez C, Deberg MA, Piccardi N, Msika P, Reginster JY, Henrotin YE. Osteoblasts from the sclerotic subchondral bone downregulate aggrecan but upregulate metalloproteinases expression by chondrocytes. This effect is mimicked by interleukin-6, -1beta and oncostatin M pre-treated non-sclerotic osteoblasts. Osteoarthritis and cartilage / OARS, Osteoarthritis Research Society. 2005;13:979-87.
- Sanchez C, Deberg MA, Piccardi N, Msika P, Reginster JY, Henrotin YE. Subchondral bone osteoblasts induce phenotypic changes in human osteoarthritic chondrocytes. Osteoarthritis and cartilage / OARS, Osteoarthritis Research Society. 2005;13:988-97.
- Sanchez C, Pesesse L, Gabay O, et al. Regulation of subchondral bone osteoblast metabolism by cyclic compression. Arthritis and rheumatism. 2012:64:1193-203.
- 45. Grano M, Galimi F, Zambonin G, et al. Hepatocyte growth factor is a coupling factor for osteoclasts and osteoblasts in vitro. Proceedings of the National Academy of Sciences of the United States of America. 1996;93:7644-8.
- Stella MC, Comoglio PM. HGF: a multifunctional growth factor controlling cell scattering. The international journal of biochemistry & cell biology. 1999;31:1357-62.
- 47. Guevremont M, Martel-Pelletier J, Massicotte F, et al. Human adult chondrocytes express hepatocyte growth factor (HGF) isoforms but not HgF: potential implication of osteoblasts on the presence of HGF in cartilage. Journal of bone and mineral research: the official journal of the American Society for Bone and Mineral Research. 2003;18:1073-81.
- Zhen G, Wen C, Jia X, et al. Inhibition of TGF-beta signaling in mesenchymal stem cells of subchondral bone attenuates osteoarthritis. Nature medicine. 2013;19:704-12.

- 49. Davidson EB, van Caam A, Vitters E, et al. TGF-β is a potent inducer of Nerve Growth Factor in articular cartilage via the ALK5-Smad2/3 pathway. Potential role in OA related pain? Osteoarthritis and Cartilage. 2015;23:478-86.
- 50. Sekiya I, Ojima M, Suzuki S, et al. Human mesenchymal stem cells in synovial fluid increase in the knee with degenerated cartilage and osteoarthritis. Journal of orthopaedic research: official publication of the Orthopaedic Research Society. 2012;30:943-9.
- 51. Barry F, Murphy M. Mesenchymal stem cells in joint disease and repair. Nature reviews. Rheumatology. 2013;9:584-94.
- 52. Scharstuhl A, Schewe B, Benz K, Gaissmaier C, Buhring HJ, Stoop R. Chondrogenic potential of human adult mesenchymal stem cells is independent of age or osteoarthritis etiology. Stem cells. 2007;25:3244-51.
- 53. Hayami T, Pickarski M, Wesolowski GA, et al. The role of subchondral bone remodeling in osteoarthritis: reduction of cartilage degeneration and prevention of osteophyte formation by alendronate in the rat anterior cruciate ligament transection model. Arthritis and rheumatism. 2004;50:1193-206.
- 54. Lampropoulou-Adamidou K, Dontas I, Stathopoulos IP, et al. Chondroprotective effect of high-dose zoledronic acid: An experimental study in a rabbit model of osteoarthritis. Journal of orthopaedic research: official publication of the Orthopaedic Research Society. 2014;32:1646-51.
- 55. Sagar DR, Ashraf S, Xu L, et al. Osteoprotegerin reduces the development of pain behaviour and joint pathology in a model of osteoarthritis. Annals of the rheumatic diseases. 2014;73:1558-65.
- 56. Pelletier JP, Kapoor M, Fahmi H, et al. Strontium ranelate reduces the progression of experimental dog osteoarthritis by inhibiting the expression

- of key proteases in cartilage and of IL-1beta in the synovium. Annals of the rheumatic diseases. 2013;72:250-7.
- 57. Davis AJ, Smith TO, Hing CB, Sofat N. Are bisphosphonates effective in the treatment of osteoarthritis pain? A meta-analysis and systematic review. PloS one. 2013:8:e72714.
- 58. Laslett LL, Dore DA, Quinn SJ, et al. Zoledronic acid reduces knee pain and bone marrow lesions over 1 year: a randomised controlled trial. Annals of the rheumatic diseases. 2012;71:1322-8.
- 59. Laslett LL, Kingsbury SR, Hensor EM, Bowes MA, Conaghan PG. Effect of bisphosphonate use in patients with symptomatic and radiographic knee osteoarthritis: data from the Osteoarthritis Initiative. Annals of the rheumatic diseases. 2014;73:824-30.
- 60. Reginster JY, Badurski J, Bellamy N, et al. Efficacy and safety of strontium ranelate in the treatment of knee osteoarthritis: results of a double-blind, randomised placebo-controlled trial. Annals of the rheumatic diseases. 2013;72:179-86.
- 61. Pelletier JP, Roubille C, Raynauld JP, et al. Disease-modifying effect of strontium ranelate in a subset of patients from the Phase III knee osteoarthritis study SEKOIA using quantitative MRI: reduction in bone marrow lesions protects against cartilage loss. Annals of the rheumatic diseases. 2015;74:422-9.
- Liu HY, Wu AT, Tsai CY, et al. The balance between adipogenesis and osteogenesis in bone regeneration by platelet-rich plasma for age-related osteoporosis. Biomaterials. 2011;32:6773-80.
- Liu HY, Huang CF, Lin TC, et al. Delayed animal aging through the recovery of stem cell senescence by platelet rich plasma. Biomaterials. 2014;35:9767-76.

- 64. Philippart P, Meuleman N, Stamatopoulos B, et al. In vivo production of mesenchymal stromal cells after injection of autologous platelet-rich plasma activated by recombinant human soluble tissue factor in the bone marrow of healthy volunteers. Tissue engineering. Part A. 2014;20:160-70.
- 65. Filardo G, Kon E, Pereira Ruiz MT, et al. Plateletrich plasma intra-articular injections for cartilage degeneration and osteoarthritis: single- versus double-spinning approach. Knee surgery, sports traumatology, arthroscopy: official journal of the ESSKA. 2012;20:2082-91.
- 66. Diego Delgado PS, Emma Muiños, Bruno Paiva, Froilán Granero, Eduardo Anitua, Sabino Padilla, Felipe Prósper, Mikel Sánchez. Phase II Clinical trial of intraosseous platelet rich plasma infiltration for severe knee osteoarthritis: preliminary results at 2 months. 2015. ICRS 2015 Chicago
- 67. Sanchez M, Anitua E, Azofra J, Andia I, Padilla S, Mujika I. Comparison of surgically repaired Achilles tendon tears using platelet-rich fibrin matrices. The American journal of sports medicine. 2007;35:245-51.
- 68. Sánchez M, Delgado D, Sánchez P, et al. Platelet Rich Plasma and Knee Surgery. BioMed research international. 2014;2014.
- 69. Evans CH, Kraus VB, Setton LA. Progress in intraarticular therapy. Nature reviews. Rheumatology. 2014;10:11-22.
- 70. Kang ML, Im Gl. Drug delivery systems for intraarticular treatment of osteoarthritis. Expert Opin Drug Deliv. 2014;11:269-82.

- 71. Anitua E, Zalduendo MM, Prado R, Alkhraisat MH, Orive G. Morphogen and proinflammatory cytokine release kinetics from PRGF-Endoret fibrin scaffolds: evaluation of the effect of leukocyte inclusion. Journal of biomedical materials research. Part A. 2015;103:1011-20.
- 72. Martino MM, Briquez PS, Guc E, et al. Growth factors engineered for super-affinity to the extracellular matrix enhance tissue healing. Science. 2014;343:885-8.
- 73. Sanchez M, Anitua E, Cugat R, et al. Nonunions treated with autologous preparation rich in growth factors. Journal of orthopaedic trauma. 2009;23:52-9.
- Kyllonen L, D'Este M, Alini M, Eglin D. Local drug delivery for enhancing fracture healing in osteoporotic bone. Acta biomaterialia. 2015;11:412-34.
- 75. Hunziker EB, Lippuner K, Keel MJ, Shintani N. An educational review of cartilage repair: precepts & practice--myths & misconceptions--progress & prospects. Osteoarthritis and cartilage / OARS, Osteoarthritis Research Society. 2015;23:334-50.
- 76. Burr DB. The importance of subchondral bone in the progression of osteoarthritis. The Journal of rheumatology. Supplement. 2004;70:77-80.
- 77. Conboy I, Freimer J, Weisenstein L. 5.526 Tissue Engineering of Muscle Tissue. In: Ducheyne P, ed. Comprehensive Biomaterials. Oxford: Elsevier; 2011:345-59.
- 78. Anitua E, Andia I, Sanchez M, et al. Autologous preparations rich in growth factors promote proliferation and induce VEGF and HGF production by human tendon cells in culture. Journal of orthopaedic research: official publication of the Orthopaedic Research Society. 2005;23:281-6.

- 79. Anitua E, Sanchez M, Merayo-Lloves J, De la Fuente M, Muruzabal F, Orive G. Plasma rich in growth factors (PRGF-Endoret) stimulates proliferation and migration of primary keratocytes and conjunctival fibroblasts and inhibits and reverts TGF-beta1-Induced myodifferentiation. Investigative ophthalmology & visual science. 2011;52:6066-73.
- 80. Anitua E, Sanchez M, De la Fuente M, Zalduendo MM, Orive G. Plasma rich in growth factors (PRGF-Endoret) stimulates tendon and synovial fibroblasts migration and improves the biological properties of hyaluronic acid. Knee surgery, sports traumatology, arthroscopy: official journal of the ESSKA. 2012;20:1657-65.
- Assirelli E, Filardo G, Mariani E, et al. Effect of two different preparations of platelet-rich plasma on synoviocytes. Knee surgery, sports traumatology, arthroscopy: official journal of the ESSKA. 2014.
- 82. Seidel MF, Herguijuela M, Forkert R, Otten U. Nerve growth factor in rheumatic diseases. Seminars in arthritis and rheumatism. 2010;40:109-26.
- 83. Wasterlain AS, Braun HJ, Dragoo JL. Contents and formulations of platelet-rich plasma. Operative Techniques in Orthopaedics. 2012;22:33-42.
- 84. Schippinger G, Fankhauser F, Oettl K, Spirk S, Hofmann P. Does single intramuscular application of autologous conditioned plasma influence systemic circulating growth factors? J Sports Sci Med. 2012;11:551-6.
- Wasterlain AS, Braun HJ, Harris AH, Kim HJ, Dragoo JL. The systemic effects of platelet-rich plasma injection. The American journal of sports medicine. 2013;41:186-93.