

# Bioinformatics Mini Project 2024

## Part 1 – The ORF Detector

### An Introduction to ORFs:

An open reading frame (ORF) is defined as a start codon followed by a downstream in-frame stop codon. The sequence of these will typically vary depending on whether the DNA itself is spliced or not (i.e. whether it contains regions of non-coding DNA). While ORFs vary in length, standard ORFs tend to be defined as being composed of at least 100 codons ([Claverie et.al, 1997](#)) and small ORFs (smORFs) are defined as being less than 100 codons in length ([Kute et.al, 2022](#)).

### The ORF Detector:

#### Python code:

```
sequence = SeqIO.read("part1_sequence.fasta", "fasta")

# Printing the sequence and its ID
print("Sequence:", sequence.seq)
print("ID:", sequence.id)

def find_all_ORFs(sequence):
    ORFs = []
    #finding reverse complement and iterating over 3 frames for both sequences
    for strand, nuc in [(+1, sequence.seq), (-1, sequence.seq.reverse_complement())]:
        for frame in range(3):
            length = 3 * ((len(sequence)-frame) // 3) # Multiple of three for codons
            for i in range(frame, length, 3):
                codon = nuc[i:i+3]
                if codon in ['ATG', 'AUG']:
                    for j in range(i+3, length, 3):
                        if nuc[j:j+3] in ['TAA', 'TAG', 'TGA', 'UAA', 'UAG', 'UGA']:
                            ORFs.append(nuc[i:j+3])
                            break
    ORFs.sort(key=len, reverse=True)
    return ORFs # Now returning a list of all ORFs, in order of size

ORFs = find_all_ORFs(sequence)

# Create a table to input results into
table = PrettyTable()

# Adding columns to the table
table.field_names = ["ORF Length", "ORF Sequence", "Amino Acid Sequence"]

# saving ORFs as fasta files and adding the translated amino acid sequence for each ORF into the table
for i, orf in enumerate(ORFs, 1):
    amino_acid_sequence = orf.translate(to_stop=True)
    table.add_row([len(orf), orf, amino_acid_sequence])
    record = SeqRecord(orf, id=f"ORF{i}", description=f"ORF{i} from sequence {sequence.id}")
    SeqIO.write(record, f"ORF{i}.fasta", "fasta")

# Print results in the table
print(table)
```

#### Imported Packages:

```
#importing libraries
from Bio.Seq import Seq
from Bio import SeqIO
from Bio.SeqRecord import SeqRecord
from prettytable import PrettyTable
```

Testing with the trial sequence:

### Trial sequence (part1\_sequence):

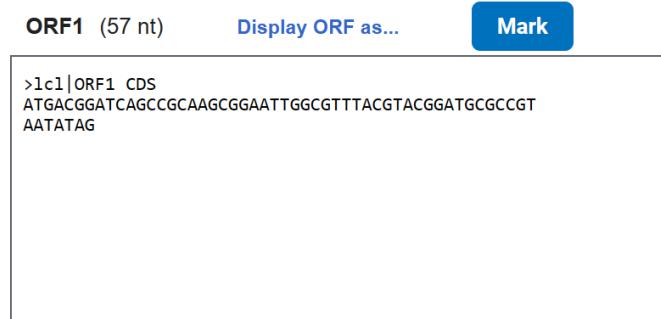
ATTGCCATGACGGATCAGCCGCAAGCGGAATTGGCGTTACGTACGGATGCGCCGTAATATAAGGCCA  
TAGAC

### **Expected outputs:**

The online ORF finder from the National Center for Biotechnology Information website ([ORFfinder Viewer - NCBI \(nih.gov\)](#)) was used to check the output of the ORF finder using the 'part1\_sequence' fasta file as a trial sequence.



### Expected ORF 1:



### Expected Amino acid sequence for ORF 1;

**ORF1** (18 aa)

[Display ORF as...](#)

**Mark**

```
>lc1|ORF1  
MTDQPQAEELAFTYGCASI
```

Expected ORF 2:

**ORF2** (54 nt)

[Display ORF as...](#)

**Mark**

```
>lc1|ORF2 CDS  
ATGGCCTATATTACGGCGCATCCGTACGTAAACGCCATTCCGCTTGCCTGA
```

Expected Amino acid sequence for ORF 2:

**ORF2** (17 aa)

[Display ORF as...](#)

**Mark**

```
>lc1|ORF2  
MAYITAHPYVNANSACG
```

### Actual output:

Actual ORF and Amino Acid output:

Sequence: ATTGCCATGACGGATCAGCCGCAAGCGGAATTGGCGTTACGTACGGATGCGCCGTAATATAGGCCATAGAC		
ID: part1_sequence		
ORF Length	ORF Sequence	Amino Acid Sequence
57	ATGACGGATCAGCCGCAAGCGGAATTGGCGTTACGTACGGATGCGCCGTAATATAG	MTDQPQAEELAFTYGCASI
54	ATGGCCTATATTACGGCGCATCCGTACGTAAACGCCATTCCGCTTGCCTGA	MAYITAHPYVNANSACG

The actual output for the ORF nucleotide sequence and Amino acid sequence fit the Expected outputs found by the NCBI ORF Finder tool. Therefore no edits needed to be made to the Python script after

this result. However, this ORF finder still had not been tested on larger sequences, so the BRCA2 cDNA sequence was input as a query (please see below).

Testing with the BRCA2 cDNA sequence:

Expected output:

It is expected that the longest ORF from this sequence will match the BRCA2 peptide sequence given in Ensemble () .

Actual output with the ORF finder:

```
#importing libraries
from Bio.Seq import Seq
from Bio import SeqIO
from Bio.SeqRecord import SeqRecord

#reading the fasta file
sequence = SeqIO.read("BRCA2_cDNA_ensemble.fa", "fasta")

def find_all_ORFs(sequence):
    ORFs = []
    #finding reverse complement and iterating over 3 frames for both sequences
    for strand, nuc in [(+1, sequence.seq), (-1, sequence.seq.reverse_complement())]:
        for frame in range(3):
            length = 3 * ((len(sequence)-frame) // 3) # Multiple of three for codons
            for i in range(frame, length, 3):
                codon = nuc[i:i+3]
                if codon in ['ATG', 'AUG']:
                    for j in range(i+3, length, 3):
                        if nuc[j:j+3] in ['TAA', 'TAG', 'TGA', 'UAA', 'UAG', 'UGA']:
                            ORFs.append(nuc[i:j+3])
                            break
        ORFs.sort(key=len, reverse=True)
    return ORFs # Now returning a list of all ORFs, in order of size

ORFs = find_all_ORFs(sequence)

print()

# Printing the longest ORF and its Length
if ORFs:
    print(f'Longest ORF: {ORFs[0]}')
    print()
    print(f'Length of longest ORF: {len(ORFs[0])}')

# Printing the total number of ORFs found
print()
print(f'Total ORFs found: {len(ORFs)}')
print()

# Saving the Longest ORF as a fasta file: BRCA2_longest_orf.fasta
if ORFs:
    longest_orf = SeqRecord(ORFs[0], id="Longest_ORF", description="Longest ORF from BRCA2")
    SeqIO.write(longest_orf, "BRCA2_longest_orf.fasta", "fasta")
```

Length of longest ORF: 10257

Total ORFs found: 429

This code saves the longest ORF out of the 429 found as a fasta file (BRCA2\_longest\_orf.fasta) and also printing it as an output (for my own debugging/viewing benefit).

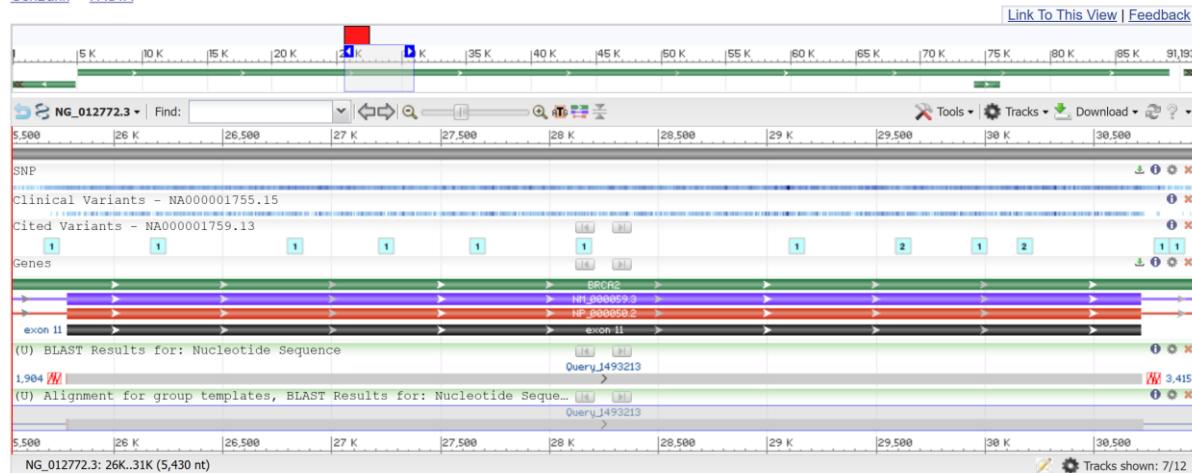
#### BLAST results for the BRCA2 gene longest ORF:

Sequences producing significant alignments		Download		Select columns		Show		100	?
		GenBank		Graphics		Distance tree of results		MSA Viewer	
Description	Scientific Name	Max Score	Total Score	Query Cover	E value	Per. Ident	Acc. Len		
<input checked="" type="checkbox"/> Homo sapiens BRCA2 DNA repair associated (BRCA2), RefSeqGene (LRG_293) on chromosome 13	Homo sapiens	9114	19079	100%	0.0	99.98%	91193	NG_012772.3	
<input checked="" type="checkbox"/> Homo sapiens zygote arrest 1 like (ZAR1L), RefSeqGene on chromosome 13	Homo sapiens	470	595	3%	2e-129	99.23%	18665	NG_017006.2	

#### **Homo sapiens BRCA2 DNA repair associated (BRCA2), RefSeqGene (LRG\_293) on chromosome 13**

NCBI Reference Sequence: NG\_012772.3

[GenBank](#) [FASTA](#)



#### Comments:

As expected, the observed output matched the BRCA2 peptide sequence given in Ensemble. Therefore, as long as no unforeseen bugs occur in the code, this script is able to detect the longest ORF and all ORFs within a sequence that is fairly long or reasonably short in length.

#### Possible extension and model limitations:

It cannot be expected that all ORFs found by the ORF finder will code for proteins. Whilst the output was expected for the human BRCA2 cDNA sequence, the result could have been different if the regular DNA sequence was used (pre-splicing). This current model of the ORF Detector cannot distinguish between introns and exons. Whilst it could be theoretically possible to implement some kind of command to bias the ORF finder against regions expressing typical intron sequence motifs (such as increased C/G repetition), (Xie et.al, 2023). these alterations would most likely have to be species specific due to the highly variable abundance of introns between species and would most likely be prone to bias (Zlotorynski et.al, 2019).

## Part 2 – Decoding Dinosaur DNA

### Introduction – Sequence 1

The goal of this section (Part 2, section 1) is to determine whether the Dinosaur DNA sequence from Michael Crichton's Jurassic Park is based on fact or fiction. To achieve this, the ORF finder coded for in Part 1 will be utilised as well as the NCBI BLAST+ online webservice.

Sequence-1: Finding 'Dinosaur' ORFs

Dinosaur DNA Sequence 1:

```
GCGTTGCTGGCGTTTCCATAGGCTCCGCCCCCTGACGAGCATCACAAAAATCGACGCGGTGG  
CGAAACCCGACAGGACTATAAAGATACCAGGCCTTCCCCCTGGAAGCTCCCTCGTGTCCGACC  
CTGCCGCTTACCGGATAACCTGTCCGCCTTCTCCTCGGAAAGCGTGGCTGCTCACGCTGTACCT  
ATCTCAGTTCGGTAGGTGCGTCAAGCTGGCTGAGTCCAACCCGGTAAAGTAGGACAGGTGCCGCAGCGCTCG  
CCTTATCCGTAACTATCGTCTTGAGTCCAACCCGGTAAAGTAGGACAGGTGCCGCAGCGCTCT  
GGGTCACTTCGGCGAGGACCGCTTCGCTGGAGATCGGCCTGCGCTTGCCTGCGGTATTCGGAATCT  
TGCACGCCCTCGCTCAAGCCTCGTCACTCCAAACGTTCGGCAGAAGCAGGCCATTATGCC  
GCATGGCGGCCGACCGCTGGCTGGCGTTCGCGACGCCAGGCTGGATGGCCTTCCCATTATGA  
TTCTTCTCGCTTCCGGCGGCCGCTTGCAGGCCATGCTGTCCAGGCAGGTAGATGACGACCATC  
AGGGACAGCTAACGGCTTACCGCCTAACCTCGATCACTGGACCGCTGATCGTCACGGCGA  
TTTATGCCACATGGACGCGTTGCTGGCGTTTCCATAGGCTCCGCCCCCTGACGAGCATCAA  
AACAAAGTCAGAGGTGGCGAAACCCGACAGGACTATAAAGATACCAGGCCTTCCCGCTTCTCA  
GCTCTCTGTTCCGACCCCTGCCGTTACGGATAACCTGTCCGCCTTCTCCCTCGGGCTTCTCA  
ATGCTCACGCTGTAGGTATCTCAGTTGGTGTAGGTGCGCTCCAGCTGACGAACCCCCCGT  
TCAGCCGACCGCTGCGCTTACCGTAACTATCGTCTTGAGTCCAACACGACTAACGGGTTG  
GCATGGATTGTAGGCGCCCTACCTTGCTGCCTCCCGCGGTGCATGGAGCCGGCCACC  
TCGACCTGAATGGAAGCCGGGCACCTCGCTAACGGCCAAGAATTGGAGCCAATCAATTCTG  
CGGAGAACTGTGAATGCGAAACCAACCCTGGCATCGCTCCGCATCTCAGCAGCCGCAC  
CGGCGCATCTCGGGCAGCGTTGGTCCT
```

Results from the ORF finder:

ORF Length	ORF Sequence	Amino Acid Sequence
327	ATGACGACCATCAGGGACAGCTCAACGGCTTTA CCAGCCTAACTCGATCACTGGACCGCTGATCGTC CGCGATTATGCCGCACATGGACGCCTGCTGGC GTTTTCCATAGGCTCCGCCCTGACGAGCATCA CAAACAAGTCAGAGGTGGCGAAACCCGACAGGAC TATAAAGATACCAGGCCTTCCCCCTGGAAAGCGCT CTCCTGTTCCGACCTGCGCTTACCGGATACCTGT CCGCCTTCTCCCTCGGGCTTCTCAATGCTCACG CTGTAGGTATCTCAGTCGGTAGGTCGTTGCTC CAAGCTGA	MTTIRDSFNGSYQPNFDHWTADR HGDLCRTWTRCWRFSIGSAPLTSI TNKSEVAKPDRTIKIPGVSPWKRSP VPTLPLTGylSAFLPSGFLNAHAV GISVRCRSFAPS
228	ATGCCGCACATGGACCGTGGCTGGCGTTTCCAT AGGCTCCGCCCTGACGAGCATACAAACAAGT CAGAGGTGGCGAAACCCGACAGGACTATAAAGAT ACCAGGCCTTCCCCCTGGAAAGCGCTCTCGTTC GACCCCTGCCGCTTACCGGATACCTGTCCGCCCTTCT CCCTTCGGGCTTCTCAATGCTCACGCTGTAGGTAT CTCAGTCGGTAG	MPHMDALLAFFHRLRPPDEHHKQ VRGGETRQDYKDTRRFPLEALSCS DPAAYRIPVRLSPFGLSQCSRCRYL SSV
219	ATGGACCGTGTGGCGTTTCCATAGGCTCCGC CCCCCTGACGAGCATACAAACAAGTCAGAGGTGG CGAAACCCGACAGGACTATAAAGATACCAAGGCGTT TCCCCCTGGAAAGCGCTCTCTGTTCCGACCTGCGC CTTACCGGATACCTGTCCGCCCTTCTCCCTCGGGC TTTCTCAATGCTCACGCTGTAGGTATCTCAGTCGG TGTAG	MDALLAFFHRLRPPDEHHKQVRG GETRQDYKDTRRFPLEALSCSDPA AYRIPVRLSPFGLSQCSRCRYLSSV
213	ATGCCGCCGCGTGCCTGGCTGGAGATGGCGGACGC GATGGCCAAGGGTTGGTTGCGCATTACAGTTCTC CGCAAGAACATTGATTGGCTCCAATTCTTGGCCGTTAG CGAGGTGCCGCCGCTTCATTCAAGGTCGAGGTGG CCCGGCTCCATGCACCGCGGGGAGGCAGACAAGGT ATAGGGCGCGCCTACAATCCATGCCAACCGTTA A	MRRVRLLEMADAMAKGWFAHSQ FSARIDWLQFLAVSEVPASIQVEV ARLHAPRGGRQGIGRRLQSMPTR
189	ATGGCGGACCGATGGCCAAGGGTTGGTTGCGCA TTCACAGTTCTCGCAAGAACATTGATTGGCTCCAATT CTTGGCCGTTAGCGAGGTGCCGCCGGCTTCATTCA GGTCGAGGTGGCCGGCTCCATGCACCGCGGGAG GCAGACAAGGTATAGGGCGCGCCTACAATCCATG CCAACCGTTAA	MADAMAKGWFAHSQFSARIDWL QFLAVSEVPASIQVEVARLHAPR GGRQGIGRRLQSMPTR
177	ATGGCCAAGGGTTGGTTGCGCATTACAGTTCTCC GCAAGAACATTGATTGGCTCCAATTCTTGGCCGTTAGC GAGGTGCCGCCGGCTTCATTCAAGGTCGAGGTGGC CCGGCTCCATGCACCGCGGGGAGGCAGACAAGGT TAGGGCGCGCCTACAATCCATGCCAACCGTTAA	MAKGWFAHSQFSARIDWLQFLAV SEVPASIQVEVARLHAPRGGRQG IGRRLQSMPTR
150	ATGACCCAGAGCGCTGCCGCACCTGTCTACTTTA CCGGGTTGGACTCAAGACGATAGTTACCGGATAAG GCGCAGCGGTGGGCTGAACGGCACACAGCCCAGC TTGGAGCGAACGACCTACACCGAACTGAGATAGGT ACAGCGTGA	MTQSAAGTCPLPGWTQDDSYRI RRSGRAERHTAQLGANDLHRTEIG TA
138	ATGGTCGTCATCTACCTGCCCTGGACAGCATGGCCTG CAACCGGGCCGCCGGAAAGCGAGAAGAACATCAA TGGGGAAAGGCCATCCAGCCTCGCTCGAACGCC AGCCCAGCGCGTGGCCGCATGCCGGCGATAA	MVVIYLPGQHGLQRGPPEARRIIM GKAIQPRVANASPARRPPCRR
93	ATGGAGCCGGGCCACCTCGACCTGAATGGAAGCCG GCGGCACCTCGCTAACGGCCAAGAACATTGGAGCCAA TCAATTCTTGGAGAGACTGTGA	MEPGHLDLNGSRRHLANGQELEPI NSCGEL
72	ATGGCCTTCCCCATTATGATTCTCTCGCTTCCGGC GGCCCGCGTGCAGGCCATGCTGTCCAGGCAGGTA G	MAFPIMILLASGGPRCRPCCPGR
72	ATGGATTGTAGGCGCCGCCCTACCTGTCTGCCT CCCCCGGGTGCATGGAGCCGGCACCTCGACCTG A	MDCRRRPIPCLPPRGAWSRATST

72	ATGCTCGTCAGGGGGCGGAGCCTATGGAAAAACGCCAGCAACCGTCCATGTGCGGCATAATCGCCGTGA	MLVRGAEPMEKRQQRVHVRHKSP
69	ATGGGAAGGCCATCCAGCCTCGCGTCGCGAACGCAAGCCCAGCGCGTCGGCCCATGCCGGCGATAA	MGKAIQPRVANASPARPPCRR
63	ATGCTGTCCAGGCAGGTAGATGACGACCATCAGGGACAGCTTCAACGGCTCTTACCAAGCCTAA	MLSRQVDDDHQQLQRLPA
63	ATGGCGGCCGACCGCCTGGCTGGCGTTCGCGACCGAGGCTGGATGGCCTCCCCATTATGA	MLSRQVDDDHQQLQRLPA
57	ATGATTCTTCTCGCTTCCGGCGGCCCGCTGCAGGCCATGCTGTCCAGGCAGGTAG	MILLASGGPRCRPCCPGR
51	ATGTGCGGCATAATGCCGTGACGATCAGCGTCAGTGATCGAAGTTAG	MCGINRRDDQRSSDRS
48	ATGGAAAAACGCCAGCAACCGTCCATGTGCGGCAATAATCGCCGTGA	MEKRQQRVHVRHKSP
45	ATGCCGGCGATAATGCCCTGCTTCTGCCGAAACGTTGGAGTGA	MPAIMACFSPKRLE
42	ATGGCCTGCAACCGGGCCGCCGGAAAGCGAGAAGAATCATAA	MACNAGRKRREES
33	ATGGCCTGCTTCTGCCGAAACGTTGGAGTGA	MACFSPKRLE
30	ATGCACCGCGGGGAGGCAGACAAGGTATAG	MHRGEADKV
15	ATGCTCACGCTGTAG	MLTL
15	ATGCCAACCGTTAA	MPTR

Results from the ORF finder:

```

#reading the fasta file
sequence = SeqIO.read("part2_Dinosaur_DNA.fasta", "fasta")

# Printing the sequence and its ID
print("Sequence:", sequence.seq)
print("ID:", sequence.id)

def find_all_ORFs(sequence):
    ORFs = []
    #finding reverse complement and iterating over 3 frames for both sequences
    for strand, nuc in [(+1, sequence.seq), (-1, sequence.seq.reverse_complement())]:
        for frame in range(3):
            length = 3 * ((len(sequence)-frame) // 3) # Multiple of three for codons
            for i in range(frame, length, 3):
                codon = nuc[i:i+3]
                if codon in ['ATG', 'AUG']:
                    for j in range(i+3, length, 3):
                        if nuc[j:j+3] in ['TAA', 'TAG', 'TGA', 'UAA', 'UAG', 'UGA']:
                            ORFs.append(nuc[i:j+3])
                            break
    ORFs.sort(key=len, reverse=True)
    return ORFs # Now returning a list of all ORFs, in order of size

ORFs = find_all_ORFs(sequence)

# Create a table to input results into
table = PrettyTable()

# Adding columns to the table
table.field_names = ["ORF Length", "ORF Sequence", "Amino Acid Sequence"]

# saving ORFs as fasta files and adding the translated amino acid sequence for each ORF into the table
for i, orf in enumerate(ORFs, 1):
    amino_acid_sequence = orf.translate(to_stop=True)
    table.add_row([len(orf), orf, amino_acid_sequence])
    record = SeqRecord(orf, id=f"ORF{i}", description=f"ORF{i} from sequence {sequence.id}")
    SeqIO.write(record, f"ORF{i}.fasta", "fasta")

# Print results in the table
print(table)

# Print the number of ORFs found
print("Number of ORFs found:", len(ORFs))

```

-----  
-----+  
Number of ORFs found: 24

### Sequence-1: ‘Dinosaur’ Protein analysis.

Protein type	E-value and coverage	comments
Hypothetical protein from Escherichia coli	E-value: 1e-10 Coverage: 33%	-Many repeat outputs
plasmid replication initiation protein, partial [Klebsiella pneumoniae]	E-value: 6e-10 Coverage: 42%	-Many repeat outputs
Hypothetical protein from Salmonella enterica	E-value: 7e-04 Coverage: 51%	-Many repeat outputs
Hypothetical protein (VEV11M) from Escherichia coli	E-value: 5e-04 Coverage: 100%	-Many repeat outputs

### Sequence-1: comments and conclusions

The high proportion of hypothetical/ partial bacterial protein sequences appears telling of a typical case of sample contamination. However, as no matches other than the repeated list of hypothetical bacterial protein sequences were returned, it seems unlikely that there was any original sample amplified in the first place.

I decided to investigate with this unknown original ‘Dinosaur’ sequence further and run some BLASTn searches on the sequence and its longest ORFs. A BLAST of the full sequence (see below) returned weak alignments of bacterial origin, solidifying to findings from the original BLASTp searches from the ORF finder output.

Descriptions	Graphic Summary	Alignments	Taxonomy	Sequences producing significant alignments									Download	Select columns	Show	100	?								
													GenBank	Graphics	Distance tree of results	MSA Viewer									
				Description	Scientific Name	Max Score	Total Score	Query Cover	E value	Per. Ident	Acc. Len	Accession													
<input checked="" type="checkbox"/>	<a href="#">select all</a>	100 sequences selected		<a href="#">Francisella tularensis LVS pOM1 tet(C) gene for tetracycline efflux MFS transporter Tet(C), complete CDS</a>	<a href="#">Francisella tular...</a>	435	661	44%	2e-116	87.80%	1391	<a href="#">NG_048174.1</a>													
<input checked="" type="checkbox"/>				<a href="#">Cloning vector reBmBac, complete sequence</a>	<a href="#">Cloning vector r...</a>	435	630	42%	2e-116	87.80%	134520	<a href="#">KU749552.1</a>													
<input checked="" type="checkbox"/>				<a href="#">Cloning vector pPolh-1629, complete sequence</a>	<a href="#">Cloning vector p...</a>	435	1330	92%	2e-116	87.80%	4975	<a href="#">KU749550.1</a>													
<input checked="" type="checkbox"/>				<a href="#">Expression vector pET151-OmyCasp3 DNA, complete sequence</a>	<a href="#">Expression vect...</a>	435	1424	98%	2e-116	87.80%	6510	<a href="#">AB477343.1</a>													
<input checked="" type="checkbox"/>				<a href="#">Vector pSEVA512S-16S-pad, complete sequence</a>	<a href="#">Vector pSEVA51...</a>	435	543	36%	2e-116	87.80%	5513	<a href="#">ON366567.1</a>													
<input checked="" type="checkbox"/>				<a href="#">Synthetic construct His-TEV-Nsp15 gene, complete cds</a>	<a href="#">synthetic construct</a>	435	1424	98%	2e-116	87.80%	6760	<a href="#">ON191567.1</a>													
<input checked="" type="checkbox"/>				<a href="#">Cloning vector pYAT7, complete sequence</a>	<a href="#">Cloning vector p...</a>	435	1036	97%	2e-116	87.80%	8694	<a href="#">KT970713.1</a>													

BLASTn searches of the three longest ORFs for this sequence also agreed with the original BLASTp searches. These results lead me to believe that either:

- 1) The sequence is fictitious
- 2) Or, somehow through storing and processing the sample decades later, the original sample has degraded and/ or has become contaminated with bacterial DNA

In this situation I would lean towards the sequence being entirely fictitious.

### Introduction – Sequence 2

In this section (Part 2, section 2) an updated sequence of possible Dinosaur DNA was provided by Mark Boguski, to improve upon the seemingly fictitious sequence from section 1 of part 2. Again, the ORF detector coded-for in Part 1 will be utilised as well as the NCBI BLAST+ online webservice.

### Sequence-2: Finding ‘Dinosaur’ ORFs

Dinosaur DNA – Sequence 2:  
GAATTCCGGAAGCGAGCAAGAGATAACTCCTGGCATCAGATACACTGGAGATAAGGACGGACGTG  
TGGCAGCTCCGCAGAGGATTCAGTGGAAAGTGCAATTACCTATCCCCTGGAGGCCATGGAGTCGTGG  
CGCTGGGGGGGGCCGGATGCGGGCTCCCCACTCCGTTCCCTGATGAAGCCGGAGCCTCCTGGGGC  
TGGGGGGGGCGAGAGGACGGAGGCGGGAGGCGGGGGCTGCTGGCCTCTACCCCCCTCAGGCCCGTGT  
TCCCTGGTGGCGACGACACGGGTACTTGGGACCCCCCAGTGGGTGCCGCCACCCAA  
ATGGAGCCCCCCCACACTACCTGGAGCTGCTGCAACCCCCCCCAGCCCCCCTGGCAGGCGCTGAGTGC  
GGGCCCTACTGCCACTCAGCAGCGGGCCCCCACCTGGCAGGCGCTGAGTGCCTGAGGCCAGCGTCA  
AAGAACCTGGAGCGACGGCAACGCCGCTGTGGCGCCGGACGGCACGGCATTACCTGTGCAAC  
TGGGCCTAGCCTGGGGCTTACCAACCGCCTCAACGGCCAGAACCGCCCGCTCATCCGCCCCAAA  
AAGCGCCTGCGGGTGAGTAAGCGCGAGGCACAGTGTGCAGCCACAGCGTAAAACGCCAGAC  
ATCCACCACCACTCTGTGGCGTCGAGCCCCATGGGGACCCGCTGTGCAACAACATTACGCCCTG  
CGGCCTACTACAAACTGCACCAAGTGAACCGCCCCCTCACGATGCGCAAAGACGGAATCCAAAC

```

CCGAAACCGCAAAGTTCTCCAAGGGTAAAAGCGGCCCGGGGGGGAAACCCCTCCG
CCACCGCGGGAGGGGGCGCTCCTATGGGGGGAGGGGGGACCCCTATGCCCGCCCC
CCCCGGCCGCCGCCCTCAAGCGACGCTCTGTACCGCTCGGCCCGTGGCTTCGGCC
ATTTCTGCCCTTGAAACTCCGGAGGGTTTTGGGGGGGGGGGGTACACGGCCCCC
CGGGGCTGAGCCCGCAGATTAAATAACTCTGACGTGGCAAGTGGCCTGAGAAGACA
GTGTAACATAATAATTGCACCTCGGAATTGCAGAGGGTCACTCCACTTGGACACAACAGGGC
TACTCGTAGGACCAGATAAGCACTTGCTCCCTGGACTGAAAAAGAAAGGATTATCTGTTGCTT
CTTGCTGACAAATCCCTGTGAAAGTAAAAGTCGGACACAGCAATCGATTATTCTCGCCTGTGTA
AATTACTGTGAATATTGAAATATATATATATATATATATCTGTATAGAACAGCCTCGGAGGCCGA
TGGACCCAGCGTAGATCATGCTGGATTGTACTGCCGAATTC

```

Results from the ORF finder:

ORF Length	ORF Sequence	Amino Acid Sequence
966	ATGGGAGCCATGGAGTCGTGGCGCTGGGGGG CCGGATGCGGGCTCCCCACTCCGTCCCTGATG AAGCCGGAGCCTCCTGGGCTGGGGGGGGCG AGAGGACGGAGGGGGGGCTGCTGGCCTCCT ACCCCCCTCAGGCCGCTGCCCTGGTGCCGT GGGCAGACACGGGTACTTGGGACCCCCCAGT GGGTGCCGCCGCCACCCAAATGGAGCCCCCCC ACTACCTGGAGCTGCTGCAACCCCCGGGGCA GCCCTCCATCCCTCCCTGGGCCCTACTGCC ACTCAGCAGCGGGCCCCCACCTGCGAGGCCCG TGAGTGCCTCATGCCAGGAAGAACTGCGGAGC GACGGCAACGCCGCTGTGGCGCCGGACGGCAC CGGGCATTACCTGTGCAACTGGGCTCAGCCTGC GGGCTCTACCACCGCCTAACGCCAGAACCGC CCGCTCATCCGCCAAAAAGGCCCTGCCGGTG AGTAAGCCGCAGGCACAGTGTGAGGCCACGAG CGTAAAACGTGCCAGACATCCACCAACTCTG TGGCGTCGCAGCCCATGGGGACCCCTCTGC AACAAACATTCAACGCCCTGCCCTACTACAAAC TGCACCAAGTGAACCGCCCCCTACGATGCGCA AAGACGGAATCCAAACCCGAAACCGCAAAGTT CCTCCAAGGGTAAAAGCGCGCCCCCGGGGG GGGAAACCCCTCCGCCACCGCGGGAGGGGGC GCTCTATGGGGGAGGGGGGACCCCTCTATG CCCCCCCCGCCGCCCTGGGCCGCC CCTCAAAGCGACGCTCTGTACGCTCTGCC GTGGTCCTTCCGGGCCATTCTGCCCTTGGAA ACTCCGGAGGGTTTTGGGGGGGGGGGGGG GTTACACGCCCGGGCTGAGCCCGAGA TTAA	MGAMEFVALGGPDAGS PTFPDDEAGAFLGLGGG ERTEAGLLASYPPSGR VSLVPWADTGLGTPQ WVPPATQMEPPHYLELL QPPRGSPPHSSGPLPL SSGPPCEARECVMARK NCGATATPLWRRDGTGH YLCNWASACGLYHRLN GQRPLIRPKKRLRVSK RAGTVCSHERENCQTST TTLWRRSPMGDPVCNNI HACGLYYKLHQVNRP MRKDGIQTRNRKVSSK GKKRRPPGGNP GGAPMGGGDPSMPPP PPPAAAPPQSDALYALG PVVLSGHFLPFGNSGGF FGGGAGGYTAPPGLSPQ I
957	ATGGAGTCGTGGCGCTGGGGGGCCGGATGCG GGCTCCCCACTCCGTCCCTGATGAAGCCGGA GCCTTCCTGGGCTGGGGGGGGCGAGAGGAC GGAGGCGGGGGGCTGCTGGCCTCCTACCCCC CTCAGGCCCGTGTCCCTGGTGCCTGGCAGA CACGGGTACTTGGGACCCCCCAGTGGGTGCC GCCGCCACCCAAATGGAGCCCCCCCAC GGAGCTGCTGCAACCCCCCGGGCAGCCCC CCATCCCTCCGCCCTACTGCCACTCAGC AGCGGGCCCCACCCCTGCGAGGCCCGTGAGTGC GTCATGCCAGGAAGAACTGCGGAGCGACGGCA ACGCCGCTGTGGCGCCGGACGGCACCGGGCAT	MEFVALGGPDAGSPTPF PDEAGAFLGLGGGERTE AGLLASYPPSGRVS PWADTGLGTPQWVPPA TQMEPPHYLELLQPPRG SPPHPSSGPLPLSSGPPP CEARECVMARKNC GATATPLWRRDGTGH YLCNWASACGLYHRLN GQRPLIRPKKRLRVSK RAGTVCSHERENCQT TTLWRRSPMGDPVCNNI HACGLYYKLHQVNRP MRKDGIQTRNRKVSSK GKKRRPPGGNP GGAPMGGGDPSMPPP PPPAAAPPQSDALYALG PVVLSGHFLPFGNSGGF FGGGAGGYTAPPGLSPQ I

	TACCTGTGCAACTGGGCCTCAGCCTGCGGGCTCT ACCACCGCCTCAACGGCCAGAACCGCCCCGCTCA TCCGCCCAAAAAGCGCCTCGGGGTGAGTAAGC GCGCAGGCACAGTGTGCAGCCACGAGCGTGAA AACTGCCAGACATCCACCACACTCTGTGGCGT CGCAGCCCCATGGGGGACCCGTCTGCAACAAAC ATTCACGCCCTGCGGCCTCTACTACAAACTGCACC AAGTGAACC GCCCCCTCACGATGCGCAAAGACG GAATCCAACCCGAAACCGAAAGTTCCCTCCA AGGGTAAAAGCGCGCCCCCGGGGG AACCCCTCCGCCACCGCGGGAGGGGGCGCTCCT ATGGGGGAGGGGGGACCCCTATGCC CCGCCGCCCGCCCGCCGCCGCCCTCAA AGCGACGCTCTGTACGCTCTCGGCCCGTGGTCC TTTCGGGCCATTCTGCCCTTGGAAACTCCGG AGGGTTTTGGGGGGGGCGGGGGTTACACGGCCCCCGGGG TGAGCCCGCAGATTAA	LYYKLHQVNRLPLTMRK DGIQTRNRKVSSKGKKR RPPGGNPSATAGGGAP MGGGGDPSMPPPPPPPA AAPPQSDALYALGPVVL SGHFLPGNSGGFFGGG AGGYTAPPGLSPQI
747	ATGGAGCCCCCCCCTACCTGGAGCTGCTGCAA CCCCCCCCGGGGCAGCCCCCCCCCATCCCTCCTCCG GGCCCTACTGCCACTCAGCAGCAGGGCCCCAC CCTGCAGGGCCGTGAGTGCCTATGCCAGGA AGAACTGCGGAGCGACGGCAACGCCCTGTGGC GCCGGGACGGCACCGGGATTACCTGTGCAACT GGGCCTAGCCTGCCGGCTCTACCACCGCCTCA ACGGCCAGAACCGCCCGCTCATCCGCCAAAAA AGCGCCTGCCGGGTGAGTAAGCGCGCAGGACAG TGTGCAGCCACGAGCGTGAAGACTGCCAGACAT CCACCACCACTCTGCGCTCGCAGCCCCATGG GGGACCCCGTCTGCAACACATTACGCCCTGCG GCCTCTACTACAAACTGCACCAAGTGAACCGCC CCCTCACGATGCGCAAAGACGGAATCAAACCC GAAACCGCAAAGTTCCCTCAAAGGGAAAAAGC GGCGCCCCCGGGGGGGAAACCCCTCCGCC CCGCGGGAGGGGGCGCTCTATGGGGGGAGGGG GGGACCCCTCTATGCC GGCCGCCGCCCTCAAAGCGACGCTCTGTA CGCTCTCGGCCCGTGGTCCTTCGGGCCATT CTGCCCTTGGAAACTCCGGAGGGTTTTGGGG GGGGGGCGGGGGGTTACACGGCCCCCGGGG TGAGCCCGCAGATTAA	MEPPHYLELLQPPRGSP PHPSSGPLLPLSSGPPPC EARECVMARKNCGATA TPLWRRDGTHYLCNW ASACGLYHRLNGQNRLP IRPKKRLRVSKRAGTV SHERENCQTSTTLWRR SPMGDPVCNNIHACGLY YKLHQVNRLPLTMRKDGI QTRNRKVSSKGKKRPP GGGNPSATAGGGAPMG GGGDPSMPPPPPPAA PPQSDALYALGPVVLSG HFLPGNSGGFFGGGAG GYTAPPGLSPQI
624	ATGGCCAGGAAGAACTGCGGAGCGACGGCAAC GCCGCTGCGCCGGGACGGCACCGGGCATT CCTGTGCAACTGGGCCTCAGCCTGCCGGCTCTA CCACCGCCTCAACGGCCAGAACCGCCGCTCAT CCGCCCCAAAAGCGCCTGCCGGGTGAGTAAGCG CGCAGGCACAGTGTGCAGCACGAGCGTGAAA ACTGCCAGACATCCACCAACTCTGTGGCGTC GCAGCCCCATGGGGGACCCCGTCTGCAACAAACA TTCACGCCCTGCCCTACTACAAACTGCACCA AGTGAACCGCCCCCTCACGATGCGCAAAGACGG AATCCAAACCGAAACCGCAAAGTTCCCTCAA GGTAAAAAGCGGGGCCCGGGGGGGGG ACCCCTCCGCCACCGCGGGAGGGGGCGCTCTA TGGGGGGAGGGGGGGACCCCTCTATGCC CGCCGCCGCCCTGCCGCCGCCCTCAA GCGACGCTCTGTACGCTCTGCCCTTGGAAACTCCGG GGGTTTTGGGGGGGGGGCGGGGGGGTTACACG GGCCCCCGGGCTGAGCCGCAGATTAA	MARKNCGATATPLWRR DGTGHYLCNWASACGL YHRLNGQNRLIRPKKR LRVSKRAGTVCSHEREN CQTSTTLWRRSPMGDP VCNNIHACGLYKHLQ VNRPLTMRKDGIQTRNR KVSSKGKKRPPGGNP SATAGGGAPMGGGDP SMPPPPPPAAAPPQSDA LYALGPVVLSGHFLPFG NSGGFFGGGAGGYTAPP GLSPQI

462	ATGCGGGCTCCCCACTCCGTTCCGTGATGAAGC CGGAGCCTCCTGGGGCTGGGGGGGGCGAGA GGACGGAGGCAGGGGGCTGCTGCCCTCCTACC CCCCCTCAGGCCGCGTGTCCCTGGTGCCGTGGG CAGACACGGGTACTTGGGGACCCCCCAGTGGG TGCCGCCCAGCCACCCAAATGGAGCCCCCCTACT ACCTGGAGCTGCTGCAACCCCCCGGGGAGGCC CCCCCATCCCTCCGGGCCCCACTGCCACT CAGCAGGGGCCACCCCTCGAGGCCGTGA GTGCGTCATGCCAGGAAGAACGCGGAGCGAC GGCAACGCCGCTGCGCCGGACGGCACCGG GCATTACCTGTGCAACTGGGCTCAGCTGC GCTCTACCACCGCCTCAACGCCAGAACCGCC GCTCATCCGCCAAAAAGGCCGTGCGGGTGA	MRAPPLRSLMKPEPSW GWGGARGRRGGCWP PTPPQAACPWCRGQTRV LWGPPSGCRPPPWSPP TTWSCCNPPGAAPPIPP GPYCHSAAGPHPARPVS ASWPGRTAERRQRRCG AGTAPGITCATGPQPAGS TTASTARTARSSAPKSAC G
456	ATGGCCGAAAGGACCACGGGGCGAGAGCGTA CAGAGCGTCGCTTGAGGGGGGGCGCGCGCG GGGGGGCGCGGGGGGGCATAGAGGGGTCCC CCCCCTCCCCCATAGGAGCGCCCCCTCCCGCGGT GGCGGAGGGTTTCCCCCCCCCGGGGGCGCCG CTTTTACCTTGGAGGAAACTTGCCTTTCGG GTTTGGATTCCGTCTTGCATCGTAGGGGGC GGTTCACTGGTGCAGTTGTAGTAGAGGCCGCA GGCGTAGATGTTGCAGACGGGGTCCCCCAT GGGGCTGCGACGCCACAGAGTGGTGGATGT CTGGCAGTTTCACGCTCGTGGCTGCACACTGTG CCTGCGCCTTACTCACCGCAGGCCCTTGG GGCGGATGAGGGCGGTTCTGGCCGGTGA GGTGGTAGAGGCCGCAGGCTGA	MARKDHGAESVQSVAL RGGGRGGRRGGHRCV PPSPHRAPSREGGGVS PPRGAPLFTLGGNFAVS GLDSVFAHREGAVHLVQ FVVEAGVNVDGVP HGAATPQSGGGCLAVFT LVAAHACALTHPQALF GADERAVLAVEAVVEPA G
435	ATGAAGCCGGAGCCTCCTGGGCTGGGGGGGG GCGAGAGGACGGAGGCAGGGGGCTGCTGGCC TCCTACCCCCCTCAGGCCGCGTGTCCCTGGTGC CGTGGGCAGACACGGGTACTTGGGGACCCCCC AGTGGGTGCCGCCGCCACCCAAATGGAGCCCC CCCACTACCTGGAGCTGCTGCAACCCCCCGGG GCAGCCCCCCCACCCCTCCGGGGCCCTACT GCCACTCAGCAGGGGCCACCCCTCGAGGC CCGTGAGTGCATGCCAGGAAGAACGCG AGCGACGCCAACGCCGCTGCGCCGGGACCG GCACCGGGCATTACCTGTGCAACTGGGCCTCAG CCTGCGGCTCTACCACGCCCTCAACGCCAGA ACCGCCCGCTCATGCCAAAAAGGCCGTGC GGGTGA	MKPEPSWGWWGARGR RGGCWPPPTPQAACP WCRGQTRVLWGPPSGC RPPPKWSPTTWSCCNP PGAAPPPIPPIPYCHSAA GPHPARPVSASWPGRTA ERRQRRCGAGTAPGITC ATGPQPGSTTASTARTA RSSAPKSACG
387	ATGGGGACCCGTCTGCAACAAACATTACGCC TGCGGCCTCTACTACAAACTGCACCAAGTGAAC CGCCCCCTCACGATGCCAAAGACGGAAATCAA ACCCGAAACGCCAAAGTTCTCCAAGGGTAA AAGCGGCCGGGGGGGGGGGGAAACCCCTCC GCCACCGCCGGAGGGGGCGCTCCTATGGGGGA GGGGGGACCCCTATGCCCTCCGGGCC CCCCGGCCGCCGCCCTCAAAGCAGC CTGTACGCTCTGCCCTGGTGTCTTCCGG ATTTTCTGCCCTTGAAACTCCGGAGGGTTTT GGGGGGGGGGCGGGGGTTACACGCC GGGGCTGAGGCCGCAGATTAA	MGDPVCNNIHACGLYY KLHQVNRPLTMRKDGIQ TRNRKVSSKGKKRPPG GGNPSATAGGGAPMGG GGDPSMPPPPPPAAPP QSDALYALGPVVLSGHF LPFGNSGGFFGGGAGGY TAPPGLSPQI
309	ATGCGCAAAGACGGAATCAAACCCGAAACCGC AAAGTTCTCCAAGGGAAAAAGCGGCC CCGGGGGGGGAAACCCCTCCGCCACCGCGGG AGGGGGCGCTCCTATGGGGGAGGGGGGACCC CTCTATGCCCTCCGGGCC GGGGCTGAGGCCGCAGATTAA	MRKDGIQTRNRKVSSK GKKRPPGGGNPSATAG GGAPMGGGDPSMPPP PPPAAAPPQSDALYALG PVVLSGHFLPFGNSGGF

	GCCCCCCCCTCAAAGCGACGCTCTGTACGCTCTCG GCCCGTGGTCCTTCGGGCCATTCTGCCCTT TGGAAACTCCGGAGGGTTTTGGGGGGGGGC GGGGGGTACACGGCCCCCCCAGGGCTGAGCCC GCAGATTAA	FGGGAGGYTAPPGLSPQ I
228	ATGTTACACTGTCTCTCAGCAAGGCCACTTGC CCACGTCAAGATTAAATCTGCGGGCTC AGCCCCGGGGGGGGCCGTGTAACCCCCCGCCCC CCCCCAAAAAACCTCCGGAGTTCCAAGGGC AGAAAATGGCCCAGAAGGACCACGGGGCGAG AGCGTACAGAGCGTCGCTTGAGGGGGGGCGGC GGCCGGGGGGGGCGCGGGGGGGCATAG	MLHCLLSKAHLPTSELL FKSAGSAPGGPCNPPPPP QKTLRSFQRAENGPKGP RGRERTERRFEGGRRPG GAAGGA
198	ATGGGGGGAGGGGGGGACCCCTATGCC CCGCCGCCCCCCCCGGCCGCCGCCCCCTCAA AGCGACGCTCTGTACGCTCTGGCCCCGTGGTCC TTTCGGGCCATTCTGCCCTTGGAAACTCCGG AGGGTTTTGGGGGGGGCGGGGGTTACAC GGCCCCCCCAGGGCTGAGCCCAGATTAA	MGGGGDPSMPPPPPPPA AAPPQSDALYALGPVVL SGHFLPFGNSGGFFGGG AGGYTAPPGLSPQI
174	ATGCCCCCCCCCGCCGCCCGGGCGCC CCCCTCAAAGCGACGCTCTGTACGCTCTGGCCC CGTGGTCCTTCGGCCATTCTGCCCTTGGAA AACTCCGGAGGGTTTTGGGGGGGGCGGGG GGTTACACGGCCCCCCCAGGGCTGAGCCCAG ATTAA	MPPPPPPAAAPPQSDAL YALGPVVLSGHFLPFGN SGFFGGAGGYTAPP LSPQI
171	ATGATCTACGCTGGGTCCATGCCGCC TGTTCTATAAGATATATATATATATATATT ACAATTACAGTAATTACACAGGGCAGAAA TAATCGATTGCTGTGTCGACTTTACCTTCACA GGGATTTCAGCAAGAACAGATAA	MIYAGSMPPRLFYTDIY IYIYIFTIFTVISHRREIID CCVRLLPFTGICQQEAN R
168	ATGTTGTTGCAGACGGGGTCCCCATGGGGCTGC GACGCCACAGAGTGGTGGATGTCTGGCAGT TTTCAGCCTCGTGGCTGCACACTGTGCC CTTACTCACCCGCAGGGCCTTTGGGGCGGATG AGCGGGCGGTTCTGGCCGTGAGGCGGTGGTAG	MLLQTGSPMGLRRHRV VVDVWQFSRSWLHTVP ARLLTRRRFLGRMSGRF WPLRRW
153	ATGCCGCCTCCGAGGCTGTTCTACAGATATATA TATATATATATATATTACAATATTACAGTAATT TCACACAGGCAGAAAATAATCAGATTGCTGTGTC GACTTTACCTTCACAGGGATTGTCAGCAAGA AGCAAACAGATAA	MPPPRLFYTDIYIYIFT IFTVISHRREIIDCCVLL PFTGICQQEANR
144	ATGGGGCTGCGACGCCACAGAGTGGTGGTGGAT GTCTGGCAGTTTCACGCTCGTGGCTGCACACTG TGCCTGCCGCTTACTCACCCGCAGGGCCTTT GGGGCGGATGAGCGGGCGGTCTGGCCGTGAG GCGGTGGTAG	MGLRRHRVVVDVWQFS RSWLHTVPARLLTRRRF LGRMSGFWPLRRW
135	ATGGGGGGGCTGCCCGGGGGGGTGCAGCAG CTCCAGGTAGTGGGGGGCTCCATTGGGTGG GGCGGCACCCACTGGGGGTCCCCAAAGTACC CGTGTCTGCCACGGCACCAGGGACACGCC TGA	MGGAAPGLQQLQVVG GLHLGGGRHPLGGPQST RVCPRHQGHAA
114	ATGCCCGGTGCCGCTCCGGGCCACAGCGCGT TGCCGTGCTCCGCAGTTCTCCTGGCCATGACG CACTCACGGGCCTCGCAGGGTGGGGGCCGCTG CTGAGTGGCAGTAG	MPGAVPAPQRCCRRAV LPGHDALTLAGWGPA AEWQ
111	ATGACGCACTCACGGGCCTCGCAGGGTGGGGC CCGCTGCTGAGTGGCAGTAGGGGCCGGAGGAG GGATGGGGGGCTGCCCGGGGGTTGCAGC	MTHSRASQGGGPLLSGS RGPEEGWGLPRGGCSS SR

	AGCTCCAGGTAG	
81	ATGTCTGGCAGTTTCACGCTCGTGGCTGCACAC TGTGCCTCGCGCTTACTCACCGCAGGCGCTTT TTGGGGCGGATGA	MSGSFHARGCTLCLRAY SPAGAFWGG
36	ATGAGCGGGCGGTTCTGGCCGTTGAGGCGGTGG TAG	MSGRFWPLRRW
18	ATGGCTCCCATGGGATAG	MAPMG
15	ATGGACCCAGCGTAG	MDPA
15	ATGCACTTCCAGTGA	MHFQ
9	ATGGGATAG	MG

Comments:

More ORFs were detected in the updated sequence than with the original ‘Dinosaur’ DNA sequence in Part 2:Section1. The ORFs returned by the ORD detector are also longer in length with this updated sequence, which could provide more information to the sequence, as long ORFs are often used to aid in initially identifying candidate protein-coding regions or functional RNA-coding regions in a DNA sequence. ([Deonier et.al, 2005](#)) However, this observation alone is not enough to suggest that the long reading frames are conclusive enough evidence for the presence of protein coding genes within the new sequence.

Code input:

```

#reading the fasta file
sequence = SeqIO.read("part2_Dino_DNA_v2.fasta", "fasta")

# Printing the sequence and its ID
print("Sequence:", sequence.seq)
print("ID:", sequence.id)

def find_all_ORFs(sequence):
    ORFs = []
    #finding reverse complement and iterating over 3 frames for both sequences
    for strand, nuc in [(+1, sequence.seq), (-1, sequence.seq.reverse_complement())]:
        for frame in range(3):
            length = 3 * ((len(sequence)-frame) // 3) # Multiple of three for codons
            for i in range(frame, length, 3):
                codon = nuc[i:i+3]
                if codon in ['ATG', 'AUG']:
                    for j in range(i+3, length, 3):
                        if nuc[j:j+3] in ['TAA', 'TAG', 'TGA', 'UAA', 'UAG', 'UGA']:
                            ORFs.append(nuc[i:j+3])
                            break
    ORFs.sort(key=len, reverse=True)
    return ORFs # Now returning a list of all ORFs, in order of size

ORFs = find_all_ORFs(sequence)

# Create a table to input results into
table = PrettyTable()

# Adding columns to the table
table.field_names = ["ORF Length", "ORF Sequence", "Amino Acid Sequence"]

# saving ORFs as fasta files and adding the translated amino acid sequence for each ORF into the table
for i, orf in enumerate(ORFs, 1):
    amino_acid_sequence = orf.translate(to_stop=True)
    table.add_row([len(orf), orf, amino_acid_sequence])
    record = SeqRecord(orf, id=f"ORF{i}", description=f"ORF{i} from sequence {sequence.id}")
    SeqIO.write(record, f"ORF{i}.fasta", "fasta")

# Print results in the table
print(table)

# Print the number of ORFs found
print("Number of ORFs found:", len(ORFs))

```

-----  
-----

Number of ORFs found: 25

### Sequence-2: ‘Dinosaur’ Protein analysis

Protein type	E-value and coverage	Comments
Erythroid transcription factor from Gallus <i>gallus</i>	E-value: 0 Coverage: 99%	-protein from the longest ORF -Gallus <i>gallus</i> = domestic chicken -Multiple hits for the same TF

### Sequence-2: comments and conclusions

The ORF finder output provided a lot of reasonably long protein sequences which come up as unknown on BLAST search, along with repetition after repetition of the sequence for Erythroid transcription factor from *Gallus gallus* in the longest sequence outputs.

First, like every good researcher, I will humour all possible explanation as to the origin of our second ‘Dinosaur’ sequence. Our current output from BLASTp is the amino acid sequence for the Erythroid transcription factor (of chicken origin), and a list of completely unrecognised sequences. According to Feduccia Alan in the journal of BioScience ([Alan et.al, pages 991-994, 2021](#)) it is well established that chickens are the closest living relatives of the theropod dinosaurs today. Therefore, it would not be unreasonable to consider whether or not this sequence could be from theropod-derived DNA. Infact, A Harvard University study led by John Asara found that Trex collagen (from fossil records) was more similar to bird collagen than any other group of animals tested; which links well to a comparative analysis of the chicken genome by the International Chicken Genome Sequencing Consortium, published in 2004 ([ICGSC, Nature432, 2004](#)), suggesting that many coding regions of the chicken genome, although shortened or interspaced with more C/G repeats over the course of evolution, are fairly unchanged. So, if chicken collagen is similar to T-rex collagen, then perhaps fibrillar collagen in chicken erythrocytes could be highly similar to fibrillar collagen in T-rex/theropod erythrocytes, and therefore maybe their erythroid transcription factors could be similar in structure too?

This seems unlikely.

Whilst it is true that, as we do not have Dinosaur DNA on file we would expect a few of our amino acid sequences to, perhaps, not return a match. And, as such, if we were (hypothetically) to be given one of the first complete sections of Dinosaur DNA to be discovered, in a Cambridge University IB MCB mini project, it would be possible to argue that what we are dealing with could potentially be Dinosaur DNA. I however have decided that the aforementioned reality is incredibly silly. Instead of jumping to such conclusions, I input the entire version 2 of the mysterious ‘Dinosaur’ nucleotide sequence into a BLASTn search.

This was my output:

Descriptions	Graphic Summary	Alignments	Taxonomy	Download	Select columns	Show 100	?					
Sequences producing significant alignments				GenBank	Graphics	Distance tree of results	MSA Viewer					
<input checked="" type="checkbox"/> select all 100 sequences selected				Description	Scientific Name	Max Score	Total Score	Query Cover	E value	Per. Ident	Acc. Len	Accession
<input checked="" type="checkbox"/>	<a href="#">Gallus gallus GATA binding protein 1 (globin transcription factor 1). (GATA1). mRNA</a>	<i>Gallus gallus</i>	1483	1483	66%	0.0	95.40%	1068	<a href="#">NM_205464.1</a>			
<input checked="" type="checkbox"/>	<a href="#">X.laevis GATA-binding protein (XGATA-2) gene. complete cds</a>	<i>Xenopus laevis</i>	628	628	25%	1e-174	98.34%	1938	<a href="#">M76564.1</a>			
<input checked="" type="checkbox"/>	<a href="#">PREDICTED: Xenopus laevis GATA binding protein 2 L homeolog (gata2L). transcript variant X3. mRNA</a>	<i>Xenopus laevis</i>	595	595	24%	1e-164	97.45%	3907	<a href="#">XM_018256579.2</a>			
<input checked="" type="checkbox"/>	<a href="#">PREDICTED: Xenopus laevis GATA binding protein 2 L homeolog (gata2L). transcript variant X2. mRNA</a>	<i>Xenopus laevis</i>	595	595	24%	1e-164	97.45%	3875	<a href="#">XM_018256580.2</a>			
<input checked="" type="checkbox"/>	<a href="#">PREDICTED: Xenopus laevis GATA binding protein 2 L homeolog (gata2L). transcript variant X1. mRNA</a>	<i>Xenopus laevis</i>	595	595	24%	1e-164	97.45%	3963	<a href="#">XM_018256578.2</a>			
<input checked="" type="checkbox"/>	<a href="#">Xenopus laevis GATA binding protein 2 L homeolog (gata2L). mRNA</a>	<i>Xenopus laevis</i>	595	595	24%	1e-164	97.45%	3899	<a href="#">NM_001090574.1</a>			
<input checked="" type="checkbox"/>	<a href="#">Gallus gallus breed Huxu chromosome 29</a>	<i>Gallus gallus</i>	531	1423	64%	3e-145	96.35%	6839231	<a href="#">CP100583.2</a>			

It is becoming more and more possible that, despite the broad dinosaur-themed claims, what we are really dealing with is a bunch of junk, non-peptide-coding DNA combined with the mRNA sequence of the *Gallus gallus* GATA binding protein 1. How anticlimactic.

Sadly, this sequence again appears to be of fictitious origins, and again is not comprehensively convincing enough to be Dinosaur DNA

## Part 3 - Olfactory receptor Hunting in Mammalian Genomes

### Introduction:

In Part 3 of this project, we were tasked with estimate the number and diversity of olfactory receptors in a group of mammals. Specifically, I chose to explore those of Humans, mice, cats, dolphins and dog (specifically that of a Great Dane).

### Searching for olfactory protein receptors:

```
>detected_olfactory_protein
MEKRNLTVVREFVLLGLPSSAEQQHLLSVLFCMYLATTGNNMLIIATIGFDSHLHSPMYFF
LSNLAFVDICFTSTVPQMVVNILTGTKTISFAGCLTQLFFFVFVNMDSSLCCVMAYDRYV
AICHPLHYTARMNLCLCVQLVAGLWLVTYLHALLHTVLIAQLSFCASNIIHHFFCDLNPLLQ
LSCSDVSFNVMIIFAVGGLLALTPLVCILVSYGLIFSTVLKITSTQGKQRAVSTCSCHLSVVVL
FYGTAIAVYFSPSSPHMPESDTLSTIMYSMVAPMLNPFIYTLRNRDMKRG
LQKMLLKCTVFQQQ
```

Python code to perform the BLAST search:

```

# Part 3

# Importing packages
from Bio import SeqIO
from Bio.Seq import Seq
from Bio.SeqRecord import SeqRecord
import subprocess
from Bio.Blast import NCBIXML

# Creating a BLAST database from the Homo_sapiens.GRCh38.pep.all.fa
subprocess.run(["makeblastdb", "-in", "Homo_sapiens.GRCh38.pep.all.fa", "-dbtype", "prot"])

# Performing a BLAST search
result = subprocess.run([
    "blastp",
    "-query", "detected_olfactory_protein.fasta",
    "-db", "Homo_sapiens.GRCh38.pep.all.fa",
    "-outfmt", "5",
    "-out", "blast_results.txt"
])

# Code to check if the BLAST search ran successfully
if result.returncode != 0:
    print("Error running BLAST search")
else:
    # Parse the BLAST results
    blast_record = NCBIXML.read(open("blast_results.txt"))

# Applying filters to filter the results
filtered_results = []
for alignment in blast_record.alignments:
    for hsp in alignment.hsps:
        # a) I need to check that each hit is significant (E-value <= 1e-30)
        # b) I need to check that the % similarity is >60%
        # c) I need to make sure that the alignment is close to full length e.g. > 290 amino acids
        if hsp.expect <= 1e-30 and hsp.identities / hsp.align_length >= 0.6 and hsp.align_length >= 290:
            # d) Now, recording which matched sequences (in the Homo_sapiens.GRCh38.pep.all.fa database) satisfy a), b), c)
            filtered_results.append(alignment)

# e) Trying to work out the most likely human gene corresponding to the provided sequence
# Printing the titles of the filtered alignments, including the gene name
for alignment in filtered_results:
    print(alignment.title)

# Sanity checks:
# Open a file in write mode
with open("filtered_results.txt", "w") as f:
    # Write the filtered results to the file
    for alignment in filtered_results:
        f.write(alignment.title + "\n")

# Print the filtered results
# Open the file in read mode
with open("filtered_results.txt", "r") as f:
    # Print the contents of the file
    print(f.read())

```

Due to system issues, I had to run the BLAST search in vscode using subroutines (see above), however this was my filtered sequence output:

```

>gnl|BL_ORD_ID|103003 ENSP00000386138.2 pep
chromosome:GRCh38:1:247757374:247758680:-1 gene:ENSG00000221888.4 transcript:ENST00000408896.4
gene_biotype:protein_coding transcript_biotype:protein_coding gene_symbol:OR1C1
description:olfactory receptor family 1 subfamily C member 1 [Source:HGNC Symbol;Acc:HGNC:8182]

MEKRNLTVVREFVLLGLPSSAEQQHLLSFLCMYLATTLGNNMLIATIGFDSHLHSPMY
FFLSNLAFVDICFTSTTVPQMVMNILTGTKTISFAGCLTQLFFFVFVNMDSSLCCVMAY
DRYVAICHPLHYTARMNLCLCVQLVAGLWLVTYHALLHTVLAQQLSFCASNIIHFFCD
LNPLLQLSCSDVSFNVMIIIFAVGGLALTPLVCILVSYGLIFSTVLKITSTQGKQRRAVST
CSCHLSVVLFYGTIAIVYFSPPSPHMPESDTLSTIMYSMVAPMLNPFIYTLRNNDMKRG
LQKMLLKCTVFQQQ

>gnl|BL_ORD_ID|103002 ENSP00000493457.1 pep
chromosome:GRCh38:1:247754846:247760556:-1 gene:ENSG00000221888.4 transcript:ENST00000641256.1
gene_biotype:protein_coding transcript_biotype:protein_coding gene_symbol:OR1C1
description:olfactory receptor family 1 subfamily C member 1 [Source:HGNC Symbol;Acc:HGNC:8182]

MEKRNLTVVREFVLLGLPSSAEQQHLLSFLCMYLATTLGNNMLIATIGFDSHLHSPMY
FFLSNLAFVDICFTSTTVPQMVMNILTGTKTISFAGCLTQLFFFVFVNMDSSLCCVMAY
DRYVAICHPLHYTARMNLCLCVQLVAGLWLVTYHALLHTVLAQQLSFCASNIIHFFCD
LNPLLQLSCSDVSFNVMIIIFAVGGLALTPLVCILVSYGLIFSTVLKITSTQGKQRRAVST
CSCHLSVVLFYGTIAIVYFSPPSPHMPESDTLSTIMYSMVAPMLNPFIYTLRNNDMKRG
LQKMLLKCTVFQQQ

>gnl|BL_ORD_ID|106110 ENSP00000305424.2 pep
chromosome:GRCh38:16:3188204:3206556:1 gene:ENSG00000168124.3 transcript:ENST00000304646.3
gene_biotype:protein_coding transcript_biotype:protein_coding gene_symbol:OR1F1
description:olfactory receptor family 1 subfamily F member 1 [Source:HGNC Symbol;Acc:HGNC:8194]

MSGTNQSSVSEFLLLGLSRQPQQQHLLFVFFLMSYLATVLGNLLIILSVSIDSCLHTPMY
FFLSNLFSVDICFTSTTVPKMLANHILETQTISFCGCLTQMFVFMFVDMDNFLAVMAY
DHFVAVCHPLHYTAKMTHQLCALLVAGLWVANLVNLHTLLMAPLSFCADNAITHFFCD
VTPLLKLCSDTHLNEVIILSEGALVMITPFLCILASYMHITCTVLKVPSTKGRWKAFST
CGSHLAVVLFYGTIAIVYFNPLSSHSAEKDTMATVLYTVVTPMLNPFIYSLRNRYLKGA
LKKVV

>gnl|BL_ORD_ID|103003 ENSP00000386138.2 pep
chromosome:GRCh38:1:247757374:247758680:-1 gene:ENSG00000221888.4 transcript:ENST00000408896.4
gene_biotype:protein_coding transcript_biotype:protein_coding gene_symbol:OR1C1
description:olfactory receptor family 1 subfamily C member 1 [Source:HGNC Symbol;Acc:HGNC:8182]

MEKRNLTVVREFVLLGLPSSAEQQHLLSFLCMYLATTLGNNMLIATIGFDSHLHSPMY
FFLSNLAFVDICFTSTTVPQMVMNILTGTKTISFAGCLTQLFFFVFVNMDSSLCCVMAY
DRYVAICHPLHYTARMNLCLCVQLVAGLWLVTYHALLHTVLAQQLSFCASNIIHFFCD
LNPLLQLSCSDVSFNVMIIIFAVGGLALTPLVCILVSYGLIFSTVLKITSTQGKQRRAVST
CSCHLSVVLFYGTIAIVYFSPPSPHMPESDTLSTIMYSMVAPMLNPFIYTLRNNDMKRG
LQKMLLKCTVFQQQ

>gnl|BL_ORD_ID|103002 ENSP00000493457.1 pep
chromosome:GRCh38:1:247754846:247760556:-1 gene:ENSG00000221888.4 transcript:ENST00000641256.1
gene_biotype:protein_coding transcript_biotype:protein_coding gene_symbol:OR1C1
description:olfactory receptor family 1 subfamily C member 1 [Source:HGNC Symbol;Acc:HGNC:8182]

MEKRNLTVVREFVLLGLPSSAEQQHLLSFLCMYLATTLGNNMLIATIGFDSHLHSPMY
FFLSNLAFVDICFTSTTVPQMVMNILTGTKTISFAGCLTQLFFFVFVNMDSSLCCVMAY
DRYVAICHPLHYTARMNLCLCVQLVAGLWLVTYHALLHTVLAQQLSFCASNIIHFFCD
LNPLLQLSCSDVSFNVMIIIFAVGGLALTPLVCILVSYGLIFSTVLKITSTQGKQRRAVST
CSCHLSVVLFYGTIAIVYFSPPSPHMPESDTLSTIMYSMVAPMLNPFIYTLRNNDMKRG
LQKMLLKCTVFQQQ

>gnl|BL_ORD_ID|106110 ENSP00000305424.2 pep chromosome:GRCh38:16:3188204:3206556:1
gene:ENSG00000168124.3 transcript:ENST00000304646.3 gene_biotype:protein_coding
transcript_biotype:protein_coding gene_symbol:OR1F1
description:olfactory receptor family 1 subfamily F member 1 [Source:HGNC Symbol;Acc:HGNC:8194]

MSGTNQSSVSEFLLLGLSRQPQQQHLLFVFFLMSYLATVLGNLLIILSVSIDSCLHTPMY
FFLSNLFSVDICFTSTTVPKMLANHILETQTISFCGCLTQMFVFMFVDMDNFLAVMAY
DHFVAVCHPLHYTAKMTHQLCALLVAGLWVANLVNLHTLLMAPLSFCADNAITHFFCD
VTPLLKLCSDTHLNEVIILSEGALVMITPFLCILASYMHITCTVLKVPSTKGRWKAFST
CGSHLAVVLFYGTIAIVYFNPLSSHSAEKDTMATVLYTVVTPMLNPFIYSLRNRYLKGA
LKKVV

```

As the above output only contained six sequences, I decided to check my output on the NCBI BLASTp online database. The filters on the online webpage are definitely more limited, so this will impact the output. Below we can see the filter options available Whilst I was able to filter for E values less than 1E-30, I was unable to impose a filter for percentage similarity. Percentage similarity is

distinctly different from query coverage, which measures how much of the query sequence is covered by the alignment. I was tempted to use percentage identity as an approximation for percentage similarity, however according to [Hebert et.al \(2003\)](#) and [Desalle et.al \(2005\)](#) BLAST percent identity scores do not necessarily equal the percent sequence similarity, especially as the sequence get more divergent and become harder to align. With the knowledge of this, I tested how the output changed when limiting percentage identity to greater than 60%, and it provided me with seven outputs – leading me to believe that the errors in my code may have been between an inability to discern between percentage similarity and percentage identity when applying my filters. In light of this, I decided to filter my outputs by E value only.

Your search is limited to records that include: Homo sapiens (taxid:9606)  
Your results are filtered to match records with expect value between 0 and 1e-30.

Job Title	Protein Sequence
RID	<a href="#">2M58K15R016</a> Search expires on 04-26 16:32 pm <a href="#">Download All</a>
Program	BLASTP <a href="#">?</a> <a href="#">Citation</a>
Database	nr <a href="#">See details</a>
Query ID	IclQuery_2588255
Description	unnamed protein product
Molecule type	amino acid
Query Length	314
Other reports	<a href="#">Distance tree of results</a> <a href="#">Multiple alignment</a> <a href="#">MSA viewer</a> <a href="#">?</a>

**Filter Results**

**Organism** only top 20 will appear  exclude  
Type common name, binomial, taxid or group name  
[+ Add organism](#)

Percent Identity  to   
E value  to  1e-30  
Query Coverage  to

**Filter** **Reset**

I then selected 250 outputs that matched my criteria, downloaded them into a text file direct from the NCBI BLAST+ webpage and converted the text file into a fasta format for MSA analysis. See below for my code:

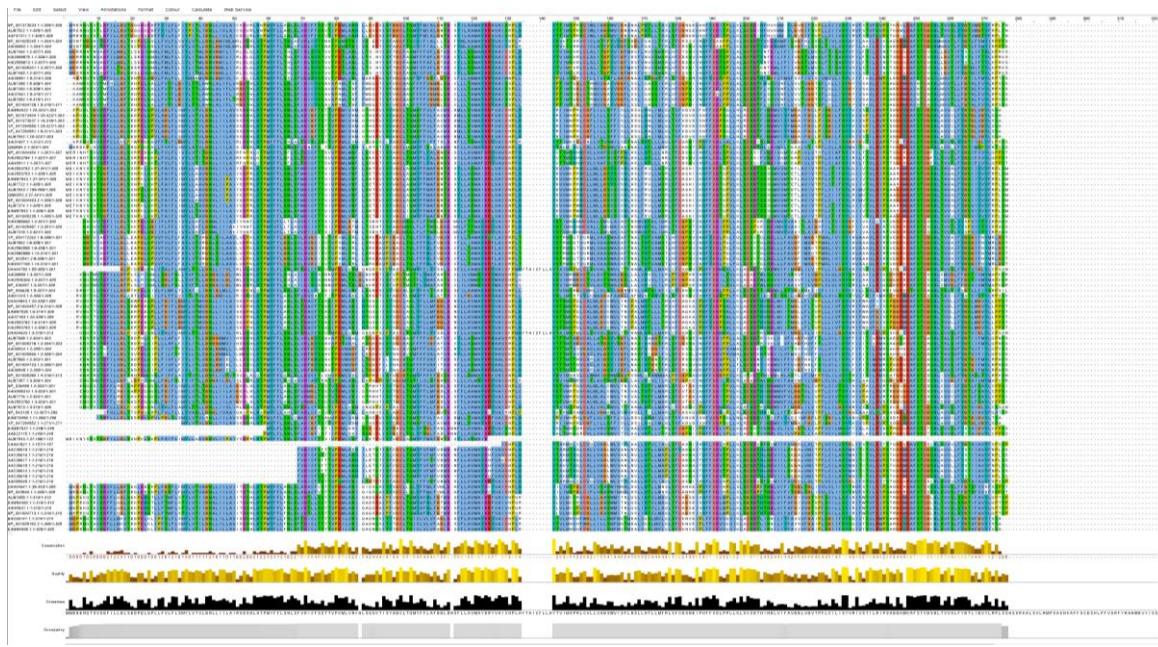
```
#Converting text files to fasta files
def txt_to_fasta(txt_file, fasta_file):
    with open(txt_file, 'r') as f:
        sequence = f.read().replace('\n', '')

    with open(fasta_file, 'w') as f:
        f.write(f">\n{sequence}")

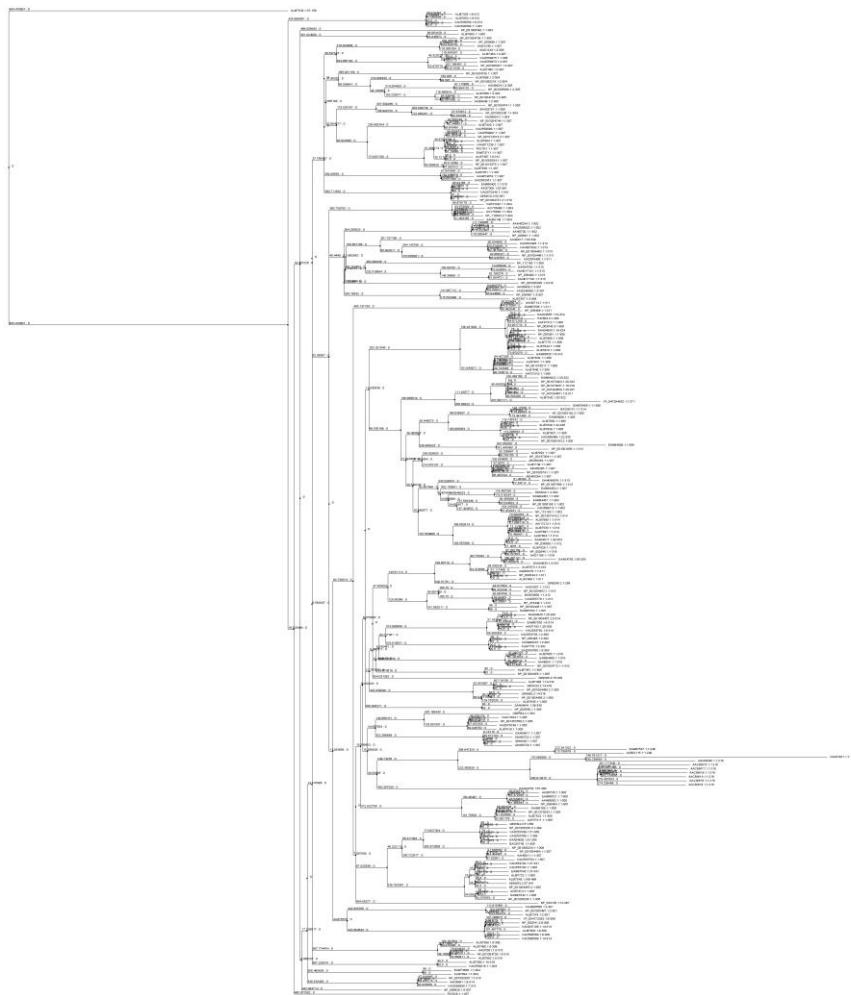
txt_file = "seqdump.txt"
fasta_file = "jalview.fasta"
txt_to_fasta(txt_file, fasta_file)
```

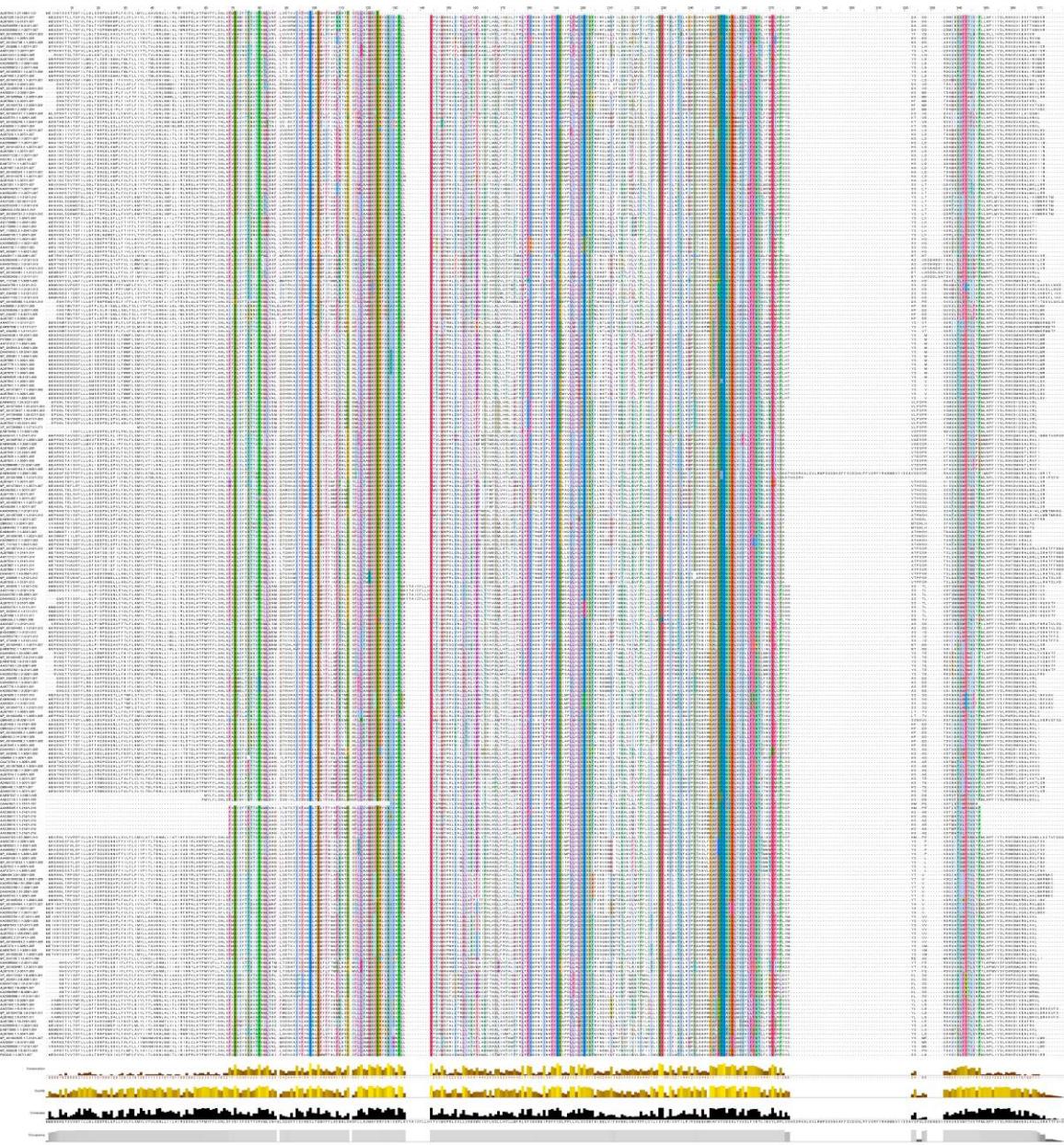
### MSA Analysis:

The sequences were analysed and the format edited when needed in jalview to ensure that the fasta file output could be read by the hmmbuild function.



Full versions:





After proofing and editing through the file in Jalview, I then downloaded my MSA fasta data to build the HMM model (and called it jalview.fa).

### The Hmm Model:

#### Expected outputs:

The values listed below are approximate, dependent on the literature cited. However, when reading through publications there appeared to be very little extreme deviations in number of olfactory receptors of the chosen mammals (although dolphin was quite hard to find references to).

- According to Nimura and Nei (2003) humans have almost 400 functional olfactory receptors ([Nimura and Nie, 2003](#)).
- According to Young et.al (2003), mice should have between 1200 and 1500 olfactory receptors ([Young et.al, 2003](#)).

- According to Quignon et.al (2003) dogs should have around 1070 olfactory receptors, although it is important to note that the average number of olfactory receptors for specific dog breeds can differ. ([Quignon et.al, 2003](#)).
- Many reports recorded dolphin olfactory receptor numbers as NaN, however Kremers et.al (2016) reported the value to be around 37, which felt sufficiently low enough to accept as an expected value. ([Kremers et.al, 2016](#)). Whilst this value might feel low for an aquatic mammal, dolphins rely more heavily on their other senses and also process smells in a completely different way to terrestrial mammals.
- In the literature, exact values for numbers of cat olfactory receptors are not given, instead the number is said to be high.

```

import matplotlib.pyplot as plt

# building HMM
!hmmbuild human.hmm jalview.fa

# cat, dog (great-dane), mouse, dolphin, human

#cat
!hmmsearch human.hmm cat.pep.all.fa > cat.out
#great dane
!hmmsearch human.hmm great_dane.pep.all.fa > great_dane.out
# mouse
!hmmsearch human.hmm Mouse.pep.all.fa > mouse.out
#dolphin
!hmmsearch human.hmm dolphin.pep.all.fa > dolphin.out
#human
!hmmsearch human.hmm human.pep.all.fa > human.out

#plotting
# using parse() filter
def parse(file):
    with open(file, 'r') as f:
        lines = f.read().split('\n') # split by new lines
        data = [line.split(',') for line in lines if line] # split by commas and filter out empty lines
        data = [[value for value in row if value] for row in data] # filter out empty strings in each row
    return len(data) # return the number of matches

species = ['human', 'cat', 'great_dane', 'mouse', 'dolphin']
counts = [parse(f"{s}.out") for s in species]

colors = ['green', 'orange', 'purple', 'pink', 'blue']

plt.bar(species, counts, color = colors)
plt.xlabel('Species')
plt.ylabel('Number of receptors')
plt.show()

```

```

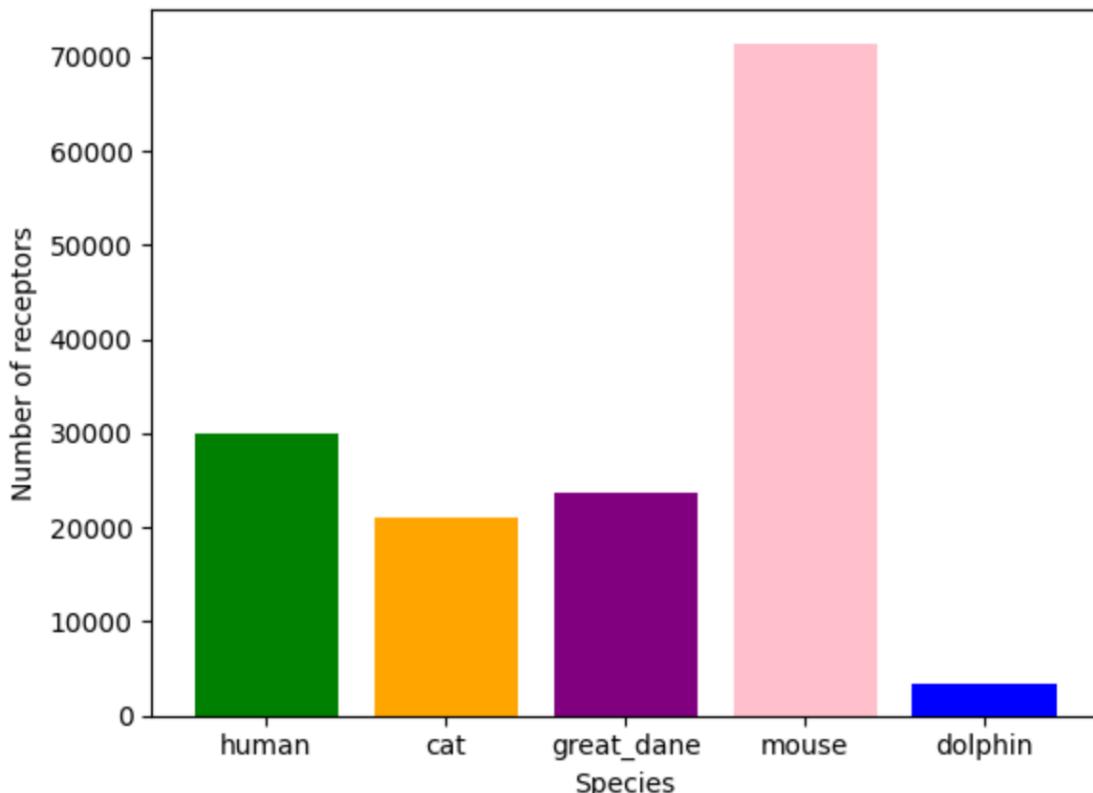
# hmmbuild :: profile HMM construction from multiple sequence alignments
# HMMER 3.3.2 (Nov 2020); http://hmmer.org/
# Copyright (C) 2020 Howard Hughes Medical Institute.
# Freely distributed under the BSD open source license.
#
# input alignment file: jalview.fa
# output HMM file: human.hmm
#
# -----



# idx name          nseq  alen  mlen eff_nseq re/pos description
#----- -----
1   jalview        250   381   307    1.23  0.591

# CPU time: 0.20u 0.00s 00:00:00.20 Elapsed: 00:00:00.20

```



```

def print_receptor_counts(species, counts):
    for s, c in zip(species, counts):
        print(f"The number of olfactory receptors in {s} is {c}.")  
  

print_receptor_counts(species, counts)

```

The number of olfactory receptors in human is 30018.  
The number of olfactory receptors in cat is 20980.  
The number of olfactory receptors in great\_dane is 23646.  
The number of olfactory receptors in mouse is 71468.  
The number of olfactory receptors in dolphin is 3316.

The model appears to over-estimate for all species proportionally, implying that perhaps if the parse() took into account sequence similarity and E-value, the overall outputs for all could have decreased. However, as they remain somewhat proportional I will not re-run the hmmer.

## References

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- Kremers et.al, 2016: <https://doi.org/10.3389/fevo.2016.00049>