Design of scalable gene panels for carrier screening

dystrophinopathies

maple syrup urine

oculocutaneous albinism

Mulibrey nanism

disease la

X-linked combined variable

combined immunodeficiency)

immunodeficiency (including severe | IL2RG



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• Severe²

• Severe²

• Severe²

• 1/3,500 males (includes de novo cases)³

Not on NBS, but on an existing panel

Present on NBS and an existing panel

• Founder effect in Mennonite population²

Present on NBS panel and an existing panel

• May be more common in Japan and India^{3,4}

• Low prevalence (110 cases described as of 2004)⁵

TRIM37 • Does not add detection rate to a disease on current panel

• Probable founder effect in Finland (85 cases reported)

Not on NBS, but on an existing panel

Not on NBS, but on an existing panel

• No known founder effect

• No known founder effect

• 1/50,000-100,0003

• 1/185,000 combined³

BCKDHA • MSUD Ib on current panel

• Moderate²

• 1/40,0003

• Moderate²

Adds detection to muscular dystrophies overall

• Does not add detection rate to a disease on current panel

• Does not add detection rate to a disease on current panel

South San Francisco, CA

Background

For every 2,000 births worldwide, nearly three children have a severe autosomal-recessive or an X-linked disorder, and only ~10% of all inherited diseases may be effectively managed with treatment.¹ Expanded carrier screening (ECS) can help mitigate the impact of such diseases by either decreasing the number of affected pregnancies or improving care for an affected child via early diagnosis. The number of diseases on an ECS panel affects both its economic viability and clinical utility; thus, panel expansions must be done judiciously. Here we report a principled method by which disease genes can be ranked to guide ECS panel expansion.

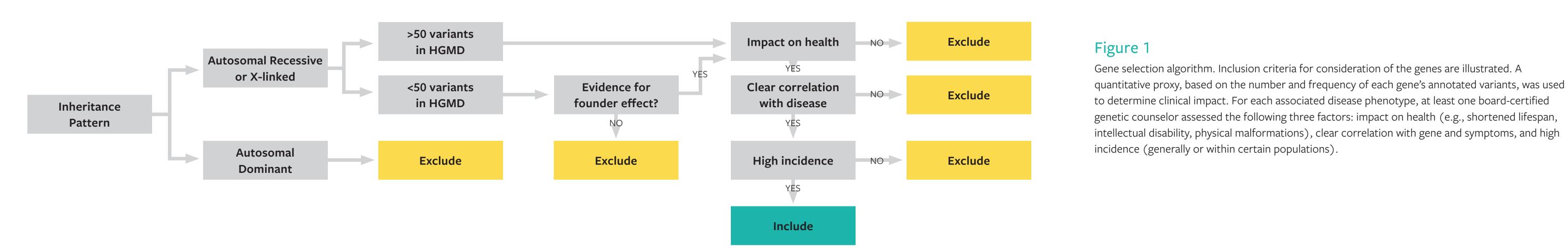
Methods

We reviewed and ranked genes obtained from commercial ECS panels and Online Mendelian Inheritance in Man (Figure 1 and Table 1). Then, screening protocols were optimized for workflow and assay cost by consideration of homology, assay compatibility, and gene size (Figure 2).

Conclusion

Collectively, the entire process described above enables a general and principled selection of severe diseases for a clinically and economically viable ECS panel that identifies the most at-risk couples.

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Diseases that were included based on the gene selection algorithm were further ranked based on: 1) disease severity, 2) disease incidence, 3) added detection rate to a specific disease phenotype already on the ECS panel, 4) added detection rate for genetic disease in a specific ethnic group, and 5) presence on the newborn screen (NBS) and/or an existing commercial ECS panel. Example rankings are listed below.

Figure 2

Two factors constrain how many genes comprise the final ECS panel: 1) the difficulty of assessing carrier status in the desired genes and 2) the aggregate number of positions being probed.

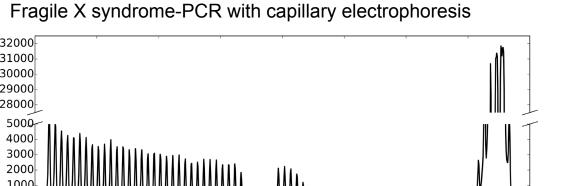
A Variant identification in genes can be challenging for a variety of reasons, including the presence of homologous regions (e.g., the CYP21A2 gene underlying 21-hydroxylase-deficient congenital adrenal hyperplasia (CAH) has a 99% identical pseudogene, CYP21A1P) or repeat sequences (e.g., CGG repeat in FMR1). Either custom NGS solutions or non-NGS assays (e.g., triplet-primed

B The size of the genome interrogated is a major limiting factor in panel design, especially for tests genes considered for inclusion on a commercial ECS panel are organized by coding-sequence length, which depicts that a single large gene can consume 40x as much NGS bandwidth as a smaller gene.

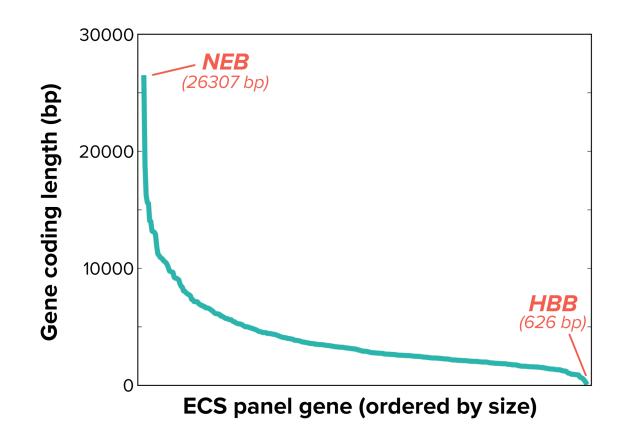
CITATIONS 1 | World Health Organization (http://apps.who.int/iris/bitstream/10665/41846/1/ WHO_TRS_865.pdf), Control of Hereditary Diseases, 1996 2 | Lazarin et al., PLoS One, 2014 (PMID: 25494330) 3 | GeneReviews 4 | Ankala et al., Hum Mutat, 2015 (PMID: 25323826) 5 | Karlberg et al., N Engl J Med, 2004 (PMID: 15590968)

A Homology associated with 21-hydroxylase-deficient CAH





B Genes by variable size of coding length



PCR followed by capillary electrophoresis for fragile X syndrome) are needed.

performing full-exon NGS. Coding-sequencing length varies greatly from gene-to-gene. Below, the