Maximum likelihood estimation and analysis with the bbmle package

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The bbmle package, designed to simplify maximum likelihood estimation and analysis in R, extends and modifies the mle function and class in the stats4 package that comes with R by default. mle is in turn a wrapper around the optim function in base R. The maximum-likelihood-estimation function and class in bbmle are both called mle2, to avoid confusion and conflict with the original functions in the stats4 package. The major differences between mle and mle2 are:

- mle2 is slightly more robust, with additional warnings (e.g. if the Hessian can't be computed by finite differences, mle2 returns a fit with a missing Hessian rather than stopping with an error)
- mle2 uses a data argument to allow different data to be passed to the negative log-likelihood function
- mle2 has a formula interface like that of (e.g.) gls in the nlme package. For relatively simple models the formula for the maximum likelihood can be written in-line, rather than defining a negative log-likelihood function. The formula interface also simplifies fitting models with categorical variables.
- bbmle defines anova, AIC, AICc, and BIC methods for mle2 objects, as well as AICtab, BICtab, AICctab functions for producing summary tables of information criteria for a set of models.

Other packages with similar functionality (extending GLMs in various ways) are aod and vgam (on CRAN), gnlr and gnlr3 in Jim Lindsey's gnlm package (http://popgen.unimaas.nl/~jlindsey/rcode.html).

1 Example

This example will use the classic data set on *Orobanche* germination from [1] (you can also use glm(...,family="quasibinomial") or the aod package to analyze these data).

1.1 Test basic fit to simulated beta-binomial data

First, generate a single beta-binomially distributed set of points as a simple test. Load the emdbook package to get functions for the beta-binomial distribution (density and random-deviate function — these functions are also available in Jim Lindsey's rmutil package).

```
> library(emdbook)
   Generate random deviates from a random beta-binomial:
> set.seed(1001)
> x1 = rbetabinom(n = 1000, prob = 0.1, size = 50, theta = 10)
  Load the package:
> library(bbmle)
   Construct a simple negative log-likelihood function:
> mtmp <- function(prob, size, theta) {
      -sum(dbetabinom(x1, prob, size, theta, log = TRUE))
+ }
  Fit the model — use data to pass the size parameter (since it wasn't hard-
coded in the mtmp function):
> m0 <- mle2(mtmp, start = list(prob = 0.2, theta = 9),
      data = list(size = 50))
> mO
Call:
mle2(minuslog1 = mtmp, start = list(prob = 0.2, theta = 9), data = list(
    size = 50))
Coefficients:
      prob
                 theta
 0.1030974 10.0758090
Log-likelihood: -2723.5
```

The summary method for mle2 objects shows the parameters; approximate standard errors (based on quadratic approximation to the curvature at the maximum likelihood estimate); and a test of the parameter difference from zero based on this standard error and on an assumption of normality.

```
> summary(m0)
```

Maximum likelihood estimation

```
Call:
```

```
mle2(minuslog1 = mtmp, start = list(prob = 0.2, theta = 9), data = list(
    size = 50))
```

Coefficients:

```
Estimate Std. Error z value Pr(z)
prob 0.1030974 0.0031624 32.601 < 2.2e-16 ***
theta 10.0758090 0.6213189 16.217 < 2.2e-16 ***
---
Signif. codes: 0 '***, 0.001 '**, 0.01 '*, 0.05 '., 0.1 ', 1
```

-2 log L: 5446.995

Construct the likelihood profile (you can apply confint directly to m0, but if you're going to work with the likelihood profile (e.g. plotting, or looking for confidence intervals at several different α values) then it is more efficient to compute the profile once):

> p0 <- profile(m0)

Compare the confidence interval estimates based on inverting a spline fit to the profile (the default); based on the quadratic approximation at the maximum likelihood estimate; and based on root-finding to find the exact point where the profile crosses the critical level.

> confint(p0)

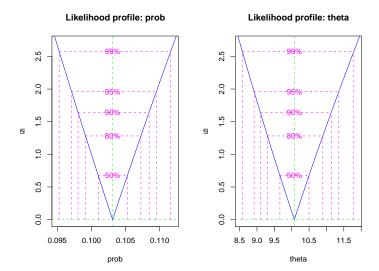
```
2.5 % 97.5 % prob 0.09709228 0.1095103 theta 8.91708211 11.3559591
```

> confint(m0, method = "quad")

> confint(m0, method = "uniroot")

All three types of confidence limits are similar. Plot the profiles:

```
> par(mfrow = c(1, 2))
> plot(p0, plot.confstr = TRUE)
```



By default, the plot method for likelihood profiles displays the square root of the the deviance (twice the difference in negative log-likelihood), so it will be V-shaped for cases where the quadratic approximation works well (as in this case). (For a better visual estimate of whether the profile is quadratic, use absVal=FALSE.)

You can also request confidence intervals calculated using uniroot, which may be more exact when the profile is not smooth enough to be modeled accurately by a spline. However, this method is also more sensitive to numeric problems.

Instead of defining an explicit function for minuslog1, we can also use the formula interface. The formula interface assumes that the density function given (1) has x as its first argument (if the distribution is multivariate, then x should be a matrix of observations) and (2) has a log argument that will return the log-probability or log-probability density if log=TRUE.

```
> m0f <- mle2(x1 \tilde{} dbetabinom(prob, size = 50, theta),
+ start = list(prob = 0.2, theta = 9))
```

It's convenient to use the formula interface to try out likelihood estimation on the transformed parameters:

```
> confint(mOcf, method = "spline")
```

```
2.5 % 97.5 % lprob -2.229963 -2.095756 ltheta 2.187948 2.429742
```

In this case the answers from uniroot and spline (default) methods barely differ.

1.2 Using real data

Get data from Crowder 1978 [1], as incorporated in the aod package:

> library(aod)

Package aod, version 1.1-24

> data(orob1)

Now construct a negative log-likelihood function that differentiates among groups:

```
> ML1 <- function(prob1, prob2, prob3, theta, x) {
+     prob <- c(prob1, prob2, prob3)[as.numeric(x$dilution)]
+     size <- x$n
+     -sum(dbetabinom(x$y, prob, size, theta, log = TRUE))
+ }</pre>
```

Results from [1]:

model	prob1	prob2	prob3	theta	sd.prob1	sd.prob2	sd.prob3	NLL
prop diffs	0.132	0.871	0.839	78.424	0.027	0.028	0.032	-34.991
full model								-34.829
homog model								-56.258

```
> m1 <- mle2(ML1, start = list(prob1 = 0.5, prob2 = 0.5,
+    prob3 = 0.5, theta = 1), data = list(x = orob1))
> m1

Call:
mle2(minuslog1 = ML1, start = list(prob1 = 0.5, prob2 = 0.5,
    prob3 = 0.5, theta = 1), data = list(x = list(dilution = c(1L,
1L, 1L, 1L, 1L, 2L, 2L, 2L, 2L, 2L, 3L, 3L, 3L, 3L),
    n = c(43L, 51L, 44L, 71L, 24L, 7L, 19L, 56L, 87L, 55L, 10L,
    13L, 62L, 104L, 51L, 11L), y = c(2L, 9L, 5L, 16L, 2L, 0L,
    17L, 43L, 79L, 50L, 9L, 11L, 47L, 90L, 46L, 9L))))
```

Coefficients:

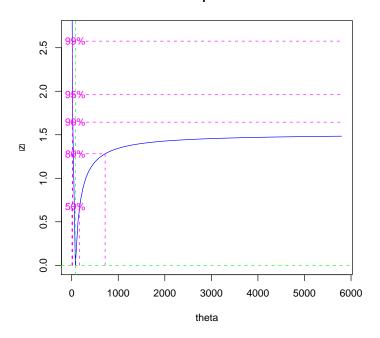
```
prob1
                prob2
                           prob3
                                       theta
 Log-likelihood: -34.99
Warning: optimization did not converge (code 1)
  The result warns us that the optimization has not converged; we also don't
match Crowder's results for \theta exactly. We can fix this by setting parscale
appropriately.
> m2 <- mle2(ML1, start = as.list(coef(m1)), control = list(parscale = coef(m1)),
      data = list(x = orob1))
> m2
Call:
mle2(minuslog1 = ML1, start = as.list(coef(m1)), data = list(
    x = list(dilution = c(1L, 1L, 1L, 1L, 1L, 1L, 2L, 2L, 2L,
    2L, 2L, 3L, 3L, 3L, 3L, 3L), n = c(43L, 51L, 44L, 71L, 24L,
   7L, 19L, 56L, 87L, 55L, 10L, 13L, 62L, 104L, 51L, 11L), y = c(2L, 104L, 51L, 11L)
    9L, 5L, 16L, 2L, 0L, 17L, 43L, 79L, 50L, 9L, 11L, 47L, 90L,
    46L, 9L))), control = list(parscale = coef(m1)))
Coefficients:
     prob1
                prob2
                           prob3
                                      theta
 Log-likelihood: -34.99
   Calculate likelihood profile:
> p2 <- profile(m2)
   Get the curvature-based parameter standard deviations (which Crowder used
rather than computing likelihood profiles):
> round(sqrt(diag(vcov(m2))), 3)
prob1 prob2 prob3 theta
 0.028 0.029 0.032 74.238
We are slightly off Crowder's numbers — rounding error?
   Crowder also defines a variance (overdispersion) parameter \sigma^2 = 1/(1+\theta).
> sqrt(1/(1 + coef(m2)["theta"]))
    theta
0.1122089
```

```
Using the delta method to get the standard deviation of \sigma:
```

```
> sqrt(deltavar(sqrt(1/(1 + theta)), meanval = coef(m2)["theta"],
      vars = "theta", Sigma = vcov(m2)[4, 4]))
[1] 0.05244163
   Another way to fit in terms of \sigma rather than \theta is to compute \theta = 1/\sigma^2 - 1
on the fly in a formula:
> m2b <- mle2(y ~ dbetabinom(prob, size = n, theta = 1/sigma^2 -
      1), data = orob1, parameters = list(prob ~ dilution,
      sigma ~ 1), start = list(prob = 0.5, sigma = 0.1))
> round(sqrt(diag(vcov(m2b))), 3)["sigma"]
sigma
0.052
> p2b <- profile(m2b)</pre>
   As might be expected since the standard deviation of \sigma is large, the quadratic
approximation is poor:
> r1 <- rbind(confint(p2)["theta", ], confint(m2, method = "quad")["theta",</pre>
> rownames(r1) <- c("spline", "quad")</pre>
> r1
            2.5 %
                     97.5 %
spline 19.81826
      -67.08021 223.9258
quad
   Plot the profile:
```

> plot(p2, which = "theta", plot.confstr = TRUE)

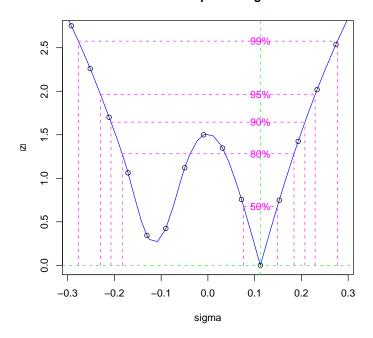
Likelihood profile: theta



What does the profile for σ look like?

> plot(p2b, which = "sigma", plot.confstr = TRUE, show.points = TRUE)

Likelihood profile: sigma



Now fit a homogeneous model:

```
> ml0 <- function(prob, theta, x) {
+    size <- x$n
+    -sum(dbetabinom(x$y, prob, size, theta, log = TRUE))
+ }
> m0 <- mle2(ml0, start = list(prob = 0.5, theta = 100),
+    data = list(x = orob1))</pre>
```

The log-likelihood matches Crowder's result:

> logLik(m0)

'log Lik.' -56.25774 (df=2)

It's easier to use the formula interface to specify all three of the models fitted by Crowder (homogeneous, probabilities differing by group, probabilities and overdispersion differing by group):

```
> m0f <- mle2(y ~ dbetabinom(prob, size = n, theta), parameters = list(prob ~
+ 1, theta ~ 1), data = orob1, start = list(prob = 0.5,
+ theta = 100))
> m2f <- mle2(y ~ dbetabinom(prob, size = n, theta), parameters = list(prob ~
+ dilution, theta ~ 1), data = orob1, start = list(prob = 0.5,</pre>
```

```
theta = 78.424))
> m3f <- mle2(y ~ dbetabinom(prob, size = n, theta), parameters = list(prob ~
      dilution, theta ~ dilution), data = orob1, start = list(prob = 0.5,
      theta = 78.424))
   anova runs a likelihood ratio test on nested models:
> anova(mOf, m2f, m3f)
Likelihood Ratio Tests
Model 1: mOf, y~dbetabinom(prob,size=n,theta): prob~1, theta~1
Model 2: m2f, y~dbetabinom(prob,size=n,theta): prob~dilution, theta~1
Model 3: m3f, y~dbetabinom(prob, size=n, theta): prob~dilution,
          theta~dilution
 Tot Df Deviance
                    Chisq Df Pr(>Chisq)
1
       2 112.515
         69.981 42.5341 2 5.805e-10 ***
           69.981 0.0008 2
                                 0.9996
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' '1
   The various ICtab commands produce tables of information criteria, option-
ally sorted and with model weights.
> AICtab(mOf, m2f, m3f, weights = TRUE, delta = TRUE, sort = TRUE)
    AIC
          df dAIC weight
m2f 78.0 4
               0.0 0.881
m3f 82.0 6
               4.0 0.119
mOf 116.5 2
            38.5 < 0.001
> BICtab(mOf, m2f, m3f, delta = TRUE, nobs = nrow(orob1),
      sort = TRUE, weights = TRUE)
    BIC
         df dBIC weight
m2f 81.1 4
              0.0 0.941
m3f 86.6 6
              5.5 0.059
mOf 118.1 2
              37.0 < 0.001
> AICctab(mOf, m2f, m3f, delta = TRUE, nobs = nrow(orob1),
      sort = TRUE, weights = TRUE)
    AICc df dAICc weight
m2f 81.6 4
              0.0 0.992
m3f 91.3 6
              9.7 0.008
mOf 117.4 2 35.8 < 0.001
```

Additions/enhancements/differences from stats4::mle

- anova method
- warnings on convergence failure
- more robust to non-positive-definite Hessian; can also specify skip.hessian to skip Hessian computation when it is problematic
- when profiling fails because better value is found, report new values
- can take named vectors as well as lists as starting parameter vectors
- added AICc, BIC definitions, ICtab functions
- added "uniroot" and "quad" options to confint
- more options for colors and line types etc etc. The old arguments are:

```
> function(x, levels, conf = c(99, 95, 90, 80, 50)/100,
+ nseg = 50, absVal = TRUE, ...) {
+ }
```

The new one is:

```
> function(x, levels, which = 1:p, conf = c(99, 95, 90,
+ 80, 50)/100, nseg = 50, plot.confstr = FALSE, confstr = NULL,
+ absVal = TRUE, add = FALSE, col.minval = "green",
+ lty.minval = 2, col.conf = "magenta", lty.conf = 2,
+ col.prof = "blue", lty.prof = 1, xlabs = nm, ylab = "score",
+ show.points = FALSE, main, xlim, ylim, ...) {
+ }
```

which selects (by character vector or numbers) which parameters to plot: nseg does nothing (even in the old version); plot.confstr turns on the labels for the confidence levels; confstr gives the labels; add specifies whether to add the profile to an existing plot; col and lty options specify the colors and line types for horizontal and vertical lines marking the minimum and confidence vals and the profile curve; xlabs gives a vector of x labels; ylab gives the y label; show.points specifies whether to show the raw points computed.

- mle.options()
- data argument
- handling of names in argument lists
- can use alternative optimizers (nlminb, constrOptim)

Bugs, wishes, to do

- BUG: mle2 fits that are obtained within a function can't be used for subsequent profiles etc. (environment issue)
- \bullet $\mathbf{WISH}:$ subset and predict
- minor **WISH**: better methods for extracting **nobs** information when possible (e.g. with formula interface)
- WISH: better documentation, especially for S4 methods
- \bullet WISH: variable-length chunks in argument list
- WISH: limited automatic differentiation (add capability for common distributions)

References

[1] Martin J. Crowder. Beta-binomial Anova for proportions. *Applied Statistics*, 27(1):34–37, 1978.