# Using R for scientific computing - ANSWERS

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### Abstract

The answers to the exercises from the document: "Using R for scientific computing" (Soetaert 2008).

Keywords: scientific computing, lecture notes, R.

This document gives the answers to the exercises in the lecture notes:

"Using R for scientific computing" (Soetaert 2008).

These notes are an introduction to  ${\sf R}$  - at beginners level. They can be found in package marelac.

In R, write:

require(marelac)

In order to make this vignette more readable, the questions are repeated.

# Chapter 1

No exercises in this chapter

# Chapter 2 - R as a scientific calculator

```
> (4/6*8-1)^(2/3)
```

[1] 2.657958

> log(20)

[1] 2.995732

> log2(4096)

[1] 12

```
> 2*pi*3
[1] 18.84956
> exp(2+cos(0.5*pi))
[1] 7.389056
> # length of 3rd side of a triangle with size 2.3 and 5.4 and angle pi/8
> sqrt(2.3^2+5.4^2-2*2.3*5.4*cos(pi/8))
[1] 3.391288
```

# Chapter 3 - computing with R-variables

```
Chapter 3.8.1
```

Use R-function mean to estimate the mean of two numbers, 9 and 17.

```
> mean(c(9,17))
```

[1] 22 32 36

# [1] 13

- Create a vector, called V, with even numbers, between 16 and 56. Do not use loops.
- Display this vector
- What is the sum of all elements of V?
- Display the first 4 elements of V
- Calculate the product of the first 4 elements of V
- Display the 4th, 9th and 11th element of V.

```
> (V<-seq(16,56,by=2)) # creates AND displays the vector

[1] 16 18 20 22 24 26 28 30 32 34 36 38 40 42 44 46 48 50 52 54 56
> # or:
> V <- 16+2*(0:20) ; V

[1] 16 18 20 22 24 26 28 30 32 34 36 38 40 42 44 46 48 50 52 54 56
> sum(V)

[1] 756
> V[1:4]

[1] 16 18 20 22
> prod(V[1:4])

[1] 126720
> V[c(4,9,11)]
```

- Create a new vector, W, which equals vector V, multiplied with 3; display its content.
- How many elements of W are smaller than 100?

```
> W<-V*3; W
 [1] 48 54 60 66 72 78 84 90 96 102 108 114 120 126 132 138
[17] 144 150 156 162 168
> W100<-W[W<100] ; length(W100)
[1] 9
> # or
> length(W[W<100])</pre>
[1] 9
   • Create a sequence that contains the values (1,1/2,1/3,1/4,\check{E},1/10)
   • Compute the square root of each element
   • Compute the square (2) of each element
   • Create a sequence with values (0/1,1/2,2/3,3/4,\check{E},9/10)
> 1/1:10
 [1] 1.0000000 0.5000000 0.3333333 0.2500000 0.2000000 0.1666667
 [7] 0.1428571 0.1250000 0.1111111 0.1000000
> sqrt(1/1:10)
 [1] 1.0000000 0.7071068 0.5773503 0.5000000 0.4472136 0.4082483
 [7] 0.3779645 0.3535534 0.3333333 0.3162278
> (1/1:10)^2
 [1] 1.00000000 0.25000000 0.11111111 0.06250000 0.04000000 0.02777778
 [7] 0.02040816 0.01562500 0.01234568 0.01000000
> (0:9)/(1:10)
                   # or : 0:9/1:10
 [1] 0.0000000 0.5000000 0.6666667 0.7500000 0.8000000 0.8333333
 [7] 0.8571429 0.8750000 0.8888889 0.9000000
```

- Create a vector, U, with 100 random numbers, uniformly distributed between -1 and 1.
- Check the range of U; all values should be within -1 and +1.
- Calculate the sum and the product of the elements of U

• How many elements of U are positive?

```
• Zero all negative values of U.
 • Sort U
> U <- runif(100,-1,1)
> range(U)
[1] -0.9983258 0.9997094
> sum(U);prod(U)
[1] -1.005794
[1] 1.84631e-34
> length(U[U>0]) # or: sum(U>0)
[1] 46
> U[U<0]<-0
> sort(U)
[56] 0.16849845 0.16989987 0.17960761 0.18460746 0.19443542
[61] 0.26244874 0.33830933 0.36658718 0.36746030 0.37451643
[66] 0.37526608 0.39202649 0.44195185 0.48391644 0.48493981
[71] 0.51121285 0.54523376 0.55396225 0.56688228 0.58353998
[76] 0.59085183 0.59276520 0.62194597 0.62906369 0.63549231
```

[81] 0.67081327 0.69988539 0.70443375 0.70572517 0.70665552 [86] 0.72074820 0.72654062 0.73149877 0.73912293 0.77413842 [91] 0.80603771 0.84760430 0.84793236 0.86264330 0.86707680 [96] 0.91146930 0.92617294 0.98571765 0.99155656 0.99970937

• Create two vectors: vector x, with the elements: 2,9,0,2,7,4,0 and vector y with the elements 3,5,0,2,5,4,6 (in that order).

- Divide all the elements of y by the elements of x.
- Select all values of y that are larger than the corresponding values of x
- Select all values of y for which the corresponding values of x are 0.
- Remove all values of y for which the corresponding values of x equal 0.
- $\bullet$  Zero all elements of x that are larger or equal than 7. Show x.

```
> x<- c(2,9,0,2,7,4,0)

> y<- c(3,5,0,2,5,4,6)

> y/x

[1] 1.5000000 0.5555556 NaN 1.0000000 0.7142857 1.0000000

[7] Inf

> x>y

[1] FALSE TRUE FALSE FALSE TRUE FALSE FALSE

> x==0

[1] FALSE TRUE FALSE FALSE FALSE TRUE

> y[y>x]

[1] 3 6

> y[x==0]

[1] 0 6
```

Chapter 3.8.2

[1] 2 0 0 2 0 4 0

> y<-y[x!=0] > x[x>=7]<-0 ; x

• Use R-function "matrix" to create a matrix with the following contents:

$$\left[\begin{array}{cc} 3 & 9 \\ 7 & 4 \end{array}\right]$$

• display it to the screen

• Use R-function "matrix" to create a matrix called "A":

$$\left[\begin{array}{cc} 3 & 9 \\ 7 & 4 \end{array}\right]$$

- Take the transpose of A.
- Create a new matrix, B, by extracting the first two rows and first two columns of A. Display it to the screen.

> A<-matrix(nrow=2,data=c(3,7,9,4)) ; A

> A<-matrix(nrow=3,data=1/1:9,byrow=TRUE) # or: 1/matrix(nrow=3,data=1:9,byrow=TRUE)
> t(A)

[3,] 0.3333333 0.1666667 0.1111111

$$> B \leftarrow A[1:2,1:2]$$
; B

# Matrix D

• Use diag to create the following matrix, called "D":

$$\left[\begin{array}{ccc} 1 & 0 & 0 \\ 0 & 2 & 0 \\ 0 & 0 & 3 \end{array}\right]$$

• Use cbind and rbind to augment this matrix, such that you obtain:

$$\left[\begin{array}{cccc}
1 & 0 & 0 & 4 \\
0 & 2 & 0 & 4 \\
0 & 0 & 3 & 4 \\
5 & 5 & 5 & 5
\end{array}\right]$$

• Remove the second row and second column of the previous matrix

```
> D <- diag(nrow=3,c(1,2,3))
> DD <- cbind(D,rep(4,3)) # or: cbind(D,4)
> DDD <- rbind(DD,rep(5,4)) # or: rbind(DD,5)
> DDD
```

> # same, in one sentence
> DD <- rbind(cbind(D,4),5)</pre>

# Chapter 3.8.3 - nematode diversity

- Select the data from station M160b (the 2nd column of Nemaspec); put these data in a vector called "dens".
- Remove from vector dens, the densities that are 0. Display this vector on the screen.
- Calculate N, the total nematode density of this station.
- Divide the values in vector dens by the total nematode density N. Put the results in vector p. The sum of all values in p should equal 1.
- Calculate S, the number of species.
- Estimate the values of diversity indices N1 and N2 and Ni, given by the following formulae:

$$N1 = e^{\sum -p_i \cdot \log_e(p_i)}$$

$$N2 = 1/(\sum p_i^2)$$

$$Ni = 1/\max(p_i)$$

• The expected number of species in a sample with size n, drawn from a population which size N, which has S species is given by:

$$ES(n) = \sum_{i=1}^{S} \left[ 1 - \frac{\binom{N-Ni}{n}}{\binom{N}{n}} \right]$$

What is the expected number of species per 100 individuals?

• Print all diversity indices to the screen, which should look like:

### > head(Nemaspec)

```
SPECIES M160a
                                 M160b
                                          M280a
                                                    M280b
                                                             M530a
                            0 6.580261 0.000000 1.120782 1.315487
1
          Acantholaimus
2 Acantholaimus elegans
                            0 0.000000 1.439706 0.000000 3.956836
                            0 0.000000 0.000000 0.000000 0.000000
3 Acantholaimus iubilus
       Acantholaimus M1
                            0 5.919719 0.000000 3.628518 0.000000
                            0 0.000000 0.000000 1.120782 0.000000
5
      Acantholaimus M10
      Acantholaimus M11
                            0 0.000000 0.000000 0.000000 0.000000
     M530b
                                         M990b
              M820a
                       M820b
                                M990a
                                                  M1220a
                                                           M1220b
1 1.727387 3.417313 3.748096 2.447545 3.728838 4.369345 4.787512
2 0.000000 0.000000 2.198407 5.080900 5.330997 3.644567 3.494481
3 1.193131 0.000000 0.000000 1.270450 1.372789 0.000000 0.000000
4 1.193131 1.155307 0.000000 5.052036 6.225598 0.000000 1.166825
5 0.000000 0.000000 0.000000 0.000000 1.115981 0.000000
6 0.000000 0.000000 0.000000 0.000000 2.285694 0.000000 0.000000
> dens <- Nemaspec[,2]</pre>
> dens <- dens[dens>0]
       <- sum(dens);
> N
> p
       <- dens/N
       <- length(p)
> NO
       <- \exp(sum(-p*log(p)))
> N1
       <- sum(p*p)^(-1)
> N2
       <-1/max(p)
> Ni
> ESS <- NO-1/choose(N,100)*sum(choose(n=(N-dens),k=100))
> c(N=N,N0=N0,N1=N1,N2=N2,Ni=Ni,ESS=ESS)
         N
                   NO
                              N1
                                         N2
                                                     Νi
                                                               ESS
576.000000 97.000000 27.782793
                                   8.364525
                                               3.162870 40.502318
```

# Chapter 4 user-defined functions

```
Chapter 4.4.1
```

```
> ## Sphere function
> Sphere <- function(radius)
+ {
+ vol <- 4/3*pi*radius^3
+ surf <- 4 *pi*radius^2
+ circ <- 2*pi*radius
+ return(list(volume=vol,surface=surf,circumference=circ))
+ }
> Sphere(6371)

$volume
[1] 1.083207e+12

$surface
[1] 510064472

$circumference
[1] 40030.17
```

### Chapter 4.4.2

The saturated oxygen concentration in water  $(molkg^{-1})$ , as function of temperature (T), and salinity (S) can be calculated by:  $SatOx = e^A$  where : A= -173.9894 + 25559.07/T + 146.4813\*  $\log(T/100)$  -22.204\*T/100 + S \* (-0.037362+0.016504\*T/100-0.0020564\*T/100\*T/100) and T is temperature in Kelvin (Tkelvin = Tcelsius+273.15).

- Make a function that implements this formula; the default values for temperature and salinity are  $20^{\circ}C$  and 35 respectively.
- What is the saturated oxygen concentration at the default conditions?
- Estimate the saturated oxygen concentration for a range of temperatures from 0 to  $30^{\circ}C$ , and salinity 35.

```
> Sat0x <- function(T=20,S=35)
+ {
+    T <- T+273.15
+    A= -173.9894 + 25559.07/T + 146.4813* log(T/100) -22.204*T/100 + S *
+    (-0.037362+0.016504*T/100-0.0020564 *T/100*T/100)
+    exp(A)
+  }
> Sat0x()
[1] 225.2346
```

# > SatOx(0:30)

```
[1] 349.6542 340.6019 331.9557 323.6924 315.7901 308.2286 300.9890
```

- [8] 294.0533 287.4051 281.0288 274.9098 269.0344 263.3897 257.9638
- [15] 252.7452 247.7235 242.8884 238.2306 233.7412 229.4118 225.2346
- [22] 221.2020 217.3070 213.5431 209.9038 206.3833 202.9759 199.6764
- [29] 196.4796 193.3808 190.3755

# Chapter 4.4.3

The Fibonacci numbers are calculated by the following relation:  $F_n = F_{n-1} + F_{n-2}$  With  $F_1 = F_2 = 1$ 

- Compute the first 50 Fibonacci numbers; store the results in a vector.
- For large n, the ratio Fn/Fn-1 approaches the "golden mean"
- What is the value of F50/F49; is it equal to the golden mean?
- When is n large enough? (i.e. sufficiently close (<1e-6) to the golden mean)

```
> Fibo<-vector()
```

- > Fibo[1:2]<-1
- > for (i in 3:50) Fibo[i]<-Fibo[i-1]+Fibo[i-2]</pre>
- > (1+sqrt(5))/2

### [1] 1.618034

> Fibo[50]/Fibo[49]

### [1] 1.618034

> Fibo[2:50]/Fibo[1:49]- (1+sqrt(5))/2

```
[1] -6.180340e-01 3.819660e-01 -1.180340e-01 4.863268e-02
```

[5] -1.803399e-02 6.966011e-03 -2.649373e-03 1.013630e-03

[9] -3.869299e-04 1.478294e-04 -5.646066e-05 2.156681e-05

[13] -8.237677e-06 3.146529e-06 -1.201865e-06 4.590718e-07

[17] -1.753498e-07 6.697766e-08 -2.558319e-08 9.771908e-09

[21] -3.732537e-09 1.425702e-09 -5.445699e-10 2.080072e-10

[25] -7.945178e-11 3.034772e-11 -1.159184e-11 4.427569e-12

[29] -1.691314e-12 6.459278e-13 -2.466916e-13 9.414691e-14

[33] -3.597123e-14 1.376677e-14 -5.329071e-15 1.998401e-15 [37] -8.881784e-16 2.220446e-16 -2.220446e-16 0.000000e+00

[41] 0.000000e+00 0.000000e+00 0.000000e+00 0.000000e+00

[45] 0.000000e+00 0.000000e+00 0.000000e+00 0.000000e+00

[49] 0.000000e+00

## Chapter 4.4.5 - nematode diversity all stations

- Make a function that will calculate the diversity indices for any data matrix.
- Calculate and show diversity on species level

Mean : 1.143 Mean : 1.047 Mean : 0.849

```
# density, each column a station
> Diversity <- function (Dens,</pre>
                        S=100) # common number of individuals on which
+
                                # to estimate expected number of species
+ {
   nstat <- NCOL(Dens)</pre>
                                                  # number of stations
   if(is.vector(Dens)) Dens <- matrix(ncol=nstat,Dens)</pre>
        <- matrix(nrow=nstat,ncol=6,data=NA)</pre>
                                                  # create matrix for results
   rownames(div) <- colnames(Dens)</pre>
   colnames(div) <- c("N", "NO", "N1", "N2", "Ninf", paste("ESS", S, sep=""))
   for (i in 1:nstat)
      dens<- Dens[,i]</pre>
      dens<- dens[dens>0] # selection of species present
          <- sum(dens)
                           # N, total density
          <- dens/N
                           # relative proportion
      p
      NO <- length(p)
                           # NO = number of species present
      N1 <- \exp(sum(-p*log(p))) # N1 = \exp(Shannon-Wiener)
      N2 <- sum(p*p)^(-1)
                                   # Na = sum(sp^a)^(1/(1-a))
      Ni <-1/max(p)
                                    # Ninf
      ESS <- NO-1/choose(N,S)*sum(choose(n=(N-dens),k=S))
      div[i,] \leftarrow c(N,N0,N1,N2,Ni,ESS)
    }
   return(div)
+ }
> summary(Nemaspec)
                                    # calculate summary characteristics
                 SPECIES
                                M160a
                                                  M160b
                                              Min. : 0.000
                     : 1
                            Min. : 0.000
Acantholaimus
Acantholaimus elegans: 1 1st Qu.: 0.000
                                              1st Qu.: 0.000
Acantholaimus iubilus: 1
                           Median : 0.000
                                              Median : 0.000
 Acantholaimus M1
                   : 1
                          Mean
                                 : 1.226
                                              Mean : 1.487
                                              3rd Qu.: 1.341
Acantholaimus M10
                     : 1 3rd Qu.: 0.000
Acantholaimus M11
                     : 1
                            Max. :182.113
                                              Max. :30.982
 (Other)
                     :464
    M280a
                     M280b
                                      M530a
                                                       M530b
Min. : 0.000 Min. : 0.000 Min. : 0.000
                                                   Min. : 0.0000
1st Qu.: 0.000 1st Qu.: 0.000 1st Qu.: 0.000
                                                   1st Qu.: 0.0000
Median: 0.000 Median: 0.000 Median: 0.000
                                                   Median : 0.0000
```

Mean : 0.8553

```
3rd Qu.: 1.183
                3rd Qu.: 1.121
                                3rd Qu.: 0.000
                                                3rd Qu.: 0.0000
Max. :54.031
                Max. :33.500
                                Max. :45.938
                                                Max. :29.2128
   M820a
                    M820b
                                     M990a
                                Min. : 0.0000
Min. : 0.0000
                Min. : 0.0000
1st Qu.: 0.0000
                 1st Qu.: 0.0000
                                1st Qu.: 0.0000
Median : 0.0000
                 Median : 0.0000
                                Median : 0.0000
Mean : 0.9234
                      : 0.9383
                                Mean : 0.9596
                 Mean
                 3rd Qu.: 0.0000
3rd Qu.: 0.0000
                                  3rd Qu.: 1.1771
Max.
      :78.3521
                 Max.
                       :38.0481
                                 Max.
                                        :47.8478
   M990b
                   M1220a
                                    M1220b
Min.
     : 0.000 Min.
                     : 0.0000
                                 Min.
                                      : 0.0000
1st Qu.: 0.000
               1st Qu.: 0.0000
                                 1st Qu.: 0.0000
Median : 0.000 Median : 0.0000
                                 Median: 0.0000
      : 1.217
               Mean
                      : 0.7213
                                 Mean
                                       : 0.7085
3rd Qu.: 1.213
                3rd Qu.: 0.0000
                                 3rd Qu.: 0.0000
Max.
      :51.271
               Max.
                      :37.5895
                                 Max. :25.6069
```

- > # remove species names
- > (divspec<-Diversity(Nemaspec[,-1]))</pre>

```
N
          NO
                    N1
                              N2
                                      Ninf
                                             ESS100
M160a 576 97 27.78279 8.364525 3.162870 40.50232
M160b 699 126 90.15358 66.778414 22.561569 60.68971
M280a 537 148 83.57356 43.779249 9.938717 59.44306
M280b 492 140 87.18598 51.988253 14.686371 61.17590
M530a 399 107 61.03511 29.166239 8.685560 54.97142
M530b 402 105 66.41245 44.308181 13.761081 54.25870
M820a 434 102 47.47238 20.190796 5.539098 48.29562
M820b 441 115 60.48626 33.178177 11.590593 52.14167
M990a 451 121 72.89122 40.725866 9.425726 57.11996
M990b 572 148 88.37314 47.535985 11.156321 61.16281
M1220a 339 106 65.53017 37.188517 9.018488 55.97925
M1220b 333 96 63.58659 41.009214 13.004317 54.90511
```

Chapter 4.4.6 - rarefaction diversity An alternative way of estimating the number of species per 100 individuals is by taking random 'subsamples' of 100 individuals and estimating the number of species from this subsample.

```
> dens <- Nemaspec[,2]
> dens <- dens[dens>0] # selection of species present
> cs <- round(dens) # rarefaction method can only work with integer numbers
> ind <- NULL # individual organisms; each one belonging to a species
> for (i in 1:length(cs)) ind <- c(ind,rep(i,times=cs[i]))
> ind100 <-sample(ind,size=100) # take 100 random individuals
> Spec <-table(ind100) # table of counts: speciesnr versus nr ind</pre>
```

```
> ESS100 <-length(Spec)  # length of Spec = number of species
> # or, three sentences combined in 1!
> length(table(sample(ind,size=100)))

[1] 38

> ESS100 <- vector()
> for (i in 1:1000) ESS100[i] <- length(table(sample(ind,size=100)))
> mean (ESS100)

[1] 40.548
```

# Chapter 5 - statistics

- Perform a hierarchic clustering of the Nemaspec dataset and plot the dendrogram
- Perform a principal component analysis (PCA) and plot the results
- repeat the PCA analysis, with the first two stations removed

```
> nemaspec <- Nemaspec[,-1]
> hc <- hclust(dist(t(nemaspec)), "ave")
> par(mfrow=c(2,2))
> plot(hc)
> plot(hc, hang = -1)
> x <- prcomp(t(nemaspec))
> biplot(x)
> x2 <- prcomp(t(nemaspec[,-(1:2)]))
> biplot(x2)
> par(mfrow=c(1,1))
```

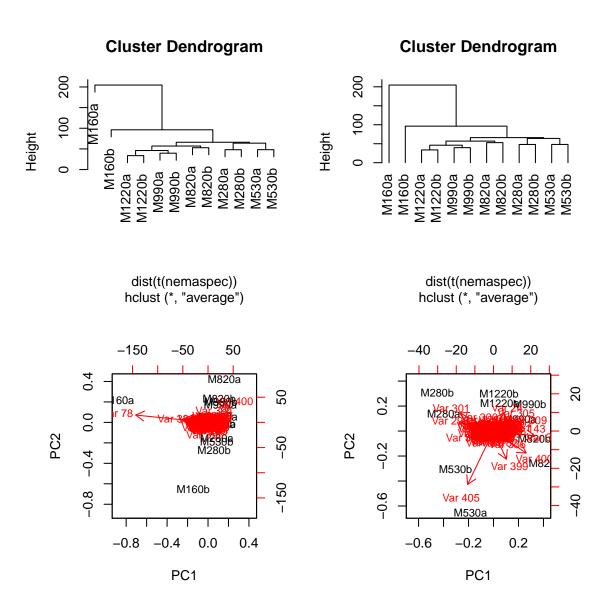


Figure 1: Cluster analysis and PCA of nematode data

# Chapter 6 - graphics

- Create a script file which draws a curve of the function  $y = x^3 sin^2(3\pi x)$  in the interval [-2, 2].
- Make a curve of the function  $y = 1/\cos(1+x^2)$  in the interval [-5,5].
- The relative importance of ammonia

$$p_{[NH_3]} = \frac{K_N}{K_N + [H^+]}$$

- Plot the relative fraction of toxic ammonia to the total ammonia concentration as a function of pH, where pH =  $-\log 10([H+])$  and for a temperature of  $30^{\circ}C$ . Use a range of pH from 4 to 9. The value of KN is  $810^{-10}$  at a temperature of  $30^{\circ}C$ .
- Add to this plot the relative fraction of ammonia at  $0^{\circ}C$ ; the value of KN at that temperature is  $810^{-11} mol kg^{-1}$ .
- For the US, the population density in 1900 (N0) was 76.1 million; the population growth can be described as:

$$N(t) = \frac{K}{1 + \left[\frac{K - N_{t0}}{N_{t0}}\right]e^{-a \cdot (t - t0)}}$$

a=0.02 yr-1, K = 500 million of people.

Actual population values are:

1900 1910 1920 1930 1940 1950 1960 1970 1980

 $76.1\ 92.4\ 106.5\ 123.1\ 132.6\ 152.3\ 180.7\ 204.9\ 226.5$ 

- Plot the population density curve as a thick line, using the US parameter values.
- Add the measured population values as points. Finish the graph with titles, labels etcĚ

```
> par(mfrow=c(2,2))
> # simple curves
> curve(x^3*sin(3*pi*x)^2,-2,2)
> curve(1/cos(1+x^2), -5, 5)
> # ammonia
> pN <- function(pH, Kn=8*10^-10) Kn/(Kn+10^-pH)
> curve(pN(x),4,9,main="fraction toxic ammonium")
> curve(pN(x,Kn=8*10^-11),4,9,add=TRUE,col="red")
> legend("topleft",lty=1,col=c("black","red"),c("30 dg","0 dg"))
> # US population
> K <- 500
> NO <- 76.1
> a <- 0.02
> curve(K/(1+((K-N0)/N0*exp(-a*(x-1900)))),1900,1980,main="US population",
        xlab="year",ylab= "million",lwd=2)
> N <- matrix(ncol=2,data=c(
```

```
+ seq(1900,1980,by=10), 76.1,92.4,106.5,123.1,132.6,152.3,180.7,204.9,226.5
                           ) )
> points(N)
```

- Have a look at the iris data; What is the class and dimension of the data set?
- Produce a scatter plot of petal length against petal width
- Repeat the same graph, using different symbol colours for the three species.
- Create a box-and whisker plot for sepal length where the data values are split into species groups
- Now produce a similar box-and whisker plot for all four morphological measurements, arranged in two rows and two columns.

### > head(iris)

```
Sepal.Length Sepal.Width Petal.Length Petal.Width Species
1
           5.1
                       3.5
                                    1.4
                                                 0.2 setosa
2
           4.9
                       3.0
                                    1.4
                                                 0.2 setosa
3
           4.7
                       3.2
                                    1.3
                                                 0.2 setosa
4
           4.6
                                    1.5
                       3.1
                                                 0.2 setosa
5
           5.0
                       3.6
                                    1.4
                                                 0.2 setosa
6
           5.4
                       3.9
                                    1.7
                                                 0.4 setosa
> class(iris)
[1] "data.frame"
> dim(iris)
[1] 150
> par(mfrow=c(2,2))
> plot(iris$Petal.Length,iris$Petal.Width,cex=1.5,pch=15,
        xlab="Petal length", ylab=" Petal width")
> plot(iris$Petal.Length,iris$Petal.Width,cex=1.5,pch=15,
       xlab="Petal length", ylab=" Petal width",
       col=c("red","blue","green")[iris$Species])
> legend("bottomright",pch=15,col=c("red","blue","green"),
       legend=levels(iris$Species))
> boxplot(Petal.Width~Species,data=iris)
> par(mfrow=c(2,2))
> boxplot(Sepal.Length~Species, data=iris,main="sepal length")
> boxplot(Sepal.Width~Species, data=iris,main="sepal width")
> boxplot(Petal.Length~Species, data=iris,main="petal length")
> boxplot(Petal.Width~Species, data=iris,main="petal width")
> mtext(outer=TRUE, side=3, line=-2, "Iris data set", cex=1.5)
```

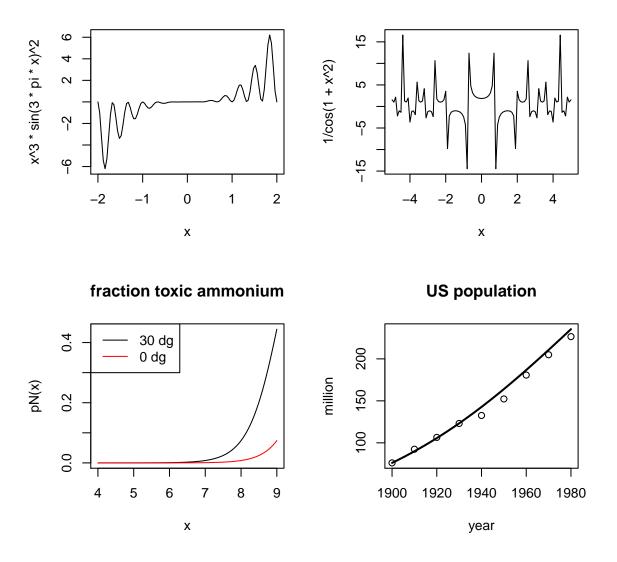
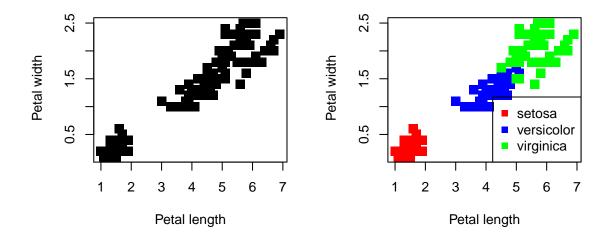


Figure 2: Use of R-function curve to plot simple functions



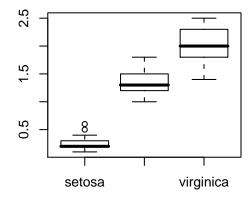


Figure 3: The iris data set

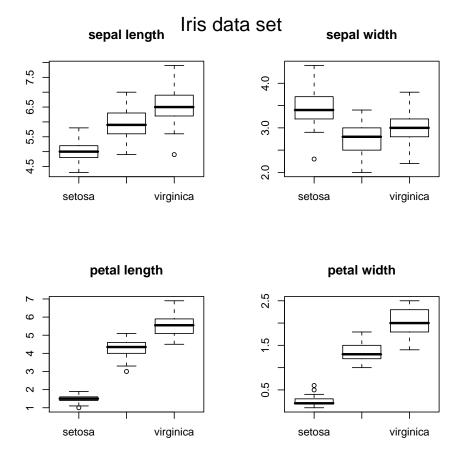


Figure 4: The iris data set

# Chapter 7 - matrix algebra

Chapter 7.1.1

• Create matrices called "A" and "B":

$$A = \left[ \begin{array}{rrr} 1 & 2 & 3 \\ 6 & 4 & 1 \\ -2 & 1 & -1 \end{array} \right]$$

$$B = \left[ \begin{array}{rrr} 1 & 4 & 7 \\ 2 & 5 & 8 \\ 3 & 6 & 9 \end{array} \right]$$

- Take the inverse of A and the transpose of A.
- Multiply A with B.
- Estimate the eigenvalues and eigenvectors of A.
- For a matrix A, x is an eigenvector, and ? the eigenvalue of a matrix A, if Ax = ?x. Test it!

```
> A <- matrix(nrow=3, data=c(1,6,-2,2,4,1,3,1,-1))
> B <- matrix(nrow=3, data=1:9)
> solve(A); t(A)
```

> A%\*%B

> eigen(A)

### \$values

[1] 6.366696+0.000000i -1.183348+2.380697i -1.183348-2.380697i

### \$vectors

```
[,1] [,2] [,3]
[1,] -0.36275602+0i -0.0725936-0.5240033i -0.0725936+0.5240033i
[2,] -0.93146469+0i -0.2632991+0.4856291i -0.2632991-0.4856291i
[3,] -0.02795726+0i  0.6441961+0.0000000i  0.6441961+0.0000000i

> ee<-eigen(A)
> A%*%ee$vectors[,1]

[,1]
[1,] -2.3095572+0i
[2,] -5.9303521+0i
[3,] -0.1779954+0i

> ee$values[1]*ee$vectors[,1]
```

# [1] -2.3095572+0i -5.9303521+0i -0.1779954+0i

Chapter 7.1.2 killer whale model

• Create a matrix, called P:

$$\begin{bmatrix} 0 & 0.0043 & 0.1132 & 0 \\ 0.9775 & 0.9111 & 0 & 0 \\ 0 & 0.0736 & 0.9534 & 0 \\ 0 & 0 & 0.0452 & 0.9804 \end{bmatrix}$$

- What is the value of the largest eigenvalue (the so-called dominant eigenvalue) and the corresponding eigenvector?.
- Create a new matrix, T, which equals P, except for the first row, where the elements are 0.
- Now estimate  $N = (I T)^{-1}$ , where I is the identity matrix.

### \$values

[1] 1.025441326 0.980400000 0.834222976 0.004835698

### \$vectors

```
[,1] [,2] [,3] [,4]
[1,] 0.06634512 0 -0.0659050 0.678780909
[2,] 0.56718211 0 0.8379894 -0.732135578
[3,] 0.57945357 0 -0.5175160 0.056807091
[4,] 0.58149491 1 0.1600233 -0.002631995
```

[4,] 40.04878 40.97062 49.48761 51.02041

# Chapter 7.1.3. System of equations

Solve the following system of linear equations for the unknown xi:

$$3x1 + 4x2 + 5x3 = 0$$

$$6x1 + 2x2 + 7x3 = 5$$

$$7x1 + x2 = 6$$

Check the results

$$> B \leftarrow c(0,5,6)$$

$$> x \leftarrow solve(A,B)$$

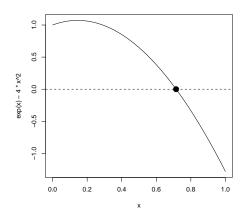


Figure 5: The root of a simple function

# Chapter 8 - roots of functions

Chapter 8.3.1 simple root of equations

- Find the root of the equation  $e^x = 4x^2$  in the interval [0,1]. First draw the function curve.
- Solve the equation  $1000 = y * (3 + x) * (1 + y)^4$  for y and with x varying over the range from 1 to 100. Plot the root as a function of x.

```
> root<-uniroot(f=function(x) exp(x)-4*x^2,interval=c(0,1))

> curve(exp(x)-4*x^2,0,1)
> abline(h=0,lty=2)
> points(root$root,0,pch=16,cex=2)

> res<-vector()
> for (x in 1:100)
+ res[x]<-uniroot (f=function(y) y*(3+x)*(1+y)^4-1000,c(-1000,1000))$root</pre>
```

Chapter 8.3.2. pCO2 rises increase acidity

> plot(1:100,res)

- Estimate the pH at equilibrium with alkalinity 2300  $molkg^{-1}$  and the current pCO2 of 360 ppm.
- Use package seacarb to estimate the dissociation constants and Henry's constants at temperature  $20^{\circ}C$ , salinity 0, and pressure 0.
- Estimate pH as a function of pco2, varying between 200 and 1250

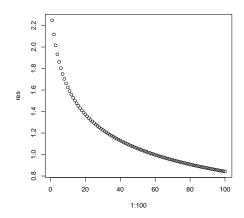


Figure 6: Roots of an equation y=f(x) for a sequence of x-values

• What is the value of pH at pCO2 = 1250

(see lecture notes for formulae)

```
> require(seacarb)
> k1 <- K1(S=0,T=20,P=0)
> k2 <- K2(S=0,T=20,P=0)
> kh <- Kh(S=0,T=20,P=0)
> nonlinfun <- function(pH,pco2=360,alk=2300e-6)</pre>
+ {
  Η
        <- 10^(-pH)
  CO2 <- pco2*kh
  HCO3 <- k1*CO2/H
  CO3 <- k2*HCO3/H
  return( HCO3+2*CO3-H*1.e6 - alk)
> uniroot(nonlinfun,interval=c(2,12),pco2=360,alk=2300,tol=1e-30)
$root
[1] 8.317286
$f.root
[1] 2.728484e-12
attr(,"unit")
[1] "mol/kg-soln"
attr(,"pH scale")
[1] "total hydrogen ion concentration"
$iter
[1] 16
```

# Effect of pCO2 on pH

Figure 7: pH as a function of pCO2

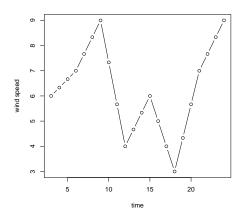


Figure 8: The interpolated wind data

# Chapter 9 - interpolating, smoothing, curve fitting

Chapter 9.1 interpolating wind data Wind velocities are: 5,6,7,9,4,6,3,7,9 at time 0, 3, E24 o'clock respectively.

- Interpolate the three-hourly measurements to hourly measurements.
- Make a plot of the interpolated values

```
> t3 <- seq(3,24,by=3)
> wind3 <- c(6,7,9,4,6,3,7,9)
> plot(approx(t3,wind3,xout=3:24),type="b" ,xlab="time",ylab="wind speed")
```

Chapter 9.2 Fitting primary production data

Primary production (pp) at different light intensities are given; Fit the resulting production estimates (pp), as a function of light intensity (ll) with the 3-parameter Eilers-Peeters equation. The primary production is calculated as:

$$pp = p \max \frac{2 \cdot (1 + \beta) \cdot I_{Iopt}}{(I_{Iopt})^2 + 2 \cdot \beta \cdot I_{Iopt} + 1}$$

where I is light and pmax,? and Iopt are parameters.

Add the best-fit line to the graph. (note: use coef to retrieve the best parameter values).

```
Formula: pp \sim pmax * 2 * (1 + b) * (l1/iopt)/((l1/iopt)^2 + 2 * b * l1/iopt + b)
   1)
Parameters:
    Estimate Std. Error t value Pr(>|t|)
pmax 10.4351 0.3171 32.909 7.93e-10 ***
               0.4353 3.676 0.00626 **
      1.5998
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 0.5445 on 8 degrees of freedom
Number of iterations to convergence: 8
Achieved convergence tolerance: 1.356e-06
> pars <- as.list(coef(fit))</pre>
> with(pars,
+ curve(pmax*2*(1+b)*(x/iopt)/((x/iopt)^2+2*b*x/iopt+1),
        add=TRUE, 1wd=2)
> title(expression (frac(pmax%*%2%*%(1+beta)%*%I/Iopt,
                  (I/Iopt)^2+2%*%beta%*%I/Iopt+1)),cex.main=0.8)
```

```
Formula: pp ~ pmax * 2 * (1 + b) * (ll/iopt)/((ll/iopt)^2 + 2 * b * ll/iopt + 1)
```

### Parameters:

```
Estimate Std. Error t value Pr(>|t|)

pmax 10.4351    0.3171 32.909 7.93e-10 ***

b    1.5998    0.4353    3.676    0.00626 **

iopt 209.6325    15.8052    13.264 9.96e-07 ***

---

Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
```

Residual standard error: 0.5445 on 8 degrees of freedom

Number of iterations to convergence: 8 Achieved convergence tolerance: 1.356e-06

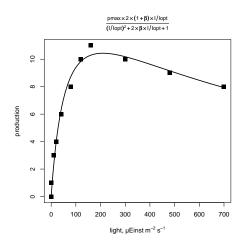


Figure 9: Primary production data with fit

# Chapter 10 differential equations

Chapter 10.1 Lotka-Volterra model

• Solves the following system of ODEs

$$\begin{array}{l} \frac{dx}{dt} = a \cdot x \cdot (1 - \frac{x}{K}) - b \cdot x \cdot y \\ \frac{dy}{dt} = g \cdot b \cdot x \cdot y - e \cdot y \end{array}$$

for initial values x=300,y=10 and parameter values: a=0.05, K=500, b=0.0002, g=0.8, e=0.03

- Make three plots, one for x and one for y as a function of time, and one plot expressing y as a function of x. Arrange these plots in 2 rows and 2 columns.
- run the model with other initial values (x=200, y=50); add the (x,y) trajectories to the phase-plane plot

```
> require(deSolve)
> model <- function (time, VAR, pars)
+ {
  with (as.list(c(VAR,pars)), {
   # the rate of change of the state variables
           <- a*x*(1-x/K)-b*x*y
   dx
           <- g*b*x*y - e*y
   dy
  return(list(c(dx,dy)))
+ }
> pars <- c(a=0.05,b=0.0002,K=500,g=0.8,e=0.03)
        <-c(x=300, y=10)
> VAR
> times <- seq(0,1000,1)
        <- as.data.frame(lsoda(VAR,times,model,pars))</pre>
> plot(out$x,out$y,type="1")
        <-c(x=200,y=50)
> out2 <- as.data.frame(lsoda(VAR,times,model,pars))</pre>
> lines(out2$x,out2$y,1ty=2)
```

Chapter 10.2 Lorenz Butterfly

Solve the Lorenz equations:

$$\begin{array}{l} \frac{dx}{dt} = -\frac{8}{3} \cdot x + y \cdot z \\ \frac{dy}{dt} = -10 \cdot (y - z) \\ \frac{dz}{dt} = -x \cdot y + 28y - z \end{array}$$

Use as initial conditions x=y=z=1; create output for a time sequence ranging from 0 to 100, and with a time step of 0.005.

```
> require(scatterplot3d)
> model<-function(t,state,parameters)</pre>
```

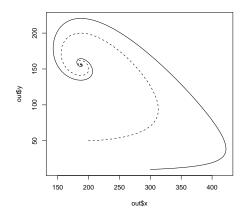


Figure 10: Result of the lotka-volterra model

```
{
   with(as.list(c(state)),{
      dx
             <- -8/3*x+y*z
             <-10*(y-z)
      dy
             <- -x*y+28*y-z
      dz
      list(c(dx,dy,dz))
                                    })
     # end of model
> state <-c(x=1, y=1, z=1)
> times <-seq(0,100,0.005)
        <-as.data.frame(lsoda(state,times,model,0))</pre>
> scatterplot3d(out$x,out$y,out$z,type="1",
          main="Lorenz butterfly",ylab="",grid=FALSE,box=FALSE)
```

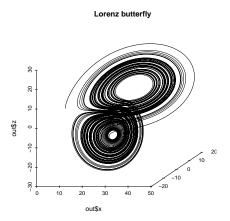


Figure 11: Results of the Lorenz model

# References

Soetaert K (2008).  $using\ R$  for  $scientific\ computing$ . NIOO-CEME, Yerseke.

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