# Mixture Analysis of the Signal Tandmobiel® Data Using the Package mixAK

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This document supplements a paper Komárek (2009) and shows an analysis of a part of Signal Tandmobiel<sup>®</sup> data introduced in Vanobbergen et al. (2000). We will estimate a joint distribution of the emergence times of six permanent teeth from one quandrant of the mouth. The dataset contains observed intervals of teeth emergence for  $4\,430$  Flemish children resulting from a longitudinal project performed in 1996-2002. The observed intervals result from annual dental examinations and are hence of the length approximately one year.

## 1 Introduction

- Due to the fact that some code (especially MCMC) is time consuming, some code chunks found in this vignette are not run when compiling the package. You should set the variable RUN.TIMECONSUMING.CODE to TRUE to run full MCMC and related code.
- Having run full MCMC and related code, setting the variable RUN.ALLOUT to TRUE will cause that all output shown in this vignette is re-created and not taken from previously computed results.
- Results based on full MCMC runs are stored (chains excluded) in Tandmob-Result.RData in ./RESULT\_OBJ directory and are used to create majority of this vignette at the time of compilation of the package. Additionally, file Tandmob-PDensBiModelK02.RData in the same directory contains estimated predictive density.

R⇒ Setting variables RUN.ALLOUT and RUN.TIMECONSUMING.CODE.

```
> RUN.TIMECONSUMING.CODE <- FALSE
> RUN.ALLOUT <- FALSE
```

R⇒ Directory to store postscript files with figures. Figures which require chains are stored in FIGKEEPDIR directory all other figures are stored in FIGDIR directory.

```
> FIGDIR <- "./figures/"
> FIGKEEPDIR <- "./figuresKeep/"</pre>
```

 $R \Rightarrow$  Directories with results computed in past. Objects with chains will be stored in directory specified by variable RESULTDIR. All other objects will be stored in directory RESULT2DIR.

```
> RESULTDIR <- "/home/komarek/RESULT_OBJ/mixAK-Tandmob-S090426/"
> RESULT2DIR <- "./RESULT_OBJ/" ### available in package as /inst/doc/RESULT_OBJ
```

 $R \Rightarrow$  Display options.

```
> options(width = 80)
```

 $R \Rightarrow$  Load results computed in past. Variable Kshow determines which value for fixed number of components K is used in section 4.

```
> Kshow <- 2
> if (RUN.ALLOUT){
    load(paste(RESULT2DIR, "Tandmob-Result.RData", sep=""))
       ## contains ModelK (without chains), PDensUniModelK
   load(paste(RESULT2DIR, "Tandmob-PDensBiModelKO", Kshow,".RData", sep=""))
       ## contains PDensBiModelK[[Kshow]]
    load(paste(RESULTDIR, "Tandmob-Model0", Kshow, ".RData", sep=""))
       ## contains Model0=ModelK[[Kshow]] (chains included)
    PDensUniModel0 <- PDensUniModelK[[Kshow]]</pre>
    PDensBiModel0 <- PDensBiModelK[[Kshow]]</pre>
+ }else{
    load(paste(RESULT2DIR, "Tandmob-Result.RData", sep=""))
       ## contains ModelK (without chains), PDensUniModelK
   load(paste(RESULT2DIR, "Tandmob-PDensBiModelKO", Kshow,".RData", sep=""))
       ## contains PDensBiModelK[[Kshow]]
   Model0 <- ModelK[[Kshow]]</pre>
   PDensUniModel0 <- PDensUniModelK[[Kshow]]</pre>
   PDensBiModel0 <- PDensBiModelK[[Kshow]]</pre>
+ }
```

R⇒ Load the package mixAK and packages coda and colorspace. Package coda is used to perform some basic convergence diagnostics, package colorspace is used to draw nicer image plots with estimated bivariate densities.

```
> library("mixAK")
> library("coda")
> library("colorspace")
```

# 2 Exploration of the data

 $R \Rightarrow$  The data are read as follows.

```
> data("Tandmob", package = "mixAK")
> data("TandmobEmer", package = "mixAK")
```

R⇒ The following table shows the number and proportion of each type of censoring for each tooth.

```
> NUM.PROP.CENS <- function(data, tanden)
+ {
    TABLE <- matrix(NA, nrow=length(tanden), ncol=3)</pre>
    rownames(TABLE) <- paste(tanden)</pre>
    colnames(TABLE) <- c("Left", "Interval", "Right")</pre>
    for (tt in 1:length(tanden)){
      ebeg <- get(data)[,paste("EBEG.", tanden[tt], sep="")]</pre>
      eend <- get(data)[,paste("EEND.", tanden[tt], sep="")]</pre>
      TABLE[tt, "Left"] <- sum(is.na(ebeg) & !is.na(eend))</pre>
      TABLE[tt, "Interval"] <- sum(!is.na(ebeg) & !is.na(eend))</pre>
      TABLE[tt, "Right"]
                           <- sum(!is.na(ebeg) & is.na(eend))</pre>
      rm(list=c("ebeg", "eend"))
    PROP.TABLE <- t(round(apply(TABLE, 1, prop.table)*100, 3))</pre>
    RET <- list(Table=TABLE, Prop.Table=PROP.TABLE)</pre>
    return(RET)
+ }
> tanden < rep(c(10, 20, 30, 40), 7) + rep(1:7, each=4)
> TAB01 <- NUM.PROP.CENS(data="Tandmob", tanden=tanden)
```

\$Table			\$Prop.Table	
Lef	t Interval	Right	Left Interval	Rig
11 246	1 1745	224	11 55.553 39.391	5.0
21 246	1 1728	241	21 55.553 39.007	5.4
31 392	6 436	68	31 88.623 9.842	1.5
41 393	4 434	62	41 88.804 9.797	1.4
12 65	1 3223	556	12 14.695 72.754	12.5
22 65	2 3240	538	22 14.718 73.138	12.1
32 192	0 2183	327	32 43.341 49.278	7.3
42 195	2 2163	315	42 44.063 48.826	7.1
13 1	2 2095	2323	13 0.271 47.291	52.4
23	4 2101	2325	23 0.090 47.427	52.4
33 2	1 3070	1339	33 0.474 69.300	30.2
43 1	8 3077	1335	43 0.406 69.458	30.1
14 3	0 2733	1667	14 0.677 61.693	37.6
24 4	2 2776	1612	24 0.948 62.664	36.3
34 2	5 2851	1554	34 0.564 64.357	35.0
44 2	5 2861	1544	44 0.564 64.582	34.8
15 1	6 1861	2553	15 0.361 42.009	57.6
25 3	0 1873	2527	25 0.677 42.280	57.0
35 2	5 1795	2610	35 0.564 40.519	58.9
45 1	6 1780	2634	45 0.361 40.181	59.4
16 380	9 554	67	16 85.982 12.506	1.5
26 380	9 555	66	26 85.982 12.528	1.4
36 382	0 535	75	36 86.230 12.077	1.6
46 373	9 604	87	46 84.402 13.634	1.9
17	0 1053	3377	17 0.000 23.770	76.2
27	1 1107	3322	27 0.023 24.989	74.9
37	1 1640	2789	37 0.023 37.020	62.9
47	1 1509	2920	47 0.023 34.063	65.9

# 3 Transformation of the data and preparation of the MCMC

R⇒ We will take teeth 1–6 of one quadrant (right maxillary teeth, i.e. teeth 11, 12, 13, 14, 15, 16). To pass the data into NMixMCMC function, 6-column matrices (called y0, y1 and censor) have to be created. Matrix y0 will contain lower limit of observed intervals in the case of interval censoring or lower limits of right-censored observations. Matrix y1 will contain upper limits of observed intervals in the case of interval censoring and NA's in the case of right censoring. Matrix censor will contain 3 for interval-censored observations and 0 for right-censored observations. All originally left-censored observations have been changed into interval-censored ones with 5 years of age as the lower limit of the observed intervals in the data.frame TandmobEmer. The reason for this is the fact that 5 years of age is clinically the minimal age when the permanent teeth emerge (Ekstrand et al., 2003). However note that this is not necessary and it is possible to consider also left-censored observation and pass them into the function NMixMCMC (use then the data.frame Tandmob).

R⇒ Preparation of the data using the original data.frame Tandmob:

```
> Emerg.min <- 5
> Tooth <- 10 + 1:6
> y0 <- Tandmob[, paste("EBEG.", Tooth, sep="")]
> y1 <- Tandmob[, paste("EEND.", Tooth, sep="")]</pre>
> y0[is.na(y0)] <- Emerg.min
     ### Left-censored changed into interval-censored with the lower limit=5
     ###
> censor <- matrix(3, nrow=nrow(y0), ncol=length(Tooth))</pre>
> censor[is.na(y1)] <- 0</pre>
> colnames(y0) <- colnames(y1) <- colnames(censor) <- Tooth
> rownames(y0) <- rownames(y1) <- rownames(censor) <- Tandmob$IDNR
R⇒ Preparation of the data using the data.frame TandmobEmer:
> Tooth <- 10 + 1:6
> y0 <- TandmobEmer[, paste("EBEG.", Tooth, sep = "")]
> y1 <- TandmobEmer[, paste("EEND.", Tooth, sep = "")]</pre>
> censor <- TandmobEmer[, paste("CENSOR.", Tooth, sep = "")]</pre>
> colnames(y0) <- colnames(y1) <- colnames(censor) <- Tooth
> rownames(y0) <- rownames(y1) <- rownames(censor) <- TandmobEmer$IDNR</pre>
R⇒ Print part of prepared data:
> print(y0[1:5, ])
                                            > print(y1[1:5, ])
   11 12
            13
                  14
                       15 16
                                                         13
                                                              14
                                                                   15
1 5.0 5.0
           8.5
                7.7
                      9.5 5.0
                                            1 7.7 7.7
                                                       9.5 8.5 10.2 7.7
2 5.0 8.4 9.3
                9.3 10.2 5.0
                                            2 7.4 9.3 10.2 10.2 10.9 7.4
3 5.0 6.5 10.5 9.5 10.5 5.0
                                                                   NA 6.5
                                            3 6.5 7.7
                                                        NA 10.5
5 7.5 8.2 10.9 10.9 10.9 6.5
                                            5 8.2 9.1
                                                        NA
                                                              NA
                                                                   NA 7.5
6 5.0 7.4 10.9 10.0 10.0 5.0
                                            6 7.4 8.4 11.8 10.9 10.9 7.4
```

 $R \Rightarrow$  Print censor indicators:

```
> print(censor[1:5, ])
```

```
11 12 13 14 15 16
   3
      3
          3
             3
1
                3
                    3
      3
         3
             3
                3
                    3
      3
          0
             3
      3
   3
          0
                    3
      3
         3
   3
             3
                3
```

 $R \Rightarrow$  Length of the MCMC simulation for all models in this document (burn-in of 10 000 iterations, additional 20 000 iterations are kept for the inference, thinning of 1:10):

```
> nMCMC <- c(burn = 10000, keep = 20000, thin = 10, info = 1000)
```

R⇒ Grid of values where we evaluate and subsequently plot the predictive densities for all models in this document (we use different range of values for different teeth to cover the range when a specific tooth emerges with probability approaching one):

```
> lygridBi <- 50
> lygridUni <- 100
> ymin < -c(4, 5, 6, 6, 6, 4)
> ymax <- c(10, 12, 17, 16, 17, 9)
> ygridBi <- list(seq(ymin[1], ymax[1], length=lygridBi),</pre>
                  seq(ymin[2], ymax[2], length=lygridBi),
                  seq(ymin[3], ymax[3], length=lygridBi),
                  seq(ymin[4], ymax[4], length=lygridBi),
                  seq(ymin[5], ymax[5], length=lygridBi),
                  seq(ymin[6], ymax[6], length=lygridBi))
 ygridUni <- list(seq(ymin[1], ymax[1], length=lygridUni),</pre>
                   seq(ymin[2], ymax[2], length=lygridUni),
                   seq(ymin[3], ymax[3], length=lygridUni),
                   seq(ymin[4], ymax[4], length=lygridUni),
                   seq(ymin[5], ymax[5], length=lygridUni),
                   seq(ymin[6], ymax[6], length=lygridUni))
```

# 4 Model with two mixture components

In this section, we will fit a mixture model with K=2 components.

R⇒ We will also compute penalized expected deviance and hence two chains will be sampled.

## 4.1 Specification of the prior distributions and MCMC simulation

 $R \Rightarrow$  The minimal specification of the prior distribution with K=2:

```
> Prior0 <- list(priorK = "fixed", Kmax = 2)</pre>
```

 $R \Rightarrow$  Running MCMC with default values for all prior parameters and K = 2 (output is shown from MCMC simulation performed by author):

```
> if (RUN.TIMECONSUMING.CODE){
    set.seed(770328)
   Model0 <- NMixMCMC(y0=y0, y1=y1, censor=censor, prior=Prior0,</pre>
                       nMCMC=nMCMC, scale=list(shift=0, scale=1), PED=TRUE)
+ }
Chain number 1
_____
MCMC sampling started on Sun Apr 26 11:25:40 2009.
Burn-in iteration 10000
Iteration 30000
MCMC sampling finished on Sun Apr 26 13:10:24 2009.
Chain number 2
MCMC sampling started on Sun Apr 26 13:10:26 2009.
Burn-in iteration 10000
Iteration 30000
MCMC sampling finished on Sun Apr 26 14:55:48 2009.
Computation of penalized expected deviance started on Sun Apr 26 14:55:50 2009.
```

 $R \Rightarrow$  The prior distribution for the function NMixMCMC was the same as with

Computation of penalized expected deviance finished on Sun Apr 26 15:18:24 2009.

Note that due to the fact that the argument scale has been specified in function NMixMCMC as scale=list(shift=0, scale=1), the MCMC has been run on the original data.

#### 4.2 Posterior inference

R⇒ Basic posterior summary of the fitted model is obtained using the command print (Model0).

R $\Rightarrow$  Quantities shown in section labeled "Penalized expected deviance" are computed from two sampled chains and have the following meaning: D.expect is  $\hat{D}_e$  from Komárek (2009), p(opt) is estimated optimism  $\hat{p}_{opt}$  computed with unit weights, PED equals D.expect + p(opt) gives the estimate of penalized expected deviance with optimism computed without the use of importance sampling. Further, wp(opt) is estimated optimism  $\hat{p}_{opt}$  computed as described in Komárek (2009), i.e., using importance sampling. Finally, wPED equals D.expect + wp(opt) gives the estimate of penalized expected deviance as described in Komárek (2009).

R $\Rightarrow$  Quantities shown in section labeled "Deviance information criteria" are computed separately from the first and the second sampled chain. They have the following meaning: DIC is deviance information criterion denoted as DIC in Komárek (2009), pD is the effective dimension  $p_D$ , D.bar is approximated posterior mean  $\overline{D}$  of the deviance and D.in.bar is  $\tilde{D}$  – deviance evaluated in the "estimate".

R $\Rightarrow$  Section "Posterior summary statistics for moments of mixture for original data" gives posterior summary statistics for  $E(Y) = m + SE(Y^*)$  and quantities derived from  $var(Y) = S var(Y^*) S'$  in the notation of Komárek (2009), separately for each generated chain.

```
> print(Model0)
```

```
2 component normal mixture estimated using MCMC
```

```
Penalized expected deviance:
```

\_\_\_\_\_

```
D.expect p(opt) PED wp(opt) wPED 74778.2223 459.2338 75237.4561 459.2338 75237.4561
```

#### Deviance information criteria:

\_\_\_\_\_

```
DIC pD D.bar D.in.bar
Chain 1 64062.40 7098.499 56963.9 49865.40
Chain 2 64054.82 7096.818 56958.0 49861.18
```

### Posterior summary statistics for moments of mixture for original data:

-----

```
Means (chain 1):
```

```
y.Mean.1 y.Mean.2 y.Mean.3
                                            y.Mean.4
                                                        y.Mean.5
                                                                   y.Mean.6
        7.00102262 8.10834928 11.29975988 10.54675810 11.51834928 6.37538449
Std.Dev. 0.01375146 0.01527585 0.02815026 0.02266548 0.03371765 0.01519845
Min.
        6.94711487 8.04651059 11.19197588 10.45744045 11.39203273 6.32343325
2.5%
        6.97411202 8.07860646 11.24651693 10.50249254 11.45442970 6.34505266
        6.99174122 8.09806241 11.28039978 10.53153059 11.49521798 6.36535623
1st Qu.
        7.00122635 8.10821390 11.29913691 10.54667520 11.51756885 6.37549987
Median
3rd Qu.
        7.01032846 8.11863886 11.31816986 10.56194177 11.54052797 6.38565697
```

```
97.5%
         7.02763640 8.13856493 11.35682278 10.59131925 11.58678065 6.40472259
         7.05491105 8.16876362 11.42465980 10.62946205 11.69510593 6.43426475
Max.
Means (chain 2):
           y.Mean.1
                    y.Mean.2
                                  y.Mean.3
                                              y.Mean.4
                                                           y.Mean.5
                                                                      y.Mean.6
         7.00110318 8.10856452 11.29943283 10.54682157 11.51832469 6.37536765
Std.Dev. 0.01377727 0.01527937 0.02809354 0.02249410 0.03370979 0.01513347
         6.94634087 8.05016462 11.19021099 10.46194276 11.37340677 6.31682037
Min.
         6.97360828 8.07906882 11.24624302 10.50315966 11.45460077 6.34575436
2.5%
         6.99195384 8.09814585 11.28019586 10.53171044 11.49548388 6.36505283
1st Qu.
Median
         7.00124567 8.10849702 11.29896640 10.54677898 11.51746522 6.37533664
3rd Qu.
         7.01036670 8.11892163 11.31795651 10.56178921 11.54061897 6.38561720
97.5%
         7.02780596 8.13858318 11.35682863 10.59134962 11.58707869 6.40522481
Max.
         7.05318510 8.16998521 11.42133369 10.63809863 11.67877121 6.44064805
Standard deviations and correlations (chain 1):
             y.SD.1 y.Corr.2.1 y.Corr.3.1 y.Corr.4.1 y.Corr.5.1 y.Corr.6.1
         0.73193686 0.69046221 0.62497557 0.53914256 0.54549361 0.51926080
Mean
Std.Dev. 0.01229001 0.01260025 0.01630977 0.01647473 0.01828935 0.02218780
         0.68391225 0.63208218 0.54483922 0.46802571 0.47003346 0.43413094
Min.
2.5%
         0.70803615 0.66512178 0.59212096 0.50633349 0.50853909 0.47457376
1st Qu.
         0.72364934 0.68219136 0.61407742 0.52819445 0.53345843 0.50452237
         0.73194636 0.69064346 0.62536820 0.53943387 0.54555324 0.51959648
Median
3rd Qu.
         0.74015803 0.69911454 0.63608019 0.55031418 0.55774004 0.53458374
         0.75630350 0.71444541 0.65611875 0.57089306 0.58103201 0.56148304
97.5%
         0.79280701\ 0.73473514\ 0.68552354\ 0.59769910\ 0.62247391\ 0.59981795
Max.
             y.SD.2 y.Corr.3.2 y.Corr.4.2 y.Corr.5.2 y.Corr.6.2
         0.93034835 0.61382150 0.47846794 0.49762726 0.42456799 1.33888038
Mean
Std.Dev. 0.01419046 0.01617137 0.01660534 0.01871340 0.02433653 0.03818642
         0.87298810 0.54590257 0.41029615 0.42165911 0.32340115 1.21146151
Min.
         0.90301097 0.58169254 0.44562835 0.46001236 0.37575223 1.27096470
2.5%
         0.92074441 0.60301670 0.46719011 0.48506071 0.40839809 1.31213939
1st Qu.
         0.93017796 0.61413383 0.47864335 0.49788239 0.42502482 1.33658615
Median
         0.93983998 0.62489415 0.48979726 0.51028896 0.44098052 1.36294137
3rd Qu.
97.5%
         0.95860917 0.64484377 0.51027175 0.53345156 0.47171558 1.41970829
         0.99100969 0.66952572 0.53414393 0.57279576 0.51080810 1.52135572
Max.
         y.Corr.4.3 y.Corr.5.3 y.Corr.6.3
                                              y.SD.4 y.Corr.5.4 y.Corr.6.4
         0.59830273\ 0.55772378\ 0.48610616\ 1.30003544\ 0.73632923\ 0.47032940
Mean
Std.Dev. 0.01931984 0.02069057 0.02664376 0.02202476 0.01427254 0.02478035
         0.52453772 0.44489211 0.36435822 1.20902995 0.66905579 0.37431619
Min.
2.5%
         0.55896787 0.51557089 0.43284903 1.25916574 0.70705975 0.42007667
         0.58558137  0.54426821  0.46847712  1.28513771  0.72696302  0.45404815
1st Qu.
         0.59869135 0.55813563 0.48645514 1.29927592 0.73684939 0.47077941
Median
         0.61180554 0.57187389 0.50421487 1.31426079 0.74607173 0.48712673
3rd Qu.
         0.63446898 \ 0.59679595 \ 0.53657647 \ 1.34559611 \ 0.76300541 \ 0.51767794
97.5%
         0.68963845 0.63646183 0.58216992 1.41443024 0.78787508 0.55395921
Max.
             y.SD.5 y.Corr.6.5
                                   y.SD.6
         1.52876064 0.50488853 0.57949181
Mean
```

```
Std.Dev. 0.04620749 0.02645607 0.01172826
Min.
         1.38098016 0.40281237 0.53351873
2.5%
         1.44630482 0.45189276 0.55641042
         1.49642788 0.48720407 0.57157770
1st Qu.
Median
         1.52566962 0.50541136 0.57943161
3rd Qu.
         1.55813809 0.52287338 0.58738606
97.5%
         1.62732978 0.55568326 0.60258691
         1.76983288 0.60195652 0.62588187
Max.
Standard deviations and correlations (chain 2):
             y.SD.1 y.Corr.2.1 y.Corr.3.1 y.Corr.4.1 y.Corr.5.1 y.Corr.6.1
Mean
         0.73193038 0.69047857 0.62495701 0.5391806 0.54571215 0.51955487
Std.Dev. 0.01222552 0.01276867 0.01649097 0.0166532 0.01822416 0.02228140
Min.
         0.68419078 0.63451577 0.55786986 0.4569567 0.45900697 0.42806345
         0.70815456 0.66441375 0.59176430 0.5054894 0.50878992 0.47500115
2.5%
1st Qu.
         0.72368188 0.68219890 0.61401213 0.5282277 0.53359237 0.50473753
         0.73197648 0.69079633 0.62528624 0.5394119 0.54592312 0.52004345
Median
3rd Qu.
         0.74000928 0.69917178 0.63606359 0.5506265 0.55817953 0.53482575
         0.75615590 0.71489740 0.65630551 0.5708424 0.58055171 0.56203937
97.5%
Max.
         0.78013331 0.73674696 0.68816049 0.6028364 0.61128646 0.60911313
             y.SD.2 y.Corr.3.2 y.Corr.4.2 y.Corr.5.2 y.Corr.6.2
Mean
         0.93040224 0.61376874 0.47847419 0.49785089 0.42464098 1.33845172
Std.Dev. 0.01432785 0.01627950 0.01667399 0.01870734 0.02419231 0.03810459
Min.
         0.87458004 0.53872991 0.41203259 0.41227712 0.31909957 1.21447630
2.5%
         0.90300818 0.58135917 0.44508801 0.46047869 0.37724587 1.27012515
         0.92051495 0.60292695 0.46737588 0.48525176 0.40841386 1.31194839
1st Qu.
         0.93014634 0.61414163 0.47862421 0.49803323 0.42492399 1.33622217
Median
         0.93990290 0.62493404 0.48986882 0.51053207 0.44128408 1.36273203
3rd Qu.
97.5%
         0.95908202 0.64499439 0.51024479 0.53369959 0.47110881 1.41874295
         0.99097289 0.67162720 0.54293518 0.57946629 0.51892806 1.54957489
Max.
         y.Corr.4.3 y.Corr.5.3 y.Corr.6.3
                                              y.SD.4 y.Corr.5.4 y.Corr.6.4
         0.59852591 0.55780945 0.48615027 1.29999036 0.73645422 0.47059316
Mean
Std.Dev. 0.01942745 0.02052465 0.02670332 0.02182824 0.01420315 0.02486793
         0.49664176 0.46816619 0.38025760 1.21477424 0.67780441 0.36529847
Min.
2.5%
         0.55908511 0.51662955 0.43317760 1.25901228 0.70757615 0.42070864
         0.58589315 0.54433447 0.46828976 1.28489900 0.72726704 0.45416075
1st Qu.
         0.59893110 0.55835707 0.48654016 1.29928296 0.73677950 0.47108515
Median
         0.61195867 0.57203660 0.50422386 1.31429359 0.74616889 0.48734453
3rd Qu.
97.5%
         0.63489268 0.59616657 0.53764807 1.34488597 0.76314732 0.51805408
Max.
         0.66965451 0.63037709 0.58425141 1.40418926 0.78786136 0.55658624
             y.SD.5 y.Corr.6.5
                                   v.SD.6
         1.52865200 0.50536940 0.57946697
Mean
Std.Dev. 0.04600838 0.02682988 0.01151379
         1.37806663 0.37240365 0.53640705
Min.
2.5%
         1.44680507 0.45133093 0.55679329
1st Qu.
         1.49652250 0.48759810 0.57181555
Median
         1.52570964 0.50590876 0.57948013
         1.55739033 0.52363467 0.58721213
3rd Qu.
```

```
97.5%
         1.62658443 0.55602929 0.60216855
Max.
         1.75117759 0.60487881 0.62841052
R⇒ Computation of the marginal (univariate) predictive densities (separately for chain 1 and
chain 2):
> if (RUN.TIMECONSUMING.CODE){
+ PDensUniModel0 <- list()
+ PDensUniModel0[[1]] <- NMixPredDensMarg(Model0[[1]], grid=ygridUni)
+ PDensUniModel0[[2]] <- NMixPredDensMarg(Model0[[2]], grid=ygridUni)
+ }
R⇒ Default plot method for the computed object (see Figure 1):
> postscript(paste(FIGDIR, "figTandmob01.ps", sep=""), width=10, height=7,
             horizontal=FALSE)
> plot(PDensUniModel0[[1]])
> dev.off()
```

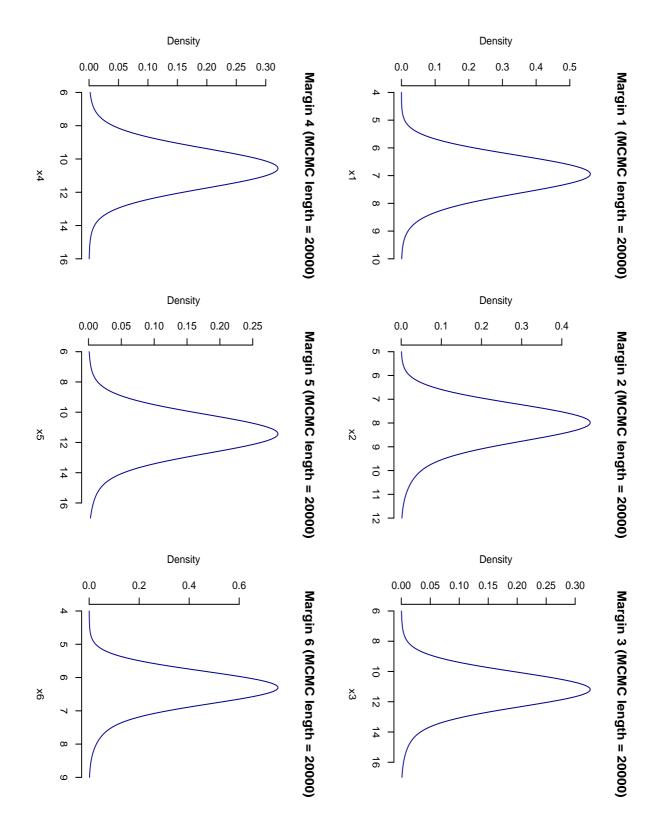
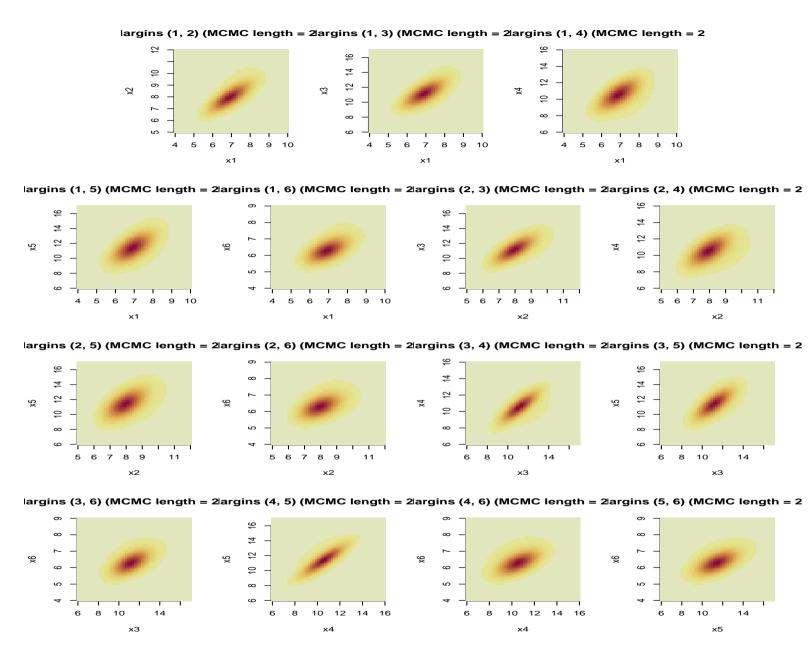


Figure 1: Default **plot** method for the marginal predictive densities based on the model with two mixture components (results from chain 1).

 $R \Rightarrow$  Computation of the joint bivariate predictive densities (for each pair and separately for chain 1 and chain 2):



Default plot method for the joint bivariate predictive densities based on the model with two mixture components (results from chain 1). 5. Figure?

## 4.3 Nicer figures of posterior predictive densities

 $R \Rightarrow$  Plot of the marginal predictive densities, results from chain 1 in red, results from chain 2 in blue, (see Figure 3):

```
> postscript(paste(FIGDIR, "figTandmob03.ps", sep=""), width=7, height=5,
             horizontal=FALSE)
> xlim <- c(4, 17)
> ylim <- c(0, 0.8)
> par(mfrow=c(2, 3), bty="n", mar=c(4, 4, 4, 0)+0.1)
> for (tt in 1:6){
   plot(PDensUniModel0[[1]]$x[[tt]], PDensUniModel0[[1]]$dens[[tt]],
         type="1", col="red", xlim=xlim, ylim=ylim,
         xlab="Age (years)", ylab="Density of emergence",
         main=paste("Tooth ", 10+tt, sep=""))
    lines(PDensUniModel0[[2]]$x[[tt]], PDensUniModel0[[2]]$dens[[tt]], col="blue")
+ }
> dev.off()
R⇒ Contour plots of the joint bivariate predictive densities for each pair, results from chain 1 (see
Figure 4):
> CH <- 1
> postscript(paste(FIGDIR, "figTandmob04.ps", sep=""), width=10, height=7,
             horizontal=FALSE)
> par(mfrow=c(3, 5), bty="n", mar=c(4, 4, 4, 0)+0.1)
> for (tt1 in 1:5){
    for (tt2 in (tt1+1):6){
      contour(PDensBiModel0[[CH]]$x[[tt1]], PDensBiModel0[[CH]]$x[[tt2]],
              PDensBiModel0[[CH]]$dens[[paste(tt1, "-", tt2, sep="")]],
              col="red", xlab="Age (years)", ylab="Age (years)",
              main=paste("Teeth ", 10+tt1, "-", 10+tt2, sep=""))
    7
+ }
> dev.off()
```

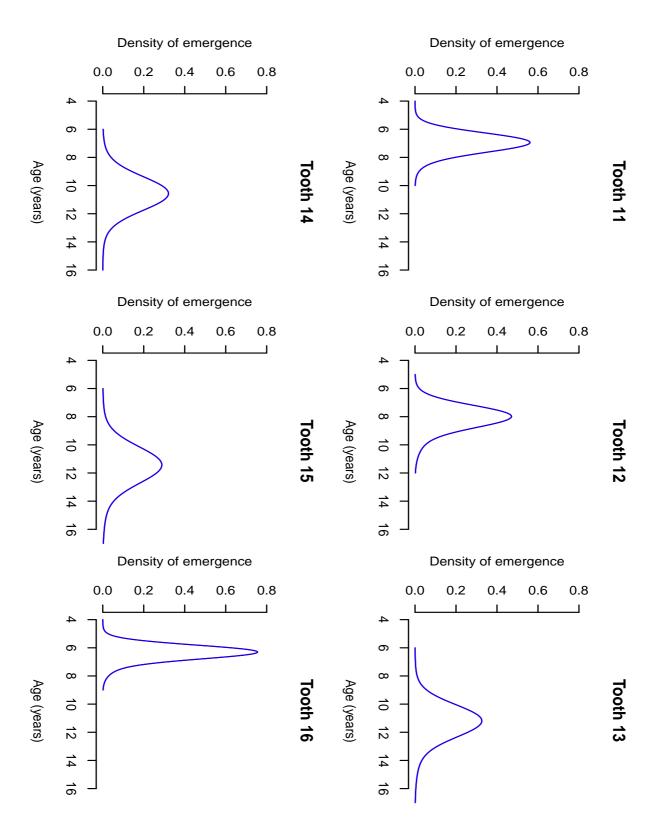
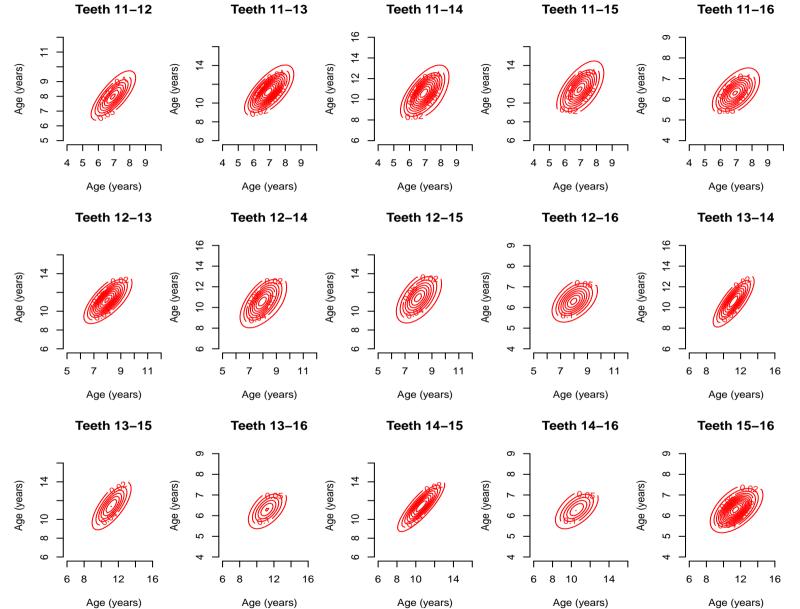


Figure 3: Marginal predictive densities based on the model with two mixture components, results from chain 1 in red, results from chain 2 in blue.



Contour plots of the joint bivariate predictive densities based on the model with two mixture components, results from chain 1. Figure 4:

 $R \Rightarrow$  Image plots of the joint bivariate predictive densities for each pair, results from chain 1 (see Figure 5):

Note that package colorspace is needed to specify the colors in the plot.

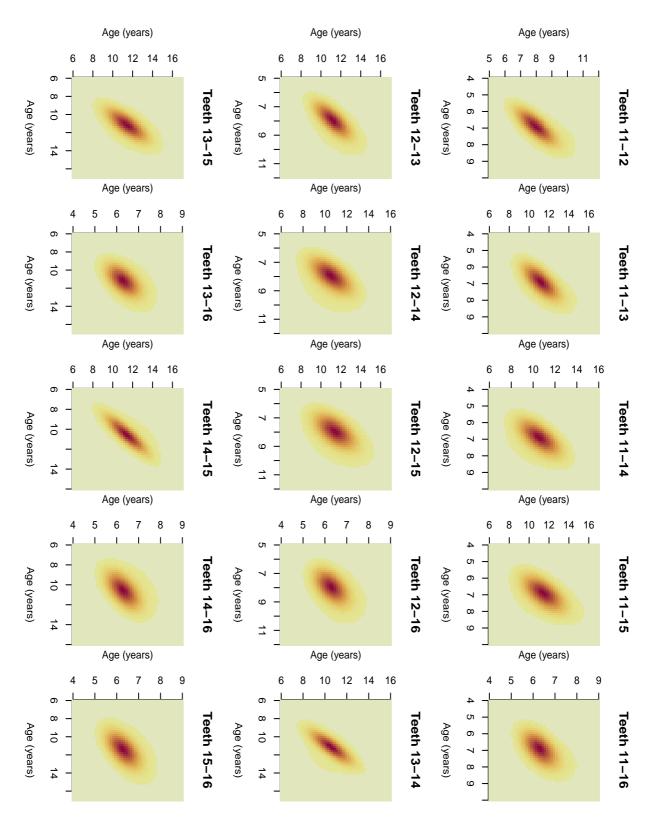


Figure 5: Image plots of the joint bivariate predictive densities based on the model with two mixture components, results from chain 1.

## 4.4 Convergence diagnostics

R⇒ Single chain convergence diagnostics using chain 1 will be shown here.

```
> CH <- 1
```

R⇒ Converting the chains into mcmc objects to be used in the package coda:

```
> if (RUN.ALLOUT){
+   start <- Model0[[CH]]$nMCMC["burn"] + 1
+   end <- Model0[[CH]]$nMCMC["burn"] + Model0[[CH]]$nMCMC["keep"]
+   chgammaInv <- mcmc(Model0[[CH]]$gammaInv, start=start, end=end)
+   chmixture <- mcmc(Model0[[CH]]$mixture, start=start, end=end)
+   chdeviance <- mcmc(Model0[[CH]]$deviance, start=start, end=end)
+ }</pre>
```

## 4.4.1 Traceplots

 $R \Rightarrow$  Traceplots for the overall means of emergence (see Figure 6):

```
> if (RUN.ALLOUT){
    postscript(paste(FIGKEEPDIR, "figTandmob06.ps", sep=""), width=7, height=10,
               horizontal=FALSE)
   1wd <- 0.5
    par(mfrow=c(2, 3), bty="n")
    traceplot(chmixture[, "y.Mean.1"], smooth=FALSE, col="darkblue",
              lwd=lwd, main="E(Tooth 11)")
    traceplot(chmixture[, "y.Mean.2"], smooth=FALSE, col="darkblue",
              lwd=lwd, main="E(Tooth 12)")
+
    traceplot(chmixture[, "y.Mean.3"], smooth=FALSE, col="darkblue",
              lwd=lwd, main="E(Tooth 13)")
    traceplot(chmixture[, "y.Mean.4"], smooth=FALSE, col="darkblue",
              lwd=lwd, main="E(Tooth 14)")
    traceplot(chmixture[, "y.Mean.5"], smooth=FALSE, col="darkblue",
+
              lwd=lwd, main="E(Tooth 15)")
    traceplot(chmixture[, "y.Mean.6"], smooth=FALSE, col="darkblue",
              lwd=lwd, main="E(Tooth 16)")
    dev.off()
+ }
```

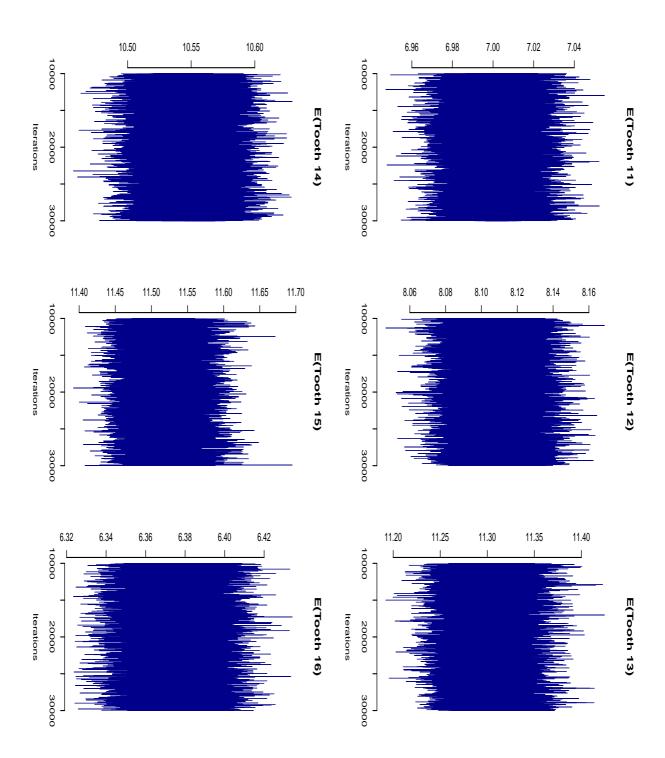


Figure 6: Model with two mixture components. Traceplots for the means of emergence.

```
R⇒ Traceplots for the standard deviations of emergence (not shown):
```

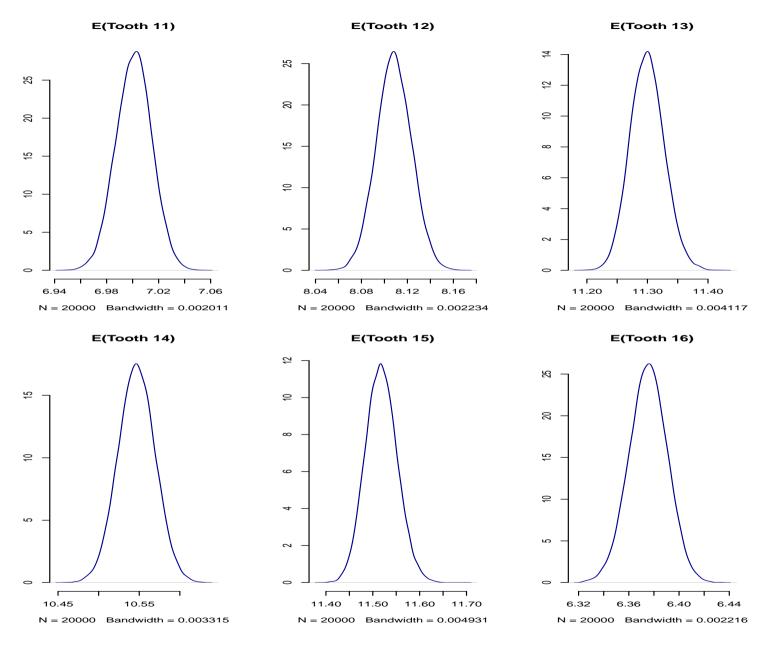
```
> if (RUN.ALLOUT){
    postscript(paste(FIGKEEPDIR, "figTandmob07.ps", sep=""), width=7, height=10,
               horizontal=FALSE)
    par(mfrow=c(2, 3), bty="n")
    traceplot(chmixture[, "y.SD.1"], smooth=FALSE, col="darkgreen",
              lwd=lwd, main="SD(Tooth 11)")
+
    traceplot(chmixture[, "y.SD.2"], smooth=FALSE, col="darkgreen",
              lwd=lwd, main="SD(Tooth 12)")
    traceplot(chmixture[, "y.SD.3"], smooth=FALSE, col="darkgreen",
+
+
              lwd=lwd, main="SD(Tooth 13)")
    traceplot(chmixture[, "y.SD.4"], smooth=FALSE, col="darkgreen",
              lwd=lwd, main="SD(Tooth 14)")
+
    traceplot(chmixture[, "y.SD.5"], smooth=FALSE, col="darkgreen",
              lwd=lwd, main="SD(Tooth 15)")
    traceplot(chmixture[, "y.SD.6"], smooth=FALSE, col="darkgreen",
              lwd=lwd, main="SD(Tooth 16)")
    dev.off()
+ }
R⇒ Traceplots for the pairwise correlations between the emergence times (not shown):
> if (RUN.ALLOUT){
    postscript(paste(FIGKEEPDIR, "figTandmob08.ps", sep=""), width=10, height=7,
               horizontal=FALSE)
    par(bty="n", mar=c(1, 1, 4, 0)+0.1)
    layout(matrix(c(1,2,3,4,5,0,6,7,8,9,0,0,10,11,12,0,0,0,13,14,0,0,0,0,15),
                  nrow=5, byrow=TRUE))
    traceplot(chmixture[, "y.Corr.2.1"], smooth=FALSE, col="darkgreen",
+
              main="Corr 2-1")
+
    traceplot(chmixture[, "y.Corr.3.1"], smooth=FALSE, col="darkgreen",
+
              main="Corr 3-1")
    traceplot(chmixture[, "y.Corr.4.1"], smooth=FALSE, col="darkgreen",
+
              main="Corr 4-1")
+
    traceplot(chmixture[, "y.Corr.5.1"], smooth=FALSE, col="darkgreen",
+
              main="Corr 5-1")
    traceplot(chmixture[, "y.Corr.6.1"], smooth=FALSE, col="darkgreen",
+
              main="Corr 6-1")
+
    traceplot(chmixture[, "y.Corr.3.2"], smooth=FALSE, col="darkgreen",
+
              main="Corr 3-2")
    traceplot(chmixture[, "y.Corr.4.2"], smooth=FALSE, col="darkgreen",
              main="Corr 4-2")
    traceplot(chmixture[, "y.Corr.5.2"], smooth=FALSE, col="darkgreen",
+
              main="Corr 5-2")
    traceplot(chmixture[, "y.Corr.6.2"], smooth=FALSE, col="darkgreen",
              main="Corr 6-2")
    traceplot(chmixture[, "y.Corr.4.3"], smooth=FALSE, col="darkgreen",
```

```
main="Corr 4-3")
    traceplot(chmixture[, "y.Corr.5.3"], smooth=FALSE, col="darkgreen",
              main="Corr 5-3")
    traceplot(chmixture[, "y.Corr.6.3"], smooth=FALSE, col="darkgreen",
              main="Corr 6-3")
    traceplot(chmixture[, "y.Corr.5.4"], smooth=FALSE, col="darkgreen",
              main="Corr 5-4")
    traceplot(chmixture[, "y.Corr.6.4"], smooth=FALSE, col="darkgreen",
              main="Corr 6-4")
+
   traceplot(chmixture[, "y.Corr.6.5"], smooth=FALSE, col="darkgreen",
              main="Corr 6-5")
    dev.off()
+ }
R⇒ Traceplots for some other parameters (not shown):
> if (RUN.ALLOUT){
    postscript(paste(FIGKEEPDIR, "figTandmob09.ps", sep=""), width=10, height=7,
               horizontal=FALSE)
    par(bty="n")
    layout(matrix(c(1,1,2,2,3,3,0,4,4,5,5,0), nrow=2, byrow=TRUE))
+
    traceplot(chgammaInv[, "gammaInv1"], smooth=FALSE,
              col="brown", main="gamma^{-1}")
    traceplot(chdeviance[, "LogLO"], smooth=FALSE,
              col="red", lwd=lwd, main="Log(L0)")
    traceplot(chdeviance[, "LogL1"], smooth=FALSE,
              col="red", lwd=lwd, main="Log(L1)")
    traceplot(chdeviance[, "dev.complete"], smooth=FALSE,
              col="red", lwd=lwd, main="D(complete)")
    traceplot(chdeviance[, "dev.observed"], smooth=FALSE,
              col="red", lwd=lwd, main="D(observed)")
    dev.off()
```

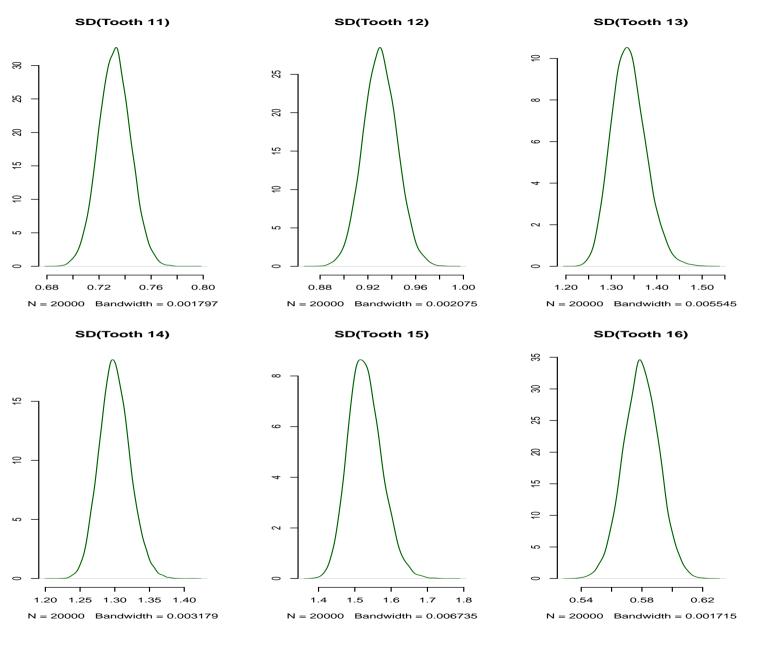
#### 4.4.2 Posterior density estimates

R⇒ Posterior density estimates for the overall means of emergence (see Figure 7):

```
> if (RUN.ALLOUT){
    postscript(paste(FIGKEEPDIR, "figTandmob10.ps", sep=""), width=7, height=10,
               horizontal=FALSE)
    par(mfrow=c(2, 3), bty="n")
    densplot(chmixture[, "y.Mean.1"], show.obs=FALSE, col="darkblue",
             main="E(Tooth 11)")
    densplot(chmixture[, "y.Mean.2"], show.obs=FALSE, col="darkblue",
+
             main="E(Tooth 12)")
+
+
    densplot(chmixture[, "y.Mean.3"], show.obs=FALSE, col="darkblue",
             main="E(Tooth 13)")
+
    densplot(chmixture[, "y.Mean.4"], show.obs=FALSE, col="darkblue",
             main="E(Tooth 14)")
+
    densplot(chmixture[, "y.Mean.5"], show.obs=FALSE, col="darkblue",
             main="E(Tooth 15)")
    densplot(chmixture[, "y.Mean.6"], show.obs=FALSE, col="darkblue",
+
             main="E(Tooth 16)")
    dev.off()
+ }
R⇒ Posterior density estimates for the standard deviations of emergence (see Figure 8):
> if (RUN.ALLOUT){
    postscript(paste(FIGKEEPDIR, "figTandmob11.ps", sep=""), width=7, height=10,
               horizontal=FALSE)
    par(mfrow=c(2, 3), bty="n")
    densplot(chmixture[, "y.SD.1"], show.obs=FALSE, col="darkgreen",
             main="SD(Tooth 11)")
    densplot(chmixture[, "y.SD.2"], show.obs=FALSE, col="darkgreen",
             main="SD(Tooth 12)")
    densplot(chmixture[, "y.SD.3"], show.obs=FALSE, col="darkgreen",
+
             main="SD(Tooth 13)")
+
    densplot(chmixture[, "y.SD.4"], show.obs=FALSE, col="darkgreen",
             main="SD(Tooth 14)")
    densplot(chmixture[, "y.SD.5"], show.obs=FALSE, col="darkgreen",
             main="SD(Tooth 15)")
    densplot(chmixture[, "y.SD.6"], show.obs=FALSE, col="darkgreen",
             main="SD(Tooth 16)")
    dev.off()
```



Posterior density estimates for the means of Figure 7: Model with two mixture components. emergence.



Posterior density estimates for the standard Model with two mixture components. Figure 8: Model with t deviations of emergence.

 $R \Rightarrow$  Posterior density estimates for the pairwise correlations between the emergence times (see Figure 9):

```
> if (RUN.ALLOUT){
   postscript(paste(FIGKEEPDIR, "figTandmob12.ps", sep=""), width=10, height=7,
               horizontal=FALSE)
    par(bty="n", mar=c(2, 1, 4, 0)+0.1)
    layout(matrix(c(1,2,3,4,5,0,6,7,8,9,0,0,10,11,12,
                    0,0,0,13,14, 0,0,0,0,15), nrow=5, byrow=TRUE))
    densplot(chmixture[, "y.Corr.2.1"], show.obs=FALSE, col="darkgreen",
+
             main="Corr 2-1")
    densplot(chmixture[, "y.Corr.3.1"], show.obs=FALSE, col="darkgreen",
             main="Corr 3-1")
+
    densplot(chmixture[, "y.Corr.4.1"], show.obs=FALSE, col="darkgreen",
             main="Corr 4-1")
+
    densplot(chmixture[, "y.Corr.5.1"], show.obs=FALSE, col="darkgreen",
             main="Corr 5-1")
    densplot(chmixture[, "y.Corr.6.1"], show.obs=FALSE, col="darkgreen",
             main="Corr 6-1")
    densplot(chmixture[, "y.Corr.3.2"], show.obs=FALSE, col="darkgreen",
+
             main="Corr 3-2")
+
    densplot(chmixture[, "y.Corr.4.2"], show.obs=FALSE, col="darkgreen",
             main="Corr 4-2")
    densplot(chmixture[, "y.Corr.5.2"], show.obs=FALSE, col="darkgreen",
             main="Corr 5-2")
    densplot(chmixture[, "y.Corr.6.2"], show.obs=FALSE, col="darkgreen",
             main="Corr 6-2")
+
    densplot(chmixture[, "y.Corr.4.3"], show.obs=FALSE, col="darkgreen",
             main="Corr 4-3")
    densplot(chmixture[, "y.Corr.5.3"], show.obs=FALSE, col="darkgreen",
             main="Corr 5-3")
+
    densplot(chmixture[, "y.Corr.6.3"], show.obs=FALSE, col="darkgreen",
             main="Corr 6-3")
+
    densplot(chmixture[, "y.Corr.5.4"], show.obs=FALSE, col="darkgreen",
             main="Corr 5-4")
    densplot(chmixture[, "y.Corr.6.4"], show.obs=FALSE, col="darkgreen",
             main="Corr 6-4")
    densplot(chmixture[, "y.Corr.6.5"], show.obs=FALSE, col="darkgreen",
             main="Corr 6-5")
    dev.off()
+ }
```

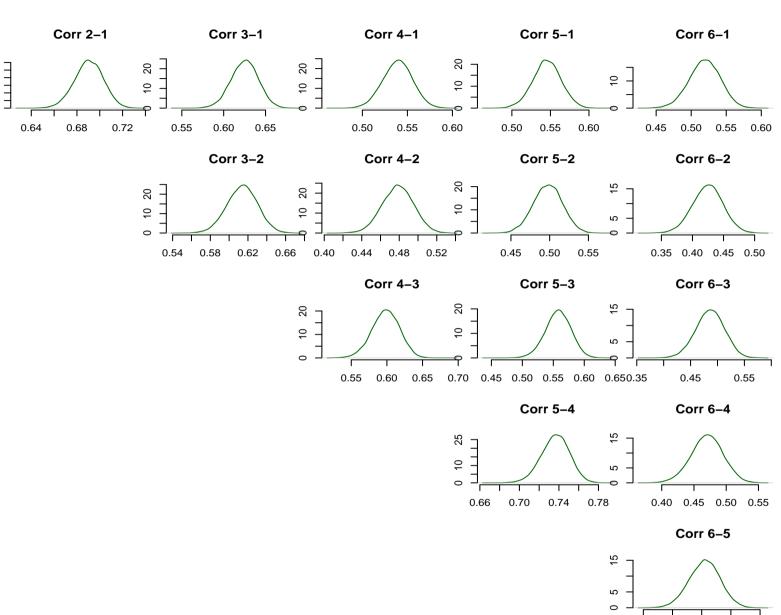


Figure 9: Model with two mixture components. Posterior density estimates for the pairwise correlations between the emergence times.

0.40

0.50

0.60

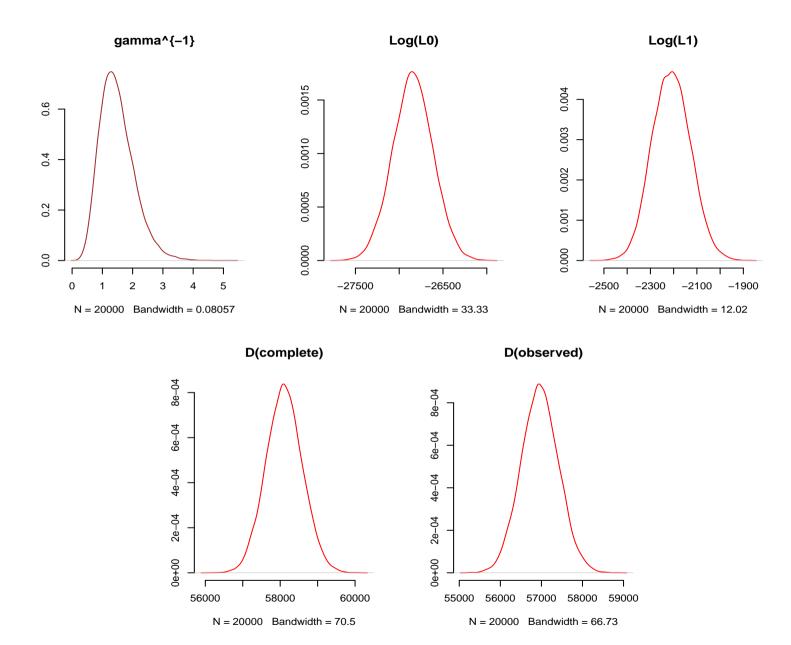
## R⇒ Posterior density estimates for some other parameters (see Figure 10):

```
> if (RUN.ALLOUT){
   postscript(paste(FIGKEEPDIR, "figTandmob13.ps", sep=""), width=10, height=7,
               horizontal=FALSE)
   par(bty="n")
   layout(matrix(c(1,1,2,2,3,3,0,4,4,5,5,0),
           nrow=2, byrow=TRUE))
    densplot(chgammaInv[, "gammaInv1"], show.obs=FALSE,
             col="brown", main="gamma^{-1}")
    densplot(chdeviance[, "LogLO"], show.obs=FALSE,
             col="red", main="Log(L0)")
    densplot(chdeviance[, "LogL1"], show.obs=FALSE,
             col="red", main="Log(L1)")
   densplot(chdeviance[, "dev.complete"], show.obs=FALSE,
             col="red", main="D(complete)")
    densplot(chdeviance[, "dev.observed"], show.obs=FALSE,
             col="red", main="D(observed)")
   dev.off()
+ }
```



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Figure 10: Model with two mixture components. Posterior density estimates for some other parameters.



## 4.4.3 Autocorrelation plots

R⇒ Autocorrelation plots for the overall means of emergence (see Figure 11):

```
> if (RUN.ALLOUT){
    postscript(paste(FIGKEEPDIR, "figTandmob14.ps", sep=""), width=7, height=10,
               horizontal=FALSE)
    par(mfrow=c(2, 3), bty="n")
    autocorr.plot(chmixture[, "y.Mean.1"], auto.layout=FALSE, ask=FALSE,
                  col="darkblue", main="E(Tooth 11)")
    autocorr.plot(chmixture[, "y.Mean.2"], auto.layout=FALSE, ask=FALSE,
                  col="darkblue", main="E(Tooth 12)")
    autocorr.plot(chmixture[, "y.Mean.3"], auto.layout=FALSE, ask=FALSE,
                  col="darkblue", main="E(Tooth 13)")
    autocorr.plot(chmixture[, "y.Mean.4"], auto.layout=FALSE, ask=FALSE,
                  col="darkblue", main="E(Tooth 14)")
    autocorr.plot(chmixture[, "y.Mean.5"], auto.layout=FALSE, ask=FALSE,
                  col="darkblue", main="E(Tooth 15)")
    autocorr.plot(chmixture[, "y.Mean.6"], auto.layout=FALSE, ask=FALSE,
                  col="darkblue", main="E(Tooth 16)")
    dev.off()
+ }
R⇒ Autocorrelation plots for the standard deviations of emergence (see Figure 12):
> if (RUN.ALLOUT){
    postscript(paste(FIGKEEPDIR, "figTandmob15.ps", sep=""), width=7, height=10,
               horizontal=FALSE)
    par(mfrow=c(2, 3), bty="n")
    autocorr.plot(chmixture[, "y.SD.1"], auto.layout=FALSE, ask=FALSE,
                  col="darkgreen", main="SD(Tooth 11)")
    autocorr.plot(chmixture[, "y.SD.2"], auto.layout=FALSE, ask=FALSE,
                  col="darkgreen", main="SD(Tooth 12)")
+
    autocorr.plot(chmixture[, "y.SD.3"], auto.layout=FALSE, ask=FALSE,
                  col="darkgreen", main="SD(Tooth 13)")
+
    autocorr.plot(chmixture[, "y.SD.4"], auto.layout=FALSE, ask=FALSE,
                  col="darkgreen", main="SD(Tooth 14)")
    autocorr.plot(chmixture[, "y.SD.5"], auto.layout=FALSE, ask=FALSE,
                  col="darkgreen", main="SD(Tooth 15)")
    autocorr.plot(chmixture[, "y.SD.6"], auto.layout=FALSE, ask=FALSE,
                  col="darkgreen", main="SD(Tooth 16)")
    dev.off()
```

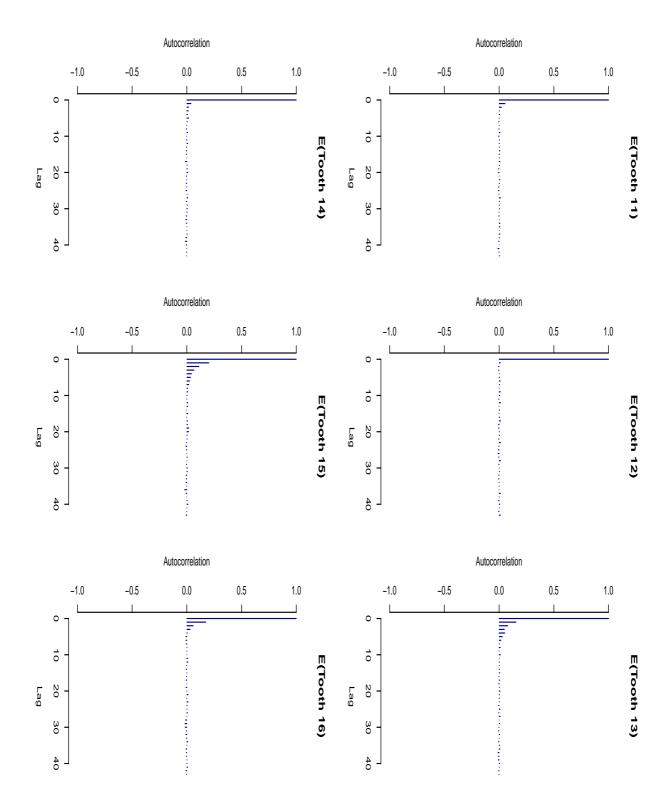


Figure 11: Model with two mixture components. Autocorrelation plots for the means of emergence.

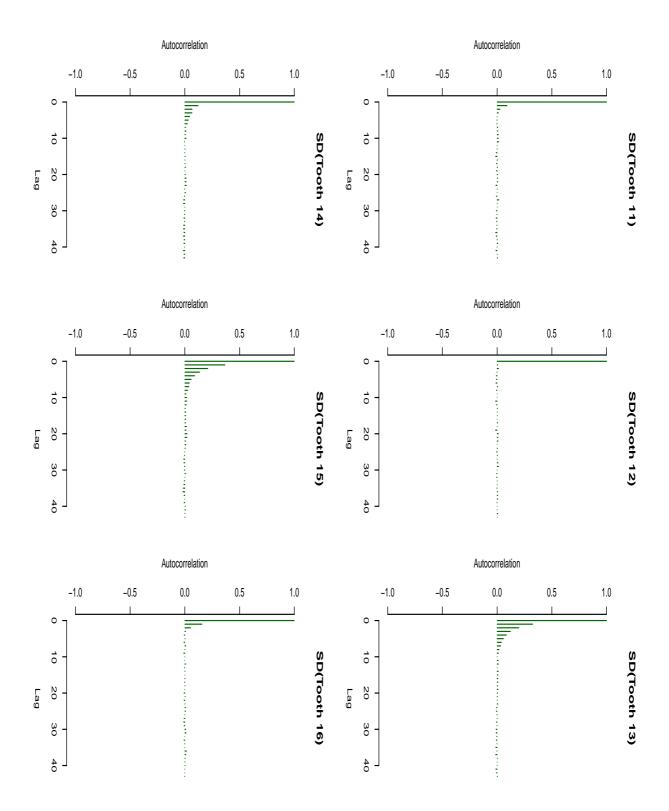


Figure 12: Model with two mixture components. Autocorrelation plots for the standard deviations of emergence.

#### R⇒ Autocorrelation plots for the pairwise correlations between the emergence times (see Figure 13):

```
> if (RUN.ALLOUT) {
    postscript(paste(FIGKEEPDIR, "figTandmob16.ps", sep=""), width=10, height=7,
               horizontal=FALSE)
    par(bty="n", mar=c(1, 1, 4, 0)+0.1)
    layout(matrix(c(1,2,3,4,5,0,6,7,8,9,0,0,10,11,12,
                    0,0,0,13,14, 0,0,0,0,15), nrow=5, byrow=TRUE))
    autocorr.plot(chmixture[, "y.Corr.2.1"], auto.layout=FALSE,
                  ask=FALSE, col="darkgreen", main="Corr 2-1")
+
    autocorr.plot(chmixture[, "y.Corr.3.1"], auto.layout=FALSE,
                  ask=FALSE, col="darkgreen", main="Corr 3-1")
    autocorr.plot(chmixture[, "y.Corr.4.1"], auto.layout=FALSE,
                  ask=FALSE, col="darkgreen", main="Corr 4-1")
    autocorr.plot(chmixture[, "y.Corr.5.1"], auto.layout=FALSE,
+
+
                  ask=FALSE, col="darkgreen", main="Corr 5-1")
    autocorr.plot(chmixture[, "y.Corr.6.1"], auto.layout=FALSE,
                  ask=FALSE, col="darkgreen", main="Corr 6-1")
+
    autocorr.plot(chmixture[, "y.Corr.3.2"], auto.layout=FALSE,
                  ask=FALSE, col="darkgreen", main="Corr 3-2")
+
    autocorr.plot(chmixture[, "y.Corr.4.2"], auto.layout=FALSE,
+
                  ask=FALSE, col="darkgreen", main="Corr 4-2")
    autocorr.plot(chmixture[, "y.Corr.5.2"], auto.layout=FALSE,
                  ask=FALSE, col="darkgreen", main="Corr 5-2")
    autocorr.plot(chmixture[, "y.Corr.6.2"], auto.layout=FALSE,
+
                  ask=FALSE, col="darkgreen", main="Corr 6-2")
+
    autocorr.plot(chmixture[, "y.Corr.4.3"], auto.layout=FALSE,
+
                  ask=FALSE, col="darkgreen", main="Corr 4-3")
    autocorr.plot(chmixture[, "y.Corr.5.3"], auto.layout=FALSE,
                  ask=FALSE, col="darkgreen", main="Corr 5-3")
    autocorr.plot(chmixture[, "y.Corr.6.3"], auto.layout=FALSE,
                  ask=FALSE, col="darkgreen", main="Corr 6-3")
    autocorr.plot(chmixture[, "y.Corr.5.4"], auto.layout=FALSE,
+
                  ask=FALSE, col="darkgreen", main="Corr 5-4")
+
    autocorr.plot(chmixture[, "y.Corr.6.4"], auto.layout=FALSE,
                  ask=FALSE, col="darkgreen", main="Corr 6-4")
    autocorr.plot(chmixture[, "y.Corr.6.5"], auto.layout=FALSE,
                  ask=FALSE, col="darkgreen", main="Corr 6-5")
    dev.off()
```

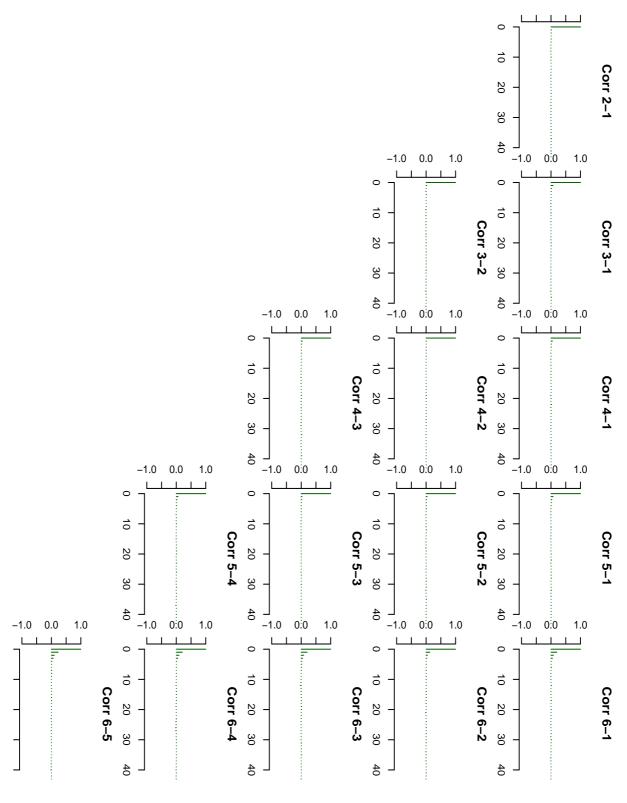


Figure 13: Model with two mixture components. Autocorrelation plots for the pairwise correlations between the emergence times.

## R⇒ Autocorrelation plots for some other parameters (see Figure 14):

```
> if (RUN.ALLOUT){
   postscript(paste(FIGKEEPDIR, "figTandmob17.ps", sep=""), width=10, height=7,
               horizontal=FALSE)
   par(bty="n")
   layout(matrix(c(1,1,2,2,3,3,0,4,4,5,5,0), nrow=2, byrow=TRUE))
    autocorr.plot(chgammaInv[, "gammaInv1"], auto.layout=FALSE,
                  ask=FALSE, col="brown", main="gamma^{-1}")
+
    autocorr.plot(chdeviance[, "LogLO"], auto.layout=FALSE,
                  ask=FALSE, col="red", main="Log(L0)")
    autocorr.plot(chdeviance[, "LogL1"], auto.layout=FALSE,
                  ask=FALSE, col="red", main="Log(L1)")
    autocorr.plot(chdeviance[, "dev.complete"], auto.layout=FALSE,
                  ask=FALSE, col="red", main="D(complete)")
    autocorr.plot(chdeviance[, "dev.observed"], auto.layout=FALSE,
                  ask=FALSE, col="red", main="D(observed)")
    dev.off()
```

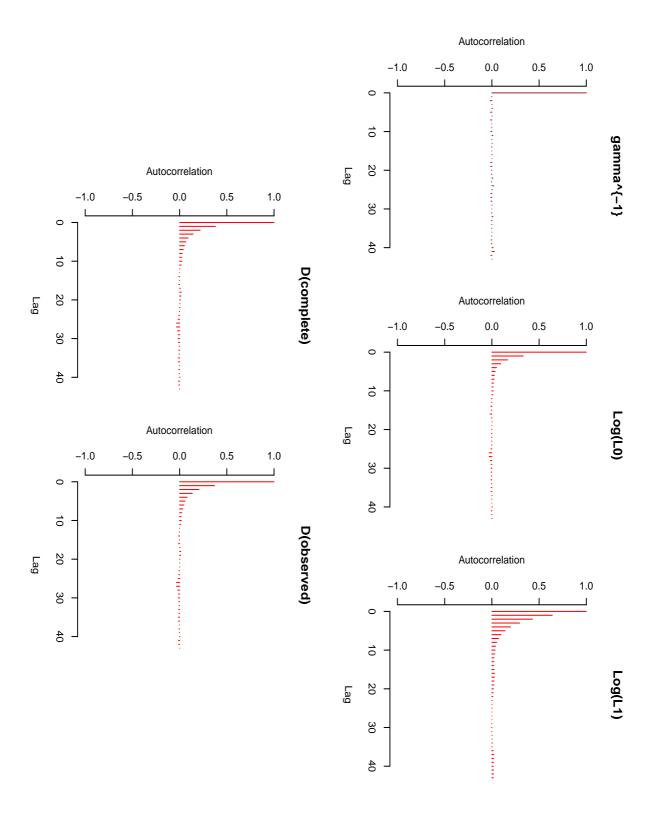


Figure 14: Model with two mixture components. Autocorrelation plots for some other parameters.

# 5 Models with different fixed numbers of components

In this section, we will fit a mixture model for  $K=1,\ldots,10$ , compare the deviance based quantities and predictive densities.

 $R \Rightarrow$  Running the MCMC simulation for K = 1, ..., 10 (output printed during the MCMC run on the screen not shown), computation of predictive densities.

R⇒ After predictive densities are computed, we remove all chains from resulting objects (to save some memory).

```
> Seed <- c(770328, 770328, 770328, 770328, 770328,
            770328, 780830, 761014, 770328, 770328)
> Keep <- c("iter", "nMCMC", "dim", "prior", "init", "RJMCMC",
            "scale", "freqK", "propK", "DIC", "moves",
+
            "summ.y.Mean", "summ.y.SDCorr",
            "summ.z.Mean", "summ.z.SDCorr")
> if (RUN.TIMECONSUMING.CODE){
   ModelK <- list()</pre>
    PDensUniModelK <- list()</pre>
   PDensBiModelK <- list()
    for (K in 1:10){
      set.seed(Seed[K])
      cat(paste("K = ", K, "\n----\n", sep=""))
      cat("Seed is", Seed[K], "\n")
      Prior0 <- list(priorK="fixed", Kmax=K)</pre>
      Model0 <- NMixMCMC(y0=y0, y1=y1, censor=censor, prior=Prior0,
                         nMCMC=nMCMC, scale=list(shift=0, scale=1), PED=TRUE)
      cat(paste("\nComputation of marginal pred. densities started on ", date(),
                "\n", sep=""))
      PDensUniModelK[[k]] <- list()</pre>
      PDensUniModelK[[k]][[1]] <- NMixPredDensMarg(ModelO[[1]], grid=ygridUni)
      PDensUniModelK[[k]][[2]] <- NMixPredDensMarg(ModelO[[2]], grid=ygridUni)
      cat(paste("Computation of marginal pred. densities finished on ", date(),
                "\n\n", sep=""))
      cat(paste("Computation of joint pred. densities started on ", date(),
                "\n", sep=""))
      PDensBiModelK[[k]] <- list()</pre>
      PDensBiModelK[[k]][[1]] <- NMixPredDensJoint2(Model0[[1]], grid=ygridBi)</pre>
      PDensBiModelK[[k]][[2]] <- NMixPredDensJoint2(Model0[[2]], grid=ygridBi)</pre>
      cat(paste("Computation of joint pred. densities finished on ", date(),
                "\n\n", sep=""))
      ModelK[[k]][[1]] <- Model0[[1]][Keep]</pre>
      ModelK[[k]][[2]] <- Model0[[2]][Keep]</pre>
      class(ModelK[[k]][[1]]) <- class(ModelK[[k]][[2]]) <- "NMixMCMC"</pre>
```

```
rm(list="Model0")
  }
+ }
R⇒ Summary of PED and DIC's for the fitted models:
> PED <- ModelK[[1]]$PED
> DIC <- list(Chain1=ModelK[[1]][[1]]$DIC, Chain2=ModelK[[1]][[2]]$DIC)
> for (k in 2:length(ModelK)){
   PED <- rbind(PED, ModelK[[k]]$PED)</pre>
   DIC[[1]] <- rbind(DIC[[1]], ModelK[[k]][[1]]$DIC)</pre>
   DIC[[2]] <- rbind(DIC[[2]], ModelK[[k]][[2]]$DIC)</pre>
+ }
> rownames(PED) <- paste("K=", 1:length(ModelK), sep="")</pre>
> rownames(DIC[[1]]) <- rownames(DIC[[2]]) <- paste("K=", 1:length(ModelK), sep="")</pre>
> print(PED)
    D.expect
               p(opt)
                            PED
                                 wp(opt)
                                               wPED
K=1 78695.46 157.6598 78853.12 157.6598 78853.12
K=2 74778.22 459.2338 75237.46 459.2338 75237.46
K=3 74923.44 934.1437 75857.58 934.1437 75857.58
K=4 75437.12 1235.8121 76672.94 1235.8121 76672.94
K=5 75632.71 1722.6202 77355.33 1722.6200 77355.33
K=6 76357.36 3171.4515 79528.81 3171.4518 79528.81
K=7 76269.30 3717.2918 79986.59 3717.2861 79986.59
K=8 76476.79 5059.1473 81535.94 5059.1495 81535.94
K=9 76668.48 5614.8044 82283.29 5614.8060 82283.29
K=10 76683.01 6620.5835 83303.59 6620.5875 83303.60
> print(DIC)
$Chain1
          DIC
                    рD
                          D.bar D.in.bar
K=1 66735.18 4699.580 62035.60 57336.02
K=2 64062.40 7098.499 56963.90 49865.40
K=3 64041.76 7190.355 56851.40 49661.05
K=4 63212.65 7061.547 56151.10 49089.55
K=5 62768.35 6957.968 55810.39 48852.42
K=6 64483.55 8535.527 55948.02 47412.50
K=7 65167.36 9111.387 56055.97 46944.59
K=8 63117.53 7186.084 55931.44 48745.36
K=9 62986.51 7055.850 55930.66 48874.82
K=10 64409.17 8468.477 55940.69 47472.21
```

```
$Chain2

DIC pD D.bar D.in.bar

K=1 66733.05 4699.827 62033.22 57333.39

K=2 64054.82 7096.818 56958.00 49861.18

K=3 64018.44 7184.664 56833.78 49649.12

K=4 63207.57 7060.128 56147.44 49087.32

K=5 62883.15 6994.865 55888.28 48893.42

K=6 64058.40 8111.954 55946.44 47834.49

K=7 62945.71 7034.567 55911.14 48876.58

K=8 64873.65 8877.754 55995.89 47118.14

K=9 63359.93 7463.778 55896.15 48432.37

K=10 63395.95 7303.974 56091.97 48788.00
```

 $R \Rightarrow$  Plots of the (univariate) predictive densities for different values of K are shown in Figures 15–17:

```
> if (RUN.ALLOUT){
    CH <- 1
    for (fig in 1:3){
      postscript(paste(FIGKEEPDIR, "figTandmob", 17+fig, ".ps", sep=""),
                 width=7, height=10, horizontal=FALSE)
      par(mfrow=c(2, 1), bty="n")
      for (tt in (2*(fig-1)+1):(2*fig)){
        plot(PDensUniModelK[[Kshow]][[CH]]$x[[tt]],
             PDensUniModelK[[Kshow]][[CH]]$dens[[tt]],
             type="n", xlab="Age (years)", ylab="Density",
             main=paste("Tooth ", 10+tt, sep=""))
        for (k in 1:10){
          lines(PDensUniModelK[[k]][[CH]]$x[[tt]],
                PDensUniModelK[[k]][[CH]]$dens[[tt]],
                col="blue")
        }
        lines(PDensUniModelK[[Kshow]][[CH]]$x[[tt]],
              PDensUniModelK[[Kshow]][[CH]]$dens[[tt]],
              col="red", lwd=2)
      }
      dev.off()
    }
+ }
```

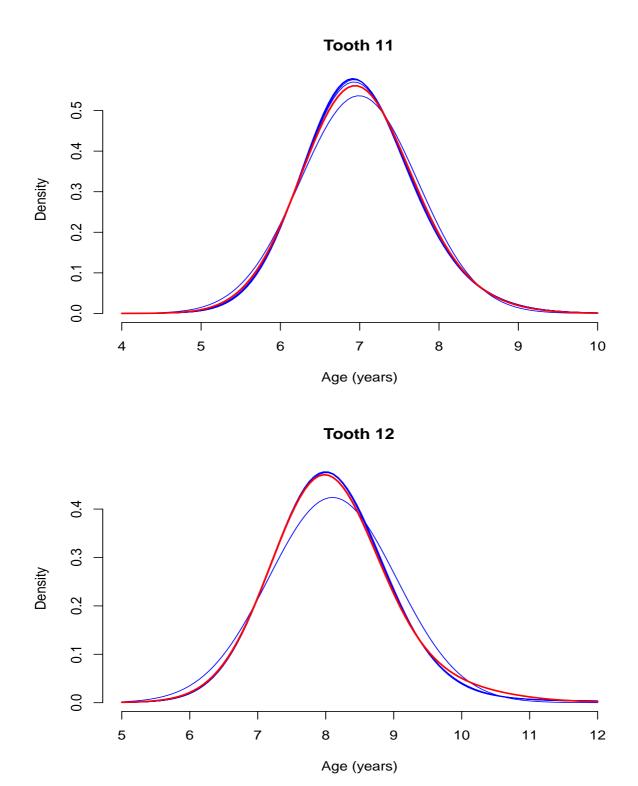


Figure 15: Teeth 11 and 12. Estimated predictive densities of emergence for different values of K ranging from 1 to 10 (blue lines) and for K = 2 (red line).

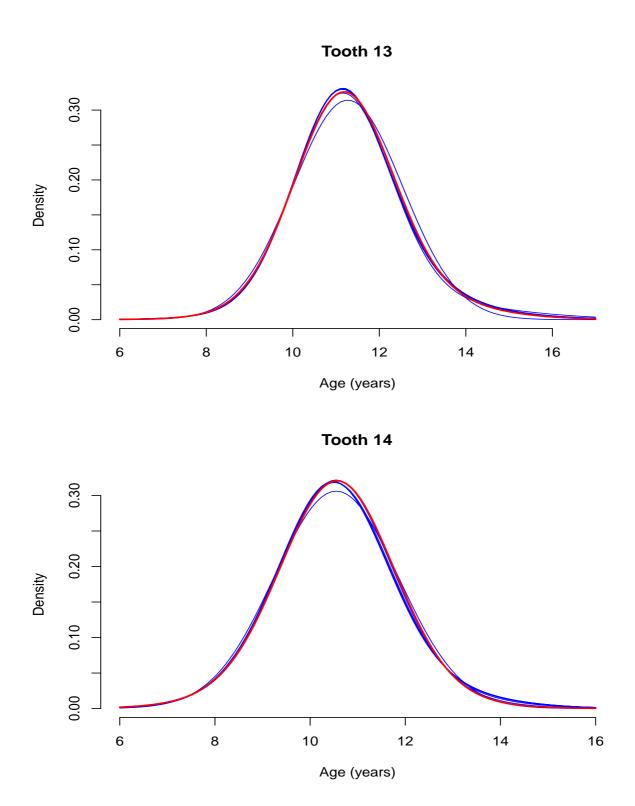


Figure 16: Teeth 13 and 14. Estimated predictive densities of emergence for different values of K ranging from 1 to 10 (blue lines) and for K = 2 (red line).

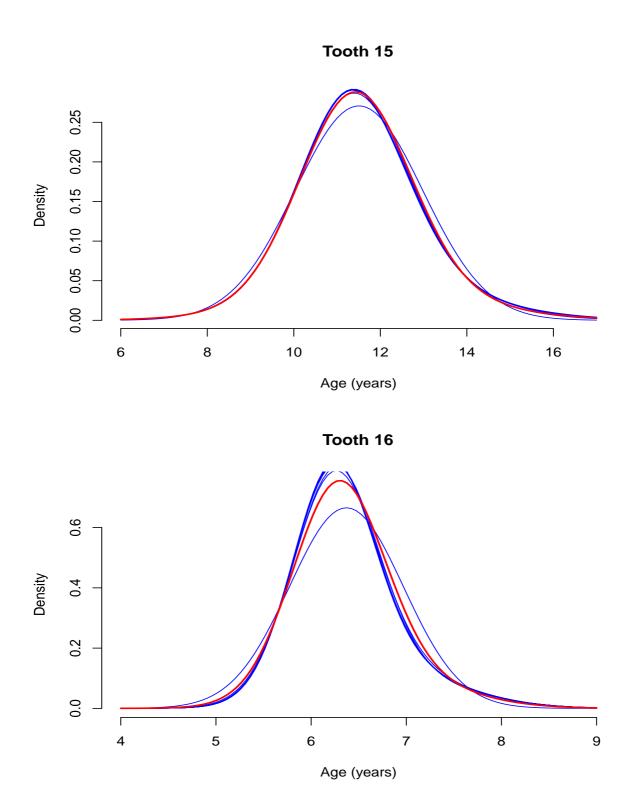


Figure 17: Teeth 15 and 16. Estimated predictive densities of emergence for different values of K ranging from 1 to 10 (blue lines) and for K = 2 (red line).

#### $R \Rightarrow$ Save results for future use:

```
> if (RUN.TIMECONSUMING.CODE){
+    save(list="Model0",
    file=paste(RESULTDIR, "/Tandmob-Model0", Kshow, ".RData", sep=""))
+
+    PDensBiModelK <- list()
+    PDensBiModelK[[Kshow]] <- PDensBiModel0
+    save(list="PDensBiModelK",
+        file=paste(RESULT2DIR, "/Tandmob-PDensBiModelK0", Kshow, ".RData", sep=""))
+
+    save(list=c("ModelK", "PDensUniModelK", "PDensBiModelK"),
        file=paste(RESULT2DIR, "/Tandmob-Result.RData", sep=""))
+ }</pre>
```

## References

- EKSTRAND, K. R., CHRISTIANSEN, J., and CHRISTIANSEN, M. E. (2003). Time and duration of eruption of first and second permanent molars: a longitudinal investigation. *Community Dentistry and Oral Epidemiology*, **31**, 344–350.
- KOMÁREK, A. (2009). A new R package for Bayesian estimation of multivariate normal mixtures allowing for selection of the number of components and interval-censored data. *Computational Statistics and Data Analysis*, **53**, To appear.
- VANOBBERGEN, J., MARTENS, L., LESAFFRE, E., and DECLERCK, D. (2000). The Signal-Tandmobiel<sup>®</sup> project a longitudinal intervention health promotion study in Flanders (Belgium): baseline and first year results. *European Journal of Paediatric Dentistry*, **2**, 87–96.