

November 3, 2021

Tutorial for setting up alchemical free energy simulations using AMBER20 with AMBER-DD boost

1 Availability of the AMBER-DD boost package

The AMBER Drug Discovery Boost package consists of the current release version of AmberTools21, which can be downloaded directly from AmberMD.org, and the specialized version of AMBER20 (the pmemd package). The AMBER20 part of AMBER Drug Discovery Boost package is only available to AMBER20 license holders. Active Amber developers can access the package directly through the Amber development GitLab repository. For other AMBER20 license holders, the primary mode for accessing the AMBER Drug Discovery Boost package is through GitLab repository set up through the laboratory for Biomolecular Simulation Research (LBSR) at Rutgers, which mirrors the AMBER Drug Discovery Boost package in the Amber development GitLab repository.

A verified AMBER20 license holder will need a GitLab account in order to access the AMBER Drug Discovery Boost GitLab repository. A new GitLab account can easily be obtained at www.GitLab.com. The user first need to send the e-mail address/username associated with the GitLab account to: Abir Ganguly at abir.ganguly@rutgers.edu or Darrin York at Darrin.York@rutgers.edu in order to be added to the AMBER Drug Discovery Boost GitLab projects. *Note*: if you have created a new GitLab account through a social media account such as Google or Facebook, you will need to manually set up your GitLab password in order for git clone to work. Once added, the user will receive three separate notification emails confirming that the user has been added to the following three projects:

```
Laboratory for Biomolecular Simulation Research / AMBER Drug Discovery Boost Laboratory for Biomolecular Simulation Research / Alchemical_FE Laboratory for Biomolecular Simulation Research / FE-ToolKit Upon this confirmation the user will be able to check out the packages as follows:

With ssh-key setup in GitLab (recommended):
git clone git@gitlab.com:RutgersLBSR/amber-drug-discovery-boost.git
git clone git@gitlab.com:RutgersLBSR/Alchemical_fe.git
git clone git@gitlab.com:RutgersLBSR/FE-ToolKit.git

Without ssh-key setup in GitLab:
git clone https://gitlab.com/RutgersLBSR/amber-drug-discovery-boost.git
git clone https://gitlab.com/RutgersLBSR/Alchemical_fe.git
git clone https://gitlab.com/RutgersLBSR/FE-ToolKit.git
The Alchemical fe folder contains the following sub-folders:
```

- Documentation containing documentation specific to AMBER Drug Discovery Boost
- *Tutorials* containing tutorials for setting up alchemical free energy simulations using AMBER Drug Discovery Boost
- Examples containing test cases for relative binding free energy (RBFE) and relative solvation free energy (RSFE) calculations
- ullet bin containing scripts to help set up alchemical free energy simulations using Amber Drug Discovery Boost

We are also in the process of putting documentation up on the Wiki site that will be updated on a regular basis - https://gitlab.com/RutgersLBSR/alchemical_fe/-/wikis/Setup-AFE_AMBER_DD_BOOST

$2\quad \text{Purpose of } setup_fe$

The $setup_fe$ script is designed to help setup the directory structure and input files that are necessary for running alchemical free energy simulations with AMBER-DD Boost. The script accepts a simplified input file named 'input' that is described in detail later in the tutorial. Briefly, for a given system, such as a specific protein target, or a collection of small molecules, a) a list of desired transformations can be provided, b) key simulation settings can be specified, and c) initial configuration files (MD equilibrated parameter (parm) and coordinate (rst) files associated with the system and specified transformations) must be provided. $setup_fe$ can then be used to generate a hierarchy of directories containing relevant parameter, coordinate, and AMBER input files and job submission scripts. $setup_fe$ can subsequently be used to analyze the free energy simulations using FE-ToolKit

3 Initial requirements of setup_fe

Following conditions must be met before running $setup_fe$ -

- I AMBERHOME needs to be set
- II cpptraj(AmberTools) needs to be installed and available in \$PATH
- III parmed (AmberTools) needs to be installed and available in \$PATH
- IV python3 needs to be installed and available in \$PATH
- V bin subdirectory within Alchemical fe folder needs to be available in \$PATH
- VI RDKit needs to be installed and python3 bindings to RDKit needs to be available in PYTHONPATH
 - RDKit can be installed with a package manager
 - Fedora sudo dnf install rdkit.x86 64 python3-rdkit.x86 64
 - Ubuntu sudo apt install python-rdkit
 - or, RDKit can be installed using conda
 - See http://www.rdkit.org/docs/Install.html#installationfordetails

VII folder containing initial structure and parameter files

• For relative binding free energy (RBFE) calculations, this folder should contain, for each intended transformation (edge), PDB file(s) of the protein-ligand(s) complex(es) and parameter (mol2, lib, fremod) files for the associated ligands and any nonstandard residues present in the protein-ligand PDB file(s).

For relative and absolute solvation free energy (RSFE, ASFE) calculations, this folder should contain, for each intended transformation, parameter (mol2, lib, fremod) files for the associated ligand(s).

For further details related to input format please refer to Table 1

4 Setup of $setup_fe$

The setup_fe script can be created by running the script makesetup_fe.sh located in the bin subdirectory of the Alchemical_fe folder.

```
./makesetup fe.sh
```

 $make setup_fe.sh$ help set the following three variables: path TOWRKDIR, path TOWFToolKit, and path TOFE ToolKit within $setup_fe$.

- I \$pathTOWRKDIR should be set to the intended working directory.
- II \$pathTOWFToolKit should be set to the path of the Alchemical fe folder.
- III \$pathTOFEToolKit should be set to the path of the FE-ToolKit folder.

By defaults, these variables are set to the path of the *current working directory*. Alternatively, these variables can be modified manually in *setup* fe.

5 Input file for $setup_fe$

setup_fe requires an input file named input that contains key settings of the alchemical free energy simulations that are going to be set up. A template input file can be generated by running the script with a flag -h or -help. A typical input file looks like the following -

```
path\_to\_input=initial\\ system=CDK2\\ translist=(1h1q\ 1h1s\ 1h1q\ 1oi9\ 1oi9\ 1h1s)\\ nlambda=11\\ protocol=unified\\ mapmethod=2\\ mapinspect=0\\ mapnetwork=true\\ boxbuild=1\\ boxbufcom=16\\ boxbufaq=20\\ ionconc=0.15
```

```
pff=ff14SB
lff=gaff2
wm = tip4pew
mdboxshape = cubic
ntrials = 3
cutoff=10
repex=true
nstlimti = 5000
numexchqti=1000
hmr=false
notrajectory=true
scalpha = 0.5
scbeta = 1.0
gti add sc=5
gti scale beta=1
gti cut=1
gti\_cut\_sc\_on=8
gti cut sc off=10
gti lam sch=1
gti ele sc=1
gti\_vdw\_sc=1
gti\_cut\_sc=2
gti\_ele\_exp=2
gti vdw exp=2
two state = false
bidirection aq=true
bidirection com=true
ticalc = rbfe
stage = analysis
setupmode = 0
partition = general - long - gpu
nnodes=1
nqpus=8
wallclock = 3-00:00:00
path to data=data
exptdata file = skip
bar = true
ccc = false
ccc ddG=true
start = 0.0
stop=100.0
check convergence=true
show all cycles = true
```

Table 1 provides a detailed description of keywords that are specific to $setup_fe$. For keywords/flags that are specific to AMBER20 and AMBER-DD Boost refer to the AMBER20 reference manual and AMBER-DD Boost Documentation.

6 File infrastructure created by setup fe

With the input file present in \$pathTOWRKDIR, $setup_fe$ can be executed as ./setup_fe

In "setup" mode, setup_fe creates a folder named "system", as defined i in the input file in \$pathTOWRKDIR. The "system" folder will have two subdirectories, setup and run. The folder setup will house the various intermediate files that were generated and used in creating the final input files. The folder run will contain independent subdirectories corresponding to each entry (transformation) in the keyword "translist" in the input file. These subdirectories that will have the same naming convention as provided in the input file, will further contain subdirectories named "com" and "aq" for RBFE calculations, or only the subdirectory "aq" for RSFE and ASFE calculations. The "com" and "aq" subdirectories correspond to complex and aqueous simulations, respectively, and will contain the final merged TI parameter and coordinate input files, template submission slurm scripts, an input folder containing relevant AMBER input files, and production sub-folders corresponding to each specified independent trial that will house the production simulation data.

In "analysis" mode, setup_fe creates a folder named results in \$pathTOWRKDIR. The results folder will contain a subdirectory data that will have a nested directory structure containing Energy and DV/DL data from the various simulations in the transformation network being analyzed. The results folder will also contain the graphmbar input file named graphmbar.inp for FE-Toolkit, a python script named gmbar.py that facilitates the generation of the graphmbar input file, the graphmbar output file named graphmbar.out generated by FE-Toolkit, and a simplified output file named out summarizing the final free energy results of the entire transformation network.

Table 1. Keywords associated with the ProFESSA workflow

Keyword	Value	Description	Example
path_to_input	string	Path to directory that contains input files. It should contain a subdirectory	path_to_input
		system.	=/home/username/afe/initial
system	string	Name of the system. A folder named system should be present in	system=CDK2
		path_to_input/, and should contain the initial structure and parameter files.	
translist	list	A list of desired transformations or edges. In the case of RBFE or RSFE	translist=(1h1q~1h1r
	of	calculations, translist should be a list in which each entry consists of two mol-	1h1q~1h1s)
	strings	names separated by the character "~", while in the case of ASFE calculations,	
		translist should be a list of molnames. Initial structure/parameter files of these	
		molnames should be provided in $path_to_input/system$ and should be named	
		as follows:	
		For RBFE calculations, each molname present in <i>translist</i> should have an asso-	
		ciated molname.pdb, representing the receptor-ligand complex structure and	
		[molname_0.mol2 molname_0.lib molname_0.frcmod], representing the lig-	
		and parameters. Parameters of additional non-standard residues, if present,	
		can be provided as [molname_1.mol2 molname_1.lib molname_1.frcmod],	
		$[molname_2.mol2\ molname_2.lib\ molname_2.frcmod]$, etc.	
		For RSFE and ASFE calculations, each molname present in <i>translist</i> should	
		have associated [molname_0.mol2 molname_0.lib molname_0.frcmod] files.	
ticalc	string	Specifies nature of calculation. Acceptable values - rbfe, rsfe, asfe.	ticalc=rbfe
nlambda	integer	Number of lambda windows in TI calculation. Acceptable values - positive	nlambda=21
		integers.	
protocol	string	Protocol for running TI simulations. Acceptable value - unified.	protocol=unified
mapmethod	integer	Specifies the algorithm using which TS/TC regions are going to be determined.	mapmethod=1
		Acceptable values - 0, 1, 2.	
		θ specifies MCS algorithm.	
		1 specifies MCS-E algorithm.	
		2 specifies MCS-E2 algorithm.	
		For a given transformation/edge molname1~molname2 in translist, an atom	
		map file, molname1~molname2.map.txt, is generated in the folder sys-	
		tem/setup.	
mapinspect	integer	Allows manual inspection of TS/TC regions. Acceptable values - 0, 1, 2.	mapinspect=true
		θ specifies no manual inspection.	
		1 specifies manual inspection.	
		2 specifies resume setup assuming manual inspection has been completed.	
		If mapinspect is set to 2, the necessary atom map files should be present in	
		system/setup folder.	
mapnetwork	string	Specifies if network-wide consistent TS/TC regions of ligands will be gener-	mapnetwork=false
		ated. Acceptable values - true, false.	
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Keyword	Value	Description	Example
boxbuild	string	Specifies if and how MD boxes will be built. Acceptable values - 0, 1, 2, skip.	boxbuild=1
	/	θ specifies to build boxes only for "aqueous" state and not for "complex" state.	
	integer	1 specifies to build boxes for both "aqueous" and "complex" states.	
		2 specifies to build boxes for both "aqueous" and "complex" states with iden-	
		tical number of water and ions.	
		skip specifies to skip box building altogether.	
		For RSFE and ASFE calculations, boxbuild θ and 1 are identical.	
boxbufcom	integer	Specifies MD box buffer for "complex" states. Relevant only for RBFE calcu-	boxbufcom=16
		lations. Acceptable values - positive integers.	
boxbufaq	integer	Specifies MD box buffer for "aqueous" states. Relevant only for RBFE calcu-	boxbufaq=21
		lations. Acceptable values - positive integers.	
ionconc	float	Specifies the MD box ion concentration in units of mol/L (M). Acceptable	ionconc=0.15
		values - positive real number.	
pff	string	Specifies protein forcefield. Acceptable values - ff14SB	pff=ff14SB
lff	string	Specifies ligand forcefield. Acceptable values - gaff, gaff2	lff=gaff2
wm	string	Specifies water model. Acceptable values - tip4pew, tip3p	wm=tip4pew
mdboxshape	string	Specifies shape of MD box. Acceptable values - cubic	mdboxshape=cubic
ntrials	integer	Specifies the number of independent trials of calculation. Acceptable values -	ntrials=10
		positive integers.	
cutoff	integer	Specifies non-bonded cutoff in TI simulations. Acceptable values - positive	cutoff=10
		integers.	
repex	string	Specifies if Hamiltonian Replica Exchange will be employed.	repex=true
nstlimti	integer	Specifies the length of production TI simulations in units of fs. Acceptable	nstlimti=5000
		values - positive integers.	
numexchgti	integer	Specifies the number of exchanges in replica exchange TI simulations. numex-	numexchgti=1000
<u> </u>		<i>chgti</i> is ignored is <i>repex</i> is set to false. Acceptable values - positive integers.	
hmr	string	Specifies if Hydrogen Mass Repartitioning will be used. Acceptable values -	hmr=false
		true, false.	
notrajectory	string	Specifies if production trajectories will be saved during TI simulations. Ac-	notrajectory=true
J J		ceptable values - true, false.	
scalpha	float	Specifies the value of AMBER DD BOOST keyword scalpha in TI simulations.	scalpha=0.5
ocarpiia	11000	Acceptable values - positive real numbers.	Secipita 0.0
scbeta	float	Specifies the value of AMBER DD BOOST keyword scheta in TI simulations.	scbeta=0.5
000000	11000	Acceptable values - positive real numbers.	555000 010
gti add sc	integer	Specifies the value of $AMBER\ DD\ BOOST$ keyword $gti\ add\ sc$ in TI simu-	gti add sc=5
501_ddd_50	meeger	lations. Acceptable values - positive integers.	gu_udd_sc o
gti_scale_beta	float	Specifies the value of AMBER DD BOOST keyword gti scale beta in TI	gti scale beta=1
	11000	simulations. Acceptable values - positive real number.	801_20000_20000 1
gti cut	integer	Specifies the value of $AMBER\ DD\ BOOST$ keyword gti_cut in TI simulations.	gti cut=1
501_Cut	Integer	Acceptable values - positive integers.	801_000—1
gti_cut_sc_on	integer	Specifies the value of AMBER DD BOOST keyword gti cut sc on in TI	gti_cut_sc_on=8
5 ¹¹ _Cut_5C_0H	mtegel	simulations. Acceptable values - positive integers.	801_000_50_011—0
	1	bilingrations, Acceptable values = positive littegers,	1

Table 1 – continued from previous page

Keyword	Value	Description	Example
gti_cut_sc_off	integer	Specifies the value of AMBER DD BOOST keyword gti cut sc off in TI	gti cut sc off=10
gti_cut_sc_on	Integer	simulations. Acceptable values - positive integers.	gti_cut_sc_oii=10
gti lam sch	integer	Specifies the value of AMBER DD BOOST keyword gti lam sch in TI sim-	gti lam sch=1
gti_iaiii_scii	Integer	ulations. Acceptable values - positive integers.	gti_iaiii_scii—i
gti_ele_sc	integer	Specifies the value of AMBER DD BOOST keyword gti_ele_sc in TI simula-	gti ele sc=1
gti_ele_sc	Integer	tions. Acceptable values - positive integers.	gti_eie_sc=1
mt:d aa	intomon	Specifies the value of AMBER DD BOOST keyword gti vdw sc in TI simu-	gti vdw sc=1
gti_vdw_sc	integer	lations. Acceptable values - positive integers.	gti_vdw_sc=1
mti out ao	intomon	Specifies the value of AMBER DD BOOST keyword gti cut sc in TI simu-	gti cut sc=1
gti_cut_sc	integer		gti_cut_sc_1
		lations. Acceptable values - positive integers.	
gti_ele_exp	integer	Specifies the value of AMBER DD BOOST keyword gti_ele_exp in TI simulations. Accordable advantable and acciding interests	gti_ele_exp=2
1	. ,	lations. Acceptable values - positive integers.	4: 1 0
gti_vdw_exp	integer	Specifies the value of AMBER DD BOOST keyword gti_vdw_exp in TI sim-	gti_vdw_exp=2
		ulations. Acceptable values - positive integers.	
twostate	string	Specifies if two state setup will be employed in TI simulations. Acceptable	twostate=true
1.11		values - true, false.	
bidirection_aq	string	Specifies if bidirectional setup will be used for "aqueous" state TI simulations.	bidirection_aq=false
		Applicable when two state is set to false. Acceptable values - true, false.	
bidirection_com	string	Specifies if bidirectional setup will be used for "complex" state TI simulations.	bidirection_aq=false
		Applicable when two state is set to false. Acceptable values - true, false.	
stage	string	Specifies the action of the script. Acceptable values - setup, analysis.	stage=setup
		setup specifies script to set up TI simulations.	
		analysis specifies script to perform analysis.	
setupmode	integer	Specifies the mode of simulation setup. Acceptable values - θ	setupmode=0
		0 sets up regular TI simulations.	
partition	string	Specifies the HPC partition on which TI runs will be performed. Acceptable	partition=gpu
		values - null, name of HPC partition.	
nnodes	integer	Specifies the number of nodes to be requested for a single set of TI simulations.	nnodes=1
		Acceptable values - positive integer.	
ngpus	integer	Specifies the number of gpus per node to be requested for a single set of TI	ngpus=8
		simulations. Acceptable values - positive integer.	
wallclock	string	Specifies the wallclock on TI jobs. Acceptable values - formatted time in	wallclock=3-00:00:00
		hours:minutes:days.	
path to data	string	Specifies the path to production runs. Default path is set to sys-	path to data=CDK2/unified/run
		tem/protocol/run	, ,
exptdatafile	string	Specifies the name of a text file containing experimental ligand binding free	exptdatafile=Expt.dat
±		energies. Acceptable values - skip, filename.	
		The text file should have two columns corresponding to molname (column 1)	
		and relative ligand binding free energy (column 2).	
	1		
bar	string	Specifies if BAR is going to be used for analysis instead of MBAR. Acceptable	bar=false

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Table 1 – continued from previous page

Keyword	Value	Description	Example
ccc	string	Specifies if cycle closure corrections are to be applied during analysis. Accept-	ccc=true
		able values - true, false.	
ccc_ddG	string	Specifies if cycle closures will be applied to "complex" - "aqueous" delta delta	$ccc_ddG=true$
		Gs instead of "complex" and "aqueous" delta Gs, independently. Acceptable	
		values - true, false.	
start	float	Specifies the percentage of data to ignore from the beginning of TI production	start=20.0
		runs. Acceptable values - float numbers ranging from 0 to 100, and less than	
		stop.	
stop	float	Specifies the percentage of data to read from the start of TI production runs.	stop=100.00
		Acceptable values - float numbers ranging from 0 to 100, and greater than	
		start.	
check_convergence	string	Specifies if check of data convergence will be carried out during analysis. Ac-	check_convergence=true
		ceptable values - true, false.	
		If check_convergence is set to true, the analysis is carried out multiple times,	
		for a range of <i>start</i> and <i>stop</i> values.	
showallcycles	string	Specifies if the output should show information on all possible cycles within	showallcycles=true
		the given transformation network. Acceptable values - true, false.	