

Building a gene annotation object

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Introduction

In this tutorial, we describe the process for making and using rich gene annotation objects to be used throughout the `crisprVerse` ecosystem. Such objects enable users to retrieve coordinates of transcripts, exons, etc. Those objects are also used by several functions in the `crisprDesign` package to add gene annotations to both gRNA on-targets and off-targets. This is what the `txObject` argument in many of the functions expect.

We will also describe the process for constructing and using a transcription start site (TSS) annotation object (`tssObject` argument in many of the functions).

Installation

See the Installation tutorial to learn how to install the packages `crisprDesign` and `crisprDesignData` required in this tutorial.

Getting started

The packages can be loaded into an R session in the usual way:

```
library(crisprDesign)
library(crisprDesignData)
```

Building gene annotation objects

In the `crisprVerse`, we represent gene annotations using `GRangesList` object, and this can be easily constructed using the commonly-used Bioconductor objects `TxDb` (see the `GenomicFeatures` package to learn more about `TxDb` objects). We will now show several ways of constructing such objects.

Building a `GRangesList` from Ensembl

We construct a gene annotation object for the human genome using the Ensembl release 104 (hg38). This can be done using the function `getTxDb` in `crisprDesign`:

```
txdb <- getTxDb(organism="Homo sapiens", release=104)
```

This may take several minutes, and note that this requires an internet connection. In case it times out, one can increase the timeout option using the following:

```
options(timeout = max(1000000, getOption("timeout")))
```

Once obtained, we can convert the object into a `GRangesList` using the function `TxDb2GRangesList` from `crisprDesign`:

```
grList <- TxDb2GRangesList(txdb)
```

We will specify that the genome is hg38:

```
GenomeInfoDb::genome(grList) <- "hg38"
```

And that's it! The `grList` object contains all of the information about the Ensembl release 104 gene model, and is ready to be used in the `crisprVerse`. Let's take a quick look at our gene annotation object:

```
names(grList)
```

```
## [1] "transcripts" "exons"          "cds"            "fiveUTRs"      "threeUTRs"
## [6] "introns"      "tss"
```

```
grList$transcripts
```

```
## GRanges object with 111751 ranges and 14 metadata columns:
```

```
##      seqnames      ranges strand |      tx_id      gene_id
##      <Rle>      <IRanges> <Rle> |      <character>      <character>
##          1 11869-14409      + | ENST00000456328 ENSG00000223972
##          1 12010-13670      + | ENST00000450305 ENSG00000223972
##          1 29554-31097      + | ENST00000473358 ENSG00000243485
##          1 30267-31109      + | ENST00000469289 ENSG00000243485
##          1 30366-30503      + | ENST00000607096 ENSG00000284332
##      .      ...      ...      .      ...      ...
##      MT      5826-5891      - | ENST00000387409 ENSG00000210144
##      MT      7446-7514      - | ENST00000387416 ENSG00000210151
##      MT 14149-14673      - | ENST00000361681 ENSG00000198695
##      MT 14674-14742      - | ENST00000387459 ENSG00000210194
##      MT 15956-16023      - | ENST00000387461 ENSG00000210196
##      protein_id      tx_type gene_symbol      exon_id exon_rank
##      <character>      <character> <character> <character> <integer>
##          <NA>      processed_transcript      DDX11L1      <NA>      <NA>
##          <NA>      transcribed_unproces..      DDX11L1      <NA>      <NA>
##          <NA>      lncRNA      MIR1302-2HG      <NA>      <NA>
##          <NA>      lncRNA      MIR1302-2HG      <NA>      <NA>
##          <NA>      miRNA      MIR1302-2      <NA>      <NA>
##      .      ...      ...      ...      ...      ...
##          <NA>      Mt_tRNA      MT-TY      <NA>      <NA>
##          <NA>      Mt_tRNA      MT-TS1      <NA>      <NA>
##      ENSP00000354665      protein_coding      MT-ND6      <NA>      <NA>
##          <NA>      Mt_tRNA      MT-TE      <NA>      <NA>
##          <NA>      Mt_tRNA      MT-TP      <NA>      <NA>
##      cds_start      cds_end tx_start      tx_end      cds_len exon_start exon_end
##      <integer> <integer> <integer> <integer> <integer> <integer> <integer>
##          <NA>      <NA>      <NA>      <NA>      <NA>      <NA>      <NA>
##          <NA>      <NA>      <NA>      <NA>      <NA>      <NA>      <NA>
##          <NA>      <NA>      <NA>      <NA>      <NA>      <NA>      <NA>
##          <NA>      <NA>      <NA>      <NA>      <NA>      <NA>      <NA>
##          <NA>      <NA>      <NA>      <NA>      <NA>      <NA>      <NA>
##      .      ...      ...      ...      ...      ...      ...
##          <NA>      <NA>      <NA>      <NA>      <NA>      <NA>      <NA>
##          <NA>      <NA>      <NA>      <NA>      <NA>      <NA>      <NA>
##          <NA>      <NA>      <NA>      <NA>      <NA>      <NA>      <NA>
##          <NA>      <NA>      <NA>      <NA>      <NA>      <NA>      <NA>
##          <NA>      <NA>      <NA>      <NA>      <NA>      <NA>      <NA>
```

```
## -----
## seqinfo: 25 sequences (1 circular) from hg38 genome
```

Building a tssObject

Building a TSS annotation object requires only one additional step after constructing the `GRangesList` object described above. This can be obtained using the function `getTssObjectFromTxObject` in `crisprDesign`:

```
tssObject <- getTssObjectFromTxObject(grList)
tssObject
```

```
## GRanges object with 52547 ranges and 5 metadata columns:
##          seqnames      ranges strand |          tx_id          gene_id
##          <Rle> <IRanges> <Rle> |    <character>    <character>
##    11402         1      65419      + | ENST00000641515 ENSG00000186092
##    11442         1     923923      + | ENST00000616016 ENSG00000187634
##    11444         1     925731      + | ENST00000342066 ENSG00000187634
##    11445         1     960584      + | ENST00000338591 ENSG00000187961
##    11446         1     960639      + | ENST00000622660 ENSG00000187961
##      ...      ...      ...      .   ...      ...
##   123058         Y    24047689      - | ENST00000382407 ENSG00000172352
##   123073         Y    24813186      - | ENST00000382365 ENSG00000187191
##   123074         Y    24813186      - | ENST00000315357 ENSG00000187191
##   123075         Y    24813186      - | ENST00000446723 ENSG00000187191
##   123080         Y    25052074      - | ENST00000382287 ENSG00000185894
##          gene_symbol      promoter      ID
##          <character>    <character>    <character>
##    11402         OR4F5 ENST00000641515 ENSG00000186092_ENST..
##    11442         SAMD11 ENST00000616016 ENSG00000187634_ENST..
##    11444         SAMD11 ENST00000342066 ENSG00000187634_ENST..
##    11445         KLHL17 ENST00000338591 ENSG00000187961_ENST..
##    11446         <NA> ENST00000622660 ENSG00000187961_ENST..
##      ...      ...      ...
##   123058         CDY1B ENST00000382407 ENSG00000172352_ENST..
##   123073         DAZ3 ENST00000382365 ENSG00000187191_ENST..
##   123074         DAZ3 ENST00000315357 ENSG00000187191_ENST..
##   123075         DAZ3 ENST00000446723 ENSG00000187191_ENST..
##   123080         BPY2C ENST00000382287 ENSG00000185894_ENST..
## -----
## seqinfo: 25 sequences (1 circular) from hg38 genome
```

Using gene annotation objects

The gene (or TSS) annotation objects described above are often necessary for the full characterization of CRISPR gRNAs as they are inputs for several of the `crisprDesign` functions, including `queryTxObject`, `queryTssObject`, `addGeneAnnotation`, `addTssAnnotation`, and `addSpacerAlignments`.

For convenience, we provide in the `crisprDesignData` package precomputed gene annotation for human and mouse:

Object name	Object class	Version	Description
txdb_human	GRangesList	Release 104	Ensembl gene model for human (hg38/GRCh38)
txdb_mouse	GRangesList	Release 102	Ensembl gene model for mouse (mm10/GRCm38)

Object name	Object class	Version	Description
tss_human	GRanges	Release 104	Ensembl-based TSS coordinates for human (hg38/GRCh38)
tss_mouse	GRanges	Release 102	Ensembl-based TSS coordinates for human (mm10/GRCm38)

Building a gene annotation object from a GFF file

If you have a General Feature Format (GFF) file from which you want to construct the gene annotation object, you can pass this to the `file` argument of the `crisprDesign` function `getTxDb`; this will create the TxDb object using the `GenomicFeatures` function `makeTxDbFromGFF`.

Reproducibility

```
sessionInfo()
```

```
## R version 4.2.1 (2022-06-23)
## Platform: x86_64-apple-darwin17.0 (64-bit)
## Running under: macOS Catalina 10.15.7
##
## Matrix products: default
## BLAS: /Library/Frameworks/R.framework/Versions/4.2/Resources/lib/libRblas.0.dylib
## LAPACK: /Library/Frameworks/R.framework/Versions/4.2/Resources/lib/libRlapack.dylib
##
## locale:
## [1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
##
## attached base packages:
## [1] stats4      stats      graphics  grDevices  utils      datasets  methods
## [8] base
##
## other attached packages:
## [1] BSgenome.Hsapiens.UCSC.hg38_1.4.4 BSgenome_1.65.2
## [3] rtracklayer_1.57.0                  Biostrings_2.65.2
## [5] XVector_0.37.0                      GenomicRanges_1.49.1
## [7] GenomeInfoDb_1.33.5                 IRanges_2.31.2
## [9] S4Vectors_0.35.1                   crisprDesignData_0.99.17
## [11] crisprDesign_0.99.133               crisprScore_1.1.14
## [13] crisprScoreData_1.1.3               ExperimentHub_2.5.0
## [15] AnnotationHub_3.5.0                 BiocFileCache_2.5.0
## [17] dbplyr_2.2.1                       BiocGenerics_0.43.1
## [19] crisprBowtie_1.1.1                  crisprBase_1.1.5
## [21] crisprVerse_0.99.8                  rmarkdown_2.15.2
##
## loaded via a namespace (and not attached):
## [1] rjson_0.2.21                        ellipsis_0.3.2
## [3] Rbowtie_1.37.0                      bit64_4.0.5
## [5] lubridate_1.8.0                     interactiveDisplayBase_1.35.0
## [7] AnnotationDbi_1.59.1                fansi_1.0.3
## [9] xml2_1.3.3                          codetools_0.2-18
```

## [11] cachem_1.0.6	knitr_1.40
## [13] jsonlite_1.8.0	Rsamtools_2.13.4
## [15] png_0.1-7	shiny_1.7.2
## [17] BiocManager_1.30.18	readr_2.1.2
## [19] compiler_4.2.1	httr_1.4.4
## [21] basilisk_1.9.2	assertthat_0.2.1
## [23] Matrix_1.4-1	fastmap_1.1.0
## [25] cli_3.3.0	later_1.3.0
## [27] htmltools_0.5.3	prettyunits_1.1.1
## [29] tools_4.2.1	glue_1.6.2
## [31] GenomeInfoDbData_1.2.8	dplyr_1.0.9
## [33] rappdirs_0.3.3	tinytex_0.41
## [35] Rcpp_1.0.9	Biobase_2.57.1
## [37] vctrs_0.4.1	crisprBwa_1.1.3
## [39] xfun_0.32	stringr_1.4.1
## [41] mime_0.12	lifecycle_1.0.1
## [43] restfulr_0.0.15	XML_3.99-0.10
## [45] zlibbioc_1.43.0	basilisk.utils_1.9.1
## [47] vroom_1.5.7	VariantAnnotation_1.43.3
## [49] hms_1.1.2	promises_1.2.0.1
## [51] MatrixGenerics_1.9.1	parallel_4.2.1
## [53] SummarizedExperiment_1.27.1	RMariaDB_1.2.2
## [55] yaml_2.3.5	curl_4.3.2
## [57] memoise_2.0.1	reticulate_1.25
## [59] biomaRt_2.53.2	stringi_1.7.8
## [61] RSQLite_2.2.16	BiocVersion_3.16.0
## [63] highr_0.9	BiocIO_1.7.1
## [65] randomForest_4.7-1.1	GenomicFeatures_1.49.6
## [67] filelock_1.0.2	BiocParallel_1.31.12
## [69] rlang_1.0.4	pkgconfig_2.0.3
## [71] matrixStats_0.62.0	bitops_1.0-7
## [73] evaluate_0.16	lattice_0.20-45
## [75] purrr_0.3.4	GenomicAlignments_1.33.1
## [77] bit_4.0.4	tidyselect_1.1.2
## [79] magrittr_2.0.3	R6_2.5.1
## [81] generics_0.1.3	DelayedArray_0.23.1
## [83] DBI_1.1.3	pillar_1.8.1
## [85] KEGGREST_1.37.3	RCurl_1.98-1.8
## [87] tibble_3.1.8	dir.expiry_1.5.0
## [89] crayon_1.5.1	utf8_1.2.2
## [91] tzdb_0.3.0	progress_1.2.2
## [93] grid_4.2.1	blob_1.2.3
## [95] digest_0.6.29	xtable_1.8-4
## [97] httpuv_1.6.5	Rbwa_1.1.0