# Systembiologie 551-1174-00L

# **Beyond Metabolism**

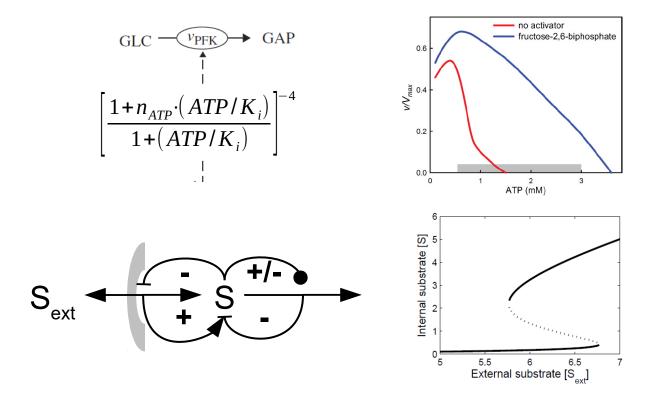
6 April, 2017
Uwe Sauer, Molecular Systems Biology
Jörg Stelling, D-BSSE

#### **Content:**

- Signaling pathways networks (US)
- Dynamic models for cell signaling (JS)
  - Principles: Src kinase
  - Application in drug discovery
- Exams (US/JS)



#### **Revisiting Dynamics: Substrate Inhibition**



- Single-enzyme regulation may appear 'meaningless' when considered without the network context.
- Wiring and dynamics are critical (negative feedback for homeostasis vs. positive feedback for dynamic memory).

#### **Example (Paradigm) for Signaling: Src Kinase**

A SARCOMA OF THE FOWL TRANSMISSIBLE BY AN AGENT SEPARABLE FROM THE TUMOR CELLS.\*

BY PEYTON ROUS, M.D.

(From the Laboratories of the Rockefeller Institute for Medical Research, New York.)



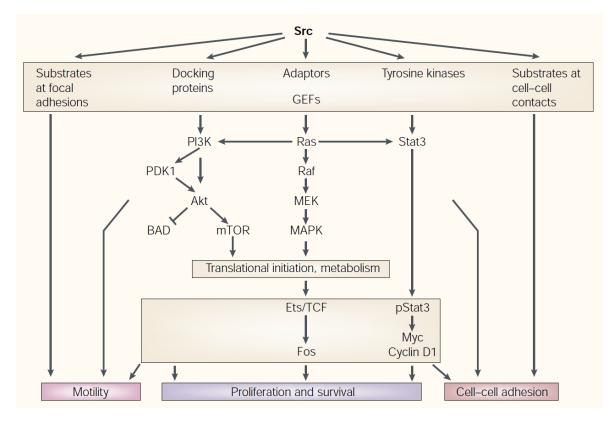
- First demonstration of virus-induced cancer.
- □ First oncogene (cellular Src protein).
- □ First tyrosine kinase (non-receptor associated).

<sup>\*</sup> Received for publication, February 9, 1911.

<sup>&</sup>lt;sup>1</sup> Peyton Rous, Jour. Exper. Med., 1910, xii, 696.

<sup>&</sup>lt;sup>2</sup> Peyton Rous, Jour. Am. Med. Assn., 1910, lv, 1805; 1911, lvi, 198.

### Src Kinase: A Key Regulator of Cell Signaling

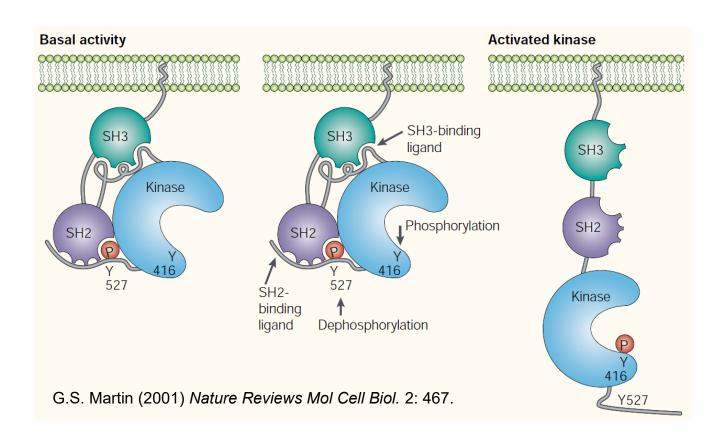


G.S. Martin (2001) Nature Reviews Mol Cell Biol. 2: 467.

 □ Src = Phosphotyrosine kinase with multiple functions in cell signaling (proteomics: ~350 potential Src substrates).

How can one protein integrate many signals?

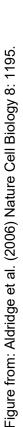
#### **Src Kinase: Mechanism of Activation**

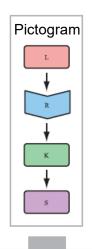


 Regulation by two key phosphorylation events (Y527: intramolecular inhibition; Y416: kinase domain activation).

How can we model Src activation dynamics?

### Models for Cell Signaling: Same Concepts





#### Reaction list

$$R+L \rightleftharpoons LR^*$$

$$k_{r_1}$$

$$LR^*+K \rightleftharpoons LR^*K$$

$$LR^*K \rightarrow LR^*+K^*$$

$$LR^*K \rightarrow LR^*+K^*$$

$$K^*+S \rightleftharpoons K^*S$$

$$K^*S \rightarrow K^*+S^*$$

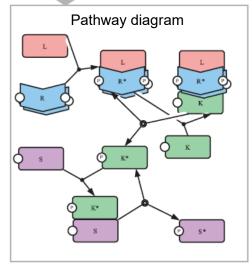
#### **Approximations**

if 
$$[S]_0 >> [K^*]_0$$
:  $\frac{d[K^*S]}{dt} \approx 0$ 

$$\frac{d[K^*]}{dt} = k_{13}[LR^*K] + k_{15}[K^*S]$$

$$-\frac{k_{15}[K^*]_0[S]}{\left(\frac{k_{14} + k_{15}}{k_{14}}\right) + [S]}$$

$$\frac{d[S^*]}{dt} = \frac{k_{15}[K^*]_0[S]}{\left(\frac{k_{14} + k_{15}}{k_{14}}\right) + [S]}$$



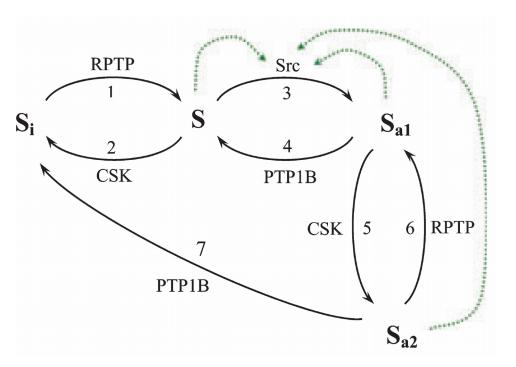
#### **Mass-action kinetics**

$$\begin{split} &\frac{d[R]}{dt} = -k_{r1}[L][R] + k_{r1}[LR^*] \\ &\frac{d[LR^*]}{dt} = k_{r1}[L][R] - k_{r1}[LR^*] - k_{r2}[LR^*][K] \\ &+ k_{r2}[LR^*K] + k_{r3}[LR^*K] \\ &\frac{d[LR^*K]}{dt} = k_{r2}[LR^*][K] - k_{r2}[LR^*K] - k_{r3}[LR^*K] \\ &\frac{d[K]}{dt} = -k_{r2}[LR^*][K] + k_{r2}[LR^*K] \\ &\frac{d[K^*]}{dt} = k_{r3}[LR^*K] - k_{r4}[K^*][S] + k_{r4}[K^*S] \\ &\frac{d[S]}{dt} = k_{r4}[K^*][S] + k_{r4}[K^*S] \\ &\frac{d[S]}{dt} = k_{r4}[K^*][S] - k_{r4}[K^*S] - k_{r5}[K^*S] \\ &\frac{d[S^*]}{dt} = k_{r5}[K^*S] \\ &\frac{d[S^*]}{dt} = k_{r5}[K^*S] \end{split}$$

$$\frac{d \mathbf{x}(t)}{dt} = f(\mathbf{x}(t), \mathbf{u}(t), \mathbf{p})$$

#### **Example: Model for Src Kinase**

- □ S<sub>i</sub>: inhibited (pY527, Y416)
- □ S: partial act. (Y527, Y416)
- $\square$  S<sub>31</sub>: active (Y527, pY416)
- $\square$  S<sub>a</sub>: active (pY527, pY416)



N. Kaimachnikow & B. Kholodenko (2009) FEBS J. 276: 4102.

- Src model with four states, seven reactions, autoactivation.
- □ States are **not** independent → Not Michaelis-Menten!

## **Example: Model for Src Kinase**

- □ S<sub>i</sub>: inhibited (pY527, Y416)
- □ S: partial act. (Y527, Y416)
- $\square$  S<sub>a1</sub>: active (Y527, pY416)
- $\square$  S<sub>a2</sub>: active (pY527, pY416)

Autocatalytic network:

$$S+S \stackrel{k_S^f}{\rightleftharpoons} S \cdot S \xrightarrow{k_S^{cat}} S + S_{a1}$$

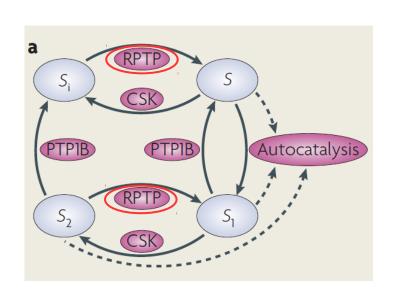
$$S_{a1} + S \stackrel{k_{a1}^f}{\rightleftharpoons} S_{a1} \cdot S \stackrel{k_{a1}^{cat}}{\longrightarrow} S_{a1} + S_{a1}$$

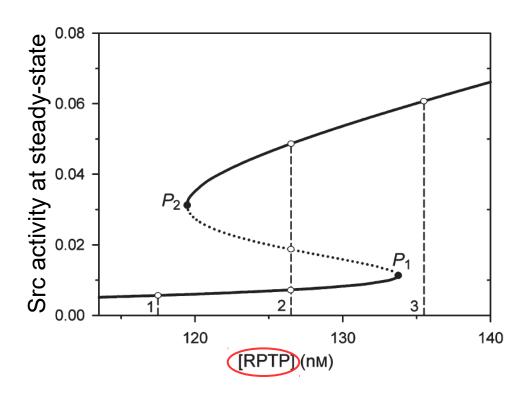
$$S_{a2} + S \stackrel{k_{a2}^f}{\rightleftharpoons} S_{a2} \cdot S \stackrel{k_{a2}^{cat}}{\longrightarrow} S_{a2} + S_{a1}$$

N. Kaimachnikow & B. Kholodenko (2009) FEBS J. 276: 4102.

- Src model with four states, using mass-action kinetics.
- □ States are **not** independent:  $v_3 = \left(\frac{k_S^{\text{cat}}}{K_S} \left[S\right] + \frac{k_{a1}^{\text{cat}}}{K_{a1}} \left[S_{a1}\right] + \frac{k_{a2}^{\text{cat}}}{K_{a2}} \left[S_{a2}\right]\right) \left[S\right]$

#### **Principle: Dynamic Memory**

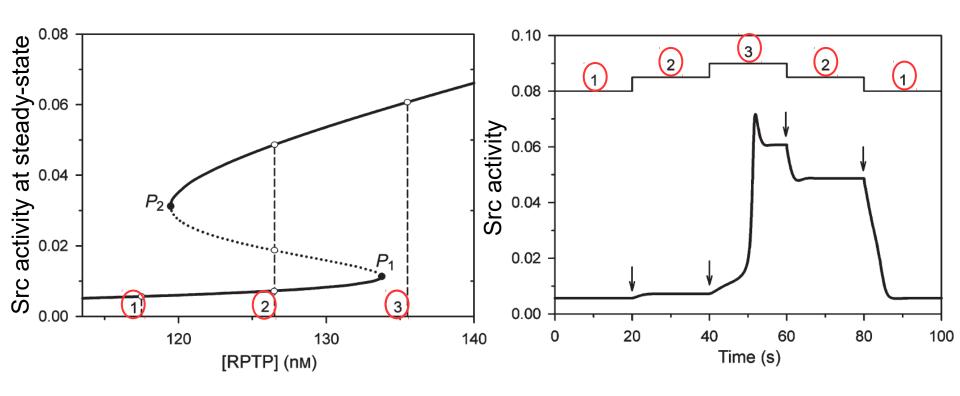




N. Kaimachnikow & B. Kholodenko (2009) FEBS J. 276: 4102.

□ Src autocatalytic activation implies positive feedback →
 Dynamic memory is possible (depending on context).

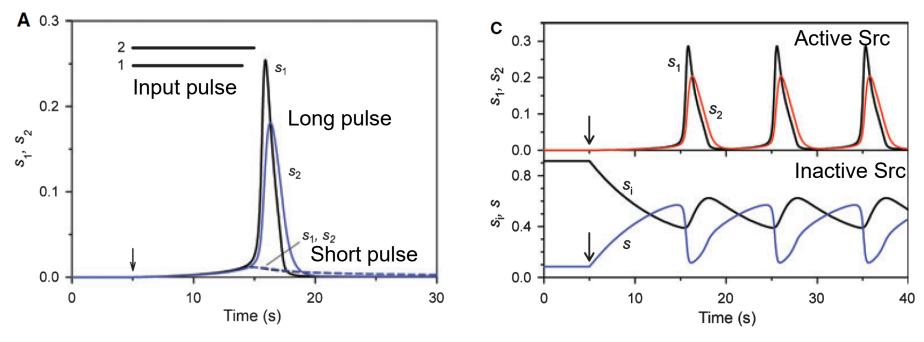
#### **Principle: Dynamic Memory**



N. Kaimachnikow & B. Kholodenko (2009) FEBS J. 276: 4102.

□ Src autocatalytic activation implies positive feedback →
 Dynamic memory is possible (depending on context).

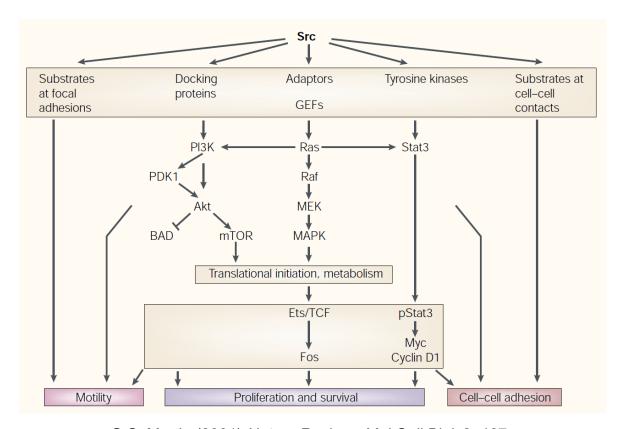
### Src Kinase: Versatile Dynamic Behaviors



N. Kaimachnikow & B. Kholodenko (2009) FEBS J. 276: 4102.

- □ Excitability (left): Src responds with pulse of activity to signals above (2) but not below (1) threshold duration.
- Oscillations (right): In the right context, kinase activity oscillates autonomously (for constant inputs).

#### **Principle: Integrated Cell Signaling**



G.S. Martin (2001) Nature Reviews Mol Cell Biol. 2: 467.

 Conclusion: Such pictures can be misleading because signals are (dynamically) separated in time (and space).

# Systembiologie 551-1174-00L

# **Beyond Metabolism**

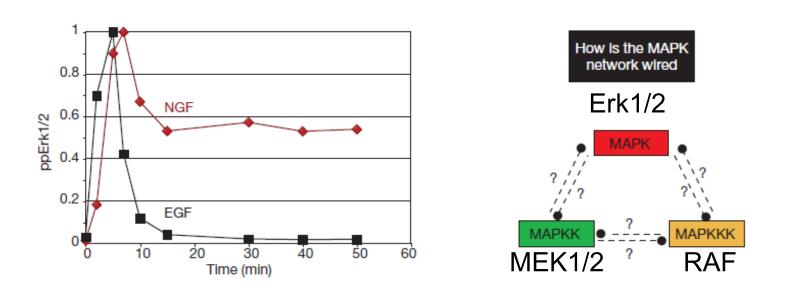
6 April, 2017
Uwe Sauer, Molecular Systems Biology
Jörg Stelling, D-BSSE

#### **Content:**

- Signaling pathways networks (US)
- Dynamic models for cell signaling (JS)
  - MAPK cascade signaling
  - Application in drug discovery
- Exams (US/JS)



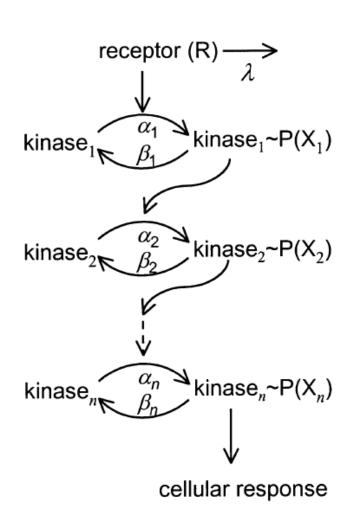
#### **Example: MAPK Signaling in Human Cells**



S. Santos et al. (2007) Nature Cell Biol. 9: 324-330.

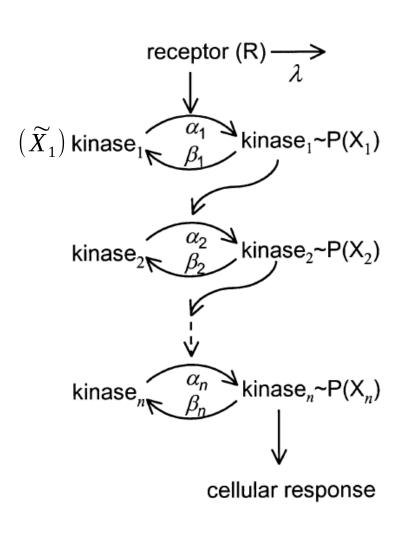
 Qualitatively different output of MAPK cascade signaling for different growth factor stimulations: NGF induces differentiation, EGF induces proliferation.

### MAPK Cascade Signaling: General Questions



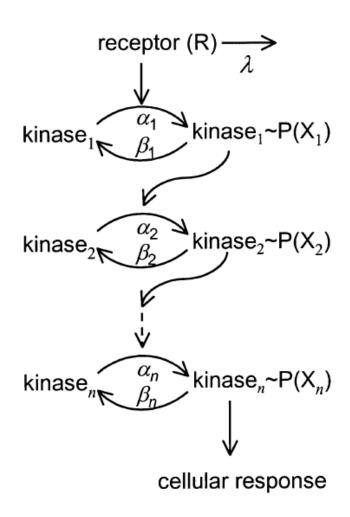
- How does a common
   pathway structure generate
   distinct dynamic behaviors
   (e.g., transient vs. sustained signaling)?
- Why do MAPK cascades have usually three levels?
- Which components influence signaling most?

### An Abstract Model of MAPK Cascade Signaling



- □ Kinase cascade with i=1...n levels (kinases).
- □ States are concentrations of:
  - Active, phosphorylated
     kinase X<sub>i</sub>
  - Inactive kinase  $\widetilde{X}_i$
- □ Parameters for kinase  $(\alpha_i)$  and phosphatase  $(\beta_i)$ .
- Input: receptor signal R(t).

### An Abstract Model of MAPK Cascade Signaling



 $\Box$  Kinase activity (i > 1):

$$\frac{dX_i}{dt} = \nu_{p,i} - \nu_{d,i} = \tilde{\alpha}_i X_{i-1} \tilde{X}_i - \beta_i X_i$$

 $\Box$  With total concentration  $C_i$ :

$$\frac{dX_i}{dt} = \alpha_i X_{i-1} \left( 1 - \frac{X_i}{C_i} \right) - \beta_i X_i$$

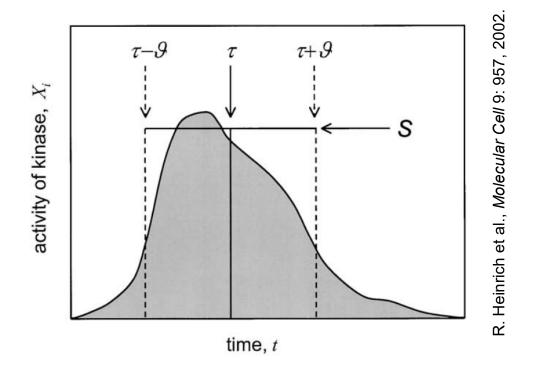
 $\square$  Activation by  $R(t) = \exp(-\lambda t)$ :

$$\frac{dX_1}{dt} = \alpha_1 R(t) \left( 1 - \frac{X_1}{C_1} \right) - \beta_1 X_1$$

Figures from: R. Heinrich et al., *Molecular Cell* 9: 957, 2002.

How to quantify signaling performance?

#### Signaling Cascades: Performance Criteria



- $\square$  Signaling time  $\tau$ : Expected time of arrival of the signal.
- $\square$  Signal duration v: Variance of the expected time.
- Signal amplitude S: Average amplitude during signaling.

#### **Signaling Cascades: Weak Activation**

 $\square$  Assume 'weak activation' of all kinases:  $X_i << C_i$ 

$$\frac{dX_i}{dt} = \alpha_i X_{i-1} \left( 1 - \frac{X_i}{C_i} \right) - \beta_i X_i \longrightarrow \frac{dX_i}{dt} = \alpha_i X_{i-1} - \beta_i X_i$$

- □ Analytic solutions for performance of the cascade:
  - Signaling time:

$$\tau = \frac{1}{\lambda} + \sum_{j=1}^{n} \frac{1}{\beta_{j}}$$

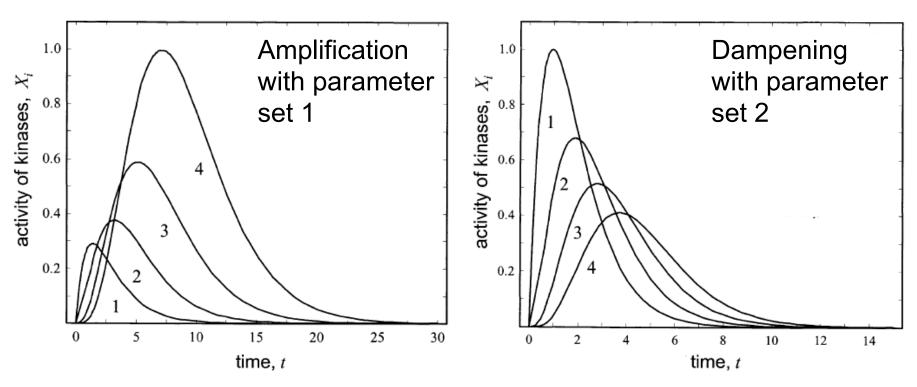
Signal duration:

$$\vartheta = \sqrt{\frac{1}{\lambda^2} + \sum_{j=1}^n \frac{1}{\beta_j^2}}$$

■ Signal amplitude:

$$S = \frac{S_0 \prod_{k=1}^n \frac{\alpha_k}{\beta_k}}{\sqrt{1 + \lambda^2 \sum_{j=1}^n \frac{1}{\beta_j^2}}}$$

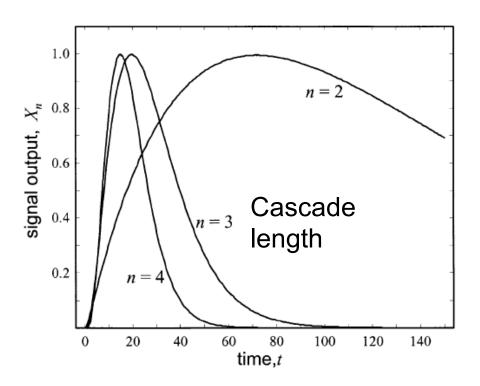
#### **Signaling Cascades: Performance**



R. Heinrich et al., Molecular Cell 9: 957, 2002.

 Qualitative dynamics: Amplification or dampening of signal traversing the cascade → Parameter-dependent.

#### **Signaling Cascades: Performance**



R. Heinrich et al., Molecular Cell 9: 957, 2002.

 Counter-intuitive finding: Cascades with more levels can propagate signals faster.

# Systembiologie 551-1174-00L

# **Beyond Metabolism**

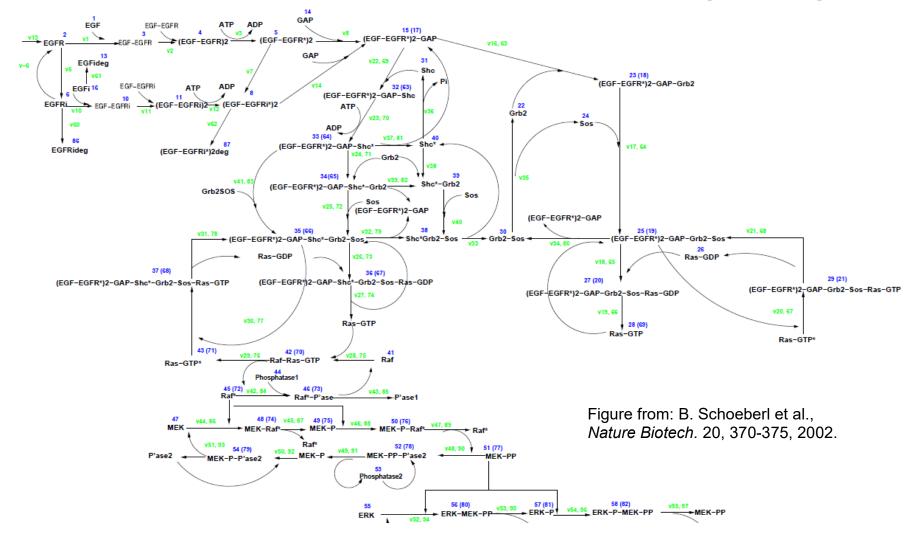
6 April, 2017
Uwe Sauer, Molecular Systems Biology
Jörg Stelling, D-BSSE

#### **Content:**

- Signaling pathways networks (US)
- Dynamic models for cell signaling (JS)
  - MAPK cascade signaling
  - Application in drug discovery
- Exams (US/JS)



#### A 'Real' Model of MAPK Cascade Signaling



□ ODE model for Epidermal Growth Factor (EGF) signaling in humans → Target discovery for anti-cancer drugs.

#### **MAPK Cascade Signaling Network**

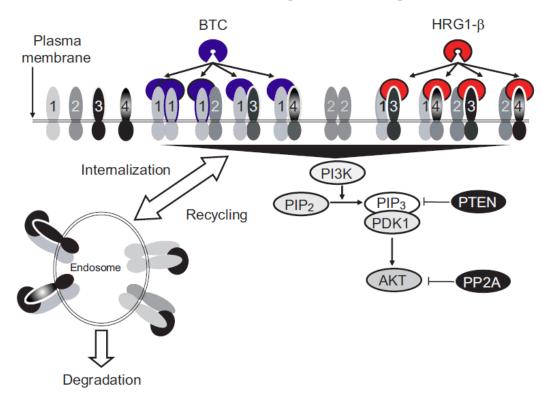
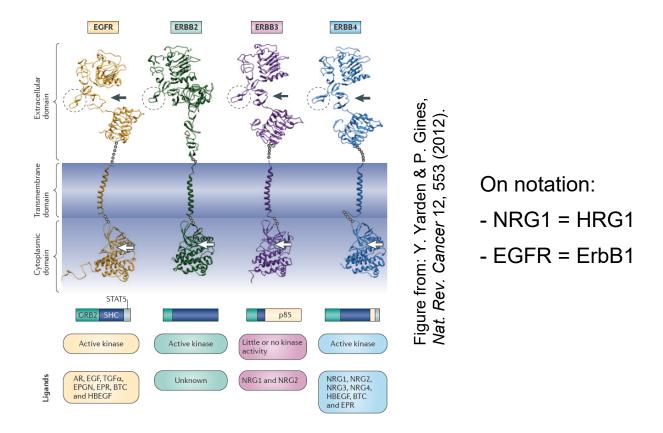


Figure from: B. Schoeberl et al., Sci. Signal. 2, ra31 (2009).

□ Signaling network with four receptors (ErbB1-4) and two ligands (BTC, Hrg1-β) → Growth signaling in cancers.

#### **MAPK Cascade Signaling Network**



- □ Signaling network with four receptors (ErbB1-4) and two ligands (BTC, Hrg1-β) → Growth signaling in cancers.
- Special case ErbB3: receptor without kinase activity.

### Model Development for ErbB Signaling

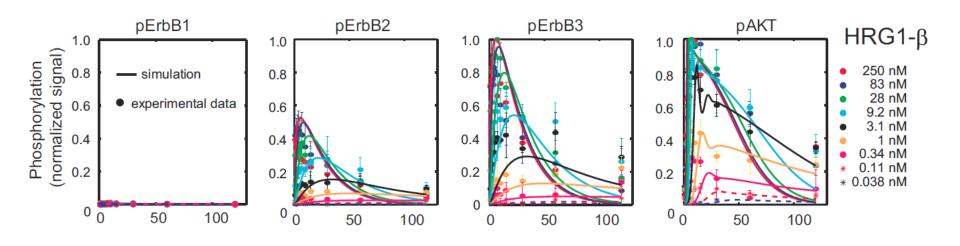


Figure from: B. Schoeberl et al., Sci. Signal. 2, ra31 (2009).

- □ ODE model with ~500 states and ~230 parameters.
- Challenge: Calibrating parameters with experiments.
- Needs: Dynamic data, different ligand doses, ...

#### **Model Development for ErbB Signaling**

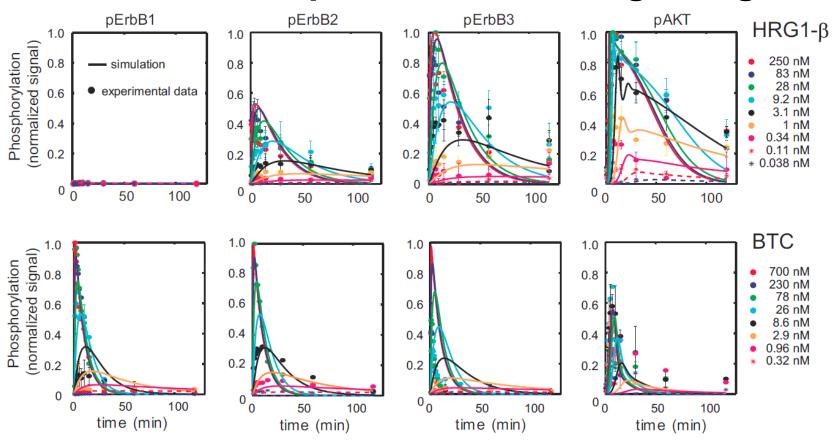
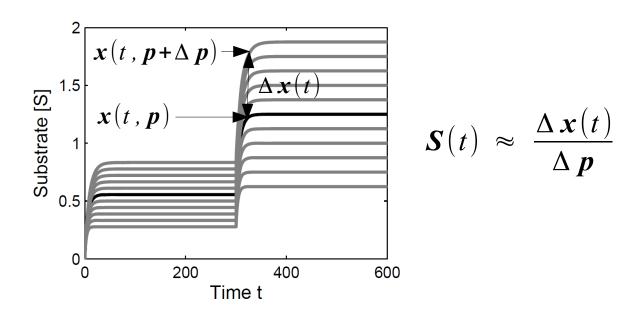


Figure from: B. Schoeberl et al., Sci. Signal. 2, ra31 (2009).

Calibrated model describes experiments reasonably.

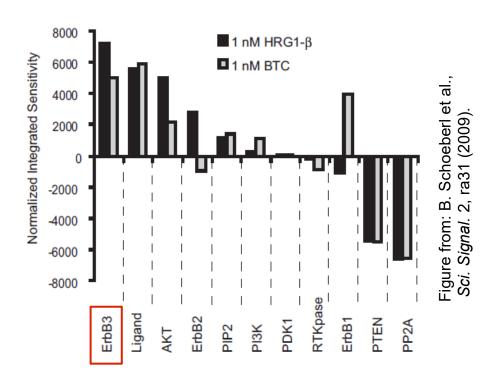
How to use the model to identify potential drug targets?

#### **Concept: Parameter Sensitivities**



□ Parameter sensitivities S(t): Changes of system states in response to (small) changes in model parameters around a reference parameter set.

#### **Model Analysis: Parameter Sensitivities**



- Question: Which perturbations (e.g. by drugs) are most effective for abolishing growth signaling?
- Surprising result: Perturbing the 'non-active'
   receptor ErbB3 is predicted to be most effective.

#### Model Application: Virtual Drug Design

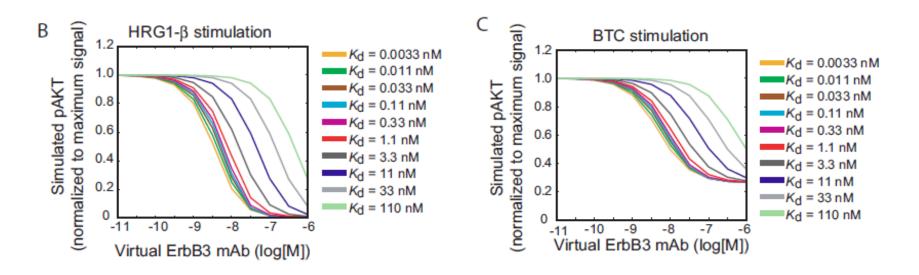


Figure from: B. Schoeberl et al., Sci. Signal. 2, ra31 (2009).

- Question: Can one design a monoclonal antibody (mAb) against ErbB3 that prevents signaling?
- □ Required mAb affinities are feasible → Produce mAb.

#### **Application in the 'Real' World**

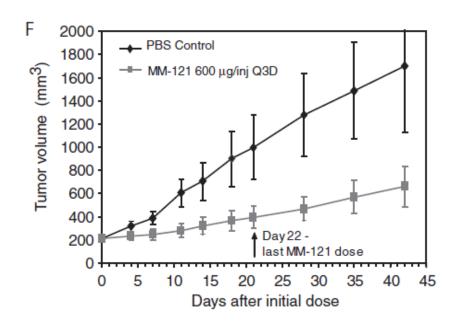
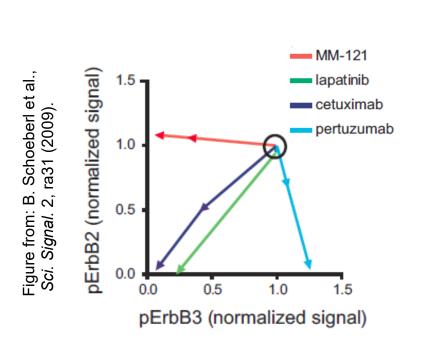


Figure from: B. Schoeberl et al., Sci. Signal. 2, ra31 (2009).

□ Validation: In vivo (mouse) experiments, monitor tumor growth with and without mAb (MM-121) treatment → Reduced tumor growth with mAb.

#### Why Is the Target ErbB3 Relevant?



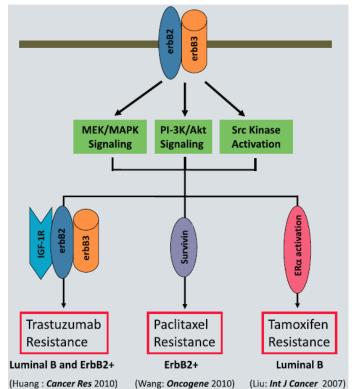


Figure from: J. Ma et al., *Molecular Cancer* 13: 105 (2014).

□ Inhibiting ErbB3 has different effects than those of established cancer drugs → One could overcome cancer resistance to drug treatments.

#### **Summary: Teaching Goals II-IV**

- Cell signalling follows the same principles as discussed for metabolism: importance of dynamics and feedback.
- Dynamic mathematical modeling generalizes to cell signalling, but assumptions need to be critically evaluated (e.g., Src kinase does not follow Michaelis-Menten).
- Large-scale dynamic models can be applied to identify potential drug targets by analyzing (parameter) sensitivities in cell signaling networks.

"All models are wrong but some models are useful."

George Box (1979)

Robustness in the strategy of scientific model building.