

In superresolution microscopy methods, like PALM, individual fluorescent molecules such as tagged proteins are detected. In ordinary microscopes, this is rather difficult because light emitted or scattered by out-of-focus molecules confer a strong background to the image. Another source of unwanted fluorescent signal is the autofluorescence of some structures such as mitochondria, chloroplasts, or autofluorescing molecules such as NADH. All these sources of light tend to blur the fluorescence from the particular molecule of interest. Here, we will introduce a technique that allows the detection of single fluorophores without detection of the background noise: Total-internal reflection fluorescence (TIRF) microscopy.

**A** only molecules in the evanescent field fluoresce

sample

cover slip

immersion oil

laser light

objective

critical angle for total internal reflection

$\sim 200$  nm

**B**

$n_a$

$n_b$

$n_a < n_b$

$\theta_1$

$\theta_2$

$\theta_c$

$\theta_1'$

Partially reflected

Totally reflected

**C**

In a TIRF microscope, the specimen is illuminated with a laser at an angle where total internal reflection occurs. This is possible, because the refraction index of the glass cover slide is greater than that of water ( $n = 1.3$ ). Because of total internal reflection, the light does not enter the sample, and the majority of fluorescent molecules are not illuminated. However, electromagnetic energy does extend, as a so-called evanescent field, for a very short distance (100 to 200 nm) beyond the surface of the cover slip and into the specimen, allowing just those molecules in the layer closest to the surface to become excited (see figure 1B). When these molecules fluoresce, their emitted light is no longer competing with out-of-focus light from the other molecules in the specimen, and can now be detected. TIRF has allowed several groundbreaking experiments, such as imaging of single motor proteins moving along microtubules.

**CAL** center for active learning