

# LeaRning R

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*All information is my personal opinion*



# Getting started

## 1. Download R

- for either Mac or Windows: <https://cran.rstudio.com/>

## 2. Install Rstudio

- scroll down to installers and download either the Windows or Mac version: <https://www.rstudio.com/products/rstudio/download/#download>

## 3. Open Rstudio

- a) type `install.packages('tidyverse')` in the bottom left panel (the console)
- b) let R install all required packages for you (can take 10 min)
- c) after the command finished type `library(tidyverse)`.
- d) If you encounter problems / errors along the way let me know and I can help you. You're all set!

# Overview

## 1. Why R?

## 2. Interacting with R

- Rstudio
- R basics
- Data workflow

—————→ What you need to know to get started

## 3. Visualizing data in R

- The ggplot approach
- Data exploration (can we beat The Economist?)

—————→ Learning to speak the *grammar of graphics*

## 4. How to use R in the lab (from easy to advanced)

—————→ Potential use cases

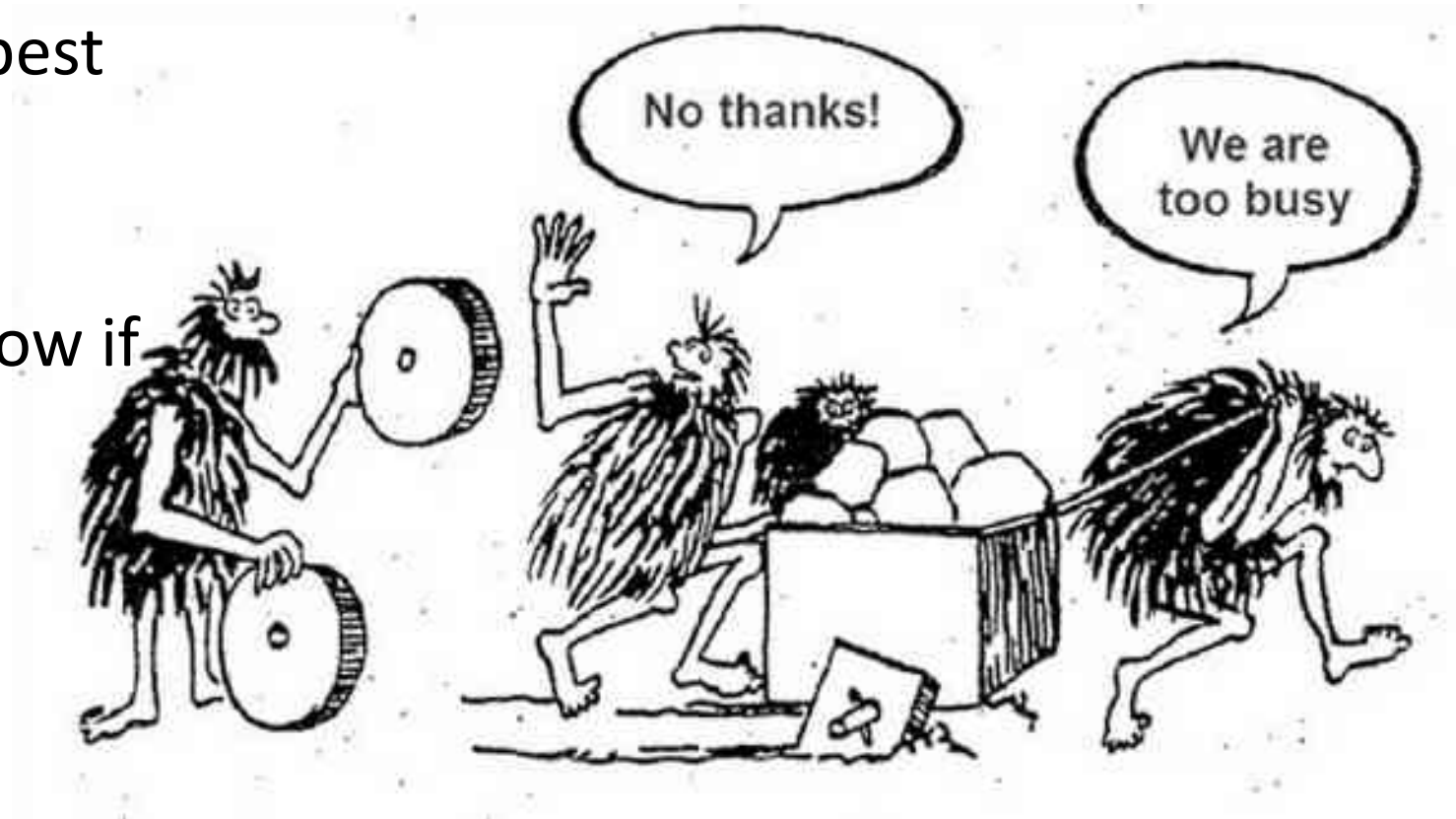
## 5. Outlook

# Why R?

*(my personal opinion)*

# Why I think learning R is a good idea?

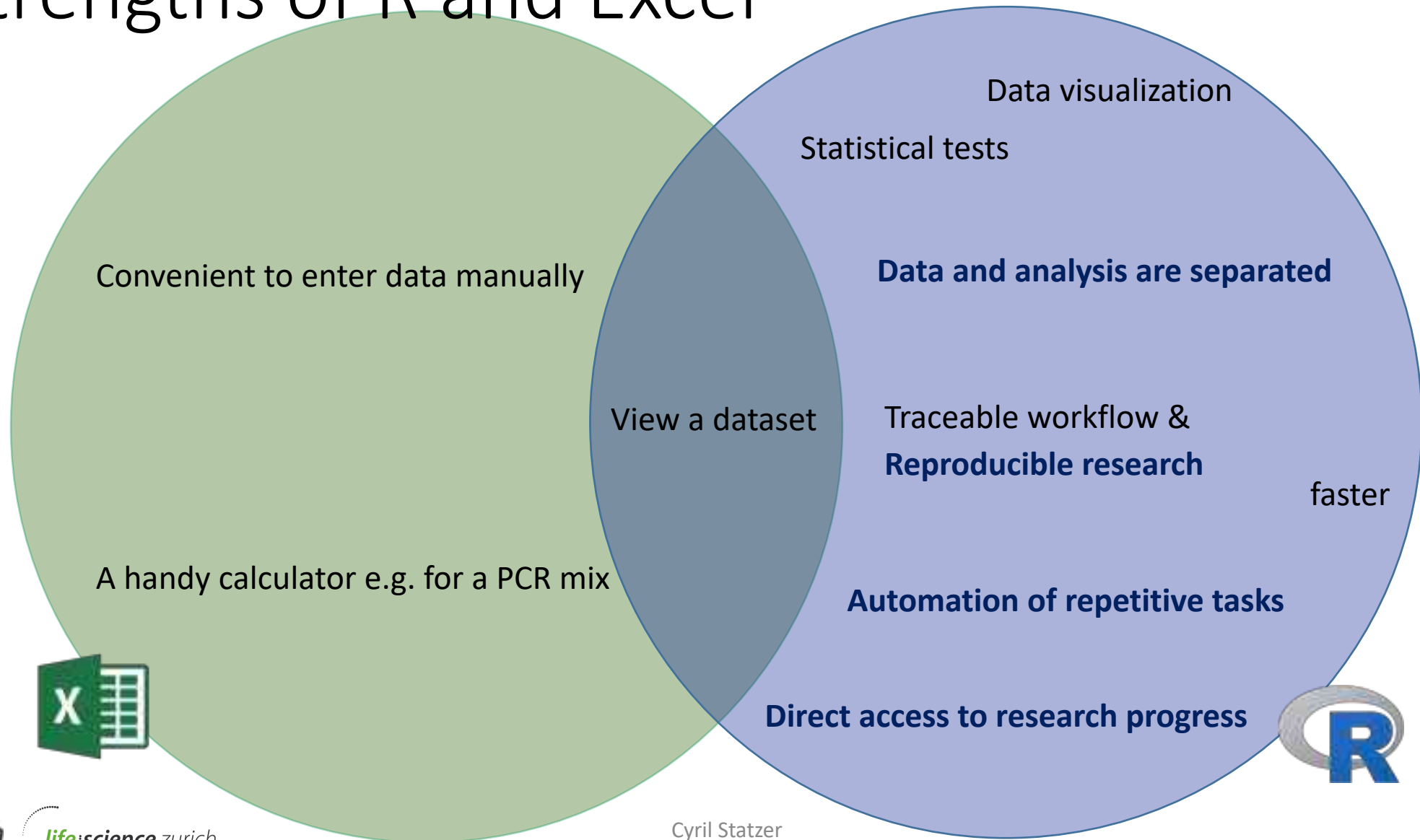
- Are you using the best available tools?
- How would you know if you were not?



Le Borgne, 2017

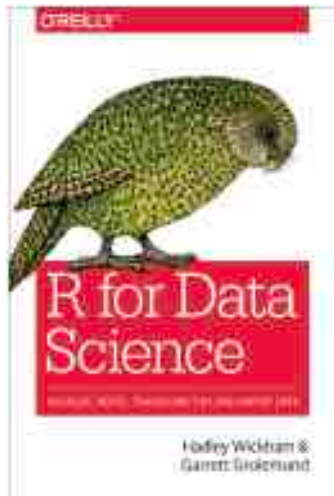
*most of us*

# Strengths of R and Excel

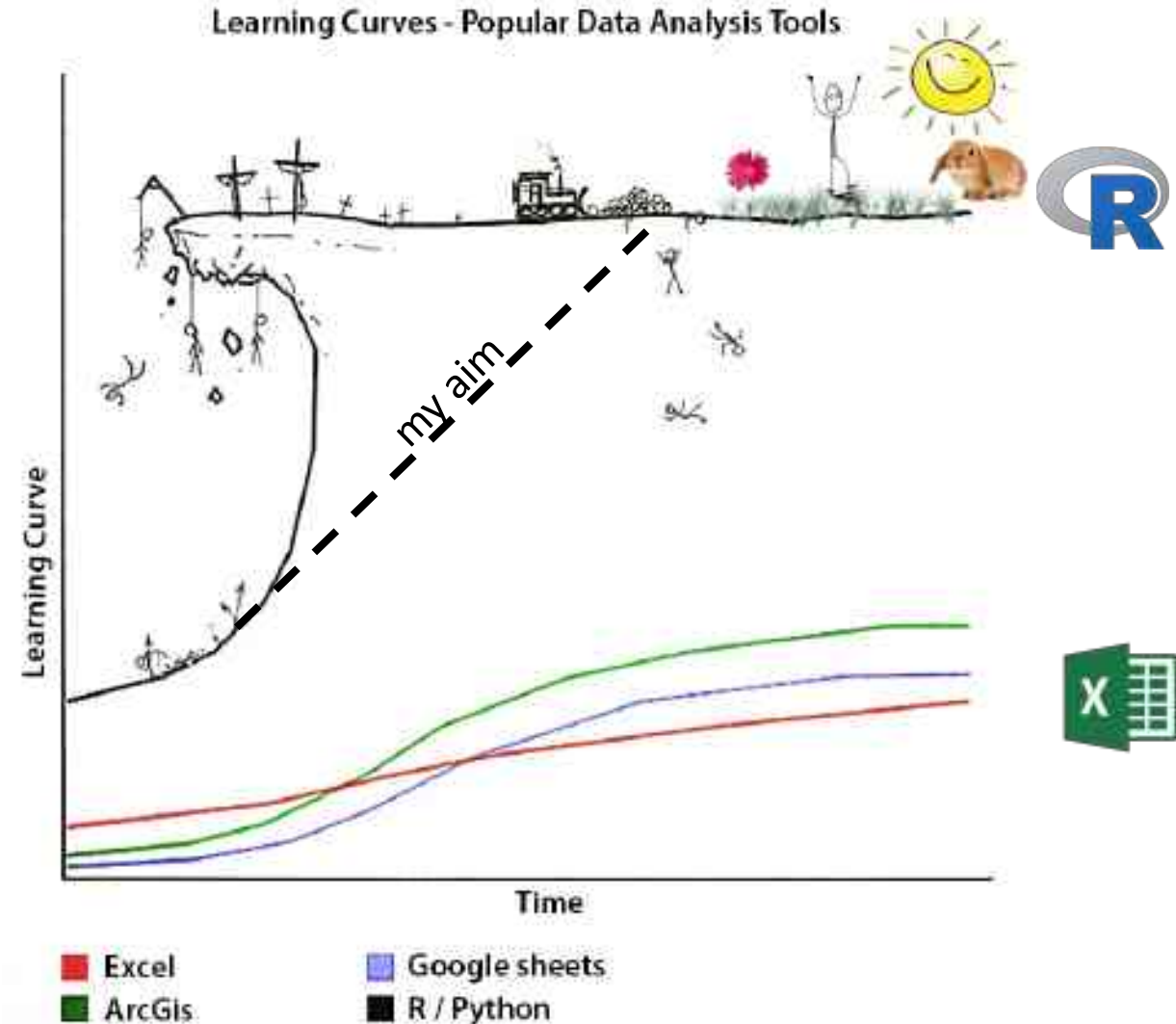


# Getting started is hard

- Initial work required but it pays off greatly
- You can get help everywhere



The best book is for free:  
<https://r4ds.had.co.nz>



## *Disclaimer*



- This presentation will become overwhelming towards the end
- We'll go from learning a new language to writing a short story in one lesson
- Try to hang on and understand the overall concepts



# How can we use R & RStudio?

# Rstudio (1/3)



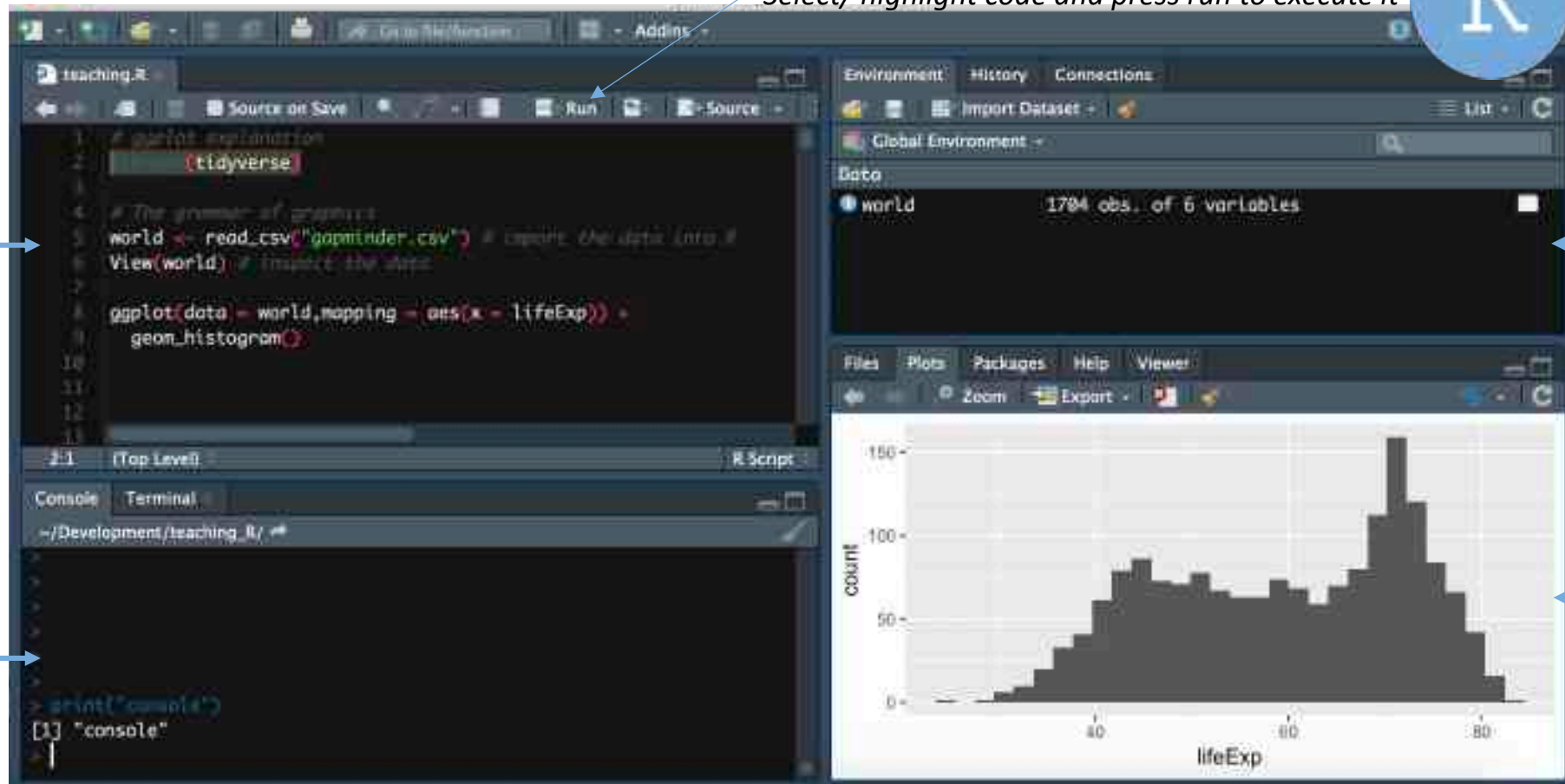
Select/ highlight code and press run to execute it

script

objects

plots

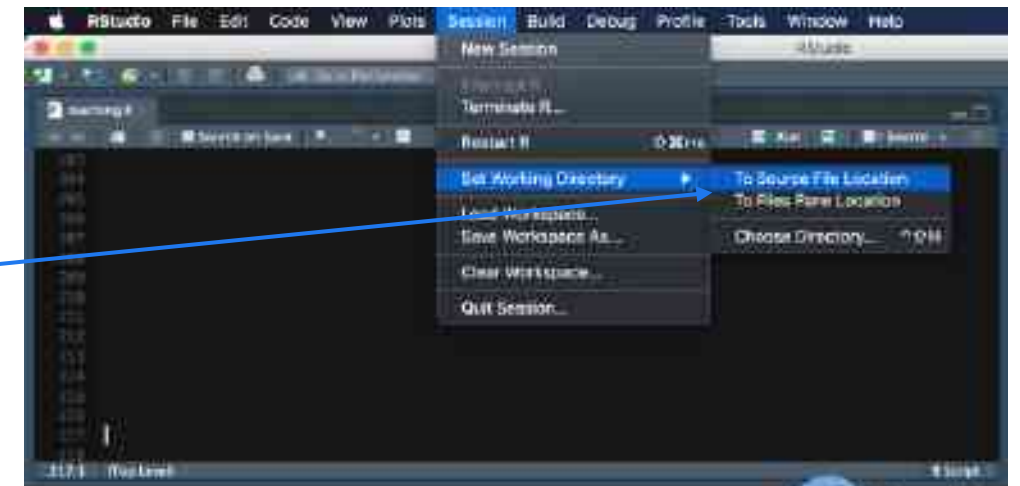
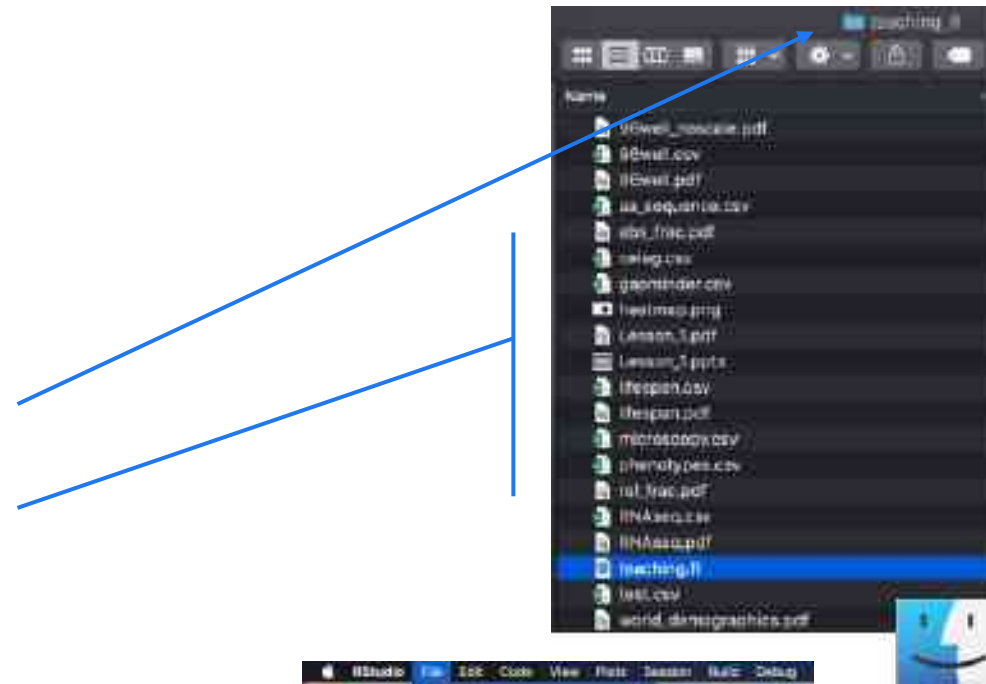
console



1.) new script, 2.) save script to folder, 3.) Set working directory to script location

# Rstudio (2/3)

1. Make a **folder** for your R project
2. Put your **data** into this folder
3. Open Rstudio, open and then **save the R script to this folder**
4. When you start Rstudio set the **Working directory** to the location of your script



# Rstudio (3/3)

1. Now R can “see” the files you want to work with:

```
> dir()
[1] "Lesson_1.pptx"      "96well_noscale.pdf"  "96well.csv"          "96well.pdf"
[5] "aa_sequence.csv"    "abs_frac.pdf"        "celeg.csv"           "gapminder.csv"
[9] "heatmap.png"       "Lesson_1.pdf"        "Lesson_1.pptx"       "lifespan.csv"
[13] "lifespan.pdf"       "microscopy.csv"      "phenotypes.csv"      "rel_frac.pdf"
[17] "RNAseq.csv"         "RNAseq.pdf"          "teaching.R"           "test.csv"
[21] "world_demographics.pdf"
> |
```

The `dir()` functions shows all the files in your folder.

2. You can directly import them by name since R knows now where to look for them

```
read_csv("gapminder.csv")
```

# R basics (1/2)



*Assignment is giving names to things. It is like labelling a box and then put anything you want into it. We call this "an object" in R.*

- **Assignment**

```
Kelvin <- 273.15
```

*"273.15 is saved in Kelvin"*

```
tempsC <- c(5, 20, 30)
```

*"tempsC has 3 numerical entries:  
5, 20 and 30"*

```
feeling <- c("cold", "medium", "hot")
```

*"feeling has 3 character entries:  
"cold", "medium" and "hot"*

- **Data types**



*The data type determines what you can do with it:  
10 / 2 works but "hello" / "world" not.*

- **Numeric:**

*Kelvin and tempsC are numeric*

*Test with: `class(Kelvin)`*

- **Character**

*feeling is character*

*Test with: `class(feeling)`*

# R basics (2/2)

- Functions

```
mean (tempsC)
```

```
18.33333
```

```
max (tempsC)
```

```
30
```

```
length (feeling)
```

```
3
```

- Multiple  
functions

```
temp <- mean (tempsC)
```

```
round (temp)
```

```
18
```

```
tempsC %>% mean () %>% round ()
```

```
18
```

# Data workflows (1/3)

```
world <- read_csv("gapminder.csv")  
world
```

	country	continent	year	lifeExp	pop	gdpPercap
	<chr>	<chr>	<int>	<dbl>	<int>	<dbl>
1	Afghanistan	Asia	1952	28.8	8425333	779.
2	Afghanistan	Asia	1957	30.3	9240934	821.
3	Afghanistan	Asia	1962	32.0	10267083	853.
4	Afghanistan	Asia	1967	34.0	11537966	836.
5	Afghanistan	Asia	1972	36.1	13079460	740.
6	Afghanistan	Asia	1977	38.4	14880372	786.
7	Afghanistan	Asia	1982	39.9	12881816	978.
8	Afghanistan	Asia	1987	40.8	13867957	852.

```
View(world)
```

Import the dataset into the object world

Display the contents of the object

This is well-structured (tidy) data!

- Each **variable** is saved in its own **column**.
- Each **observation** is saved in its own **row**.

Browse the table like in Excel

%>% is called a pipe, passing the data to the next function

== is equal ("=" in mathematics)

# Data workflows (2/3)



Take the **world** dataset **and** filter it for the Americas

```
americas <- world %>%  
  filter(continent == "Americas")
```

	country	continent	year	lifeExp	pop	gdpPerCap
	<chr>	<chr>	<int>	<dbl>	<int>	<dbl>
1	Argentina	Americas	1952	62.5	17876956	5911.
2	Argentina	Americas	1957	64.4	19610538	6857.
3	Argentina	Americas	1962	65.1	21283783	7133.
4	Argentina	Americas	1967	65.6	22934225	8053.
5	Argentina	Americas	1972	67.1	24729799	9443.
6	Argentina	Americas	1977	68.5	26983828	10079.
7	Argentina	Americas	1982	69.9	29341374	8998.
8	Argentina	Americas	1987	70.8	31620918	9140.



Take the **world** dataset **and** filter it for Asia **and** select the specified columns **and** arrange them by descending life expectancy.

```
Asia_lifexp <- world %>%  
  filter(continent == "Asia") %>%  
  select(country, year, lifeExp) %>%  
  arrange(desc(lifeExp))
```

	country	year	lifeExp
	<chr>	<int>	<dbl>
1	Japan	2007	82.6
2	Hong Kong, China	2007	82.2
3	Japan	2002	82
4	Hong Kong, China	2002	81.5
5	Israel	2007	80.7
6	Japan	1997	80.7
7	Hong Kong, China	1997	80
8	Singapore	2007	80.0



# Data workflows (3/3)



Take the **world** dataset **and** group it by continent and year and for every group **then** summarize the life expectancy by taking the mean **and then** arrange the output by year and continent

```
lifeexp_by_continent <- world %>%  
  group_by(continent, year) %>%  
  summarize(mean_life_exp = mean(lifeExp)) %>%  
  arrange(year, continent)
```


	continent <chr>	year <int>	mean_life_exp <dbl>
1	Africa	1952	39.1
2	Americas	1952	53.3
3	Asia	1952	46.3
4	Europe	1952	64.4
5	Oceania	1952	69.3
6	Africa	1957	41.3
7	Americas	1957	56.0
8	Asia	1957	49.3

*If you "speak" R you can directly "talk" to your data and tell it what you want to do.*

# How can we visualize data in R?

# Data visualization with ggplot

```
world <- read_csv("gapminder.csv")
```

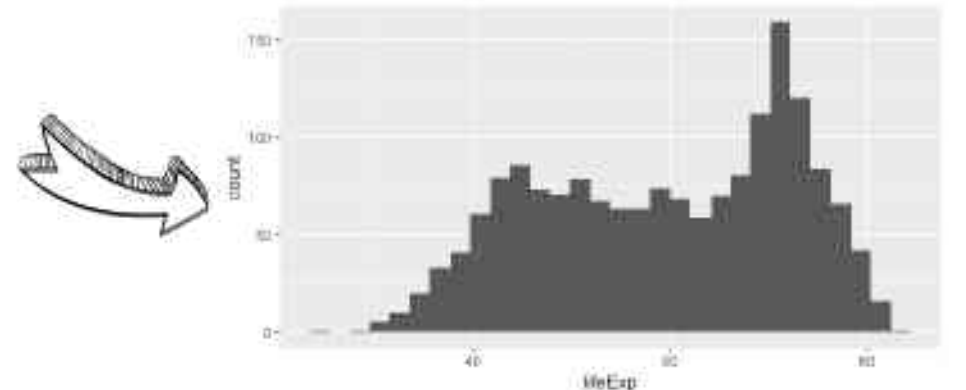


	A	B	C	D	E	F
1	country	continent	year	lifeExp	pop	gdpPercap
2	Afghanistan	Asia	1952	28.801	8425333	779.445315
3	Afghanistan	Asia	1957	30.332	9240934	820.85303
4	Afghanistan	Asia	1962	31.997	10267083	853.10071
5	Afghanistan	Asia	1967	34.02	11537966	836.197138
6	Afghanistan	Asia	1972	36.088	13079460	739.981106
7	Afghanistan	Asia	1977	38.438	14880372	
8	Afghanistan	Asia	1982	39.854	12881816	9

<https://www.gapminder.org/data/documentation/>




```
ggplot(data = world, mapping = aes(x = lifeExp)) +  
  geom_histogram()
```



# Data visualization with ggplot

```
world <- read_csv("gapminder.csv")
```



	A	B	C	D	E	F
1	country	continent	year	lifeExp	pop	gdpPercap
2	Afghanistan	Asia	1952	28.801	8425333	779.445315
3	Afghanistan	Asia	1957	30.332	9240934	820.85303
4	Afghanistan	Asia	1962	31.997	10267083	853.10071
5	Afghanistan	Asia	1967	34.02	11537966	836.197138
6	Afghanistan	Asia	1972	36.088	13079460	739.981106
7	Afghanistan	Asia	1977	38.438	14880372	
8	Afghanistan	Asia	1982	39.854	12881816	9

<https://www.gapminder.org/data/documentation/>



*What*

```
ggplot(data = world,
```

*Where*

```
mapping = aes(x = lifeExp))
```

+

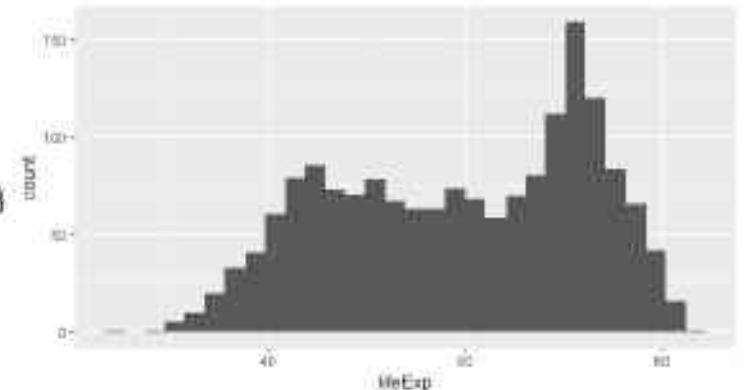
```
geom_histogram()
```



*How*



"I want to plot the **world dataset** with the **life expectancy** mapped to the **x-axis** using a **histogram**"



*What*

*Where*

```
plot <- ggplot(data = world, mapping = aes(x = continent, y = lifeExp))
```

*How*

plot



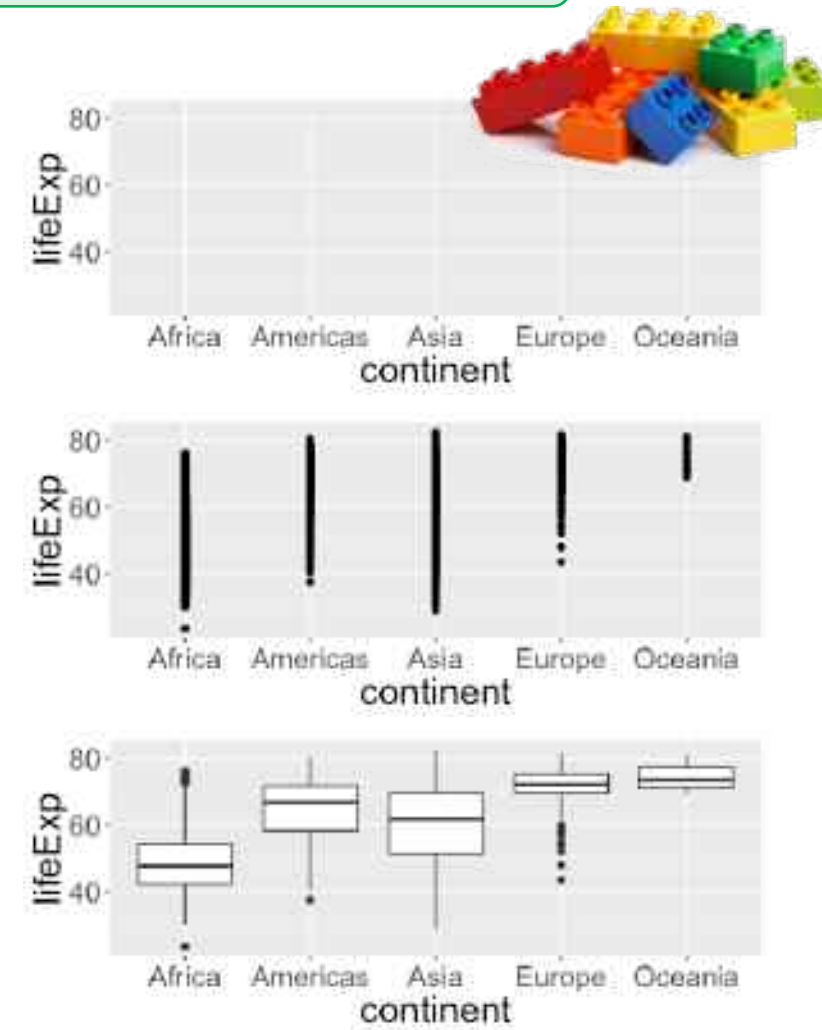
*How*

plot + geom\_point()



*How*

plot + geom\_boxplot()



"I want to plot the **world dataset** with the **continents mapped to the x-axis** and the **life expectancy to the y-axis** using **points, boxplots or more**"

*What*

```
plot <- ggplot(data = world,
```

*Where*

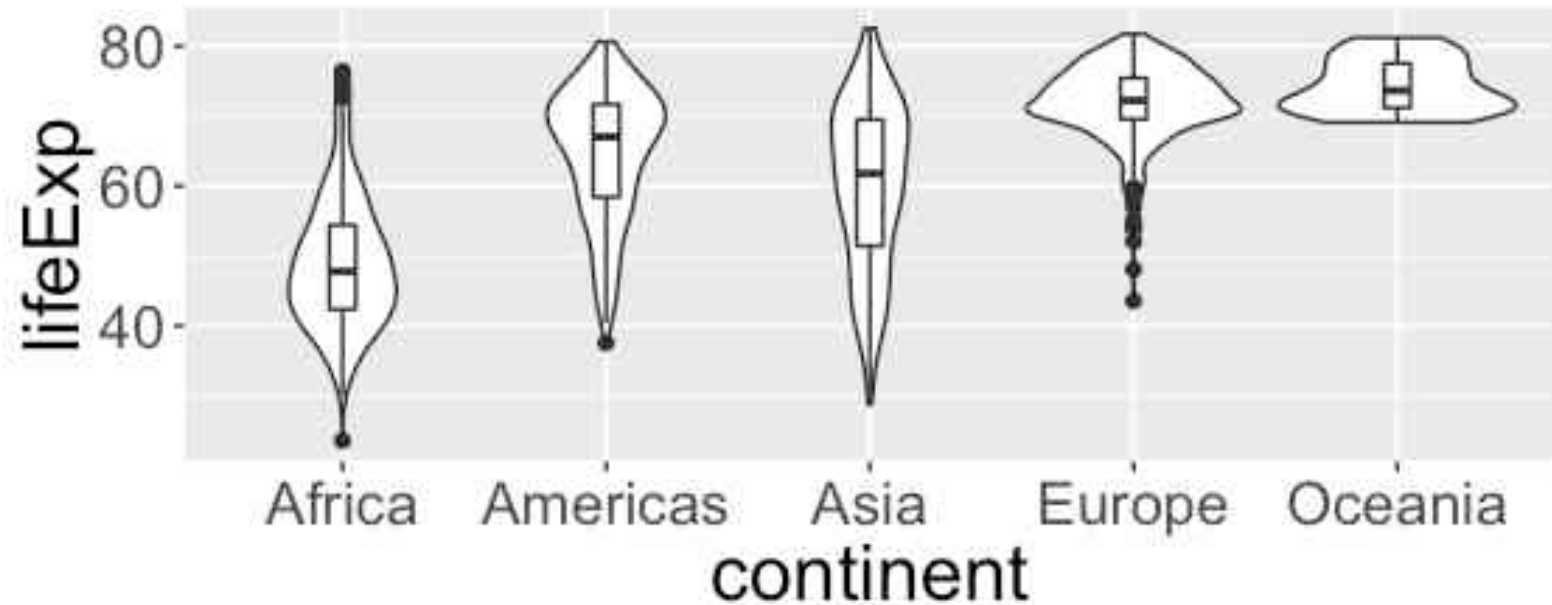
```
mapping = aes(x = continent, y = lifeExp))
```

*How*

```
plot + geom_violin() + geom_boxplot(width = 0.1)
```

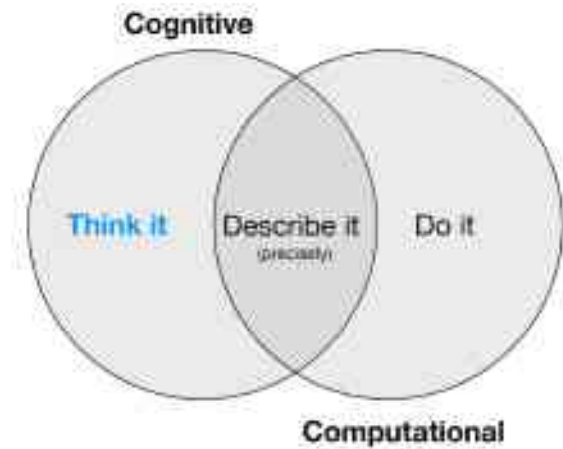


**All geoms are combinable!**



*"I want to plot the **world dataset** with the **continents** mapped to the x-axis and the **life expectancy** to the y-axis using **points, boxplots or more**"*

# ggplot syntax



ggplot ( **what dataset**, **map parameters to plot dimensions** ) +  
**what to draw**

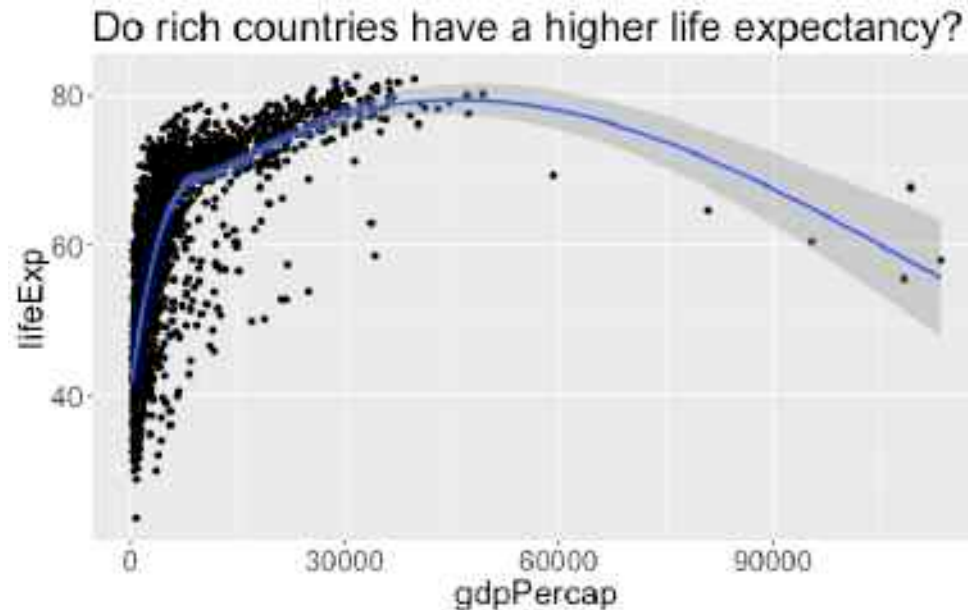
- Let's start asking questions!
  - How did the world change? How the continents and individual countries?
  - How are GDP and lifespan linked? Is this the same across continents?

# Data exploration



Do rich countries have a higher life expectancy?

```
ggplot(data = world, mapping = aes(x = gdpPercap, y = lifeExp)) +  
  geom_point() +  
  geom_smooth() +  
  ggtitle("Do rich countries have a higher life expectancy?")
```



Up to 50000 gdpPercap yes, above  
life expectancy decreases again





Reviewer: is this continent-specific?

# Data exploration



Do rich countries have a higher life expectancy?

```
ggplot(data = world, mapping = aes(x = gdpPercap, y = lifeExp)) +  
  geom_point() +  
  geom_smooth() +  
  ggtitle("Do rich countries have a higher life expectancy?")
```



?



How would you  
change the code?

?



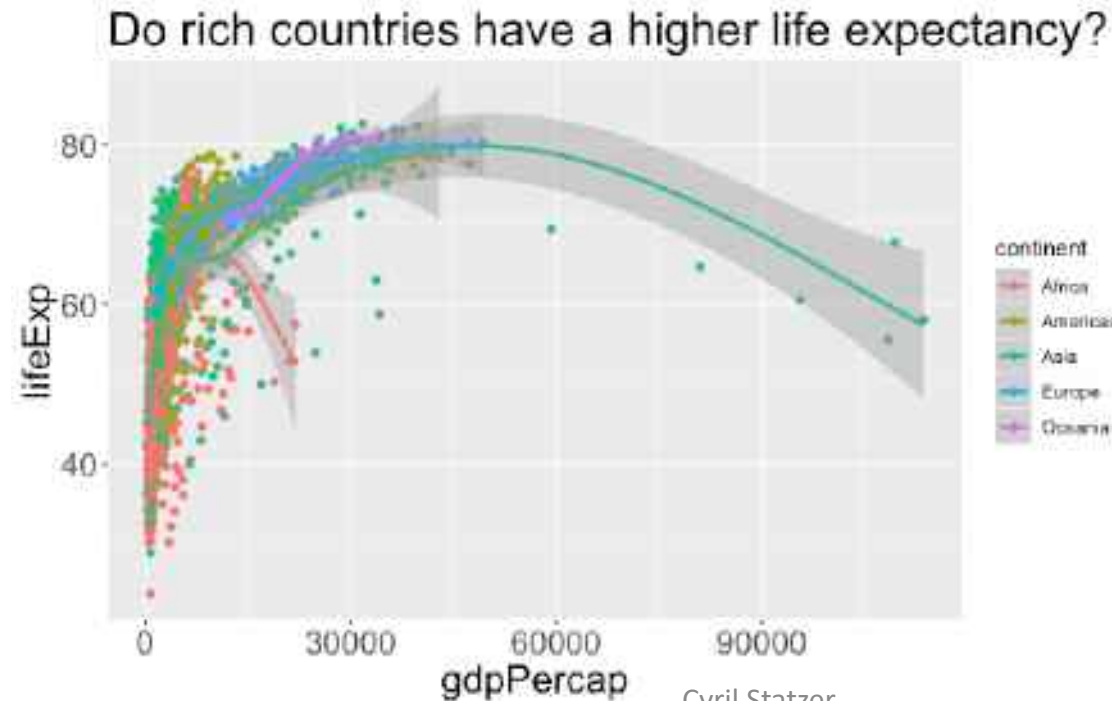
Reviewer: is this continent-specific?

# Data exploration



## Do rich countries have a higher life expectancy?

```
ggplot(data = world, mapping = aes(x = gdpPercap, y = lifeExp, color = continent)) +  
  geom_point() +  
  geom_smooth() +  
  ggtitle("Do rich countries have a higher life expectancy?")
```



life expectancy increases monotonously with gdpPercap in Oceania, Europe and the Americas

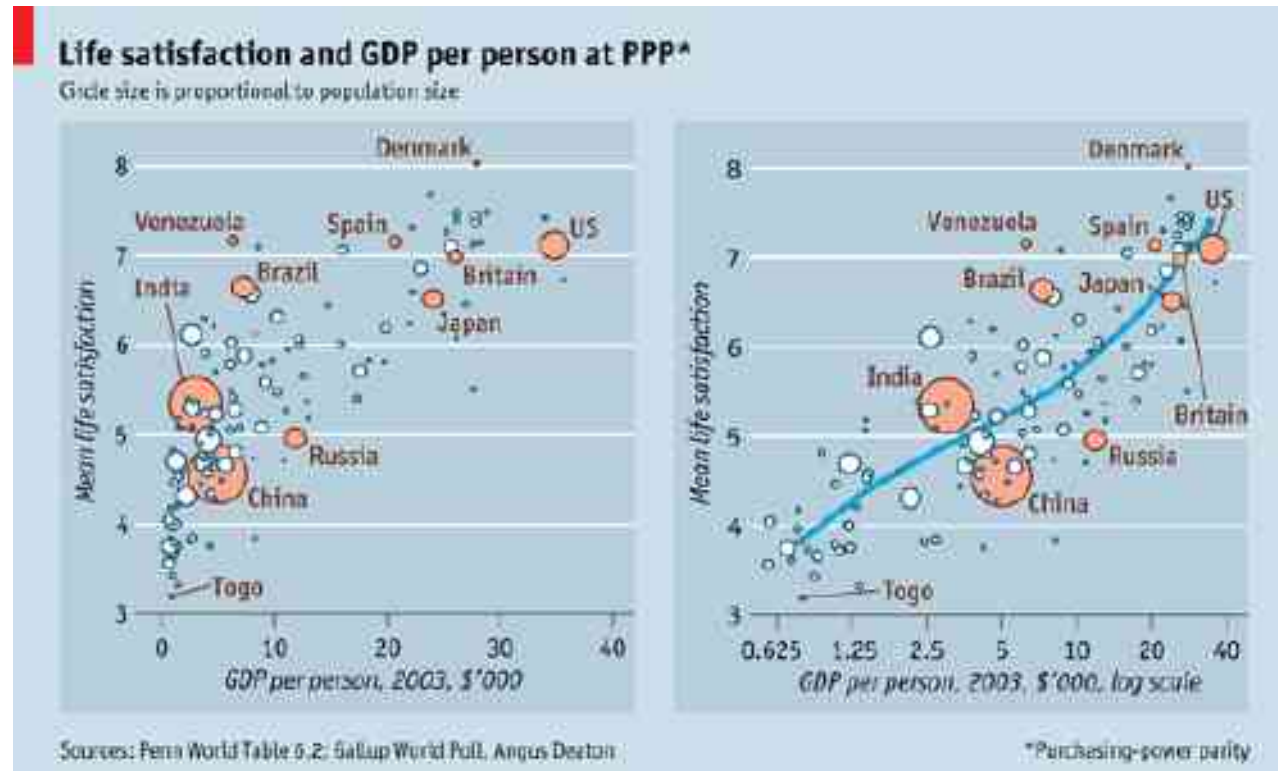


Up to 10000 gdpPercap yes, above life expectancy decreases again in Africa

Up to 50000 gdpPercap yes, above life expectancy decreases again in Asia

# Outlook

Can we do better than The Economist?



The Economist

They looked at:  
**GDP vs. life satisfaction  
in 2003**

We will compare  
**GDP vs. life expectancy  
in 2002**

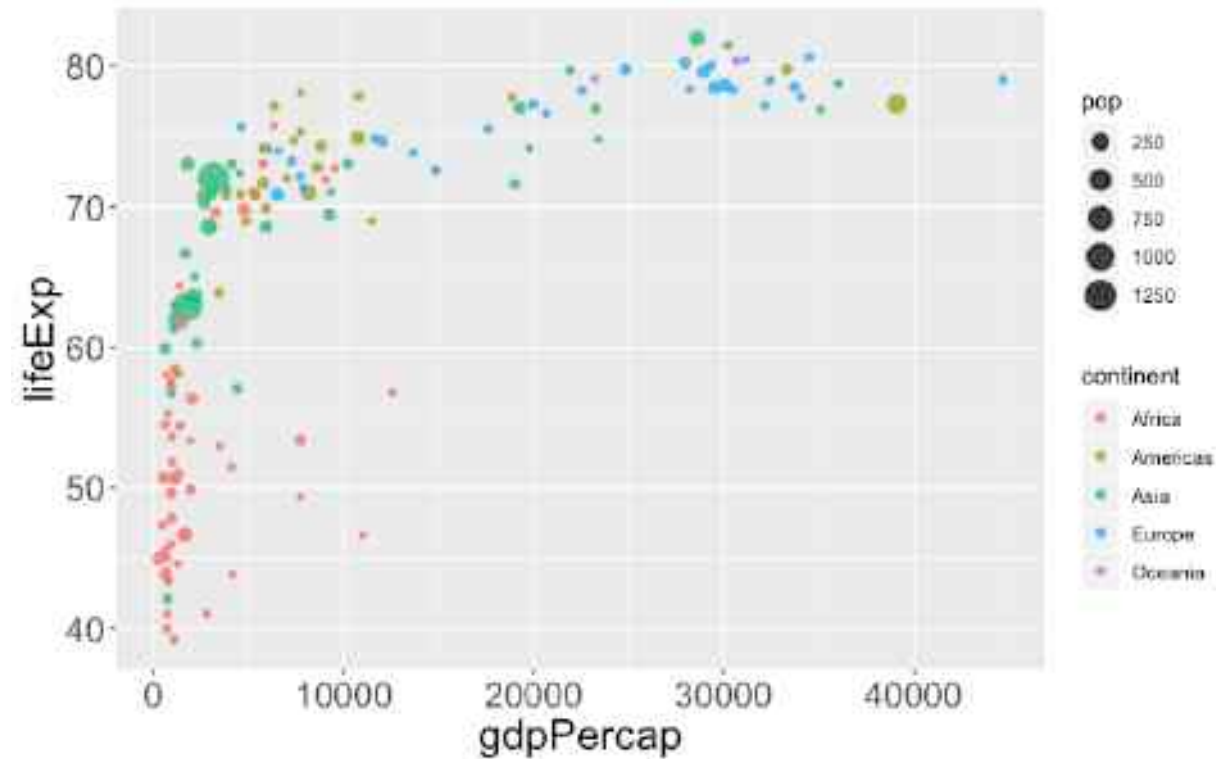
# Outlook

## Life expectancy vs. GDP in 2002

```
world_2002 <- world %>%  
  filter(year == 2002) %>%  
  mutate(pop=pop/1000000)
```

```
plot <- ggplot(data = world_2002,  
  aes(x= gdpPercap,  
      y= lifeExp,  
      size = pop,  
      color = continent)) +  
  geom_point(alpha=0.8)
```

plot



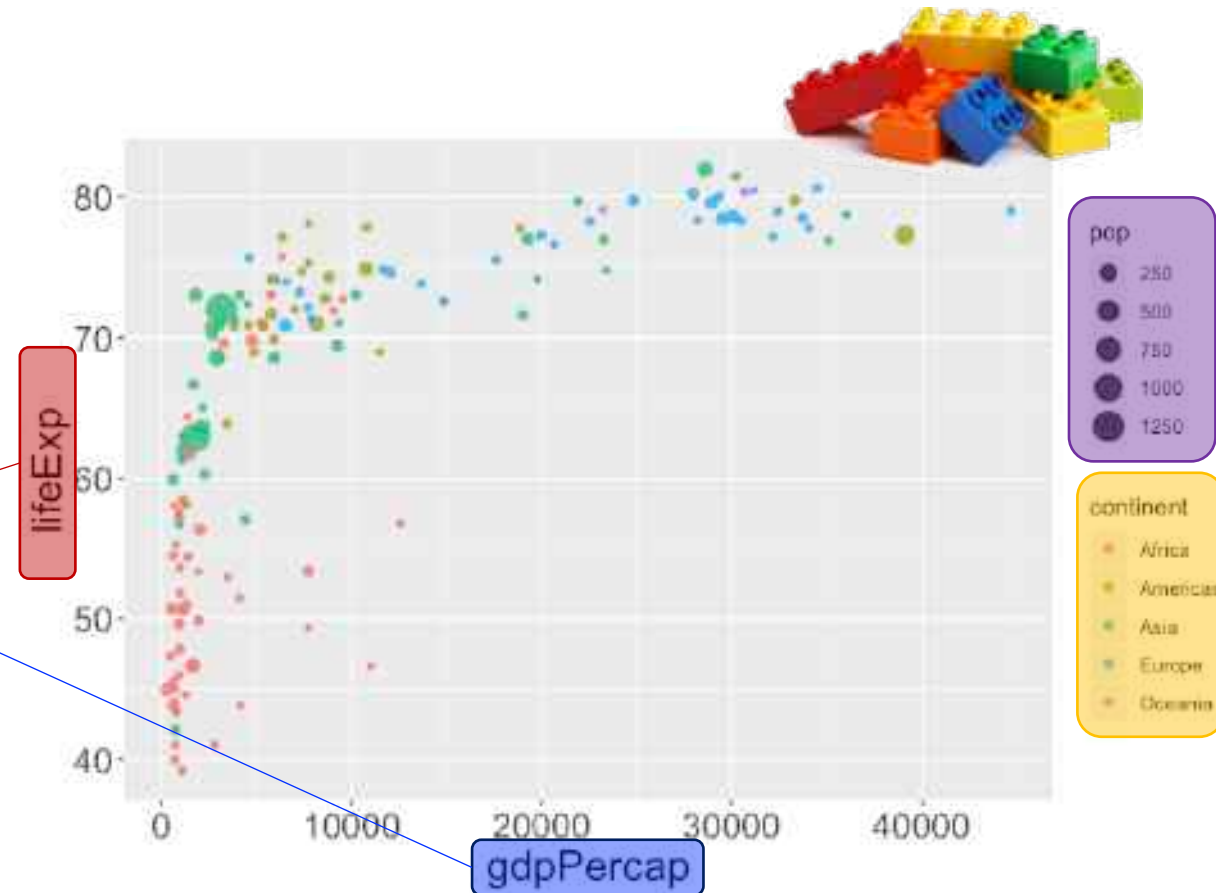
# Outlook

## Life expectancy vs. GDP in 2002

```
world_2002 <- world %>%  
  filter(year == 2002) %>%  
  mutate(pop=pop/1000000)
```

```
plot <- ggplot(data = world_2002,  
  aes(x= gdpPerCap,  
      y= lifeExp,  
      size = pop,  
      color = continent)) +  
  geom_point(alpha=0.8)
```

plot



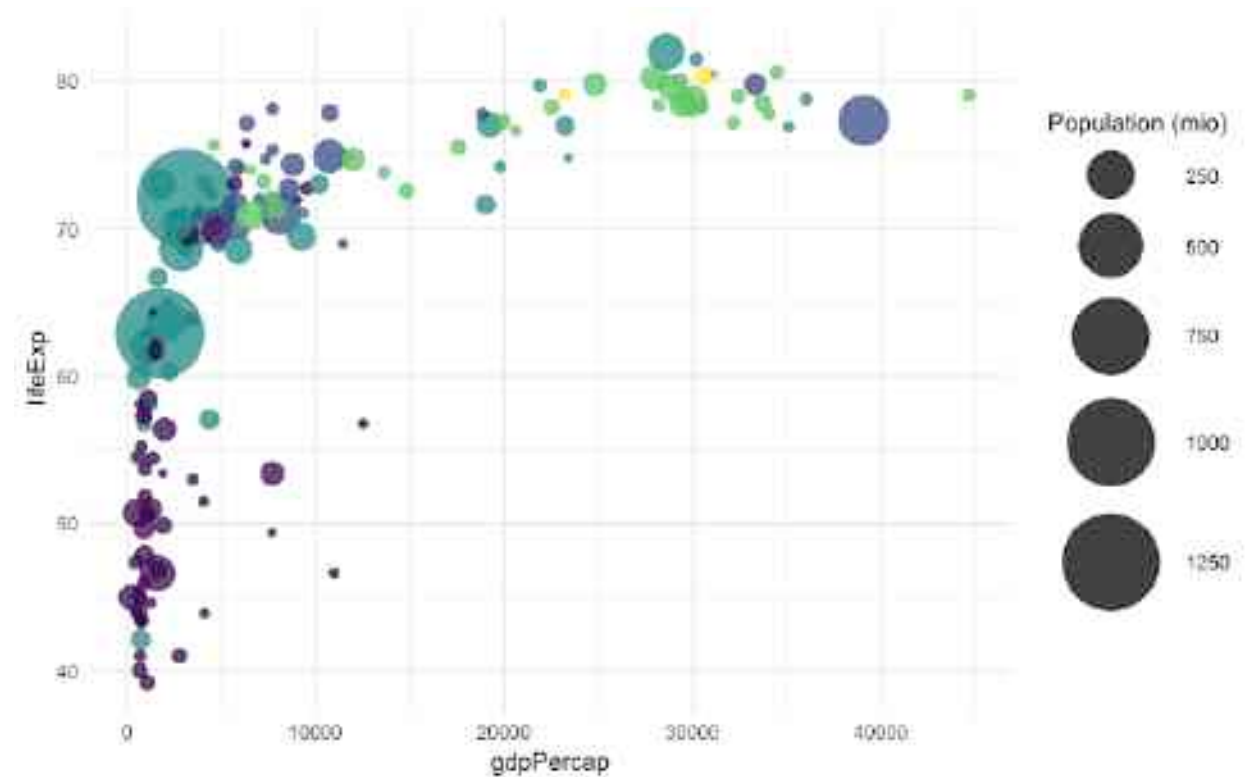
# Outlook

## Life expectancy vs. GDP in 2002

```
plot +  
  scale_size(range = c(1, 20),  
             name="Population (mio)") +  
  scale_color_viridis(discrete=TRUE,  
                     guide=FALSE) +  
  theme_minimal()
```



*“styling the plot”*



*Next level: what is the life expectancy  
in the wealthiest African countries?*

...better

# World demographics in 2002





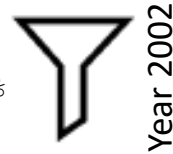
# Outlook

## Life expectancy vs. GDP in 2002



```
library(viridis)
library(ggforce)
library(ggrepel)
```

```
world_2002 <- world %>%
  filter(year == 2002) %>%
  mutate(pop=pop/1000000)
```



Year 2002

```
africa_leading <- world_2002 %>%
  filter(continent == "Africa",
         gdpPercap > 3000)
```



Labelling Africa

Logic of the plot

8 lines of code

```
plot <- ggplot(data = world_2002, aes(x=gdpPercap, y=lifeExp, size = pop, color = continent, label = country)) +
  geom_point(alpha=0.8) +
  scale_size(range = c(1, 20), name="Population (mio)") +
  scale_color_viridis(discrete=TRUE, guide=FALSE) +
  facet_zoom(x = continent == "Africa") +
  geom_label_repel(data = africa_leading, size = 3) +
  theme_bw() +
  labs(title = "World demographics in 2002", x = "GDP per capita [US dollars per inhabitant]", y = "Life expectancy [years]")
```

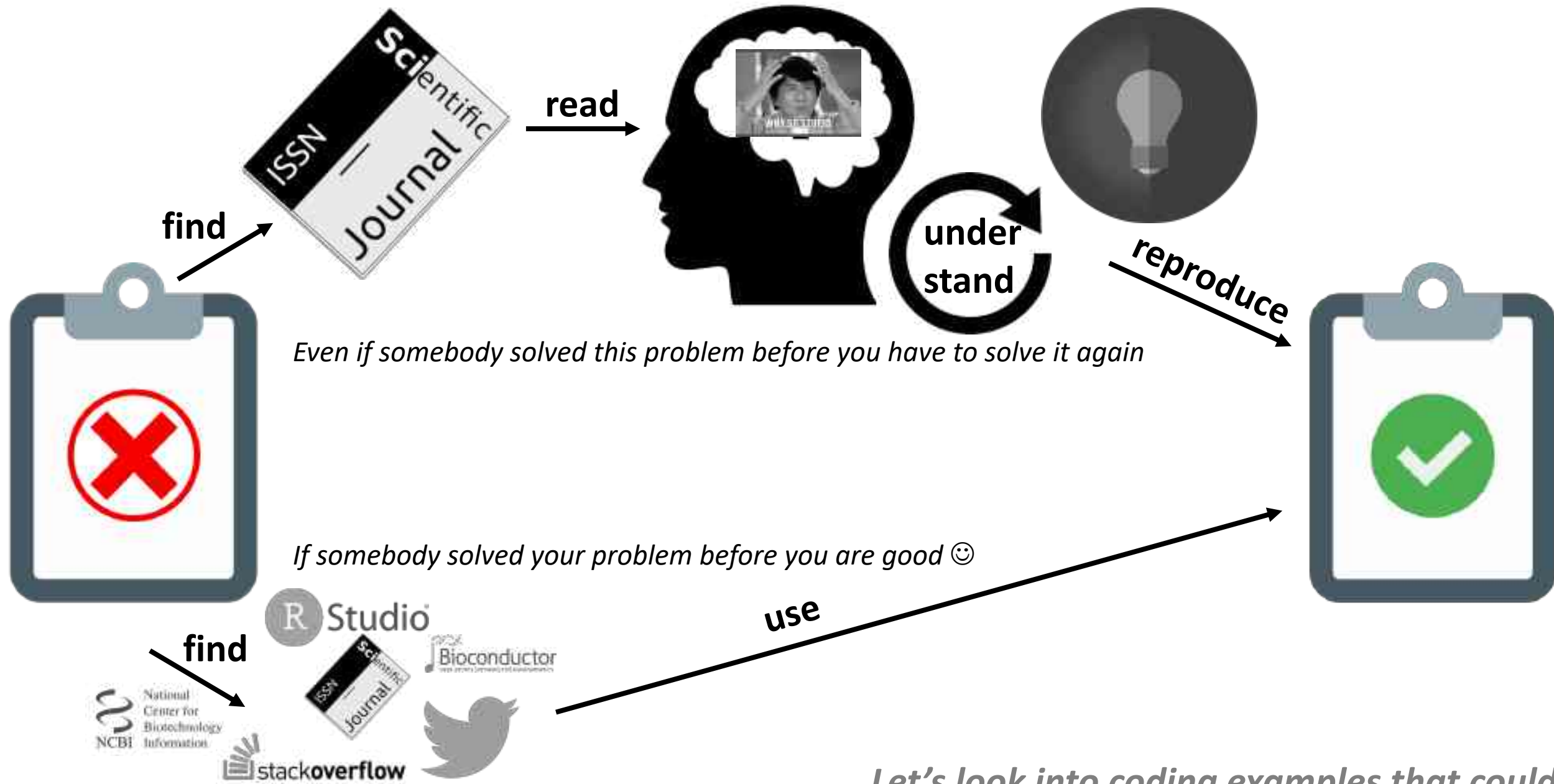
Styling

Labels

plot



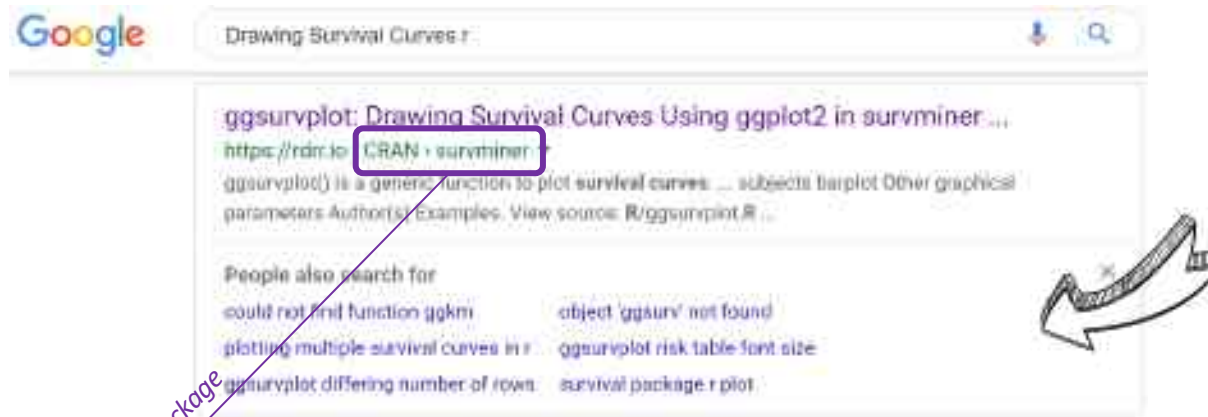
# R in the lab



lifespan.csv

# Example 1: plotting lifespans

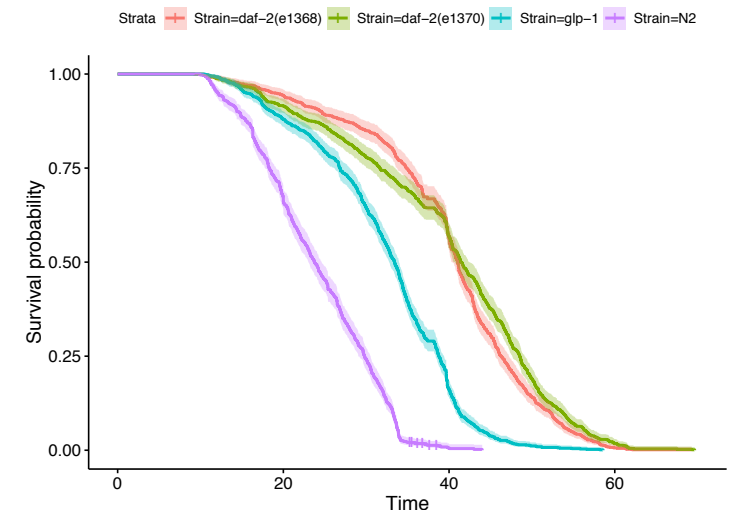
	A	B	C	D
1	Repeat	Strain	Censored	Time
2	Coruscant	daf-2(e1368)	1	10.370289
3	Andromeda	daf-2(e1368)	1	10.664641
4	Coruscant	daf-2(e1368)	1	10.953623
5	Bespin	daf-2(e1368)	1	11.011678
6	Bespin	daf-2(e1368)	1	11.074178
7	Bespin	daf-2(e1368)	1	11.074178
8	Andromeda	daf-2(e1368)	1	11.074178



```
library(tidyverse)
library(survminer)
library(survival)

data <- read_csv(file = "lifespan.csv")

fit<- survfit(Surv(Time, Censored) ~ Strain, data = data)
ggsurvplot(fit = fit, data = data)
```



plot  
import  
literature

# Example 2: plotting microscopy scoring



microscopy.csv

	A	B
1	category	Frequency
2	None	51
3	Low	170
4	Medium	15
5	High	5
6	Very high	1



library(tidyverse)  
library(waffle)

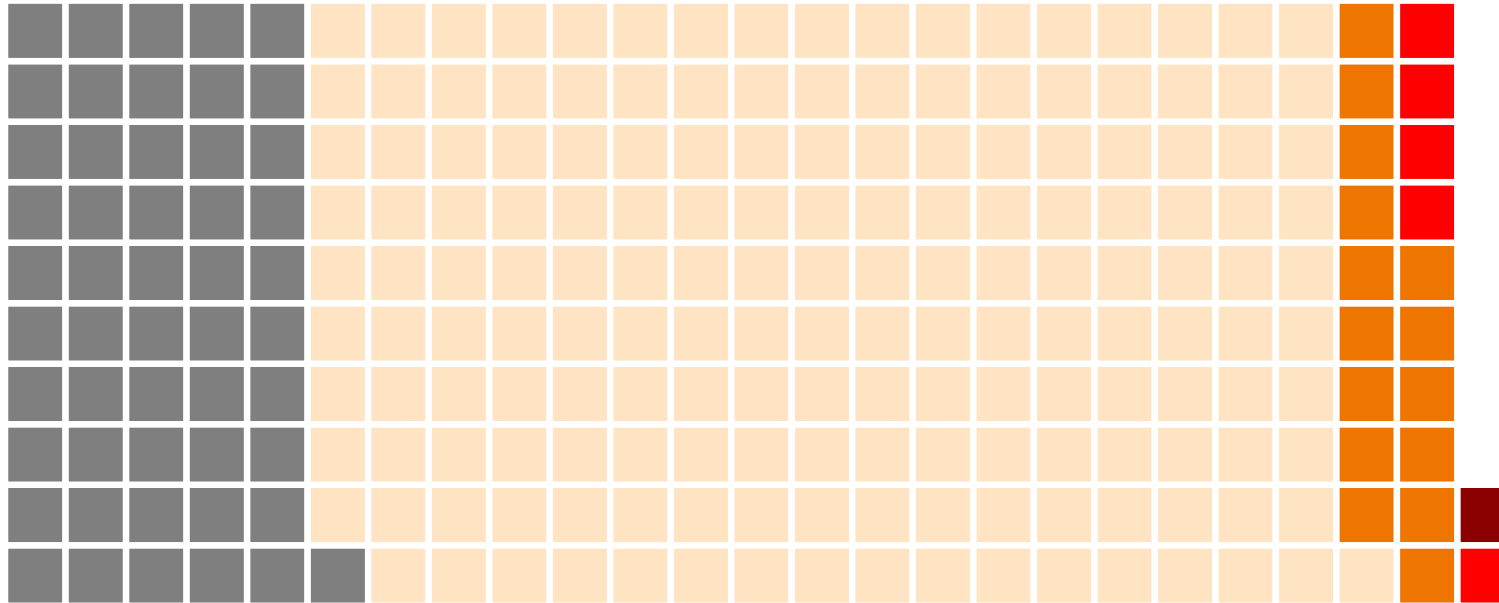
package

colors <- c("grey50", "bisque1", "darkorange2", "red", "darkred")

data <- read\_csv(file = "microscopy.csv")  
absolute <- data\$Frequency  
names(absolute) <- data\$category

waffle(absolute, rows = 10, size = 1, colors = colors, legend\_pos = "bottom", title = "Absolute observations")  
relative <- round(absolute/sum(absolute) \* 25)  
waffle(relative, rows = 1, size = 1, colors = colors, legend\_pos = "bottom", title = "Relative fractions")

## Absolute observations



*microscopy.csv*

	A	B
1	category	Frequency
2	None	51
3	Low	170
4	Medium	15
5	High	5
6	Very high	1



## Relative fractions



None
  Low
  Medium
  High
  Very high

# Example 3: Merging CHIP & RNA-seq data



gene	CHIP_count
<i>daf-2</i>	0.96
<i>emb-9</i>	1.80
<i>hlh-30</i>	1.60



gene	RNA_FC	RNA_pval
<i>cdc-40</i>	1.01	0.20
<i>pmp-30</i>	0.89	0.07
<i>Y11D7A.11</i>	1.13	0.60
<i>hlh-30</i>	1.43	0.04
<i>gst-4</i>	2.04	0.0001
<i>daf-2</i>	0.87	0.002



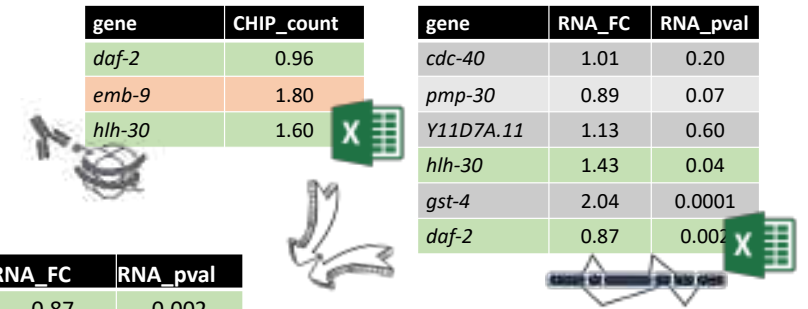
gene	CHIP_count	RNA_FC	RNA_pval
<i>daf-2</i>	0.96	0.87	0.002
<i>emb-9</i>	1.8	NA	NA
<i>hlh-30</i>	1.6	1.43	0.04
<i>cdc-40</i>	NA	1.01	0.2
<i>pmp-30</i>	NA	0.89	0.07
<i>Y11D7A.11</i>	NA	1.13	0.6
<i>gst-4</i>	NA	2.04	1.00E-04

15000 – 60000 rows in each dataset

# Example 3: Merging CHIP & RNA-seq data

```
chip <- read_csv("chip.csv")  
rnaseq <- read_csv("rna_seq.csv")  
full_join(chip, rnaseq, by = "gene")
```

	gene	CHIP_count	RNA_FC	RNA_pval
	<chr>	<dbl>	<dbl>	<dbl>
1	daf-2	0.96	0.87	0.002
2	emb-9	1.8	NA	NA
3	hlh-30	1.6	1.43	0.04
4	cdc-40	NA	1.01	0.2
5	pmp-30	NA	0.89	0.07
6	Y11D7A.11	NA	1.13	0.6
7	gst-4	NA	2.04	0.0001



# Common code patterns



- Packages («literature you use»)

```
library(tidyverse)
```

*general packages*

```
library(survminer)  
library(waffle)  
library(survival)
```

*specialized packages*

- Import your data

```
read_csv()
```

```
waffle()
```

```
full_join()
```

- Perform computation / plotting

```
survfit()
```

```
ggsurvplot()
```

*Most of your problems have already been solved by somebody! There is no need to drag squares around in powerpoint to make a waffle chart or to manually merge datasets in excel, let R do it for you*




# Advanced functions

- `as.numeric()` interpret as a number
- `unlist()` remove indexing
- `paste()` combine characters
- `str_replace_all()` similar to Cmd Find Replace
- `is.na()` Is data missing? YES or NO
- `!` Invert logic
- `separate` Split characters on pattern

# Advanced 1: Parsing strings

aa\_sequence.csv

	A	B	C	D
1	1	msqnnkrqvq	hnhemsndvc	plplpprgap
2	61	rklhvivid	qkvqknrvr	emalkdvqkv
3	121	dvaldvvt	hqqpslcyhi	gvresmgqsy
4	181	yihhqdsnql	qsydknnndd	dskppfartn
5	241	fmsdlrkare	itdgdqkndy	ldkmtrldn
6	301	dlkripdclk	vvdtpviryq	yafalnrrnk
7	361	dvclagriy	kdkfiasnye	dreslnsaie



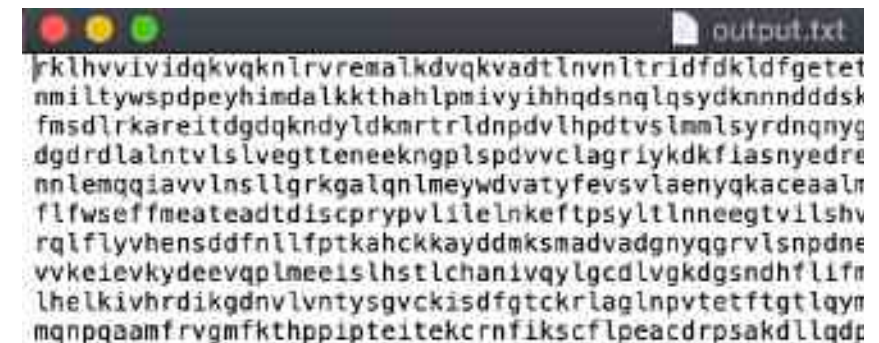
```
library(tidyverse)
```

```
input <- read_csv("aa_sequence.csv")
```

```
str <- input %>% unlist() %>% paste(., collapse=" ")  
str_proc <- str_replace_all(string = str, pattern = "[[:digit:]],[:space:]]", replacement = "")
```

```
write_lines(x = str_proc, path = "./output.txt")
```

output.txt



```
rklhvividqkvqknrvremalkdvqkvadtlnvnltridfkdldfgetet  
nmilywspdppeyhmdalkkthahlpmyihhqdsnqlqsydknnnddsk  
fmsdlrkareitdgdqkndyldkmtrldnnpdvlhpdvslmmsyrdnqnyg  
dgdrdlalntvlsvegtteneekngplspdvclagriykdkfiasnyedre  
nnlemqgiavvlnslgrkgalqnlmeywdvatyfevsvlaenyqkaceaaln  
flfwseffmeateadtdiscprypvlilelnkeftpsyltnneegtvilshv  
rqlflyvhensddfllfptkahckkayddmksmadvadgnyqgrvlsnpdne  
vvkeievkydeevqplmeeislhstlchanivqylgcdlvkgdgsndhflfr  
lhelkivhrdikgdnlvntysgvckisdftgckrlaglnpvtetftgtlqyr  
mqnpqaamfrvgmfkthppipteitekcnfikscflpeacdrpsakdllqdp
```

More advanced

# Advanced 2: data cleaning (RNAseq)

import *literature*

```
library(tidyverse)

data <- read_csv(file = "RNAseq.csv")
```

*cleaning*

```
data <- data %>% mutate(logFC = as.numeric(logFC)) %>% filter(!is.na(logFC))
readable <- data %>% separate(col = Gene_id,into = c("Junk","ID"),sep = "_")
readable <- readable %>% filter(!is.na(ID))
```

*plot*

```
ggplot(data = data, aes(x = logFC,y = -log10(padj))) +
  geom_point(alpha = 0.5)
```



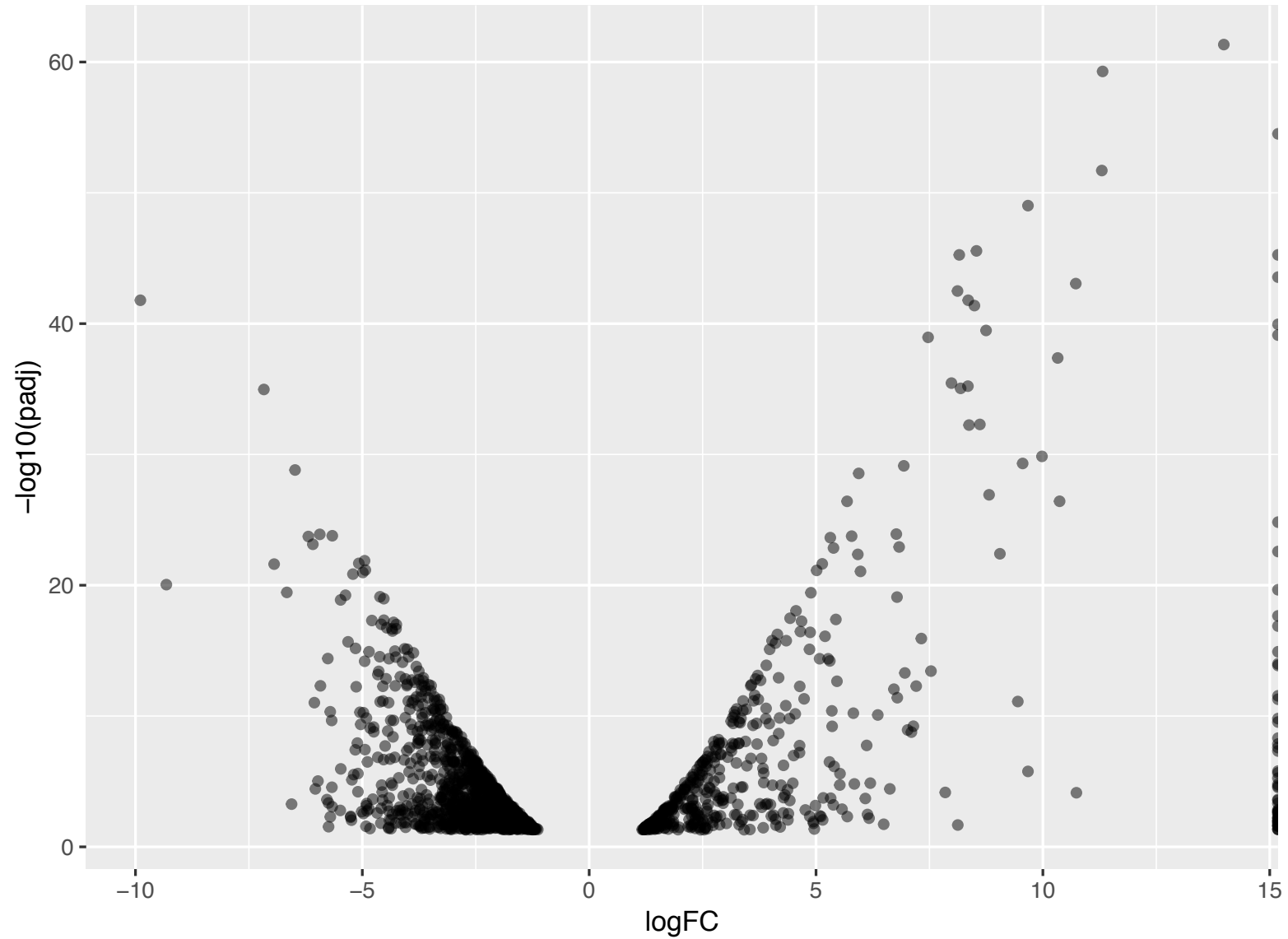
	A	B	C
1	Gene_id	logFC	padj
2	Parent=CDS_C01A2.6_wp188	4.9872	0.0006972
3	Parent=CDS_C05E4.12_wp221	8.1874	8.87E-36
4	Parent=CDS_C08E3.11_wp252	Inf	5.67E-06
5	Parent=CDS_C33C12.4_wp82	-3.0136	0.022033
6	Parent=CDS_C40A11.9_wp179	#NAME?	0.012781
7	Parent=CDS_C52E2.8	Inf	0.017116
8	Parent=CDS_C53A5.11_wp261	Inf	2.21E-18
9	Parent=CDS_D2062.11_wp21	8.5365	2.75E-46
10	Parent=CDS_F15D3.8_wp104	Inf	0.02

RNAseq.csv



Parent=Transcript_R02E12.2b.1	→	R02E12.2b.1
Parent=Pseudogene_Y105C5B.30	→	Y105C5B.30
Parent=CDS_R06A4.3		R06A4.3

## Volcano plot



# 96-well assay

```
library(tidyverse)
library(ggplot2)
library(magrittr)
library(scatterpie)
```

```
lets<- c("H","G","F","E","D","C","B","A")
df <- read_csv("96well.csv")
df <- df %>% mutate(region = paste0(Row,Column), Row = match(Row, lets))
df[,4:9] <- df[,4:9] / 80
```

```
p <- ggplot() +
  geom_point(data=expand.grid(seq(1, 12), seq(1,8)), aes(x=Var1, y=Var2),
             color="grey80", fill="white", shape=21, size=6) +
  geom_scatterpie(aes(x= Column, y= Row, r = Total, group = region), data=df,
                 cols=c("Extremely high","High", "Intermediate", "Low", "None" ))
```

```
p + scale_x_discrete(name ="Columns", limits=c(1:12)) +
  scale_y_discrete(name ="Rows", limits=lets) +
  scale_fill_manual(values = c("red","darkgreen","darkolivegreen3","darkseagreen1","grey")) +
  coord_fixed(ratio=12/12) + facet_grid(Day ~ .) +
  ggtitle("Promotor activity (day 1 and 8)")
```

	Position	Age	Intensity						
	A	B	C	D	E	F	G	H	I
1	Row	Column	Day	Extremely high	High	Intermediate	Low	None	Total
2	A	1	8	0	3	2	6	0	11
3	A	2	8	0	0	4	11	0	15
4	A	3	8	0	1	4	3	0	8
5	A	4	8	0	3	6	5	0	14
6	A	5	8	0	3	4	10	0	17
7	A	6	8	0	4	0	7	0	11
8	A	7	8	0	4	2	1	0	
9	A	8	8	0	1	0	4	0	



literature

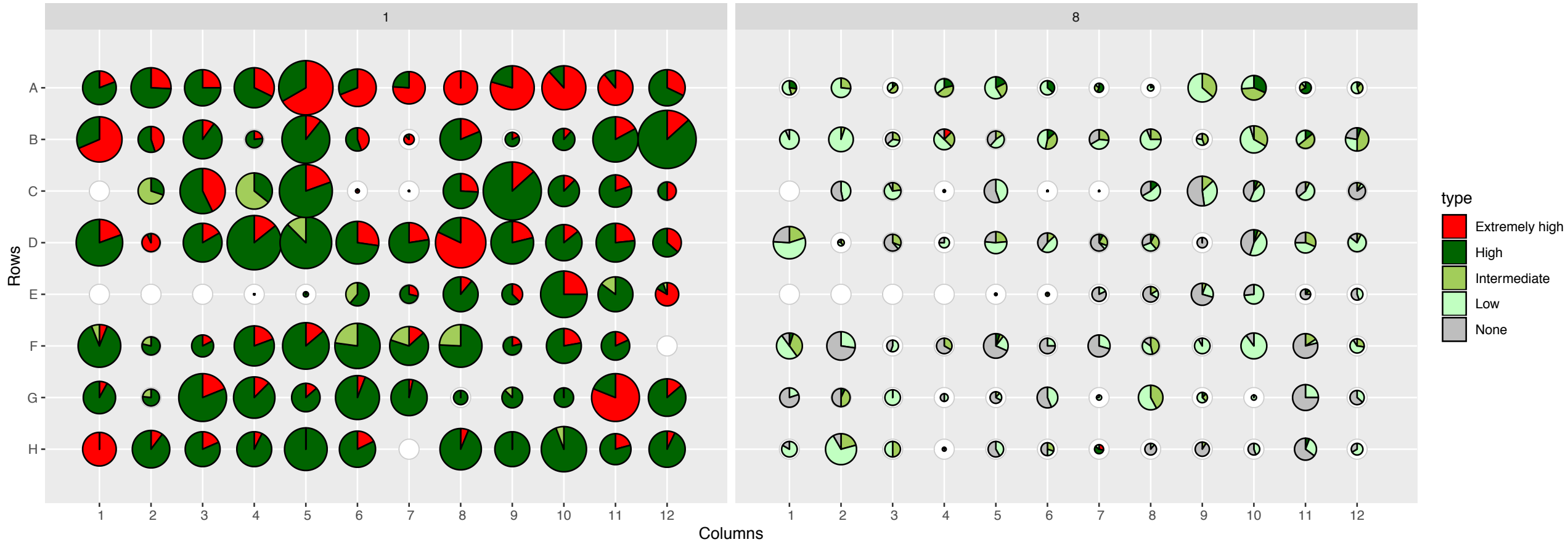
Import & clean

plot

Tweak plot

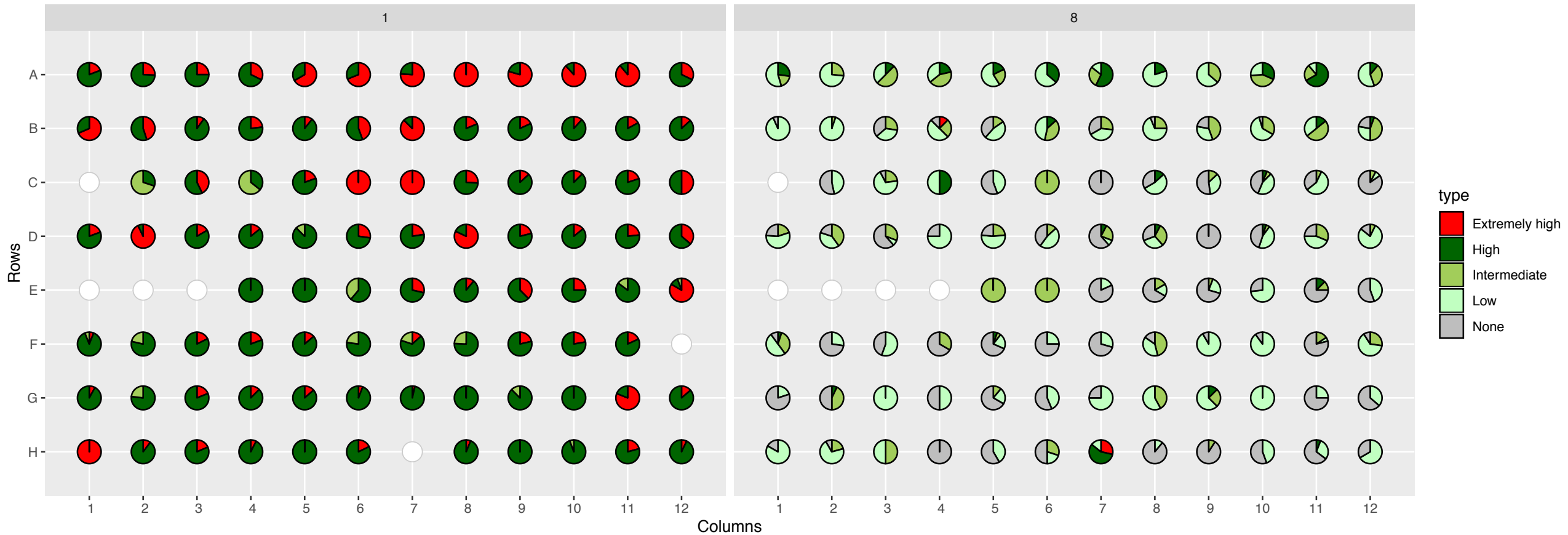
aes(x= Column, y= Row, r = **Total**, group = region)

Promotor activity (day 1 and 8)



aes(x= Column, y= Row,  
group = region)

Promotor activity (day 1 and 8)



# Summary: R for data science

- R is a fantastic tool and a huge time saver
- While intimidating at first all scripts have common patterns that are very repetitive (think of Lego blocks)
- R makes your analysis reusable & reproducible
- You can get help everywhere!



# Real world example

## proteomics analysis

- **Real dataset**  
*(uncleaned)*
- **Other people's code snippets**  
*(base R, different styles)*
- **Analysis as Rmarkdown**
- ***Time investment: ca. 10 – 15 h***  
*(95% googling stuff)*

# RMarkdown

- **Code is in the chunks**  
(light grey background)
- **Text in between chunks**  
(black background).
- **Hierarchy matters** like in a regular R script

```
1 ---
2 title: "Proteomics analysis workflow in R"
3 author: "Andrea Fossati & Cyril Statzer"
4 date: "r Sys.Date()"
5 output: pdf_document
6 ---
7 [r setup, echo = FALSE]
8 knitr::opts_chunk$set(echo=FALSE)
9 ---
10
11 \tableofcontents
12 \newpage
13
14 [r]
15 # install libraries, if you need, only run this block once otherwise leave it commented
16 # install.packages("ggplot2")
17 # install.packages("reshape2")
18 # install.packages("RColorBrewer")
19 # install.packages("ggpubr")
20 # install.packages("dplyr")
21 # install.packages("factoextra")
22 # install.packages("factoextra")
23 # install.packages("tidyverse")
24 ---
25
26 [r, message=FALSE]
27 # load libraries into your workspace, this is needed everytime you restart R or edit your document
28 library(reshape2)
29 library(RColorBrewer)
30 library(ggpubr)
31 library(ggplot2)
32 library(factoextra)
33 library(FactoMineR)
34 library(phenix)
35 library(tidyverse)
36 ---
37
38 [r]
39 # define the functions and global variables you want to use to make your life easier during the analysis
40 input_file_name <- "proteinGroups.txt"
41
42
43 subset_grep <- function(df, quant) {
44   tmp <- df[grep(quant, names(df))]
45   tmp$id <- df$Majority.protein.IDs
46   return(tmp)
47 }
```

# RMarkdown

- **A:** Produces a html or pdf
- **B:** Evaluate one block by pressing play.
- **C:** evaluate all above by pressing the middle button.
- **D:** You can include LaTeX functions (for pdf)

```
1 ---
2 title: "Proteomics analysis workflow in R"
3 author: "Andrea Fossati & Cyril Statzer"
4 date: "r Sys.Date()"
5 output: pdf_document
6 ---
7 [r setup, echo = FALSE]
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15 # install libraries, if you need, only run this block once otherwise leave it uncommented
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20 # install.packages("dplyr")
21 # install.packages("factoextra")
22 # install.packages("factoextra")
23 # install.packages("factoextra")
24
25
26 [r, message=FALSE]
27 # load libraries into your workspace, this is needed everytime you restart R or edit your document
28 library(reshap2)
29 library(RColorBrewer)
30 library(ggpubr)
31 library(ggrepel)
32 library(factoextra)
33 library(FactoMineR)
34 library(phenix)
35 library(tidyverse)
36
37
38 [r]
39 # define the functions and global variables you want to use to make your life easier during the analysis
40 input_file_name <- "proteinGroups.txt"
41
42
43 subset_grep <- function(df, quant) {
44   tmp <- df[grep(quant, names(df))]
45   tmp$id <- df$Majority.protein.IDs
46   return (tmp)
47 }
```



# Markdown structure

- Break up the code into meaningful chunks.
- I would recommend one chunk for one logical step or one plot
- Add explanatory text so you (also in 6 months 🤖) and the reader of the document knows what you did.
- Use Hashtags to generate section headers (and other markup commands)
- You can also evaluate R code in the text.

```
# Data import pre-processing
```

```
```{r}
df<-read.delim(input_file_name)
df_lfq<-filter_conrev(subset_grep(df, 'LFQ'))
rownames(df_lfq)<-df_lfq$id
df_lfq$id=NULL
```
```

In this analysis we are using the ``r input_file_name`` file as input for our workflow. The original file has ``r nrow(df)`` rows and ``r ncol(df)`` columns.

## Data import pre-processing

In this analysis we are using the proteinGroups.txt file as input for our workflow. The original file has 507 rows and 145 columns.

- This chunk imports the file and performs pre-processing
- The reader is informed about the dimensions of the file in the text

```
## Apply normalization
{r}
names(df_lfq_log) <- c(sprintf("Gfp_%d", 1:3), sprintf("Shtag_%d", 1:3), sprintf("Vanadate5_%d", 1:3), sprintf("Vanadate30_%d", 1:3))
all_median <- apply(df_lfq_log, 2, function(x) {median(x, na.rm = T)})
med_norm <- as.data.frame(t(apply(df_lfq_log, 1, function(x) {x - all_median + median(all_median)})))

```{r, warning=FALSE, message=FALSE}
med_norm <- med_norm %>%
  as_tibble(rownames = "ID") %>%
  separate(sep = "\\|", col = ID, into = c("prefix", "UNIPROT", "Gene_name")) %>%
  select(prefix, Gene_name) %>%
  column_to_rownames("UNIPROT")
med_norm %>% head()
```

Apart from the median normalization used here by subtracting the median of every individual group and adding the median of all groups there is large palette of other normalization strategies to use (you can also check out the slides from our session on Friday). For example you could use min/max, quantile, mean and others. The `med_norm` object that is generated here is used as the basis for the remaining part of the script.

### Apply normalization

##	Gfp_1	Gfp_2	Gfp_3	Shtag_1	Shtag_2	Shtag_3	Vanadate5_1
## A0A096LP49	17.95327	NA	NA	NA	NA	NA	NA
## A0A1B0GWK0	NA	NA	NA	NA	NA	NA	NA
## A4UGR9	NA	NA	NA	NA	NA	NA	NA
## A6NDX5	NA	NA	NA	NA	NA	NA	NA
## A6NKD9	NA	NA	NA	NA	NA	NA	NA
## A8MU93	NA	NA	NA	NA	NA	NA	NA

##	Vanadate5_2	Vanadate5_3	Vanadate30_1	Vanadate30_2	Vanadate30_3
## A0A096LP49	NA	NA	NA	NA	NA
## A0A1B0GWK0	NA	18.64669	NA	NA	NA
## A4UGR9	NA	NA	NA	NA	NA
## A6NDX5	NA	NA	NA	19.38634	NA
## A6NKD9	19.19695	18.49152	NA	19.84689	NA
## A8MU93	16.64703	NA	NA	NA	NA

Apart from the median normalization used here by subtracting the median of every individual group and adding the median of all groups there is large palette of other normalization strategies to use (you can also check out the slides from our session on Friday). For example you could use min/max, quantile, mean and others. The `med_norm` object that is generated here is used as the basis for the remaining part of the script.

# Concept of missing values (NA)

- R represents missing values by the symbol **NA** (**N**ot **A**vailable)

```
y <- c(1, NA, 3)
```

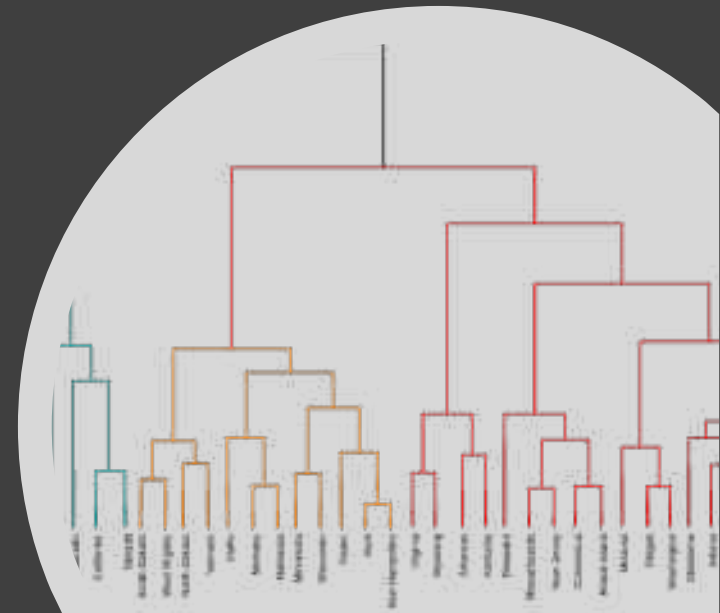
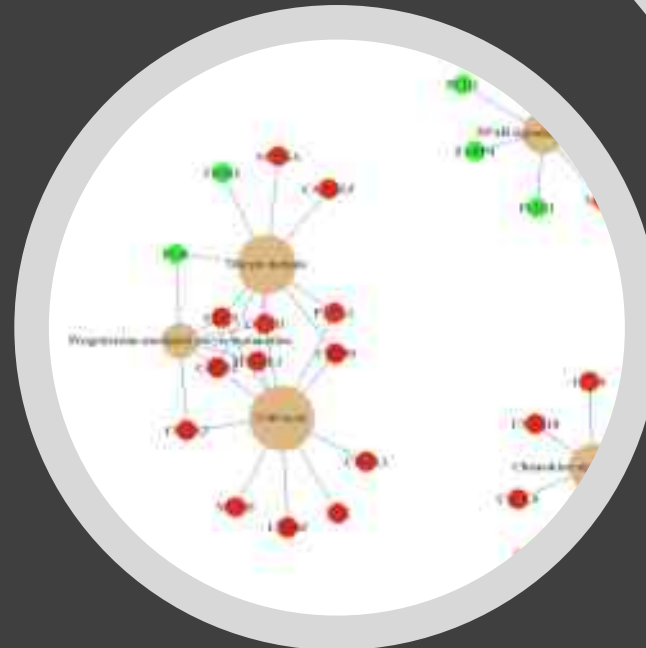
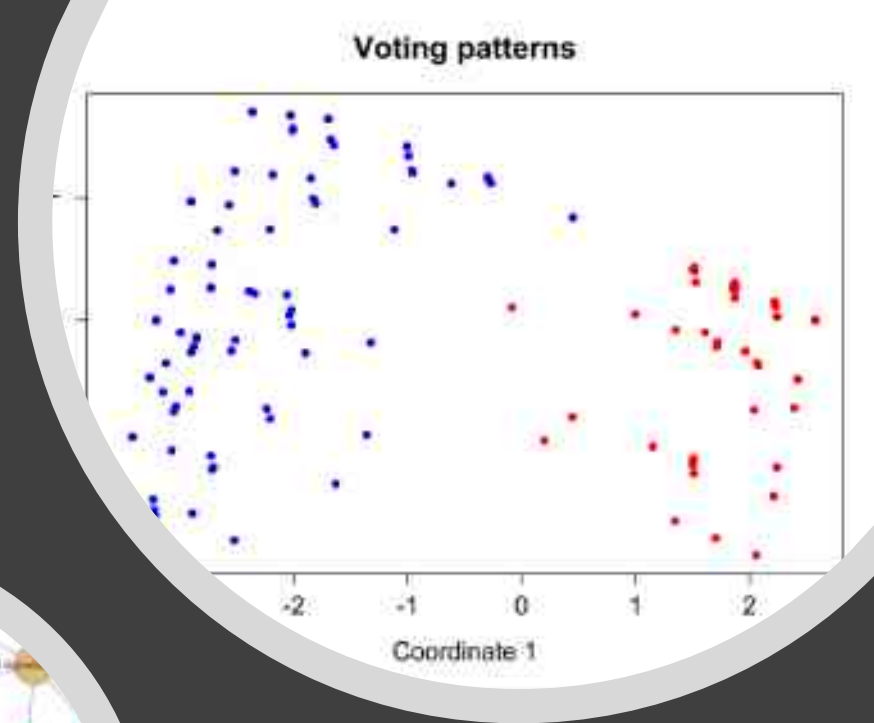
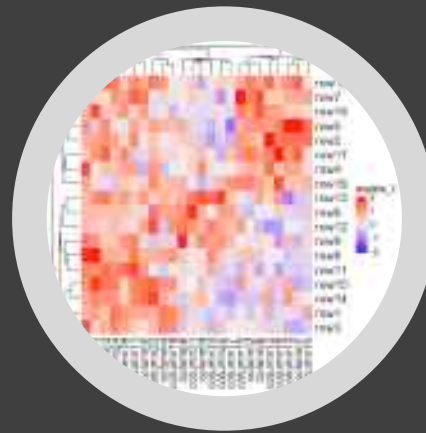
```
y %>% is.na()  
FALSE, TRUE, FALSE
```

```
y %>% mean()  
NA
```

```
y %>% mean(na.rm = TRUE)  
2
```

# Visualizing data

- Heatmaps
- KEGG pathways
- MDS, Hierarchical clustering
- PCA
- ... see last section of the markdown  
«Example of potential addition»





# Outlook

# Online resources for specific questions



plot cool stuff with ggplot r

Google Search

I'm Feeling Lucky



**stackoverflow**

<https://stackoverflow.com>



**STHDA**

Statistical tools for high-throughput data analysis

<http://www.sthda.com/english/>

# Online resources for learning



<https://www.datacamp.com>



The best book is for free:  
<https://r4ds.had.co.nz>

Data Visualization Cheat Sheet

The ggplot2 package lets you make beautiful and consistent plots of your data. It implements the grammar of graphics, so you can build up your plots. See <https://www.rstudio.com/resources/cheatsheets/> for the detailed overview. Updated 2018.

Download



<https://www.rstudio.com/resources/cheatsheets/>

